nature portfolio

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Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our Editorial Policies and the Editorial Policy Checklist.

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

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n/a	Confirmed
	The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
	🗷 A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
x	The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.
x	A description of all covariates tested
×	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
x	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>
x	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
x	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
×	Estimates of effect sizes (e.g. Cohen's <i>d</i> , Pearson's <i>r</i>), indicating how they were calculated
,	Our web collection on statistics for biologists contains articles on many of the points above.

Software and code

Policy information about availability of computer code

Serial EM 3.7 and EPU 2.1 are used to collect cryo-EM data. Data collection

Data analysis

MotionCor2 1.1.0; GCTF 1.06; RELION-3.1; Chimera 1.13; Coot 0.8.6.1; Phenix 1.19; Pymol 1.8.6.0; Prism 8.2.1; CryoSPARC 3.2; Clustal Omega server; ResMap 1.1.4.

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio guidelines for submitting code & software for further information.

Data

Policy information about availability of data

All manuscripts must include a data availability statement. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our policy

PDB coordinates has been deposited into the PDB database. EM map has been deposited into the EMDB database. All data will be available upon publication.

Human resear	ch participants		
Policy information ab	out studies involving human research participants and Sex and Gender in Research.		
Reporting on sex and	gender N/A		
Population characteris	stics N/A		
Recruitment	N/A		
Ethics oversight	N/A		
Note that full information	n on the approval of the study protocol must also be provided in the manuscript.		
Field-spec	rific reporting		
Please select the one	below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.		
Life sciences	Behavioural & social sciences		
Tot a reference copy of the	accument with an sections, see <u>nature.com/accuments/m-reporting-summary-nat.pur</u>		
Life scienc	ces study design		
All studies must disclo	ose on these points even when the disclosure is negative.		
m	the number of replicates are reported in the figure legends. Sample size was determined based on the variation of the standard error of the nean (SEM), which is represented in the figures. At least three biological repeats were performed so that the SEM was within at least 10% of the mean, the exact number of replicates are indicated in the figure legends.		
Data exclusions N	o data were excluded from the analyses.		
	ach data point in enzymatic assay was measured in at least 3 independent experiments, the exact number of replicates are reported in the gure legends.		
	o randomization is needed for the enzymatic assay as the assay do not have unknown covariates. For example, the comparison between WT and mutants in enzymatic activity, there is no feasible unknown covariate that we can minimize by randomizing the experimental groups.		
Blinding	tants were tested and analyzed blindly to avoid bias.		
We require information	for specific materials, systems and methods from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.		
Materials & expe			
n/a Involved in the s	n/a Involved in the study X ChIP-seq		
Eukaryotic ce			
	y and archaeology MRI-based neuroimaging		

Palaeontology and archaeology Animals and other organisms

Dual use research of concern

Clinical data

Eukaryotic cell lines

Policy information about <u>cell lines and Sex and Gender in Research</u>

High Five Cells (invitrogen) Cell line source(s)

Authentication No further authentication was performed for commercially available cell lines. Authentication

Mycoplasma contamination The cell lines were not tested for mycoplasma contamination.

Commonly misidentified lines (See <u>ICLAC</u> register)