

Supporting Information for

Generation of zero-valent sulfur from dissimilatory sulfate reduction in sulfate-reducing microorganisms

Shanquan Wang, Qihong Lu, Zhiwei Liang, Xiaoxiao Yu, Mang Lin, Bixian Mai, Rongliang Qiu, Wensheng Shu, Zhili He, Judy D. Wall

Shanquan Wang
Email: wangshanquan@mail.sysu.edu.cn

This PDF file includes:

Figures S1 to S8
Tables S1 to S6
SI References

A

Sulfate reducing bacteria	Abbr.	Collection no.	Origin	Gram-positive/negative	Temp.	Electron donor	Electron acceptor	Medium
<i>Desulfovibrio vulgaris</i> Hildenborough	DvH	DSM 644	Wealden clay	Gram-negative	Mesophilic	DL-Lactate	Sulfate	LS4D
<i>Desulfoformulus ruminis</i> DL	DLT	DSM 2154	Rumen of hay-fed sheep	Gram-positive*	Mesophilic	DL-Lactate	Sulfate	LS4D
<i>Desulfomicrobium baculatum</i> X	DMB	DSM 4028	Manganese ore	Gram-negative	Mesophilic	DL-Lactate	Sulfate	LS4D
<i>Pseudodesulfovibrio indicus</i> I2	PDI	DSM 101483	Deep-sea serpentinized peridotite	Gram-negative	Mesophilic	DL-Lactate	Sulfate	LS4D
<i>Thermodesulfatator autotrophicus</i> S606	TDA	DSM 101864	Deep-sea chimney wall	Gram-negative	Thermophilic	Hydrogen	Sulfate	SO4PNsalts
<i>Halodesulfovibrio marinesediminis</i> C/L 2	HDM	DSM 17456	Coastal sediment	Gram-negative	Mesophilic	DL-Lactate	Sulfate	LS4D

Note: * *Desulfoformulus ruminis* DL has Gram-positive-type cell wall structure

B

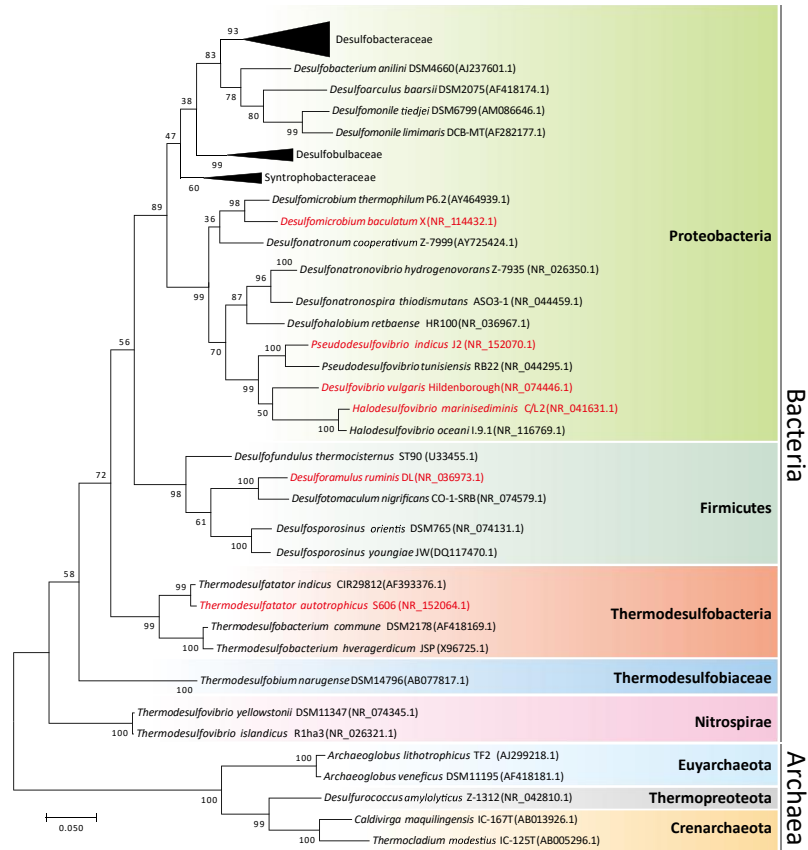


Fig. S1. Phylogenetic and physiological information on the isolated sulfate reducing microorganisms (SRMs) selected in this study. A, Basic phylogenetic and physiological information on the selected SRMs; **B,** Phylogenetic tree based on the full-length 16S rRNA gene sequences of representative SRMs. The SRMs selected in this study are in red font. The sequences were obtained from the NCBI database and the tree was created using MEGA 7 software. The bootstrap values at nodes are based on 1,000 replicates. The scale bar represents a 5 bp substitution per 100 nucleotides.

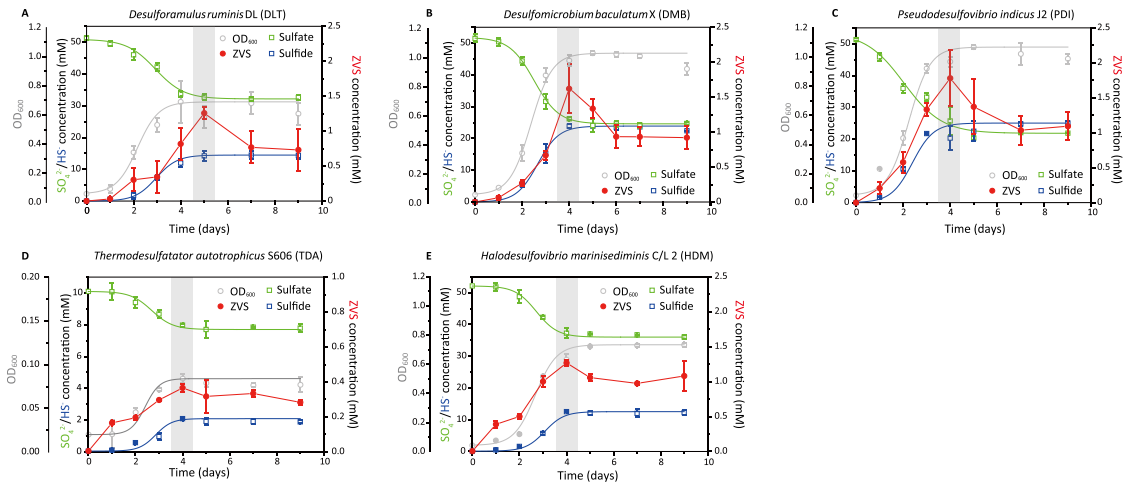


Fig. S2. Dissimilatory sulfate reduction and ZVS generation in five pure cultures of SRMs. **A**, *Desulfotomaculum ruminis* DL (DLT); **B**, *Desulfomicrobium baculatum* X (DMB); **C**, *Pseudodesulf vibrio indicus* J2 (PDI); **D**, *Thermodesulfatator autotrophicus* S606 (TDA); **E**, *Halodesulf vibrio marinisediminis* C/L 2 (HDM). The peak time for accumulation of the highest concentrations of total ZVS (ZVS peak time) in the five SRM cultures were shaded in grey. Error bars represent standard deviations of triplicate cultures.

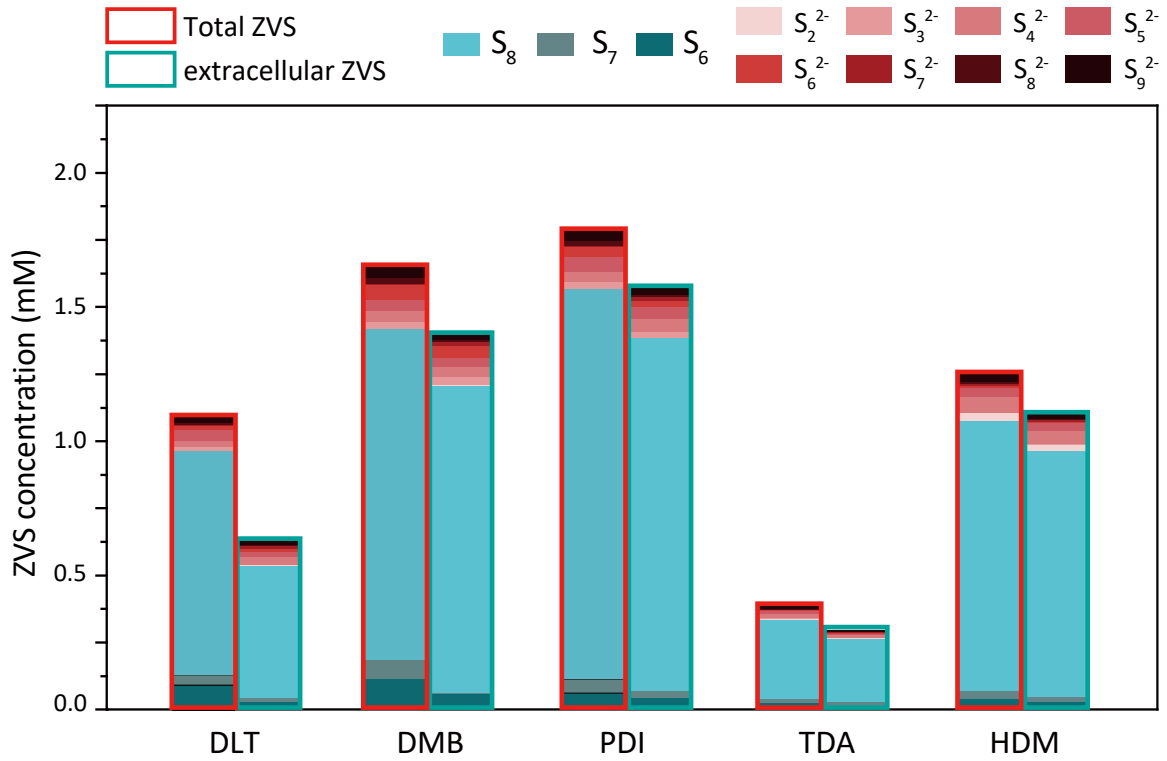


Fig. S3. The total/extracellular ZVS generation of five SRM pure cultures under their optimum growth conditions.

