

Supplementary Materials

Influence of the modifiers in polyol method on magnetically induced hyperthermia and biocompatibility of ultrafine magnetite nanoparticles

Adrian Radoń^{1,2,*}, Agnieszka Włodarczyk³, Łukasz Sieroń³, Magdalena Rost-Roszkowska⁴, Łukasz Chajec⁴, Dariusz Łukowiec¹, Agnieszka Ciuraskiewicz², Piotr Gębara⁵, Stanisław Waclawek⁶, Aleksandra Kolano-Burian²

¹ Faculty of Mechanical Engineering, Silesian University of Technology, Konarskiego 18 a St., 44-100 Gliwice, Poland

² Łukasiewicz Research Network - Institute of Non-Ferrous Metals, Sowinskiego 5 St., 44-100 Gliwice, Poland

³ Department of Medical Genetics, Faculty of Medical Sciences in Katowice, Medical University of Silesia, Medyków 18, 40-752 Katowice, Poland

⁴ Institute of Biology, Biotechnology and Environmental Protection, University of Silesia in Katowice, Bankowa 9, 40-007 Katowice, Poland

⁵ Department of Physics, Częstochowa University of Technology, Armii Krajowej 19, 42-200 Czestochowa, Poland

⁶ Institute for Nanomaterials, Advanced Technologies and Innovation, Technical University of Liberec, Studentská 1402/2, 461 17, Liberec 1, Czech Republic

*corresponding author: adrian.radon@imn.lukasiewicz.gov.pl

Table S1. Comparison of the values of aggregates diameter measured using the DLS technique (D_{DLS}) and zeta potential values (ζ potential) determined for the 3 mg/ml water dispersions

Sample	D_{DLS} (nm)	ζ potential (mV)
Fe ₃ O ₄ - URO	120.17±0.34	7.57±0.35
Fe ₃ O ₄ - NH ₄ HCO ₃	272.87±5.77	4.32±1.70
Fe ₃ O ₄ - PEG	395.57±12.60	8.86±1.26

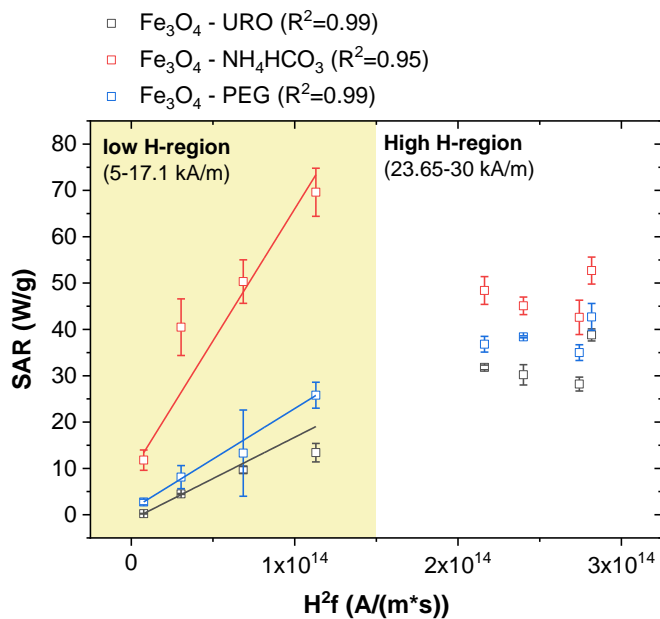


Figure S1. Relationship between SAR and H^2f determined for Fe₃O₄-URO, Fe₃O₄- NH₄HCO₃ and Fe₃O₄-PEG NPs with linear relationship determined for low H-region according to the LRT

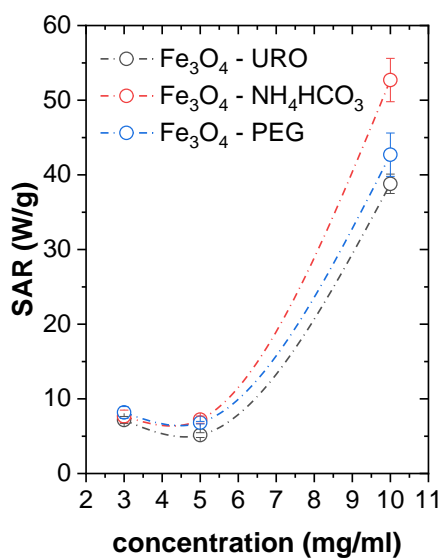


Figure S2. Relationship between SAR and colloids concentrations measured for Fe₃O₄-URO, Fe₃O₄-NH₄HCO₃ and Fe₃O₄-PEG NPs

