

Supporting Information

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Orai1 is an Entotic Ca²⁺ Channel for Non-Apoptotic Cell Death, Entosis in Cancer Development

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Supporting Information

Orai1 is an entotic Ca^{2+} channel for non-apoptotic cell death, entosis in cancer development.

Ah Reum Lee¹ and Chan Young Park¹ *

Fig. S1

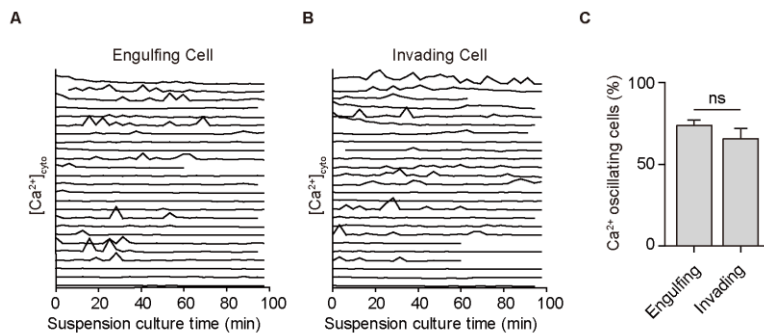


Figure S1. Entotic Ca^{2+} oscillations in engulfing and invading cells.

(A and B) Graphs of normalized GCaMP6s ratio in engulfing (A) and invading (B) cells. ($n > 22$)
(C) Quantification of Ca^{2+} oscillating cells from (A and B). Significance was determined using unpaired two-tailed t -test. ns, not significant.

Fig. S2

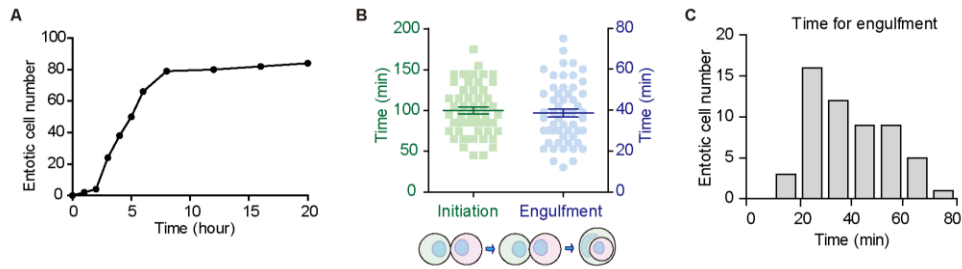


Figure S2. Time kinetics of the initiation of internalization and engulfment.

(A) Quantification of entotic cells using time-lapse imaging conducted over 20 h (n = 84). (B and C) Time to entosis initiation and engulfment. Engulfment was initiated after 1 - 2 h of suspension (n = 54) (B). Complete engulfment takes 30 - 60 min after the initiation (n = 55) (B and C).

Fig. S3

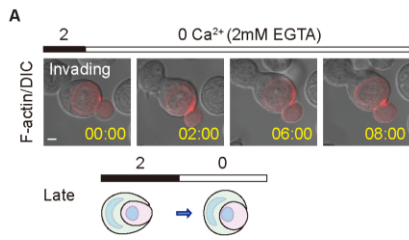


Figure S3. Entotic cells in Ca²⁺ withdrawal condition.

(A) Time-lapse images of entotic cells under Ca²⁺ withdrawal (2 → 0 mM). Cherry-Lifeact-labeled cell morphology. Scale bar = 5 μm.

Fig. S4

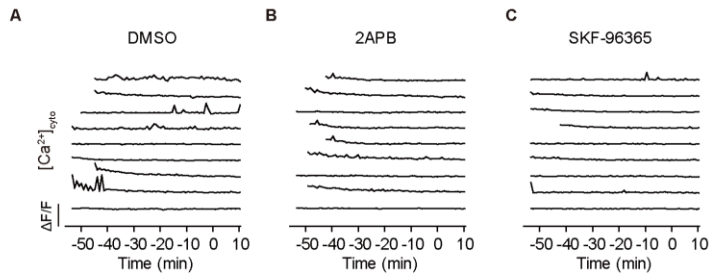


Figure S4. Ca²⁺ oscillations in SOCE inhibitors.