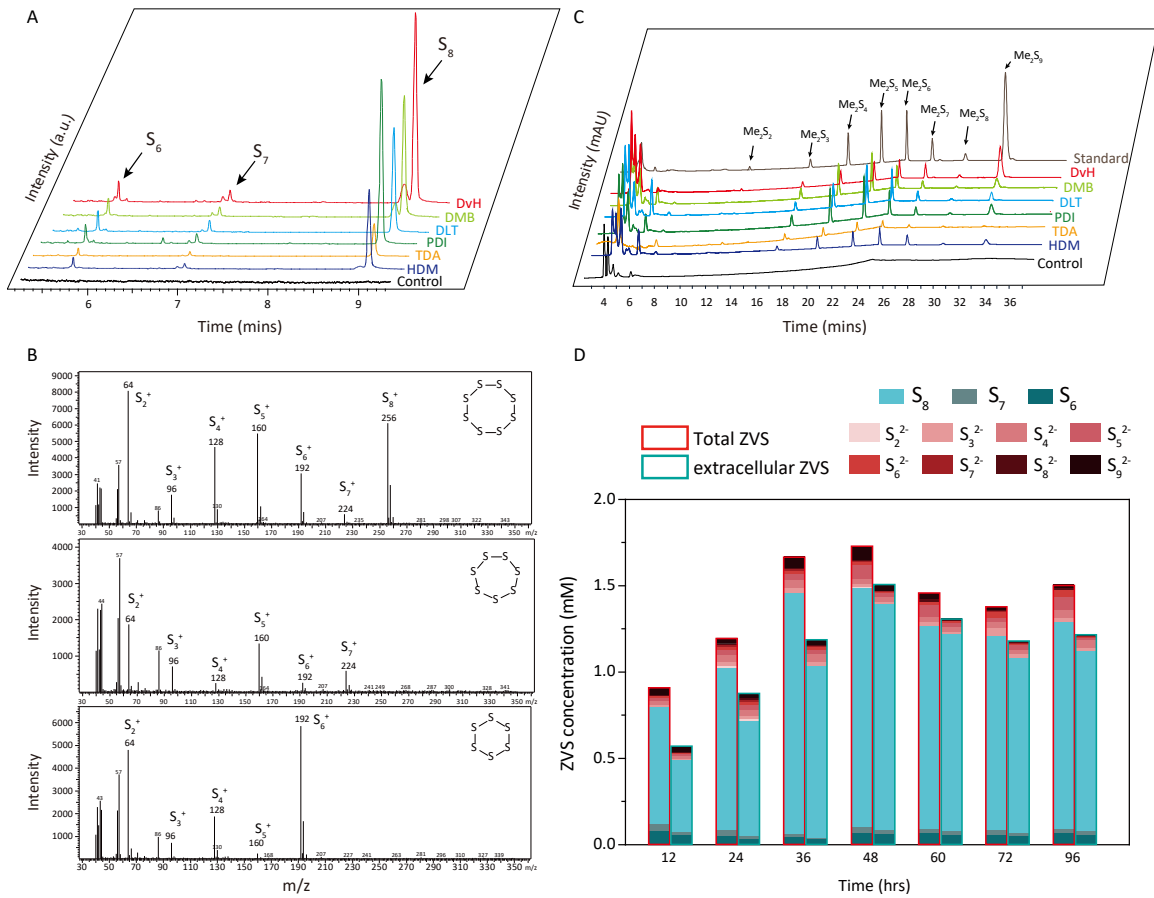


Fig. S4. Identification and characterization of DSR-derived ZVS composition. **A**, GC-MS chromatogram and **B**, mass spectrum of major elemental sulfur species in the six SRM pure cultures. **C**, HPLC chromatogram of polysulfide species in the six SRM pure cultures. **D**, Temporal change in compositional profiles of total/extracellular ZVS in DvH.

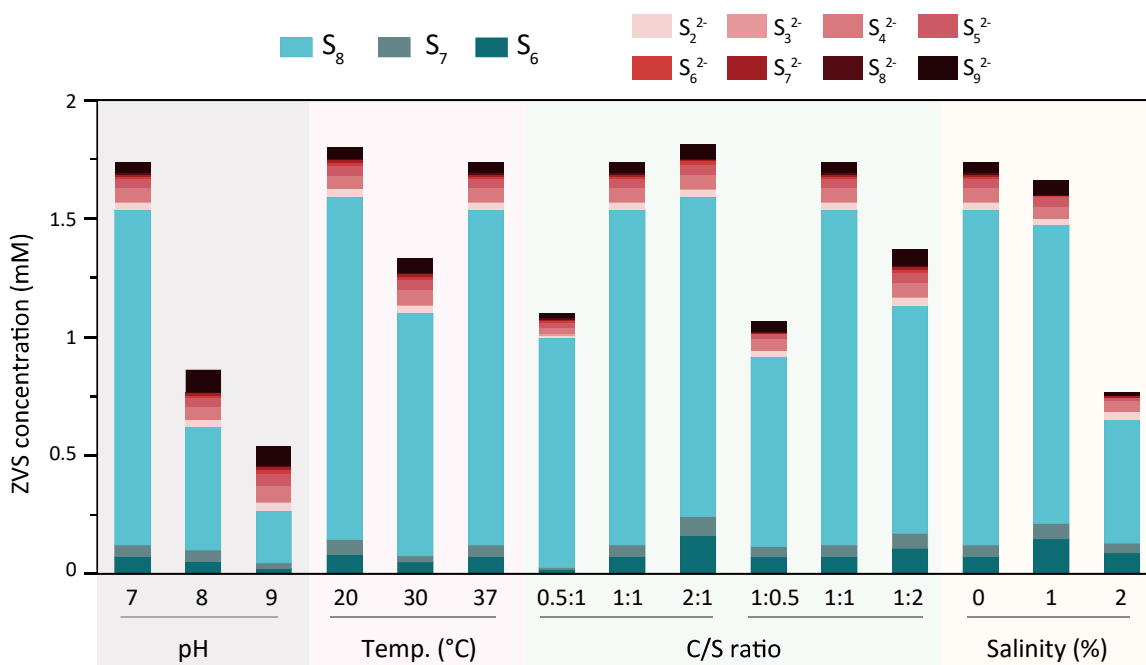


Fig. S5. Compositional profiles of ZVS in DvH cultures under different growth conditions, including different pH (7, 8, and 9), temperature (20, 30, and 37°C), lactate(C) to sulfate(S) ratio (0.5:1, 1:1, 2:1, 1:0.5, 1:1, and 1:2), and Salinity (0, 1, and 2%).

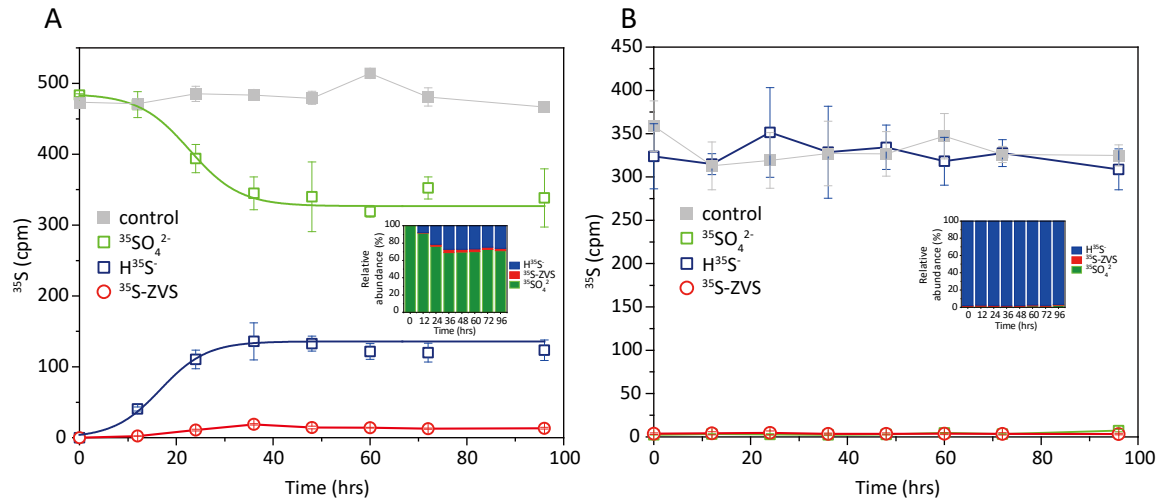


Fig. S6. Temporal changes in ^{35}S -activities of different sulfur species and their relative abundance in DvH cultures fed with ^{35}S -sulfate (A) and ^{35}S -sulfide (B).

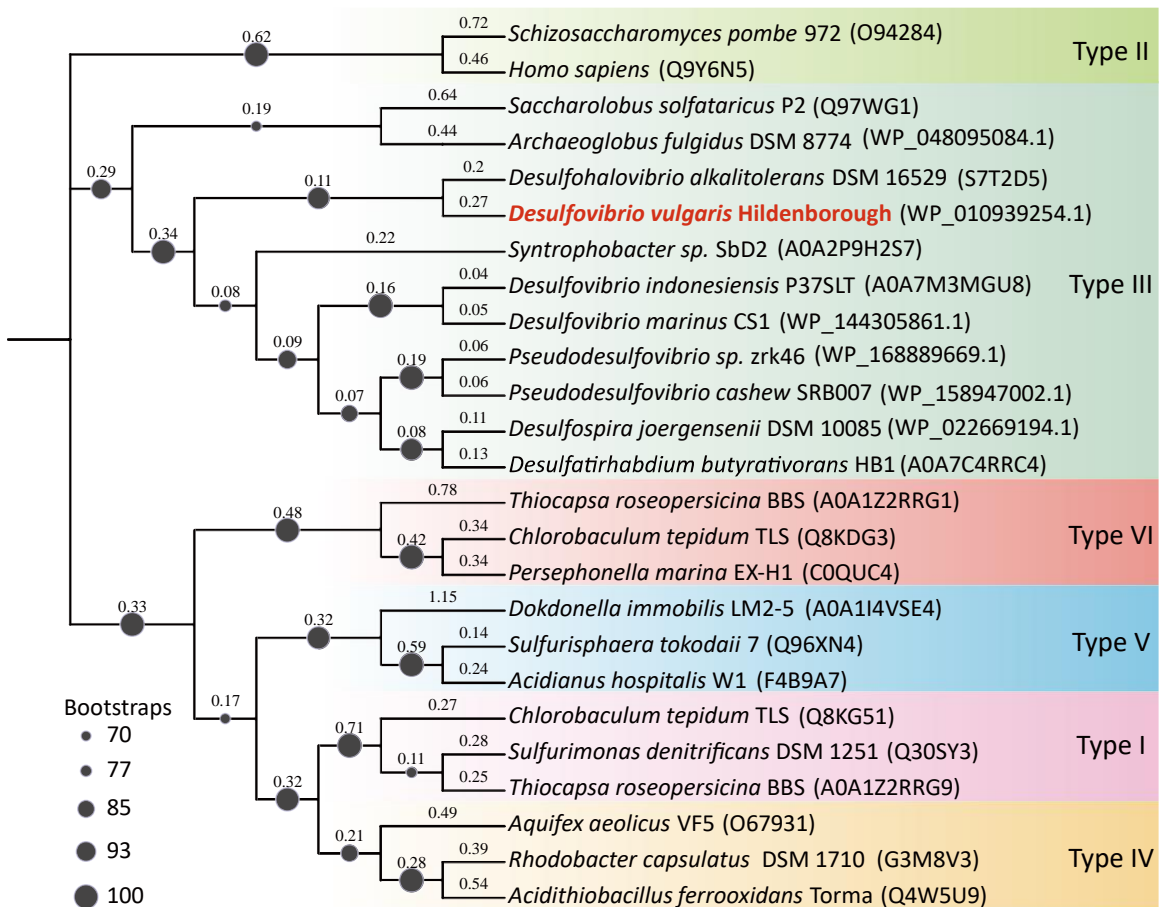


Fig. S7. Phylogeny of *sqr* homologue gene sequences from both SRMs and sulfur-oxidizing bacteria (SOB). Deduced amino acid sequences of the *sqr* homologue genes are analyzed here, and the phylogenetic tree is inferred and reconstructed under the maximum likelihood criterion using MEGA 7. Bootstrap values (%) >70 are indicated at the base of each node (black nots with different sizes). The numbers along branch represent the tree scale of each branch.