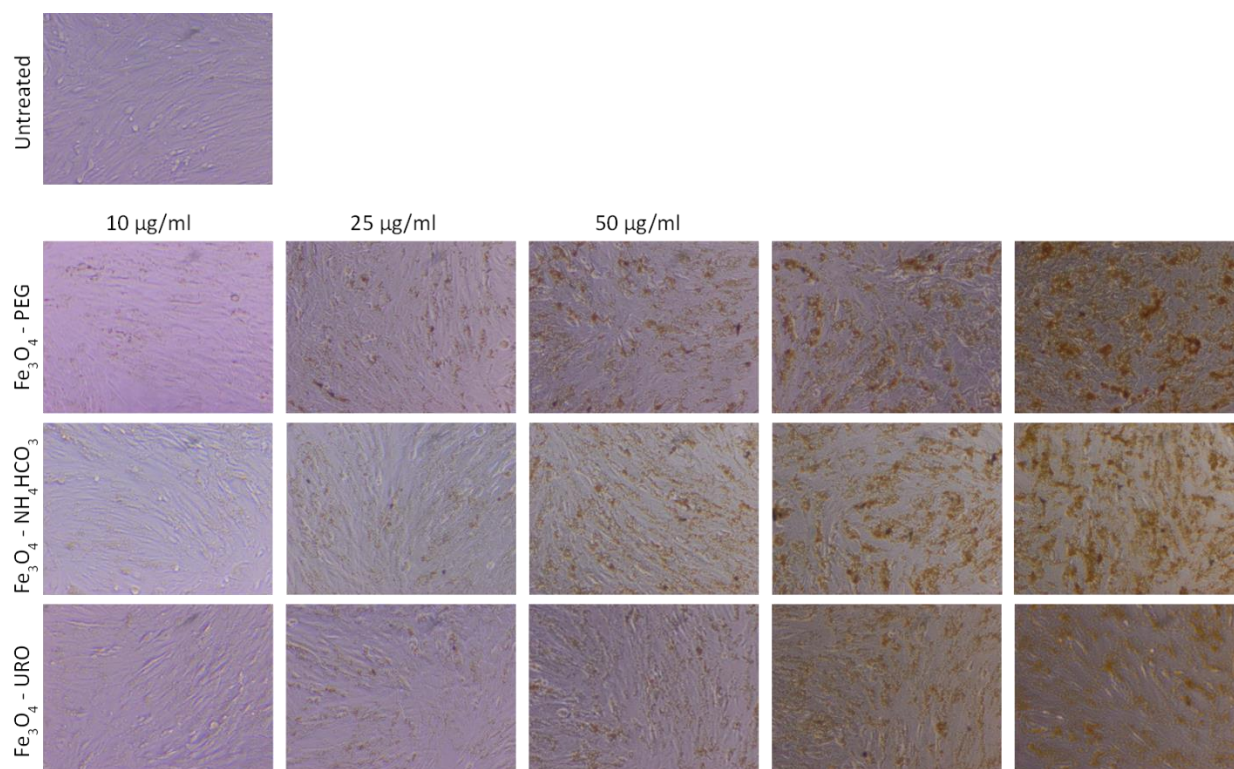


Figure S3. Picture of cells culture from the light microscope. Nanoparticles settled down on the cells layer.