(A to C) Graphs of normalized GCaMP6s ratio in SOCE inhibitors. DMSO (A, n = 28), 50 μM 2-APB (B, n = 19) and 10 μM SKF96365 (C, n = 24) were added on the culture media. The time-point marking the complete cell-in-cell structure was set to 0. Images were taken at 30 s intervals.

Fig. S5

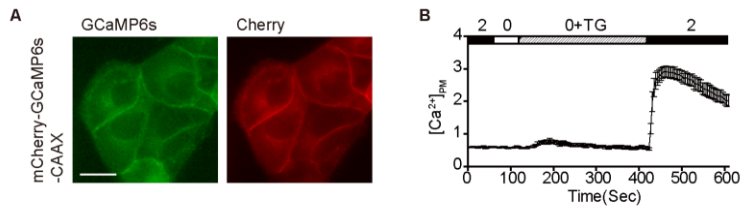


Figure S5. Ca^{2+} signals in plasma membrane.

(A) Fluorescence images of mCherry-GCaMP6s-CAAX. Scale bar = 20 μ m. **(B)** Thapsigargin (TG)-induced Ca^{2+} influx in MCF-7 cells monitored using GCaMP6s-CAAX/Cherry Fluorescence ratio.

Fig. S6

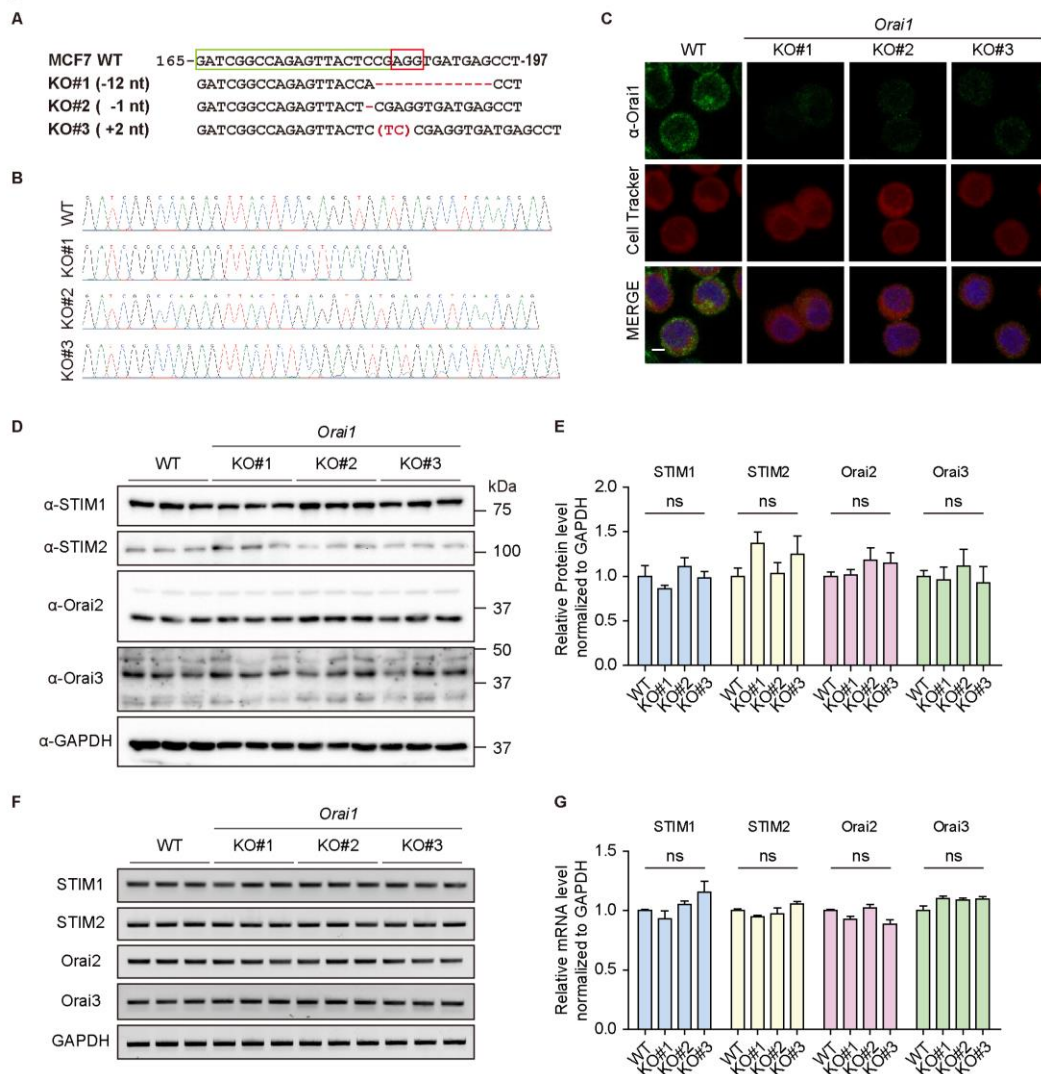


Figure S6. Genetic deletion of *Orai1* using CRISPR-Cas9 system.

(A and B) Sequences of *Orai1* KO MCF7 cells. Alignment of human *Orai1* sequences from *Orai1* KO cell lines used in this study. (C) Immunofluorescence images of endogenous *Orai1* (green). Non-labeled *Orai1* WT cells were mixed with Cell-Tracker Red labeled *Orai1* KO cell lines. (D and F) Protein (D) and mRNA (F) expression levels of other SOCE components: STIM1, STIM2, *Orai2*, and *Orai3* in *Orai1* KO cell lines. GAPDH was used as endogenous control. (E and G) Relative protein (E, from D) and mRNA (G, from F) expression level of SOCE components normalized with GAPDH. Statistical analysis was performed using one-way ANOVA followed by Dunnett's test. ns, not significant.

Fig. S7

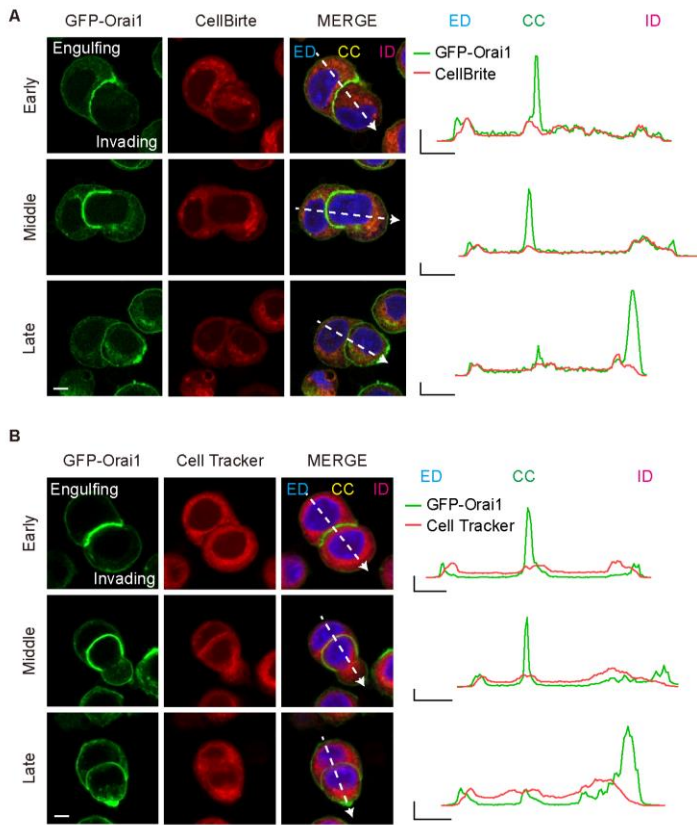


Figure S7. The polarized distribution of Orai1 during entosis.

(A and B) Fluorescence images of GFP-Orai1 expressing MCF7 cells stained with CellBrite PM marker (A, red, PM) or CellTracker (B, red, cytosol). Line scan analysis of relative GFP-Orai1 (green) and PM or cytosol marker (red) signals along the white arrow in the merge image. X axis: 5 μ m, Y axis: 1 (A.U.). Scale bar = 5 μ m. ED, Engulfing cell distal region; CC, cell-cell contact site; ID, Invading cell distal region.

Fig. S8

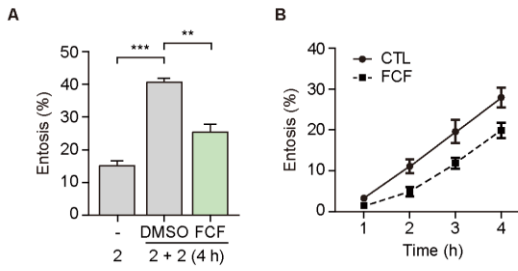


Figure S8. SEPTIN inhibitor suppresses entosis.