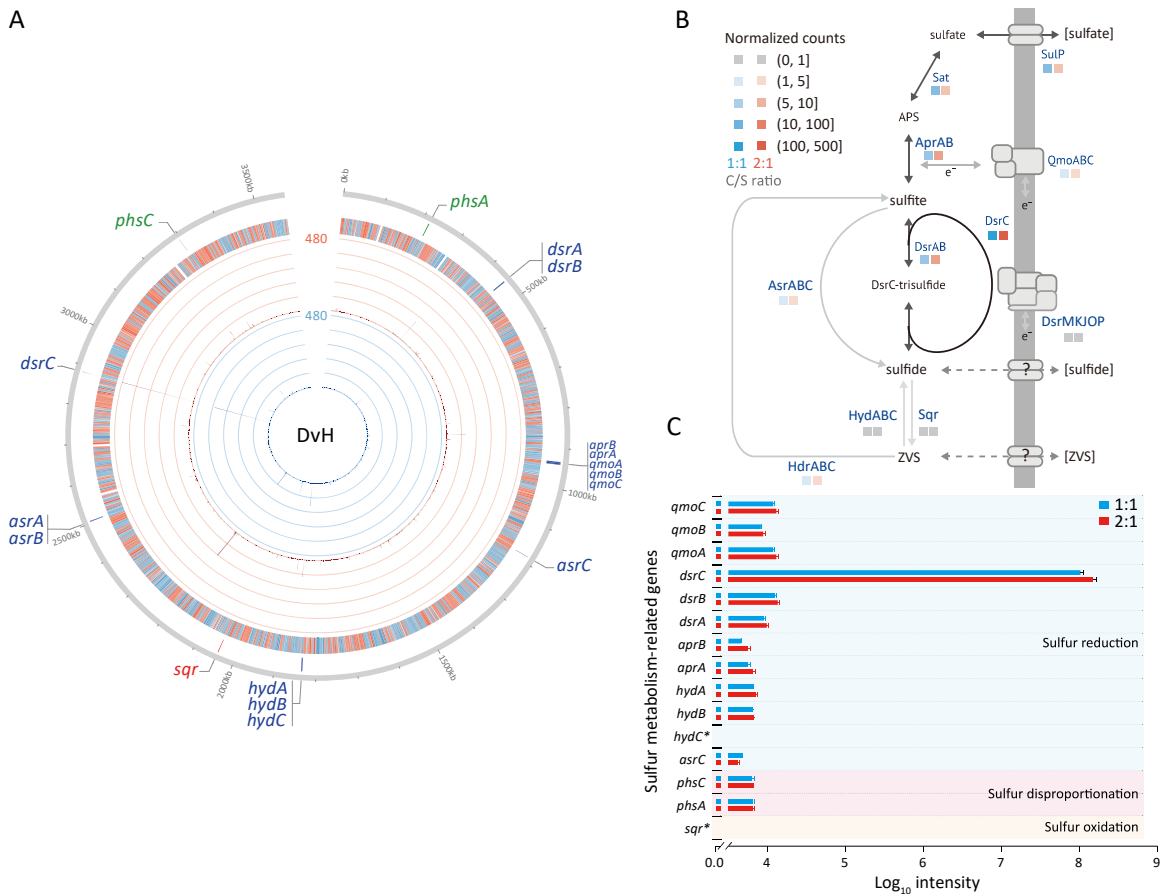


Fig. S8. Transcriptional analyses of DvH under different growth conditions. **A**, Circos plot showing the transcription levels of all genes for DvH cultures fed with two different molar ratios of lactate to sulfate, i.e., C/S ratios of 1:1 and 2:1. Tracks from inside to outside: track 1 in blue, DvH with the C/S ratio of 1:1; track 2 in red, DvH with the C/S ratio of 2:1; track 3, predicted ORFs in DvH genome (colors: red, forward strand; blue, reverse strand); track 4, predicted sulfur metabolism-related genes (colors: blue, dissimilatory sulfate reduction-related genes; red, sulfur oxidation-related genes; green, sulfur disproportionation-related genes). **B**, Transcription levels of genes encoding the sulfur metabolism-related enzymes in DvH cultures under the two growth conditions. Normalized counts were calculated on the basis of mean change in cycle threshold (Δ Ct) compared to the *recA*. **C**, Translation of key sulfur metabolism-related genes in DvH cultures under the two growth conditions. Protein relative abundances were normalized using *RecA* as the reference. Enzymes (i.e., Sqr and HydC) with translation levels under the detection limit were marked with asterisks (*). Sat, sulfate adenylyltransferase; Apr, adenosine-5'-phosphosulfate reductase; Dsr, dissimilatory sulfite reductase; DsrMKJOP, transmembrane complex; Asr, anaerobic sulfite reductase; Sqr, sulfide-quinone reductase; Qmo, quinone-interacting membrane bound oxidoreductase; Hdr, heterodisulfide reductase; Hyd, heterotrimeric electron-bifurcating hydrogenase; SulP, Sulfate permease.

Table S1. Information on genes with a higher transcription level in DvH cultures with the C/S ratio of 2:1 relative to them of DvH cultures with the C/S ratio of 1:1.

Locus_tag	Start ^a	End	Strand	Length	Annotation	KO assigned	Normalized counts (NC)			P-value
							1:1	2:1	2:1/1:1 ratio	
DVU_RS06585	1466090	1466626	-	537	Bacterioferritin	K03594	4.79	19.43	4.05	0.001
DVU_RS03670	864023	864259	+	237	Hypothetical protein	None	27.94	102.41	3.67	0.006
DVU_RS00100	27468	27635	+	168	Transcriptional regulator	None	9.87	29.60	3.00	0.057
DVU_RS03665	863403	863747	-	345	Hypothetical protein	None	7.89	21.42	2.71	0.026
DVU_RS13930	3071634	3071930	+	297	Hypothetical protein	None	21.76	48.32	2.22	0.011
DVU_RS06695	1495384	1496415	-	1032	OmpA family protein	K03286	8.57	18.80	2.20	0.011
DVU_RS14475	3222373	3222768	+	396	MucR family transcriptional regulator	None	12.68	26.46	2.09	0.000
DVU_RS14535	3236521	3236745	-	225	Rubredoxin	None	8.77	17.90	2.04	0.001
DVU_RS09905	2189295	2189708	-	414	Universal stress protein	None	12.87	24.94	1.94	0.025
DVU_RS06940	1554879	1555124	-	246	Hypothetical protein	None	11.34	21.22	1.87	0.041
DVU_RS01985	455770	456411	+	642	Septal ring lytic transglycosylase RlpA family protein	K03642	11.81	20.48	1.73	0.000
DVU_RS10905	2422628	2422822	+	195	Heavy-metal-associated domain-containing protein	K07213	56.10	96.94	1.73	0.031
DVU_RS14490	3226519	3226902	-	384	MucR family transcriptional regulator	None	17.84	29.33	1.64	0.001
DVU_RS11375	2532470	2532925	-	456	Glycine zipper 2TM domain-containing protein	K06077	22.30	36.03	1.62	0.000
DVU_RS09920	2197893	2198753	-	861	4Fe-4S binding protein	None	109.95	174.79	1.59	0.018
DVU_RS08105	1781209	1781433	-	225	DUF2188 domain-containing protein	None	15.07	22.90	1.52	0.000
DVU_RS04350	1010561	1010887	-	327	ATP synthase F0 subunit C	K02110	20.57	31.18	1.52	0.028
DVU_RS03450	813205	813657	+	453	Hypothetical protein	None	15.56	23.21	1.49	0.016
DVU_RS12420	2765700	2766248	-	549	Hypothetical protein	None	29.77	43.56	1.46	0.002
DVU_RS03785	884382	885779	-	1398	Outer membrane homotrimeric porin	None	16.24	23.31	1.44	0.008
DVU_RS01515	350418	350609	-	192	Ferredoxin	K05337	95.07	136.25	1.43	0.022
DVU_RS14905	3327925	3328311	+	387	Cytochrome c3 family protein	None	34.89	49.94	1.43	0.001
DVU_RS08625	1881750	1882058	-	309	C-type cytochrome	K08738	48.85	64.10	1.31	0.040

Table S1. Information on genes with a higher transcription level in DvH cultures with the C/S ratio of 2:1 relative to them of DvH cultures with the C/S ratio of 1:1. (Continued)

Locus_tag	Start ^a	End	Strand	Length	Annotation	KO assigned	Normalized counts (NC)			P-value
							1:1	2:1	2:1/1:1 ratio	
DVU_RS09040	1980 943	1981 416	-	474	Purine-binding chemotaxis protein CheW	K03408	18.39	23.82	1.30	0.00
DVU_RS13015	2883 462	2883 776	+	315	TusE/DsrC/DsvC family sulfur relay protein	K23077	369.83	472.75	1.28	0.058
DVU_RS10470	2314 522	2314 788	+	267	Hypothetical protein	None	16.39	20.18	1.23	0.068
DVU_RS01970	4524 27	4526 60	+	234	Protein DsvD	None	69.03	83.60	1.21	0.037
DVU_RS15420	3452 150	3452 335	-	186	Ferredoxin family protein	None	270.33	319.14	1.18	0.061
DVU_RS13295	2943 651	2944 172	+	522	Hypothetical protein	None	17.65	20.30	1.15	0.140
DVU_RS02440	5774 21	5776 81	-	261	30S ribosomal protein S15	K02956	21.71	24.70	1.14	0.215
DVU_RS03965	9268 07	9270 37	-	231	KH domain-containing protein	K06960	23.36	26.06	1.12	0.020
DVU_RS03950	9245 60	9249 04	-	345	50S ribosomal protein L19	K02884	17.99	20.02	1.11	0.196
DVU_RS03970	9271 24	9273 60	-	237	30S ribosomal protein S16	K02959	26.86	29.68	1.11	0.060
DVU_RS16690	1016 065	1016 208	+	144	Hypothetical protein	None	30.09	32.74	1.09	0.094
DVU_RS07410	1650 124	1650 630	+	507	Ferritin	K02217	21.33	22.48	1.05	0.091
DVU_RS14585	3244 295	3244 795	+	501	Peptidoglycan-associated lipoprotein Pal	K03640	60.38	61.54	1.02	0.333

Note: a, start and end position of each gene was based on the genome information of *Desulfovibrio vulgaris* Hildenborough (accession: NC_002937.3)

Table S2. Isolated and characterized strains of ZVS-metabolizing microorganisms including ZVS-reducing-microorganisms (S⁰RM) and ZVS-disproportionating-microorganisms (S⁰DM).

Species of S ⁰ RM	Phylum	Source	Electron donor & Carbon source	References
<i>Desulfuromonas acetoxidans</i>	Proteobacteria	Sea water	Acetate, ethanol, propanol	1
<i>Desulfuromonas acetexigens</i>	Proteobacteria	Freshwater sediment	Acetate	2
<i>Hippea maritima</i>	Proteobacteria	Marine hot vent	Acetate, H ₂	3
<i>Geobacter sulfurreducens</i>	Proteobacteria	Freshwater sediment	Acetate	4
<i>Pelobacter carbinolicus</i>	Proteobacteria	Freshwater sediment	Ethanol, H ₂	5
<i>Desulfurella acetivorans</i>	Proteobacteria	Hot spring	Acetate	6
<i>Acidianus infernus</i>	Crenarchaeota	Marine hot vent	Glucose, acetate, butyrate, H ₂	7
<i>Sulfolobus ambivalens</i>	Crenarchaeota	Solfataric waterhole	CO ₂ , H ₂	8
<i>Pyrodictium occultum</i>	Crenarchaeota	Hot sea floor	CO ₂ , H ₂	9
<i>Pyrobaculum neutrophilum</i>	Crenarchaeota	Hot spring	CO ₂ , H ₂	9
<i>Pyrobaculum maritimus</i>	Crenarchaeota	Hot sea water	CO ₂ , H ₂	9
<i>Stygiolobus azoricus</i>	Crenarchaeota	Solfataric field	CO ₂ , H ₂	10
Species of S ⁰ DM	Phylum	Source	Carbon source	References
<i>Dissulfurimicrobium hydrothermale</i>	Proteobacteria	Hydrothermal pond	Succinate	11
<i>Dissulfurispira thermophila</i>	Nitrospirota	Hot spring	CO ₂	12
<i>Dissulfurirhabdus thermomarina</i>	Proteobacteria	Hydrothermal vents	CO ₂	13
<i>Dissulfuribacter thermophilus</i>	Proteobacteria	Hydrothermal vents	CO ₂	14
<i>Caldimicrobium thiodismutans</i>	Thermodesulfobacteria	Hot spring	CO ₂	15
<i>Desulfocapsa sulfoexigens</i>	Proteobacteria	Marine sediment	Acetate	16
<i>Desulfurella amilsii</i>	Proteobacteria	River sediment	Acetate	17
<i>Dethiobacter alkaliphilus</i>	Firmicutes	Soda Lake	Acetate	18
<i>Pantoea agglomerans</i>	Proteobacteria	Marine sediment	Acetate	19
<i>Thermosulfurimonas dismutans</i>	Thermodesulfobacteria	Hydrothermal vents	CO ₂	20
<i>Thermosulfuriphilus ammonigenes</i>	Thermodesulfobacteria	Hydrothermal vents	CO ₂	21

Table S3. Gibbs free energy of reactions involving the dissimilatory reduction of sulfate and ZVS.

Equations	ΔG° , kJ/mol of	
	Reaction	Electron
$SO_4^{2-} + 2Lactate^- \rightarrow 2Acetate^- + HS^- + 2HCO_3^- + H^+$	-160.67	-20.08
$SO_4^{2-} + 2/3Lactate^- \rightarrow HS^- + 2HCO_3^- + 1/3H^+$	-85.2	-10.65
$SO_4^{2-} + 3/2Lactate^- + 1/2H^+ \rightarrow 3/2Acetate^- + S_0 + 3/2HCO_3^- + H_2O$	-130.725	-21.78
$SO_4^{2-} + 1/2Lactate^- + H^+ \rightarrow S_0 + 3/2HCO_3^- + H_2O$	-74.13	-12.36
$Acetate^- + 4S_0 + 4H_2O \rightarrow 4HS^- + 2HCO_3^- + 5H^+$	-3.29	-6.58
$SO_4^{2-} + 2Lactate^- + 2H^+ \rightarrow 2Acetate^- + H_2S(g) + 2CO_2(g) + 2H_2O$	-175.95	-21.99
$SO_4^{2-} + 2/3Lactate^- + 8/3H^+ \rightarrow H_2S(g) + 2CO_2(g) + 2H_2O$	-100.4	-12.55
$SO_4^{2-} + 3/2Lactate^- + 2H^+ \rightarrow 3/2Acetate^- + S_0 + 2/3CO_2(g) + 5/2H_2O$	-130.93	-21.82
$SO_4^{2-} + 1/2Lactate^- + 5/2H^+ \rightarrow S_0 + 2/3CO_2(g) + 5/2H_2O$	-74.33	-12.39
$Acetate^- + 4S_0 + H^+ + 2H_2O \rightarrow 4H_2S(g) + 2CO_2(g)$	-39.08	-19.54

Table S4. 16S rRNA gene amplicon sequencing-based metadata of global marine/terrestrial sites with presence of S⁰MMs (i.e., S⁰RM and S⁰DM) and SRMs.

No	Type	Location	Coordinates	S ⁰ RB		S ⁰ DB		SRB		Accession	Ref.
				Taxonomy	R.A. (%) ^b	Taxonomy	R.A. (%)	Taxonomy	R.A. (%)		
1	Soil	Antarctica	62.16S, 58.96W	<i>Geobacter</i>	6.16E-03	N.A.	N.A.	<i>Desulforhopalus</i> <i>Desulfobacterium</i>	5.70E-01	SRR16996395	22
2	Soil	Antarctica	74.76S, 164.01E	<i>Desulfuromonas</i>	2.75E-03	N.A.	N.A.	<i>Desulforhopalus</i> <i>Desulfobacterium</i> <i>Desulfobulbus</i>	1.57E+00	SRR16996404	22
3	Soil	Antarctica	62.18S, 58.97W	<i>Geobacter</i> <i>Desulfuromonas</i>	9.95E-03	N.A.	N.A.	<i>Desulforhopalus</i> <i>Desulfobacterium</i> <i>Desulfobacterium</i>	5.27E-01	SRR16996409	22
4	Soil	Antarctica	72.31S, 170.26E	<i>Desulfuromonas</i>	2.33E-02	<i>Pantoea</i>	4.89E-03	<i>Desulforhopalus</i> <i>Desulfobacterium</i> <i>Desulfobacterium</i>	8.43E+00	SRR16996416	22
5	Soil	Antarctica	77.21S, 166.44E	<i>Geobacter</i> <i>Desulfuromonas</i>	7.06E-04	N.A.	N.A.	<i>Desulforhopalus</i> <i>Desulfobacterium</i> <i>Desulfobacterium</i>	2.08E-02	SRR16996428	22
6	Soil	Antarctica	64.89S, 62.87W	<i>Geobacter</i>	2.03E-02	N.A.	N.A.	<i>Desulfosarcina</i> <i>Desulfosporosinus</i>	1.56E-02	SRR17050022	22
7	Soil	Antarctica	68.49S, 78.10E	<i>Geobacter</i> <i>Desulfuromonas</i>	1.05E-02	N.A.	N.A.	<i>Desulfosporosinus</i>	3.28E-02	SRR16979831	22
8	Soil	Petersham, U.S.	42.5 N, 72.19 W	<i>Geobacter</i>	2.84E-03	N.A.	N.A.	<i>Desulfosporosinus</i>	8.51E-03	c	23
9	Soil	Wheelman, U.S.	39.99N, 105.37 W	<i>Pelobacter</i> <i>Geobacter</i>	1.13E-02	<i>Pantoea</i>	7.99E-02	<i>Desulfosporosinus</i>	1.88E-03	c	23
10	Soil	Swaziland	46.63N, 10.37E	<i>Geobacter</i>	1.24E-01	N.A.	N.A.	<i>Desulfosporosinus</i> <i>Desulfobulbus</i>	6.10E-03	c	23
11	Soil	Australia	33.47 S, 145.55E	<i>Geobacter</i> <i>Pelobacter</i>	3.47E-01	N.A.	N.A.	<i>Desulfobulbus</i> <i>Desulfosporosinus</i>	1.24E-01	c	23
12	Soil	Muztag Ata glacier, China	38.26N, 75.00E	<i>Geobacter</i>	4.87E-02	<i>Pantoea</i>	1.03E-02	<i>Desulfosporosinus</i>	2.80E-02	SRR8156999	24
13	Soil	Barrow Peninsula, US	71.33N, 156.11W	<i>Geobacter</i>	8.06E-02	<i>Desulfocapsa</i> <i>Pantoea</i>	6.69E-02	<i>Desulfatiglans</i> <i>Desulfosporosinus</i>	5.43E+00	SRR9680643	25
14	Soil	Kirs, Russia	59.23N, 52.23E	<i>Geobacter</i> <i>Desulfuromonas</i>	1.70E-01	N.A.	N.A.	<i>Desulfobulbus</i> <i>Desulfococcus</i> <i>Desulfosarcina</i>	2.56E+00	ERR1779761	26
15	Soil	Al Farafra Desert, Egypt	26.24N, 25.35E	<i>Geobacter</i> <i>Desulfuromonas</i>	1.00E-01	N.A.	N.A.	<i>Desulfobulbus</i> <i>Desulfuromonas</i> <i>Thermodesulfobivrio</i>	1.59E+00	ERR1779766	26

Table S4. 16S rRNA gene amplicon sequencing-based metadata of global marine/terrestrial sites with presence of S⁰MMs (i.e., S⁰RM and S⁰DM) and SRMs. (Continued)

No	Type	Location	Coordinates	S ⁰ RB		S ⁰ DB		SRB		Accession	Ref.
				Taxonomy	R.A. (%)	Taxonomy	R.A. (%)	Taxonomy	R.A. (%)		
16	Soil	Poltava, Ukraine	49.50N, 34.70E	<i>Geobacter Desulfuro monas</i>	1.44E-01	N.A.	N.A.	<i>Desulfobulbus</i> <i>Thermodesulfobulbus</i>	3.98E+00	ERR1779774	26
17	Soil	Nalut Province, Libya	30.05N, 10.11E	<i>Geobacter Desulfuro monas</i>	5.24E-02	N.A.	N.A.	<i>Desulfobulbus</i> <i>Desulfococcus</i>	2.97E-01	ERR1779777	26
18	Marine sediment	Caspian Sea	39.74 N, 50.48 E	<i>Geobacter</i>	8.00E-03	<i>Desulfocapsa</i>	6.62E-04	<i>Desulfobulbus</i> <i>Desulfococcus</i> <i>Desulfomonile</i>	6.61E-01	SRR1583209	27
19	Marine sediment	Caspian Sea	39.99 N, 51.50 E	<i>Geobacter Desulfuro monas</i>	4.37E-02	<i>Desulfocapsa</i>	2.95E-04	<i>Desulfobulbus</i> <i>Desulfococcus</i> <i>Desulfosarcina</i> <i>Desulfomonile</i>	6.55E-01	SRR1583215	27
20	Marine sediment	Caspian Sea	40.04 N, 51.34 E	<i>Desulfuro monas</i>	5.62E-02	<i>Desulfocapsa</i>	4.56E-03	<i>Desulfobulbus</i> <i>Desulfococcus</i> <i>Desulfosarcina</i>	4.16E-01	SRR1583242	27
21	Marine sediment	Greenland Sea	77.32 N, 15.39 E	<i>Geobacter Desulfuro monas</i>	3.49E-01	<i>Desulfoporosinus</i> <i>Desulfocapsa</i>	2.52E-02	<i>Desulfobulbus</i> <i>Desulfococcus</i> <i>Desulfosarcina</i>	6.76E+00	SRR7969004	28
22	Marine sediment	French Guiana	5.54 N, 53.17W	<i>Geobacter Desulfuro monas</i> <i>Pelobacter</i>	1.78E+00	<i>Desulfocapsa</i>	1.13E-02	<i>Desulfobulbus</i> <i>Desulfococcus</i> <i>Desulfosarcina</i>	3.23E+00	SRR027243	29
23	Marine sediment	French Guiana	5.54 N, 52.11W	<i>Desulfuro monas</i> <i>Pelobacter</i>	1.70E-01	<i>Desulfocapsa</i>	1.38E-02	<i>Desulfobulbus</i> <i>Desulfococcus</i> <i>Desulfosarcina</i>	3.30E+00	SRR027245	29
24	Marine sediment	Kern River, England	51.87 N, 0.93 E	<i>Geobacter Desulfuro monas</i> <i>Pelobacter</i>	1.06E-01	<i>Desulfocapsa</i>	5.62E-02	<i>Desulfobulbus</i> <i>Desulfococcus</i> <i>Desulfosarcina</i>	3.21E+00	SRR027362	29
25	Marine sediment	Schimonnik Island, Netherlands	53.49 N, 6.13 E	<i>Desulfuro monas</i>	1.01E-01	<i>Desulfocapsa</i>	7.17E-02	<i>Desulfobulbus</i> <i>Desulfococcus</i> <i>Desulfosarcina</i>	4.11E-01	SRR027371	29
26	Marine sediment	La Parguera Natural Reserve, Puerto Rico	17.87 N, 67.04 W	<i>Pelobacter</i>	5.83E-02	<i>Desulfocapsa</i>	1.46E-02	<i>Desulfobulbus</i> <i>Desulfococcus</i>	1.17E-01	SRR027679	29
27	Marine sediment	Peninsula Valdes, Argentina	42.42 S, 64.11 W	<i>Desulfuro monas</i> <i>Pelobacter</i>	5.75E-02	<i>Desulfocapsa</i>	1.44E-02	<i>Desulfobulbus</i> <i>Desulfococcus</i> <i>Desulfosarcina</i>	2.97E-01	SRR027686	29

Table S4. 16S rRNA gene amplicon sequencing-based metadata of global marine/terrestrial sites with presence of S⁰MMs (i.e., S⁰RM and S⁰DM) and SRMs. (Continued)

No	Type	Location	Coordinates	S ⁰ RB		S ⁰ DB		SRB		Accession	Ref.
				Taxonomy	R.A. (%)	Taxonomy	R.A. (%)	Taxonomy	R.A. (%)		
28	Marine sediment	Talcahuano, Chile	36.69 S, 73.07 W	<i>Desulfuromonas</i>	1.27E-02	<i>Desulfocapsa</i>	4.13E-02	<i>Desulfobulbus</i> <i>Desulfobacter</i>	2.38E+00	SRR027975	29
29	Marine sediment	Talcahuano, Chile	36.64 S, 73.04 W	<i>Geobacter</i>	2.51E-02	<i>Desulfocapsa</i>	2.19E-02	<i>Desulfobulbus</i> <i>Desulfococcus</i> <i>Desulfobacterium</i>	1.42E+00	SRR027976	29
30	Marine sediment	Arabian sea	10.10 N, 59.29 E	<i>Geobacter</i>	4.76E-02	<i>Desulfocapsa</i>	4.04E-03	<i>Desulfobulbus</i> <i>Desulfococcus</i>	1.62E-01	ERR1779755	26
31	Marine sediment	Arabian sea	10.43 N, 57.46 E	<i>Geobacter</i> <i>Desulfuromonas</i>	8.36E-02	<i>Desulfocapsa</i>	4.19E-03	<i>Desulfobulbus</i> <i>Desulfococcus</i>	2.93E-01	ERR1779758	26
32	Marine sediment	Gulf of Aden	15.51 N, 44.97 E	<i>Geobacter</i> <i>Desulfuromonas</i>	6.25E-02	<i>Desulfocapsa</i>	4.39E-02	<i>Desulfobulbus</i> <i>Desulfococcus</i> <i>Desulfosarcina</i>	1.80E+00	ERR1779761	26
33	Marine sediment	Atlantic Ocean	52.16 S, 14.17 E	<i>Geobacter</i> <i>Desulfuromonas</i>	3.21E-02	<i>Desulfocapsa</i>	3.94E-02	<i>Desulfatignans</i> <i>Desulforhopalus</i> <i>Desulfobulbus</i>	3.06E-01	S_PRJE B23821	30
34	Marine sediment	Bering Sea	55.64 N, 179.46 E	<i>Pelobacter</i>	3.30E-01	<i>Pantoea</i>	4.77E-02	<i>Desulfatignans</i> <i>Desulfobulbus</i> <i>Desulfococcus</i>	3.88E+00	SRR1631774	31
35	Marine sediment	Pacific Ocean	4.88 N, 141.75 W	<i>Geobacter</i>	6.91E-03	N.A.	N.A.	<i>Desulfotrigonis</i>	4.87E-03	SRR12576035	32
36	Marine sediment	Pacific Ocean	13.76 N, 116.45 W	<i>Geobacter</i>	8.38E-03	N.A.	N.A.	<i>Desulfotrigonis</i>	8.38E-03	SRR1257636	32
37	Marine sediment	Forlandsundet	78.59 N, 11.39 E	<i>Desulfuromonas</i>	1.23E-01	<i>Desulfocapsa</i>	4.28E-01	<i>Desulforhopalus</i> <i>Desulfobulbus</i> <i>Desulfococcus</i>	9.15E+00	SRR4237752	33
38	Marine sediment	Atlantic Ocean	33.48 N, 54.16 W	<i>Desulfuromonas</i>	7.13E-03	<i>Desulfocapsa</i>	6.19E-02	<i>Desulfatignans</i> <i>Desulfobulbus</i>	1.56E+00	SRR6396235	34
39	Marine sediment	Okinawa Trough	30.53 N, 128.00 E	<i>Geobacter</i> <i>Desulfuromonas</i>	7.25E-02	<i>Desulfocapsa</i>	4.86E-03	<i>Desulfatignans</i> <i>Desulfobulbus</i> <i>Thermodesulfobivrio</i>	3.61E+00	SRR3416391	35
40	Marine sediment	Okinawa Trough	29.84 N, 127.63 E	<i>Geobacter</i> <i>Desulfuromonas</i>	2.23E-02	<i>Desulfocapsa</i>	2.52E-03	<i>Desulfatignans</i> <i>Desulfobulbus</i>	3.08E+00	SRR3416505	35

Table S4. 16S rRNA gene amplicon sequencing-based metadata of global marine/terrestrial sites with presence of S⁰MMs (i.e., S⁰RM and S⁰DM) and SRMs. (Continued)

No	Type	Location	Coordinates	S ⁰ RB		S ⁰ DB		SRB		Accession	Ref.
				Taxonomy	R.A. (%)	Taxonomy	R.A. (%)	Taxonomy	R.A. (%)		
41	Marine sediment	Indian Ocean	26.94 S, 67.32 E	<i>Geobacter</i>	2.19E-03	<i>Pantoea</i>	1.83E-03	<i>Desulfovibrio</i> <i>Desulfobulbus</i>	2.16E+00	SRR5007323	36
42	Marine sediment	Indian Ocean	26.76 S, 67.71 E	<i>Geobacter</i>	1.10E-03	<i>Pantoea</i>	7.34E-04	<i>Desulfovibrio</i>	3.13E-01	SRR5007326	36
43	Marine sediment	South China Sea	16.89 N, 112.35 E	<i>Geobacter</i>	3.50E-02	<i>Desulfocapsa</i>	4.07E-03	<i>Desulfovibrio</i> <i>Desulfococcus</i> <i>Desulfatiglas</i>	6.02E-01	SRR101434571	37
44	Marine sediment	South China Sea	16.97 N, 112.19 E	<i>Geobacter</i>	2.46E-02	<i>Pantoea</i>	2.00E-03	<i>Desulfovibrio</i> <i>Desulfococcus</i> <i>Desulfatiglas</i>	5.79E-01	SRR101434831	37
45	Marine sediment	North Pacific	47.45 N, 127.76 W	<i>Desulfuromonas</i>	5.30E-03	<i>Pantoea</i>	2.01E-03	<i>Desulfatiglas</i> <i>Desulfobulbus</i>	1.03E+00	DRR179137	38
46	Marine sediment	Gulf of Mexico	28.09 N, 89.02 W	<i>Desulfuromonas</i>	7.83E-03	<i>Pantoea</i>	7.57E-03	<i>Desulfatiglas</i>	1.43E+00	DRR179159	38
47	Marine sediment	North Pacific	33.22 N, 136.72 E	<i>Desulfuromonas</i>	5.14E-03	<i>Pantoea</i>	8.53E-03	<i>Desulfatiglas</i> <i>Desulfosporosinus</i>	5.09E-01	DRR179195	38
48	Marine sediment	Baltic Sea	58.62 N, 18.25 E	<i>Desulfuromonas</i>	1.61E-02	<i>Pantoea</i>	1.20E-02	<i>Desulfatiglas</i> <i>Desulfococcus</i>	5.48E-01	DRR179296	38
49	Marine sediment	Baltic Sea	55.46 N, 15.47 E	<i>Desulfuromonas</i>	3.44E-03	<i>Pantoea</i>	9.06E-02	<i>Desulfatiglas</i>	4.78E-01	DRR179305	38
50	Marine sediment	Bay of Bengal	14.00 N, 84.82 E	<i>Desulfuromonas</i>	5.26E-03	<i>Pantoea</i>	5.26E-03	<i>Desulfatiglas</i> <i>Desulfobulbus</i> <i>Desulfomonile</i>	1.09E+00	DRR179324	38
51	Marine sediment	North Atlantic ocean	33.48 N, 54.16 W	<i>Desulfuromonas</i>	3.07E-03	N.A.	N.A.	<i>Desulfatiglas</i> <i>Desulfobulbus</i>	1.90E-01	DRR179368	38
52	Marine sediment	Pacific Ocean	9.11 S, 80.58 W	<i>Desulfuromonas</i>	6.72E-03	<i>Desulfocapsa</i>	2.94E-03	<i>Desulfatiglas</i> <i>Desulfobulbus</i>	3.77E+00	DRR181262	38
53	Hot spring sediment	Tibet, China	28.52 N, 91.88 E	<i>Geobacter</i> <i>Sulfobolus</i>	3.06E-02	<i>Caldimicrobium</i> <i>Desulfurella</i>	2.79E-02	<i>Thermodesulfobivrio</i> <i>Desulfosporosinus</i>	1.19E+00	SRR5313425	39
54	Hot spring sediment	Tibet, China	29.62 N, 85.74 E	<i>Geobacter</i> <i>Sulfobolus</i>	2.92E-02	<i>Caldimicrobium</i> <i>Desulfurella</i>	1.48E-02	<i>Thermodesulfobivrio</i> <i>Desulfosporosinus</i>	3.82E-01	SRR5313441	39

Table S4. 16S rRNA gene amplicon sequencing-based metadata of global marine/terrestrial sites with presence of S⁰MMs (i.e., S⁰RM and S⁰DM) and SRMs. (Continued)

No	Type	Location	Coordinates	S ⁰ RB		S ⁰ DB		SRB		Accession	Ref.
				Taxonomy	R.A. (%)	Taxonomy	R.A. (%)	Taxonomy	R.A. (%)		
55	Hot spring sediment	Tibet, China	31.61 N, 91.83 E	<i>Geobacter Sulfolobus</i>	3.15E-02	<i>Caldimicrobium Dethiobacter</i>	8.27E-02	<i>Thermodesulfobium Desulfobium Desulfobium</i>	1.62E-01	SRR5313461	39
56	Hot spring sediment	Tibet, China	28.25 N, 91.81 E	<i>Geobacter Sulfolobus</i>	1.93E-01	<i>Caldimicrobium Dethiobacter</i>	3.63E-02	<i>Desulfotomaculum Thermodesulfobium Desulfobium</i>	1.01E+00	SRR5313524	39
57	Hot spring sediment	Yunnan, China	24.95 N, 98.44 E	<i>Geobacter Sulfolobus</i>	7.76E-01	<i>Caldimicrobium Desulfurella</i>	5.65E-01	<i>Thermodesulfobium Thermodesulfobium Desulfobium</i>	6.18E-01	CRR144043	40
58	Hot spring sediment	Iceland	64.68N, 21.40W	<i>Geobacter</i>	5.52E-03	<i>Caldimicrobium Dethiobacter</i>	2.79E-03	<i>Desulfobulbus Desulfobium Thermodesulfobium</i>	1.58E-03	SRR11066918	41
59	Hot spring sediment	Iceland	64.13N, 20.30 W	<i>Geobacter</i>	1.59E-03	<i>Caldimicrobium Dethiobacter</i>	1.05E-02	<i>Desulfobulbus Desulfobium Thermodesulfobium</i>	8.73E-03	SRR11066925	41
60	Hot spring sediment	New Zealand	38.05 S, 176.36 E	<i>Geobacter</i>	3.36E-01	<i>Desulfurella</i>	1.35E-02	<i>Thermodesulfobium Thermodesulfobium</i>	1.62E-01	SRR129639161	42
61	AMD ^d sediment	Shaoguan, China	25.04 N, 113.66 E	<i>Geobacter Desulfuromonas</i>	9.31E-03	<i>Dethiobacter</i>	3.52E-02	<i>Desulfomonile Desulfosporosinus Desulfotomaculum</i>	3.57E-01	SRR8455939	43
62	AMD sediment	Guizhou, China	25.80N, 107.61E	<i>Geobacter Desulfuromonas</i>	2.85E+00	<i>Desulfocapsa Dethiobacter</i>	1.28E-01	<i>Desulfomonile Thermodesulfobium Desulfobium Desulfobulbus</i>	3.92E+00	SRR12726967	44
63	AMD sediment	Shaoguan, China	24.55 N, 113.74 E	<i>Geobacter</i>	2.82E-03	<i>Desulfocapsa</i>	2.04E-03	<i>Desulfobulbus Desulfomonile Desulfosporosinus</i>	5.18E-01	SRR12737887	44

Table S4. 16S rRNA gene amplicon sequencing-based metadata of global marine/terrestrial sites with presence of S⁰MMs (i.e., S⁰RM and S⁰DM) and SRMs. (Continued)

No	Type	Location	Coordinates	S ⁰ RB		S ⁰ DB		SRB		Accession	Ref.
				Taxonomy	R.A. (%)	Taxonomy	R.A. (%)	Taxonomy	R.A. (%)		
64	AMD sediment	Guangxi, China	25.14N, 107.50E	<i>Geobacter</i>	4.60E-02	<i>Dethiobacter Pantoea</i>	1.07E-02	<i>Desulfobulbus Desulfomonile Desulfosporosinus</i>	9.42E-02	SRR1274 4846	44
65	AMD sediment	Shangrao, Jiangxi, China	29.00 N, 117.72 E	<i>Geobacter</i>	3.86E-03	<i>Dethiobacter</i>	1.94E-03	<i>Desulfosporosinus Desulfotomaculum</i>	1.23E-02	SRR1274 5062	44
66	AMD sediment	Fankou, Guangdong, China	25.04 N, 113.66 E	<i>Geobacter</i>	1.42E-03	N.A.	N.A.	<i>Desulfobulbus Desulfomonile</i>	5.68E-02	SRR1276 8936	44
67	AMD sediment	Huangshaping, Hunan, China	25.67N, 112.70E	<i>Geobacter</i>	4.71E-03	<i>Dethiobacter Pantoea</i>	4.68E-03	<i>Desulfotomaculum Desulfosporosinus</i>	1.15E-02	SRR1276 9161	44
68	AMD sediment	Jiuan, Guizhou, China	26.52 N, 106.57 E	<i>Geobacter Desulfuromonas</i>	3.87E-03	<i>Dethiobacter Pantoea</i>	3.97E-03	<i>Desulfotomaculum Desulfosporosinus</i>	8.43E-03	SRR1276 9163	44
69	AMD sediment	Longli, Guizhou, China	26.38 N, 106.97 E	<i>Geobacter</i>	2.00E-01	<i>Desulfocapsa Pantoea</i>	5.67E-03	<i>Desulfobulbus Desulfobulbus</i>	8.92E-02	SRR1276 9170	44
70	AMD sediment	Longshenao, Guangxi, China	24.83 N, 107.56 E	<i>Geobacter</i>	3.23E-02	<i>Desulfocapsa</i>	8.32E-04	<i>Desulfobulbus Desulfobulbus</i>	3.90E-02	SRR1276 9176	44
71	AMD sediment	Manshan, Anhui, China	31.67 N, 118.62 E	<i>Geobacter</i>	1.56E-02	<i>Desulfocapsa</i>	9.94E-04	<i>Desulfobulbus Desulfomonile Desulfosporosinus</i>	2.62E-02	SRR1280 8556	44
72	AMD sediment	Shuikoushan, Hunan, China	26.53 N, 112.60 E	<i>Geobacter Desulfuromonas</i>	7.31E-01	<i>Desulfocapsa Desulfurella Dethiobacter Pantoea</i>	5.08E-02	<i>Desulfotomaculum Desulfobulbus Desulfomonile Desulfosporosinus</i>	7.48E-01	SRR1280 8561	44
73	AMD sediment	Tongling, Anhui, China	30.94 N, 117.98 E	<i>Geobacter</i>	1.28E-02	<i>Desulfocapsa</i>	4.66E-03	<i>Desulfobulbus Desulfomonile Desulfosporosinus</i>	2.23E-02	SRR1280 8565	44
74	AMD sediment	Wuhu, Anhui, China	31.08 N, 118.25 E	<i>Geobacter</i>	3.33E-03	<i>Desulfocapsa</i>	1.12E-03	<i>Desulfobulbus Desulfobulbus</i>	4.13E-02	SRR1281 6447	44
75	AMD sediment	Wuyi, Guangxi, China	25.14 N, 107.50 E	<i>Geobacter</i>	6.32E-01	<i>Desulfocapsa Dethiobacter Pantoea</i>	4.01E-02	<i>Desulfobulbus Desulfobulbus Desulfomonile</i>	9.75E-01	SRR1281 6452	44

Table S4. 16S rRNA gene amplicon sequencing-based metadata of global marine/terrestrial sites with presence of S⁰MMs (i.e., S⁰RM and S⁰DM) and SRMs. (Continued)

No	Type	Location	Coordinates	S ⁰ RB		S ⁰ DB		SRB		Accession	Ref.
				Taxonomy	R.A. (%)	Taxonomy	R.A. (%)	Taxonomy	R.A. (%)		
76	AMD sediment	Yunfu, Guangdong, China	22.96 N, 112.01E	<i>Geobacter</i>	4.77E-03	<i>Dethiobacter</i>	3.39E-03	<i>Desulfotomaculum Desulfomonile Desulfosporosinus</i>	1.24E-02	SRR12816507	44
77	AMD sediment	Yongping, Jiangxi, China	28.19 N, 117.76 E	<i>Geobacter</i>	4.43E-03	<i>Dethiobacter Pantoea</i>	4.40E-03	<i>Desulfotomaculum Desulfosporosinus</i>	9.87E-03	SRR12816510	44
78	AMD sediment	Zhijin, Guizhou, China	26.60 N, 105.72 E	<i>Geobacter</i>	1.35E-02	<i>Pantoea Desulfurella</i>	4.94E-03	<i>Desulfotomaculum Desulfosporosinus</i>	2.81E-02	SRR12834021	44
79	AMD sediment	Yinshan, Jiangxi, China	28.97N, 117.60 E	<i>Geobacter</i>	6.21E-03	<i>Dethiobacter Pantoea</i>	6.31E-03	<i>Desulfotomaculum Desulfosporosinus</i>	1.83E-02	SRR12834185	44
80	AMD sediment	Rio Tinto, Spain	37.59N, 6.55W	<i>Geobacter</i>	1.87E-02	N.A.	N.A.	<i>Desulfobulbus Desulfosporosinus</i>	1.11E-02	QFIRB012AB	45
81	AMD sediment	Rio Tinto, Spain	37.31N, 6.82W	<i>Geobacter Desulfuromonas</i>	3.51E+00	N.A.	N.A.	<i>Desulfotomaculum Desulfobulbus Desulfosporosinus Desulfobacterium Desulfosporosinus Desulfovibrio</i>	1.29E+01	QFIRB010AC	45
82	Freshwater sediment	Enshi, Hubei, China	31.04 N, 110.4 E	<i>Geobacter Desulfuromonas</i>	1.18E-01	<i>Desulfocapsa</i>	1.06E-03	<i>Desulfobulbus Desulfobacterium Desulfosporosinus Desulfovibrio</i>	1.78E-01	SRR3944723	46
83	Freshwater sediment	Changshou, Jiangsu, China	31.77 N, 120.96 E	<i>Geobacter Desulfuromonas</i>	1.05E-01	<i>Desulfocapsa</i>	1.12E-03	<i>Desulfobulbus Desulfomonile Desulfobulbus Desulfobacterium Desulfomonile Desulfosporosinus Desulfovibrio</i>	1.09E-01	SRR3944727	46
84	Freshwater sediment	Yibing, Sichuan, China	28.8 N, 104.42 E	<i>Geobacter Desulfuromonas</i>	2.38E-01	<i>Desulfocapsa</i>	1.27E-03	<i>Desulfobulbus Desulfobacterium Desulfomonile Desulfosporosinus Desulfovibrio</i>	2.45E-01	SRR3944737	46
85	Freshwater sediment	Guangan, Sichuan	30.35 N, 106.28 E	<i>Geobacter Desulfuromonas</i>	3.97E-01	<i>Desulfocapsa</i>	1.48E-03	<i>Desulfobulbus Desulfobacterium Desulfomonile Desulfosporosinus Desulfovibrio</i>	4.69E-01	SRR3944744	46
86	Freshwater sediment	Beipei, Chongqing	29.81 N, 106.46 E	<i>Geobacter Desulfuromonas</i>	3.51E-01	<i>Desulfocapsa</i>	1.06E-03	<i>Desulfobulbus Desulfomonile Desulfobulbus Desulfobacterium Desulfomonile Desulfosporosinus Desulfovibrio</i>	2.70E-01	SRR3944753	46
87	Freshwater sediment	Panzihua, Sichuan	26.57 N, 101.7 E	<i>Geobacter Desulfuromonas</i>	7.55E-01	<i>Desulfocapsa</i>	2.75E-03	<i>Desulfobulbus Desulfovibrio</i>	2.91E-01	SRR3944771	46
88	Freshwater sediment	Wushen, Chongqing	29.33 N, 107.76 E	<i>Geobacter Desulfuromonas</i>	4.05E-01	<i>Desulfocapsa</i>	3.35E-03	<i>Desulfobulbus Desulfovibrio</i>	1.02E-01	SRR3944777	46

Table S4. 16S rRNA gene amplicon sequencing-based metadata of global marine/terrestrial sites with presence of S⁰MMs (i.e., S⁰RM and S⁰DM) and SRMs. (Continued)

No	Type	Location	Coordinates	S ⁰ RB		S ⁰ RB		SRB		Accession	Ref.
				Taxonomy	R.A. (%)	Taxonomy	R.A. (%)	Taxonomy	R.A. (%)		
89	Freshwater sediment	Zhengjiang, Jiangsu	32.18 N, 119.66 E	<i>Geobacter Desulfuro monas</i>	1.37E-01	<i>Desulfocapsa Pantoea</i>	4.13E-01	<i>Desulfobulbus Desulfomonile</i>	1.13E-01	SRR3944998	46
90	Freshwater sediment	Yichang, Hubei	30.77 N, 111.31 E	<i>Geobacter Desulfuro monas</i>	9.66E-02	<i>Desulfocapsa</i>	4.90E-03	<i>Desulfobacterium Desulfomonile Desulfobacterium Desulfobulbus</i>	3.00E-01	SRR3944785	46
91	Freshwater sediment	Dongtinghu, Hunan	29.42 N, 113.13 E	<i>Geobacter Desulfuro monas</i>	2.72E-01	<i>Pantoea</i>	7.06E-03	<i>Desulfomonile Desulfobacterium Desulfobulbus</i>	1.46E-01	SRR3944798	46
92	Freshwater sediment	Yiyang, Hunan	29.07 N, 112.29 E	<i>Geobacter Desulfuro monas</i>	1.33E-01	<i>Desulfocapsa</i>	8.85E-03	<i>Desulfomonile Desulfosporosinus Desulfobulbus</i>	3.40E-01	SRR3944806	46
93	Freshwater sediment	Changde, Hunan	28.91 N, 112.06 E	<i>Geobacter Desulfuro monas</i>	1.71E-01	<i>Desulfocapsa</i>	1.11E-02	<i>Desulfomonile Desulfosporosinus</i>	2.10E-01	SRR3944815	46
94	Hydrothermal vent sediment	Atlantic Ocean	37.84 N, 31.51 W	<i>Hippea Desulfuro monas</i>	7.81E-02	<i>Desulfocapsa</i>	2.50E-02	<i>Desulfovibrio Desulfobulbus</i>	7.43E-02	ERR10783031	47
95	Hydrothermal vent sediment	Atlantic Ocean	37.81 N, 31.58 W	<i>Geobacter</i>	1.75E-02	<i>Dissulfuribacter</i>	8.62E-02	<i>Desulfobulbus</i>	1.75E-02	ERR9954321	47
96	Hydrothermal vent sediment	Pacific Ocean, Dorado Outcrop	9.08 N, 87.10 W	<i>Geobacter Desulfuro monas Pelobacter</i>	8.47E-02	<i>Dethiobacter</i>	4.69E-05	<i>Desulfovibrio Desulfobulbus Desulfobacter Thermodesulfobacterium</i>	8.65E-02	SRR6715662	48
97	Hydrothermal vent sediment	Guayman Basin, Mexico	27.26 N, 111.50 W	<i>Geobacter Desulfuro monas</i>	1.32E-01	<i>Desulfocapsa</i>	5.34E-03	<i>Desulfobacterium Desulfobulbus</i>	1.15E+00	SRR11568399	49
98	Hydrothermal vent sediment	Guayman Basin, Mexico	27.35 N, 111.28 W	<i>Geobacter Desulfuro monas</i>	7.40E-02	<i>Desulfocapsa</i>	5.23E-01	<i>Desulfococcus Desulfosarcina Desulfobulbus</i>	3.20E+00	SRR19201761	50
99	Hydrothermal vent sediment	Pacific Ocean	20.05 S, 176.13 W	<i>Hippea</i>	1.35E-01	<i>Desulfocapsa</i>	4.58E-02	<i>Desulfobulbus Thermodesulfator</i>	1.26E+01	SRR027260	51
100	Hydrothermal vent sediment	Atlantic Ocean	38.23 N, 26.63 W	<i>Desulfuro monas</i>	2.93E-02	<i>Desulfocapsa Dissulfuribacter</i>	1.04E-01	<i>Desulfobacter</i>	1.17E02	SRR027592	51

Note: a, major genera of S⁰RM, S⁰DM, and SRMs were provided based on the taxonomy information of each sample. The following SRMs were included to calculate the relative abundance of SRMs: *Desulfotomaculum*, *Desulfovibrio*, *Desulfobulbus*, *Desulfobacter*, *Desulfobacterium*, *Desulfococcus*, *Desulfosarcina*, *Thermodesulfobacterium*, *Thermodesulfobacterium*, *Archaeoglobus*, *Thermodesulfator*, *Desulfosporomusa*, *Desulfohalobium*, *Desulfocaldus*, *Desulfomicrobium*, *Desulfonatronovibrio*, *Desulfonatronum*, *Desulfomonile*, *Desulfarculus*, *Desulfosporosinus*, *Thermodesulfobium*, *Thermocladium*. See Table S2 for the list of S⁰RM and S⁰DM.

b, relative abundance (R.A.) of S⁰RM, S⁰DM, and SRMs were calculated by using following equation: Relative abundance (%) = $I_{si} / \sum N_{si} \times 100$, where I_{si} was the total sequencing reads of S⁰RM, S⁰DM, and SRMs, and $\sum N_{si}$ was the total sequencing reads of the sample.

c, sequencing data for sample 8-11 were provided on the following website: <https://figshare.com/s/82a2d3f5d38ace925492>

d, Acid mine drainage

Table S5. Metagenome sequencing-based metadata of global marine/terrestrial sites with the presence of both S⁰RM and SRMs.

No	Type	Location	Coordinates	<i>psrA/dsrC</i> ^a	Accession	Reference
1	Marine sediment	Atlantic Ocean	30.34S, 7.81W	1.63E-03	ERR3363723	52
2	Marine sediment	Atlantic Ocean	34.79S, 18.96W	1.17E-02	ERR3363725	52
3	Marine sediment	Atlantic Ocean	32.53S, 13.55W	1.81E-03	ERR3363726	52
4	Marine sediment	Pacific Ocean	39.8S, 139.8W	3.94E-03	SRR2605792	53
5	Marine sediment	Pacific Ocean	39.8S, 139.8W	4.14E-03	SRR2605793	53
6	Marine sediment	Pacific Ocean	66.4S, 140.5W	1.59E-03	SRR5486138	54
7	Marine sediment	Pacific Ocean	66.4S, 140.5W	1.70E-03	SRR5486139	54
8	Marine sediment	Ross Sea	75.20 S, 176.59 W	1.71E-03	SRR10156264	55
9	Marine sediment	Ross Sea	72.46 S, 104.36 W	3.41E-03	SRR10156265	55
10	Marine sediment	Pacific Ocean	71.45 S, 102.16 W	2.07E-03	SRR10156269	55
11	Marine sediment	Pacific Ocean	70.48 S, 95.20 W	2.84E-03	SRR10156270	55
12	Marine sediment	Ross Sea	75.50 S, 166.30 W	3.13E-03	SRR10156272	55
13	Marine sediment	Philippine Sea	9.86 N, 138.66 E	1.50E-03	SRR7467837	56
14	Marine sediment	Philippine Sea	9.65 N, 138.82 E	1.69E-03	SRR7467840	56

a, the ratio of *psrA/dsrC* represent the ratio between S⁰RM and SRMs, where the reads of *psrA* and *dsrC* was employed to represent abundance of S⁰RM and SRMs, respectively. *psrA*: polysulfide reductase subunit A; *dsrC*: dissimilatory sulfite reductase subunit C.

Table S6. Primers for qPCR amplification of target genes.

Target gene or primer name	Forward primer	Reverse primer	Length	Reference
<i>recA</i>	GCCCTGTTTCGACATCCTCT A	GGAGTCATGCCAAGGTGT TC	222	57
<i>dsrC</i>	GCTGAAGTCACTTACAAGG GC	CCGGTGTTCTTCGAGAGG AT	218	58
Geo196F/5 35R	GAATATGCTCCTGATTC	TAAATCCGAACAACGCTT	340	59
DSV691F/8 26R	CCGTAGATATCTGGAGGA ACATCAG	ACATCTAGCATCCATCGTT TACAGC	136	60
<i>sqr</i>	ACGACTACGACTGGCTTGT G	GCAGCATCTTCTTGCGAA GG	141	This study

SI References

1. N. Pfennig, H. Biebl, *Desulfuromonas acetoxidans* gen. nov. and sp. nov., a new anaerobic, sulfur-reducing, acetate-oxidizing bacterium. *Arch. Microbiol.* 110, 3–12 (1976).
2. K. Finster, F. Bak, N. Pfennig, *Desulfuromonas acetexigens* sp. nov., a dissimilatory sulfur-reducing eubacterium from anoxic freshwater sediments. *Arch. Microbiol.* 161, 328–332 (1994).
3. M.L. Miroshnichenko, F.A. Rainey, M. Rhode, E.A. Bonch-Osmolovskaya, *Hippea maritima* gen. nov., sp. nov., a new genus of thermophilic, sulfur-reducing bacterium from submarine hot vents. *Inter. J. Syst. Bacteriol.* 49, 1033–1038 (1999).
4. F. Caccavo, D.J. Lonergan, D.R. Lovley, M. Davis, J.F. Stolz, M.J. McInerney, *Geobacter sulfurreducens* sp. nov., a hydrogen- and acetate-oxidizing dissimilatory metal-reducing microorganism. *Appl. Environ. Microbiol.* 60, 3752–3759 (1994).
5. D.R. Lovley, E.J. Phillips, D.J. Lonergan, P.K. Widman, Fe(III) and S⁰ reduction by *Pelobacter carbinolicus*. *Appl. Environ. Microbiol.* 61, 2132–2138 (1995).
6. E.A. Bonch-Osmolovskaya, T.G. Sokolova, N.A. Kostrikina, G.A. Zavarzin, *Desulfurella acetivorans* gen. nov. and sp. nov.—a new thermophilic sulfur-reducing eubacterium. *Arch. Microbiol.* 153, 151–155 (1990).
7. A. Seegerer, A. Neuner, J.K. Kristjansson, K.O. Stetter, *Acidianus infernus* gen. nov., sp. nov., and *Acidianus brierleyi* comb. nov.: facultatively aerobic, extremely acidophilic thermophilic sulfur-metabolizing archaeobacteria. *Inter. J. Syst. Bacteriol.* 36, 559–564 (1986).
8. W. Zillig, S. Yeats, I. Holz, A. Bock, F. Gropp, M. Rettenberger, S. Lutz, Plasmid-related anaerobic autotrophy of the novel archaeobacterium *Sulfolobus ambivalens*. *Nature* 313, 789–791 (1985).
9. F. Fischer, W. Zillig, K.O. Stetter, G. Schreiber, Chemolithoautotrophic metabolism of anaerobic extremely thermophilic archaeobacteria. *Nature* 301, 511–513 (1983).
10. A.H. Seegerer, A. Trincone, M. Gahrtz, K.O. Stetter, *Stygiolobus azoricus* gen. nov., sp. nov. represents a novel genus of anaerobic, extremely thermoacidophilic archaeobacteria of the order Sulfolobales. *Inter. J. Syst. Bacteriol.* 41, 495–501 (1991).
11. S. Yvenou, M. Allieux, A. Slobodkin, G. Slobodkina, M. Jebbar, K. Alain, Genetic potential of *Dissulfurimicrobium hydrothermale*, an obligate sulfur-disproportionating thermophilic microorganism. *Microorganisms* 10, 60 (2022).
12. K. Umezawa, H. Kojima, Y. Kato, M. Fukui, *Dissulfurispira thermophila* gen. nov., sp. nov., a thermophilic chemolithoautotroph growing by sulfur disproportionation, and proposal of novel taxa in the phylum Nitrospirota to reclassify the genus *Thermodesulfovibrio*. *Syst. Appl. Microbiol.* 44, 126184 (2021).
13. M. Allieux, S. Yvenou, G. Slobodkina, A. Slobodkin, Z. Shao, M. Jebbar, K. Alain, Genomic characterization and environmental distribution of a thermophilic Anaerobe *Dissulfurirhabdus thermomarina* SH388T involved in disproportionation of sulfur compounds in shallow sea hydrothermal vents. *Microorganisms* 8, 1132 (2020).
14. T.V. Kolganova, N.A. Kostrikina, E.A. Bonch-Osmolovskaya, 2013. *Dissulfuribacter thermophilus* gen. nov., sp. nov. a novel thermophilic autotrophic sulfur-disproportionating deeply-branching delta-proteobacterium from a deep-sea hydrothermal vent of the Eastern Lau Spreading Center. *Int. J. Syst. Evol. Microbiol.* 63, 1967–1971 (2013).
15. H. Kojima, K. Umezawa, M. Fukui, *Caldimicrobium thiodismutans* sp. nov., a sulfur-disproportionating bacterium isolated from a hot spring, and emended description of the genus *Caldimicrobium*. *Int. J. Syst. Evol. Microbiol.* 66, 1828–1831 (2016).
16. T.M. Frederiksen, K. Finster, Sulfite-oxido-reductase is involved in the oxidation of sulfite in *Desulfocapsa sulfoexigens* during disproportionation of thiosulfate and elemental sulfur. *Biodegradation* 14, 189–98 (2003).
17. A.P. Florentino, C. Brienza, A.J.M. Stams, I. Sánchez-Andrea, *Desulfurella amilsii* sp. nov., a novel acidotolerant sulfur-respiring bacterium isolated from acidic river sediments. *Int. J. Syst. Evol. Microbiol.* 66, 1249–1253 (2016).
18. D.Y. Sorokin, T.P. Tourova, M. Mußmann, G. Muyzer, *Dethiobacter alkaliphilus* gen. nov. sp. nov., and *Desulfurivibrio alkaliphilus* gen. nov. sp. nov.: two novel representatives of reductive sulfur cycle from soda lakes. *Extremophiles* 12, 431–439 (2008).

19. A.Y. Obraztsova, C.A. Francis, M.T. Bradley, Sulfur disproportionation by the facultative anaerobe *Pantoea agglomerans* SP1 as a mechanism for chromium(VI) reduction. *Geomicrobiol. J.* 19, 121–132 (2002).
20. A.I. Slobodkin, A.L. Reysenbach, G.B. Slobodkina, R.V. Baslerov, N.A. Kostrikina, I.D. Wagner, E.A. Bonch-Osmolovskaya, *Thermosulfurimonas dismutans* gen. nov., sp. nov. a novel extremely thermophilic sulfur disproportionating bacterium from a deep-sea hydrothermal vent. *Int. J. Syst. Evol. Microbiol.* 62, 2565–2571 (2012).
21. G.B. Slobodkina, A. Reysenbach, T.V. Kolganova, A.A. Novikov, E.A. Bonch-Osmolovskaya, A.I. Slobodkin. *Thermosulfuriphilus ammonigenes* gen. nov., sp. nov., a thermophilic, chemolithoautotrophic bacterium capable of respiratory ammonification of nitrate with elemental sulfur. *Int. J. Syst. Evol. Microbiol.* 67, 3474–3479 (2017).
22. M. Ji, W. Kong, H. Jia, M. Delgado-Baquerizo, T. Zhou, X. Liu, B.C. Ferrari, L. Malard, C. Liang, K. Xue, T.P. Makhallanyane, Y. Zhu, Y. Wang, D.A. Pearce, D. Cowan, Polar soils exhibit distinct patterns in microbial diversity and dominant phylotypes. *Soil Biol. Biochem.* 11, 108550 (2022).
23. M. Delgado-Baquerizo, A.M. Oliverio, T.E. Brewer, A. Benavent-Gonzalez, D.J. Eldridge, R.D. Bardgett, F.T. Maestre, B.K. Singh, N. Fierer, A global atlas of the dominant bacteria found in soil. *Science* 359, 320–325 (2018).
24. A. Khan, W. Kong, M. Ji, L. Yue, Y. Xie, J. Liu, B. Xu, Disparity in soil bacterial community succession along a short time-scale deglaciation chronosequence on the Tibetan Plateau. *Soil Ecol. Lett.* 2, 83-92 (2020).
25. M. Ji, W. Kong, C. Liang, T. Zhou, H. Jia, X. Dong, Permafrost thawing exhibits a greater influence on bacterial richness and community structure than permafrost age in arctic permafrost soils. *Cryosphere* 14, 3907-3916 (2020).
26. D. Probandt, K. Knittel, H.E. Tegetmeyer, S. Ahmerkamp, M. Holtappels, R. Amann, Permeability shapes bacterial communities in sublittoral surface sediments. *Environ. Microbiol.* 19, 1584–1599 (2017).
27. N. Mahmoudi, M.S. Robeson 2nd, H.F. Castro, J.L. Fortney, S.M. Techtmann, D.C. Joyner, C.J. Paradis, S.M. Pfiffner, T.C. Hazen, Microbial community composition and diversity in Caspian Sea sediments. *FEMS Microbiol. Ecol.* 91, 1–11 (2015).
28. J. Buongiorno, L.C. Herbert, L.M. Wehrmann, A.B. Michaud, K. Laufer, H. Roy, B.B. Jorgensen, A. Szykiewicz, A. Faiia, K.M. Yeager, K. Schindler, K.G. Lloyd, Complex microbial communities drive iron and sulfur cycling in Arctic Fjord sediments. *Appl. Environ. Microbiol.* 85, e00949-19 (2019).
29. S.E. Ruff, J.F. Biddle, A.P. Teske, K. Knittel, A. Boetius, A. Ramette, Global dispersion and local diversification of the methane seep microbiome. *Proc. Nat. Acad. Sci. USA* 112, 4015–4020 (2015).
30. G. Varliero, C. Bienhold, F. Schmid, A. Boetius, M. Molari, Microbial diversity and connectivity in deep-sea sediments of the South Atlantic Polar Front. *Front. Microbiol.* 10, 665 (2019).
31. R.A. Walsh, J.B. Kirkpatrick, R. Pockalny, J. Sauvage, A.J. Spivack, R.W. Murray, M.L. Sogin, S. D'Hondt, Relationship of bacterial richness to organic degradation rate and sediment age in subseafloor sediment. *Appl. Environ. Microbiol.* 82, 4994–4999 (2016).
32. E.K. Wear, M.J. Church, B.N. Oreutt, C.N. Shulze, M.V. Lindh, C.R. Smith, Bacterial and archaeal communities in polymetallic nodules, sediments, and bottom waters of the Abyssal Clarion-Clipperton Zone: Emerging patterns and future monitoring considerations. *Front. Marine Sci.* 8, 480 (2021).
33. X. Fang, T. Zhang, J. Li, N. Wang, Z. Wang, L. Yu, Bacterial community pattern along the sediment seafloor of the Arctic fjorden (Kongsfjorden, Svalbard). *Antonie van Leeuwenhoek* 112, 1121–1136 (2019).
34. Z. Kerrigan, J.B. Kirkpatrick, S. D'Hondt, Influence of 16S rRNA hypervariable region on estimates of bacterial diversity and community composition in seawater and marine sediment. *Front. Microbiol.* 10, 1640 (2019).
35. L.Wang, M. Yu, Y. Liu, J. Liu, Y. Wu, L. Li, J. Liu, M. Wang, X. Zhang, Comparative analyses of the bacterial community of hydrothermal deposits and seafloor sediments across Okinawa Trough. *J. Marine Syst.* 180, 162-72 (2018).

36. R.K. Sinha, K.P. Krishnan, F.A. Thomas, M.B. Binish, M. Mohan, P.J. Kurian, Polyphasic approach revealed complex bacterial community structure and function in deep sea sediment of ultra-slow spreading Southwest Indian Ridge. *Ecol. Indi.* 96, 40-51 (2019).
37. B. Zhang, Y. Li, S. Xiang, Y. Yan, R. Yang, M. Lin, X. Wang, Y. Xue, X. Guan, Sediment microbial communities and their potential role as environmental pollution indicators in Xuande Atoll, South China Sea. *Front. Microbiol.* 11, 1011 (2020).
38. T. Hoshino, H. Doi, G. Uramoto, L. Wormer, R.R. Adhikari, N. Xiao, Y. Morono, S. D'Hondt, K. Hinrichs, F. Inagaki, Global diversity of microbial communities in marine sediment. *Proc. Nat. Acad. Sci. USA* 117, 27587–27597 (2020).
39. Y. Zhang, G. Wu, H. Jiang, J. Yang, W. She, I. Khan, W. Li, Abundant and rare microbial biospheres respond differently to environmental and spatial factors in Tibetan Hot Springs. *Front. Microbiol.* 9, 2096 (2018).
40. Q. He, S. Wang, W. Hou, K. Feng, F. Li, W. Hai, Y. Zhang, Y. Sun, Y. Deng, Temperature and microbial interactions drive the deterministic assembly processes in sediments of hot springs. *Sci. Total Environ.* 772, 145465 (2021).
41. P.T. Podar, Z. Yang, S.H. Björnsdóttir, M. Podar, Comparative analysis of microbial diversity across temperature gradients in Hot Springs from Yellowstone and Iceland. *Front. Microbiol.* 11, 1625 (2020).
42. C. Sriaporn, K.A. Campbell, M.J. Van Kranendonk, K.M. Handley, Genomic adaptations enabling *Acidithiobacillus* distribution across wide-ranging hot spring temperatures and pHs. *Microbiome* 9, 135 (2021).
43. S.M. