Supplementary Transplantation-related Procedures

In the autologous group, the conditioning regimen was based on i. melphalan (200 or 140 mg/sm, respectively MAC and RIC conditioning) on day -2 for multiple myeloma patients; ii. on carmustine (400 mg/sm) on day -6 and thiotepa 5 mg/kg/12 hours on days -5 and -4 for patients affected by primary central nervous system lymphoma (PCNSL); and iii. on the BEAM scheme (carmustine 300 mg/sm on day -7, cytarabine 200 mg/sm/12 hours, etoposide 200 mg/sm on days -6, -5, -4, -3 and melphalan 140 mg/sm on day -2) for patients suffering from Hodgkin and non Hodgkin lymphomas other than PCNSL. For patients affected by neurological disease, conditioning was highly immunosuppressive, using BEAM plus anti-thymocyte globulin (ATG)¹. The graft source was PBSCs, mobilized with subcutaneous granulocyte-colony-stimulating factor, collected by leukapheresis and cryopreserved. Filgrastim was administered from Day +1 after autologous transplant until neutrophil recovery (II day of N > 1000/mcl). In the allogeneic group, PBSCs were mobilized with subcutaneous granulocyte-colonystimulating factor and collected by leukapheresis and infused without any ex-vivo manipulation in the majority of transplants. Bone marrow was collected from the posterior superior iliac crests and infused according to standard guidelines. Cord blood units were infused intravenously or intra bone. Starting from January 2019, patients transplanted with a CD3⁺ graft content above 300*10⁶/kg routinely received one single administration of ATG on day 5 at the dose of 5 mg/kg with the aim of minimizing GVHD risk². Starting from May 2020, we added the use of granulocyte-colony-stimulating-factors in all transplant recipients, given as a single peg-filgrastim dose or daily filgrastim until engraftment.

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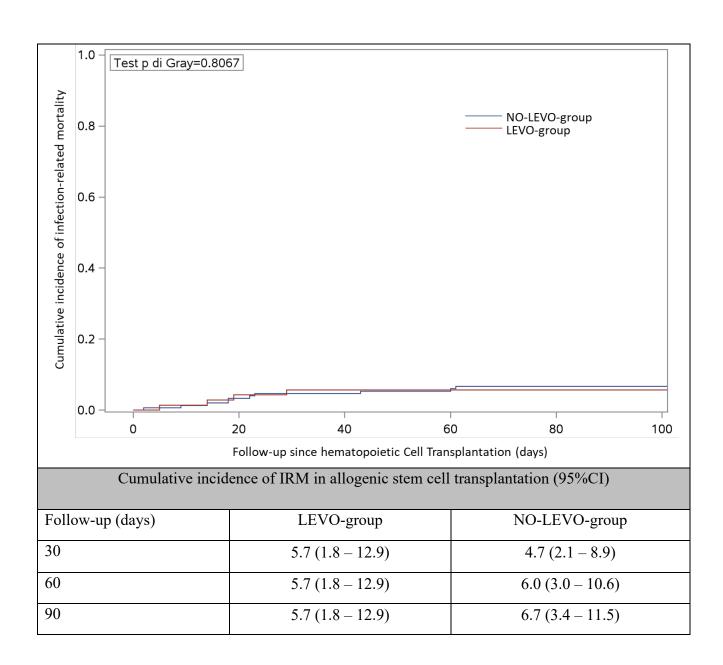
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Supplementary Table 1. Detailed description of the more represented types of antibiotic escalation in autologous stem cell transplantation.

Use of levofloxacin prophylaxis	Broad- spectrum antibiotics at first FN episode	DAP or VAN	AMK or GEN	MEM	Frequency	Percent	Cumulative frequency	Cumulative percent
NO	PTZ	0	0	1	6	12.77	11	23.40
NO	PTZ	1	0	0	12	25.53	23	48.94
NO	PTZ	1	0	1	11	23.40	34	72.34
YES	PTZ	1	0	1	2	4.26	43	91.49
YES	PTZ+VAN	0	0	1	2	4.26	45	95.74
YES	CAZ	0	0	1	2	4.26	47	100.00

Legend: 0=no; 1=yes

Supplementary Table 2. Detailed description of the more represented types of antibiotic escalation in allogeneic HSCT.