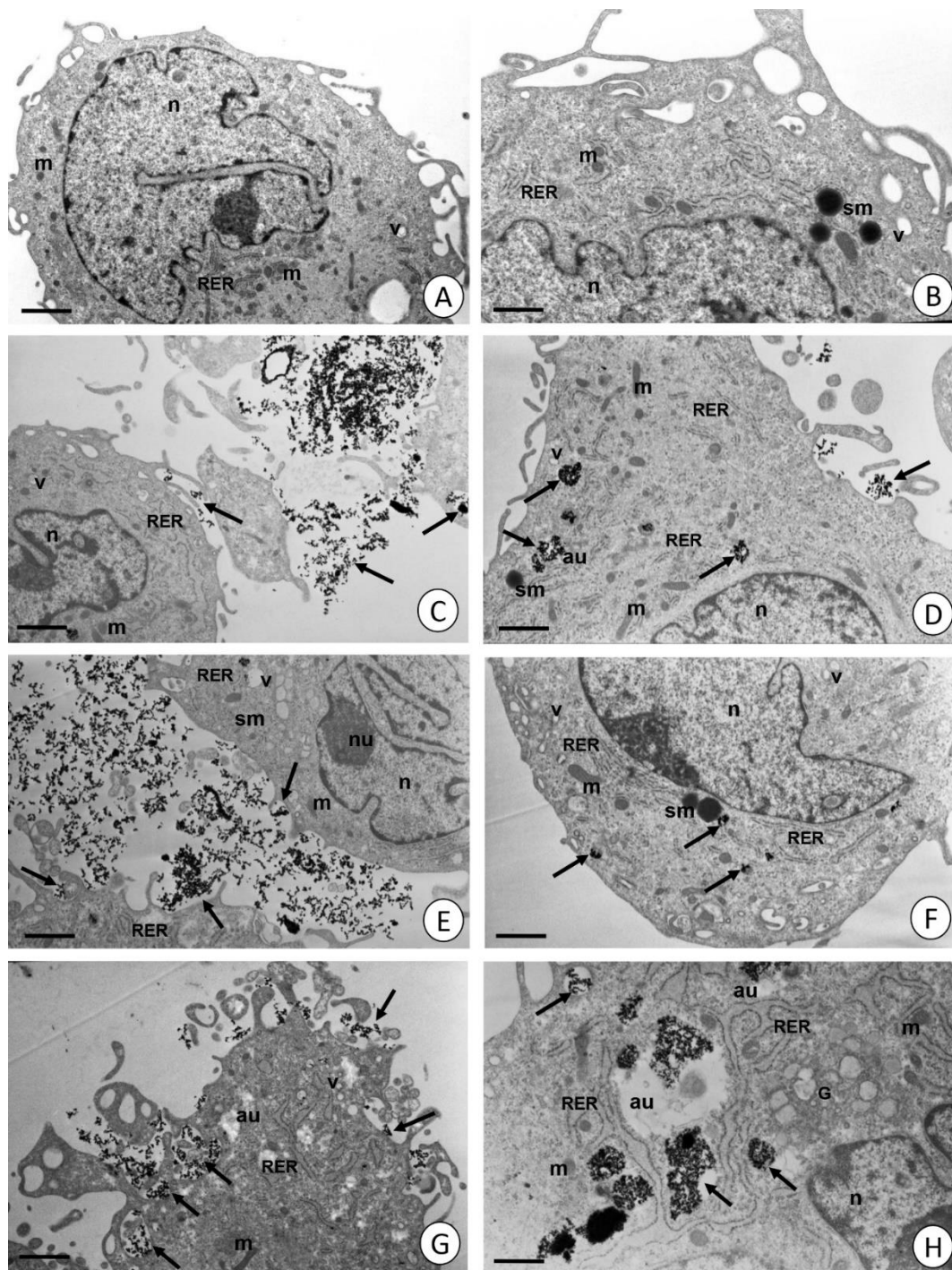


Figure S4. Fibroblasts visible in TEM. (A-B) 0 – 24h control group. (C-D) Fe_3O_4 - PEG – 24h experimental group. (E-F) Fe_3O_4 - NH_4HCO_3 – 24h experimental group. (G-H) Fe_3O_4 - URO – 24h experimental group. Nuclei (n), nucleolus (nu), mitochondria (m), cisterns of RER (RER), storage material (sm), Golgi complexes (G), vacuoles (v), autophagosomes (au), electron-dense granules (arrows). (A) Scale bar = 1.74 μm . (B) Scale bar = 0.87 μm . (C) Scale bar = 1.71 μm . (D) Scale bar = 1.44 μm . (E) Scale bar = 1.66 μm . (F) Scale bar = 1.14 μm . (G) Scale bar = 1.82 μm . (H) Scale bar = 1.04 μm .