(A) Quantification of internalized cells suspended in FCF. Cells were pre-suspended for 2 h and then mixed with FCF and cultured for an additional 2 h. Data represent mean \pm SEM of triplicate experiments ($n > 200$ in each experiment). Significance was determined using unpaired two-tailed t -test. *** $p < 0.001$, ** $p < 0.01$ (B) Time-dependent quantification of entotic cells with or without FCF. MCF7 cells were cultured in suspension for 4 h at 1 h intervals. Data represent mean \pm SEM of triplicate experiments ($n > 300$ in each experiment).

Fig. S9

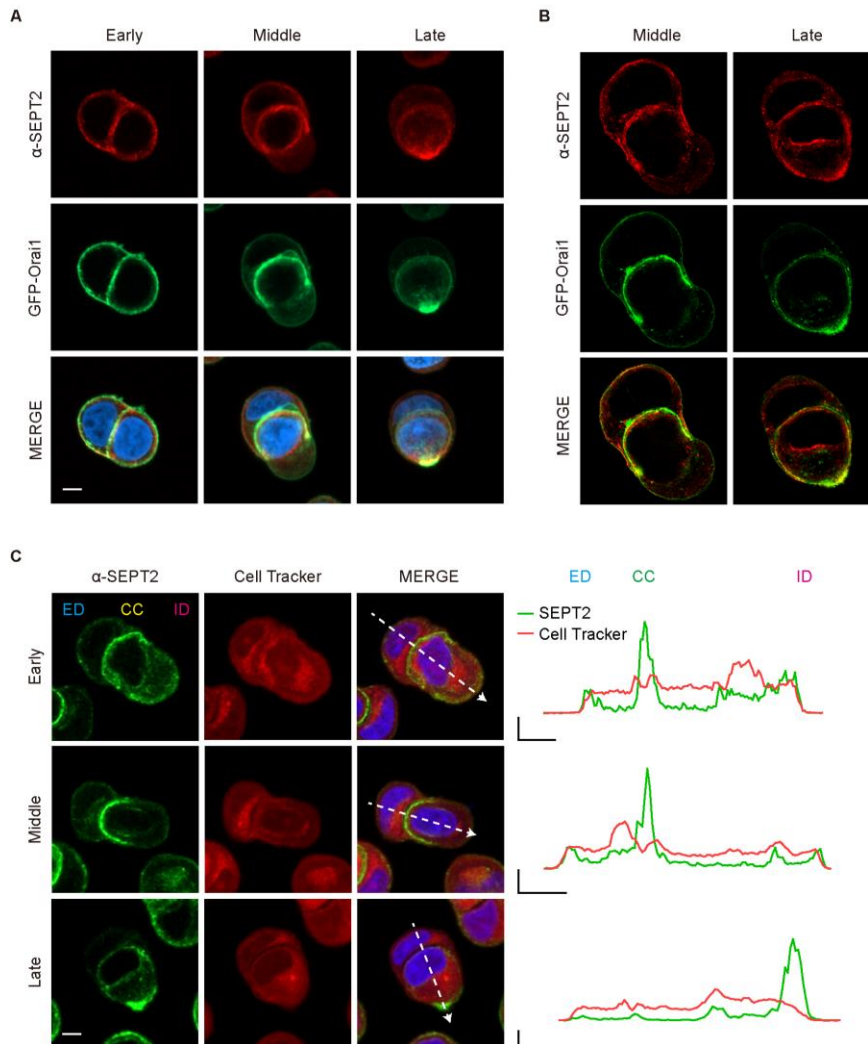


Figure S9. SEPT2 co-localizes with Orai1.

(A) 3D projection of immunofluorescence images showing endogenous SEPT2 (red) and GFP-Orai1 (green) (B) Super-resolution microscopy of the localization of SEPT2 (red) and GFP-Orai1 (green) during entosis. (C) Immunofluorescent images of suspended cells taken after 2.5 h of culturing show endogenous SEPT2 (green) and CellTracker (red, cytosol marker). Line graphs show SEPT2 (green) and cytosol marker (red) intensities for the indicated line scans. X axis: 5 μ m, Y axis: 1 (A.U.). Scale bar = 5 μ m. ED, Engulfing cell distal region; CC, cell-cell contact site; ID, Invading cell distal region.

Fig. S10

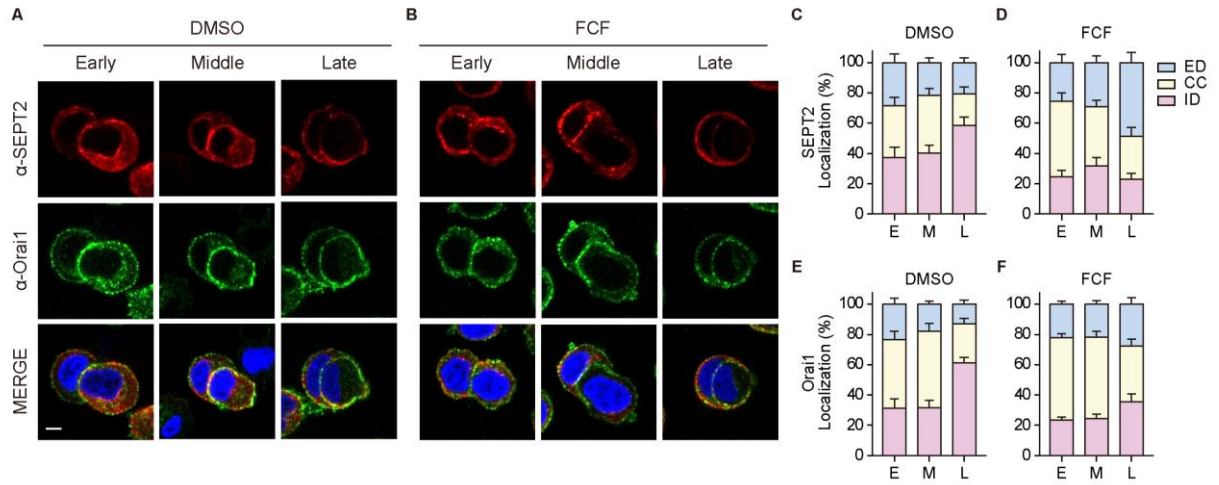


Figure S10. SEPTIN inhibitor suppresses Orai1 organization.

(A and B) Immunofluorescence images showing endogenous SEPT2 (red) and endogenous Orai1 (green) in MCF7 cells cultured for 2.5 h in DMSO (A) or FCF (B, 50 μ M). Scale bar = 5 μ m. (C to F) Bar graphs showing the distribution of SEPT2 (C, DMSO; D, FCF) and Orai1 (E, DMSO; F, FCF). n = 34 in the DMSO group; n = 29 in the FCF group. ED, Engulfing cell distal region; CC, cell-cell contact site; ID, Invading cell distal region. E, Early; M, Middle; L, Late.

Fig. S11

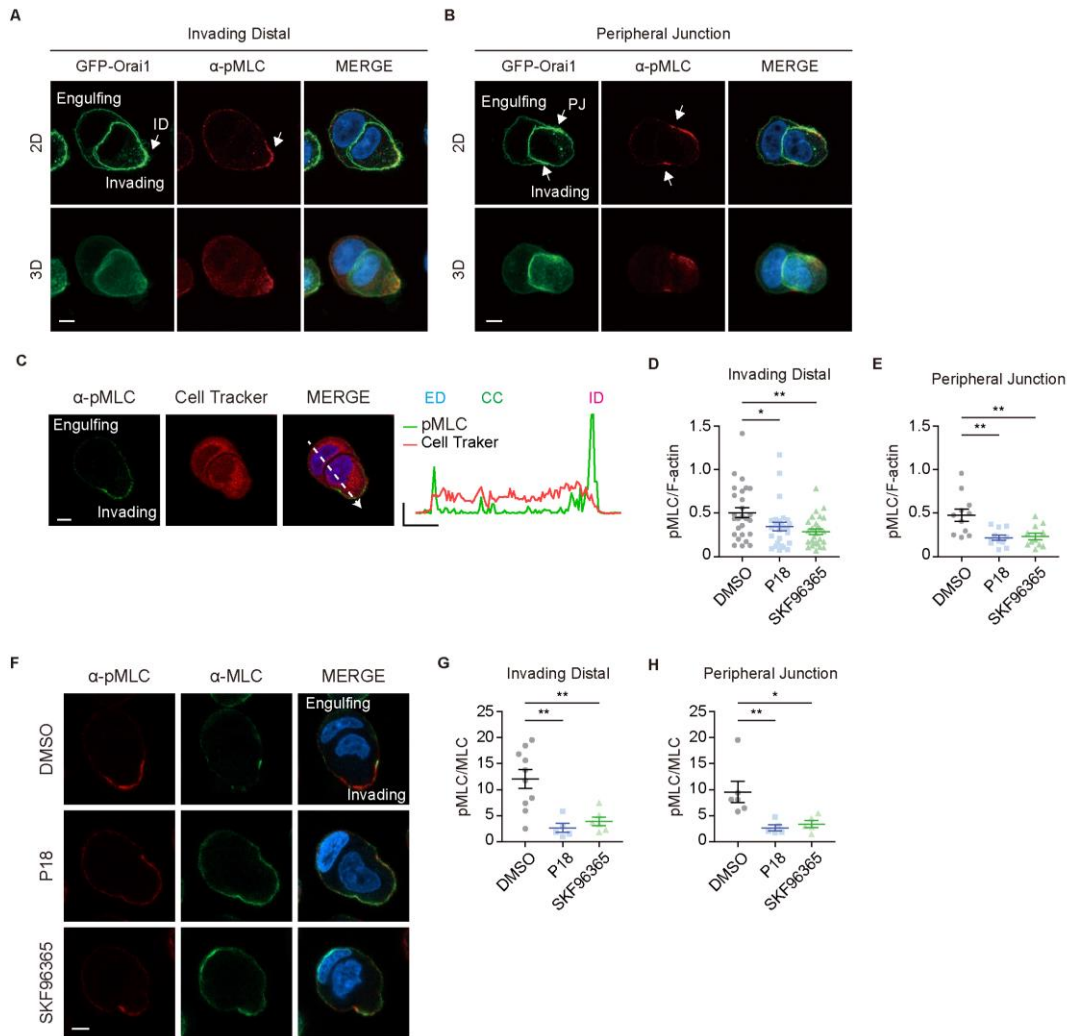


Figure S11. MLC phosphorylation is induced by Orai1 at invading cell distal region and peripheral junction.

(A and B) Immunofluorescence images showing endogenous pMLC (red) in GFP-Orai1 expressing MCF7 cells cultured in suspension for 2.5 h. pMLC is enriched at ID (A) and PJ (B). (C) Immuno-fluorescent images of suspended cells taken after 2.5 h of culturing show endogenous pMLC (green) and CellTracker (red, cytosol marker). Line graphs show relative pMLC (green) and cytosol marker (red) intensities for the indicated line scans. X axis: 5 μ m, Y axis: 5 (A.U.). (D and E) Quantification of pMLC/F-actin intensity ratio at ID (D, from Figure6E, n=27 in each group) and PJ (E, from Figure6F, n=11 in each group). (F) Immuno-fluorescence images of suspended cells after 2.5 h of culturing show endogenous pMLC (red) and MLC (green) in MCF7 cells. (G and H) Quantification of pMLC/MLC intensity ratio at ID (G, n=10, 5, 5) and PJ (H, n=6, 5, 5). Scale bar = 5 μ m. ED, Engulfing cell distal region; CC, cell-cell contact site; ID, Invading cell distal region; PJ, peripheral junction. Significance was determined using unpaired two-tailed *t*-test. ****p* < 0.001, ***p* < 0.01, **p* < 0.05.

Movie S1

Cytosolic Ca²⁺ oscillations in Cherry-GCaMP6s expressing MCF7 cells. Scale bar = 5 μm.

Movie S2

Plasma membrane Ca²⁺ oscillations in Cherry-GCaMP6s-CAAX expressing engulfing MCF7 cell. Scale bar = 5 μm.

Movie S3

Plasma membrane Ca²⁺ oscillations in Cherry-GCaMP6s-CAAX expressing invading MCF7 cell. Scale bar = 5 μm.

Movie S4

Plasma membrane Ca²⁺ oscillations in Cherry-GCaMP6s-CAAX expressing *Orai1* WT MCF7 cells. Scale bar = 20 μm.

Movie S5

Plasma membrane Ca²⁺ oscillations in Cherry-GCaMP6s-CAAX expressing *Orai1* KO MCF7 cells. Scale bar = 20 μm.

Movie S6

Orai1 localization in GFP-Orai1 expressing MCF7 cells. Scale bar = 5 μm.