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Use of levofloxacin prophylaxis	Broad- spectrum antibiotics at first FN episode	DAP or VAN	AMK or GEN	MEM	Frequency	Percent	Cumulative frequency	Cumulative percent
NO	MEM	0	1	-	2	1.69	5	4.24
NO	MEM	1	0	-	4	3.39	9	7.63
NO	PTZ	0	0	1	19	16.10	32	27.12
NO	PTZ	0	1	1	5	4.24	38	32.20
NO	PTZ	1	0	0	14	11.86	52	44.07
NO	PTZ	1	0	1	17	14.41	69	58.47
NO	PTZ	1	1	1	3	2.54	72	61.02
YES	CAZ	1	0	1	4	3.39	81	68.64
YES	MEM	1	0	-	4	3.39	86	72.88
YES	PTZ	0	0	1	11	9.32	104	88.14
YES	PTZ	0	1	1	2	1.69	106	89.83
YES	PTZ	1	0	1	12	10.17	118	100.00

Legend: 0=no; 1=yes

Supplementary Table 3. Patients' distribution according to ESBL BSI or CR BSI or acute GHVD occurrence within 100-day since allogeneic HSCT.

Use of levofloxacin prophylaxis	ESBL BSI <100 days	CR BSI <100 days	Acute GVHD	Frequency	Percent	Cumulative frequency	Cumulative percent
NO	0	0	0	107	50.95	107	50.95
NO	0	0	1	32	15.24	139	66.19
NO	1	0	0	1	0.48	140	66.67
NO	1	0	1	3	1.43	143	68.10
NO	1	1	1	1	0.48	144	68.57
YES	0	0	0	53	25.24	197	93.81
YES	0	0	1	11	5.24	208	99.05
YES	0	1	1	1	0.48	209	99.52
YES	1	0	0	1	0.48	210	100.00

Legend: 0=no; 1=yes

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Supplementary Table 4. Etiology of Gram positive bacteria pre-engraftment bloodstream infections and antimicrobial resistance in ASCT and allo-HSCT recipients, according to levofloxacin prophylaxis administration.

Blood cultures' isolates	Total bacterial isolates from blood cultures								
characteristics	Group A	Group B	p-Value	Group C	Group D	p- Value			
Gram positive bacteria	0	10	-	18	37	-			
Staphylococcus spp	0 (0%)	5 (50%)	-	4 (22%)	17 (46%)	0.139			
S. aureus	0 (0%)	1 (20%)	-	0 (0%)	17 (46%)	-			
 MR FQs resistant CoNS MR FQs resistant Enterococcus spp E. faecalis E. faecium 	0 (0%) 0 (0%) 0 (0%) 0 (0%) 0 (0%) 0 (0%)	1 (100%) 0 (0%) 4 (80%) 2 (100%) 4 (100%) 2 (20%) 0 (0%) 2 (100%)	-	0 (0%) 0 (0%) 4 (100%) 4 (100%) 4 (100%) 14 (78%) 5 (36%) 9 (64%)	0 (0%) 0 (0%) 17 (100%) 17 (100%) 17 (100%) 13 (35%) 3 (23%) 9 (69%)	0.004			
- VR-Enterococci	0 (0%)	0 (0%)		2 (14%)	5 (38%) @				
Viridans streptococci	0 (0%)	2 (20%)	-	0 (0%)	3 (8%)	0.543			
Others	0 (0%)	1 (10%)	-	0 (0%)	4 (11%) §	0.291			

Legend: Group A, autologous stem cell transplantation LEVO-group; Group B, autologous stem cell transplantation NO-LEVO-group; Group C, allogeneic hematopoietic stem cell transplantation LEVOgroup; Group D, allogeneic hematopoietic stem cell transplantation NO-LEVO-group; FQs, fluoroquinolones; MR, methicillin-resistant; CoNS, Coagulase negative staphylococci; VR, vancomycin resistant.

^{@1} Enterococcus caselliflavus intrinsically resistant to vancomycin

^{#1} Micrococcus luteus

^{§1} Clostridium ramosum, 1 Corynebacterium aurimucosum, 2 Streptococcus gallolyticus

ADDITIONAL REFERENCES

- 76 [1] Sharrack B, Saccardi R, Alexander T, *et al.* Autologous haematopoietic stem cell transplantation and other 77 cellular therapy in multiple sclerosis and immune-mediated neurological diseases: updated guidelines and 78 recommendations from the EBMT Autoimmune Diseases Working Party (ADWP) and the Joint and the 79 Joint Accreditation Committee of EBMT and ISCT (JACIE). Bone Marrow Transplant. 2020 80 Feb;55(2):283-306.
- 81 [2] Mussetti A, De Philippis C, Carniti C, *et al.* CD3+ graft cell count influence on chronic GVHD in 82 haploidentical allogeneic transplantation using post-transplant cyclophosphamide. Bone Marrow 83 Transplant. 2018 Dec;53(12):1522-1531.