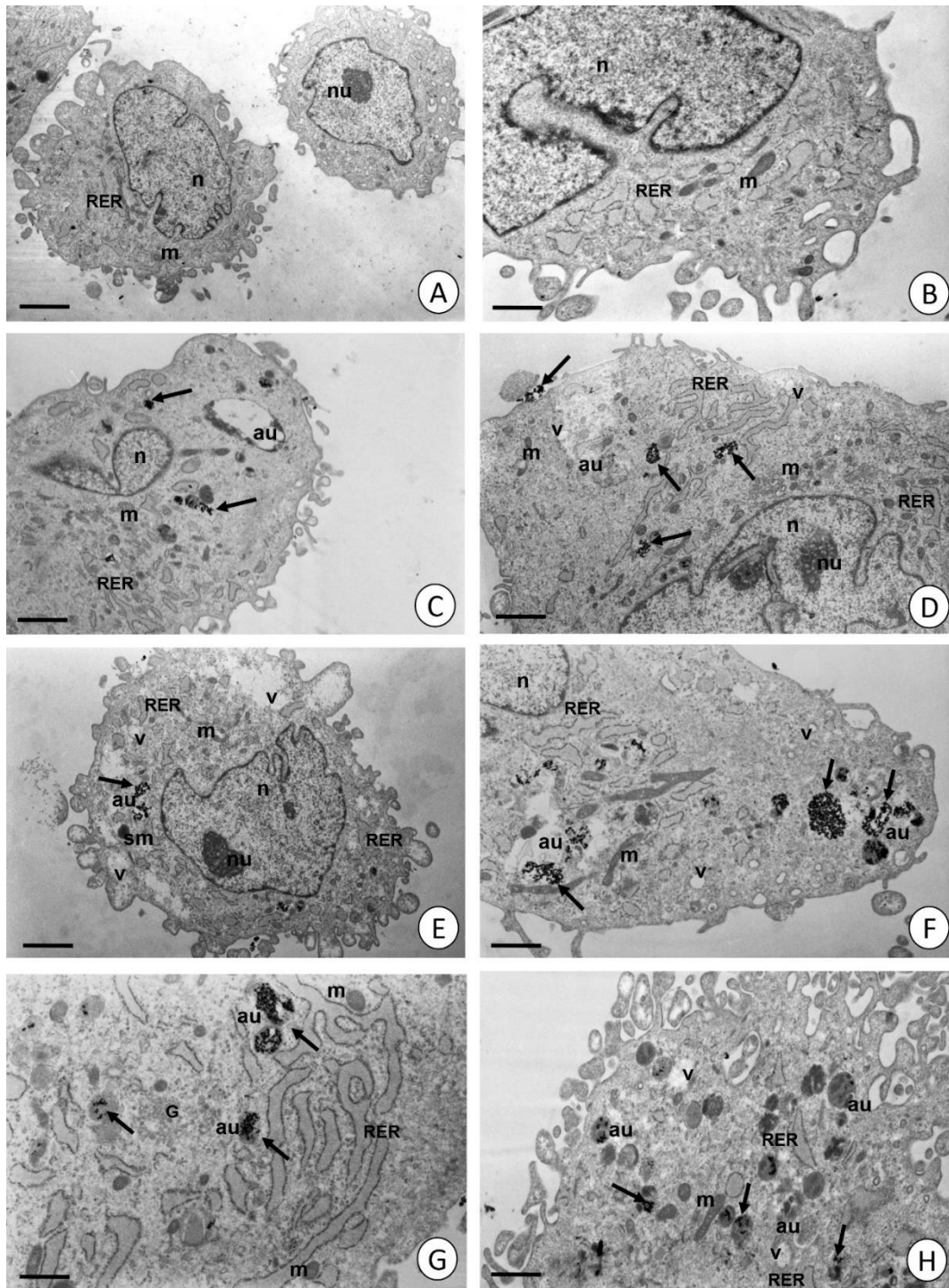


Figure S5. Fibroblasts visible in TEM. **(A-B)** 0 – 48h control group. **(C-D)** Fe_3O_4 - PEG – 48h experimental group. **(E-F)** Fe_3O_4 - NH_4HCO_3 – 48h experimental group. **(G-H)** Fe_3O_4 - URO – 48h experimental group. Nuclei (n), nucleolus (nu), mitochondria (m), cisterns of RER (RER), Golgi complexes (G), storage material (sm), vacuoles (v), autophagosomes (au), electron-dense granules (arrows). **(A)** Scale bar = 2.28 μm . **(B)** Scale bar = 1.10 μm . **(C)** Scale bar = 1.84 μm . **(D)** Scale bar = 1.82 μm . **(E)** Scale bar = 2.28 μm . **(F)** Scale bar = 1.42 μm . **(G)** Scale bar = 1.22 μm . **(H)** Scale bar = 0.88 μm .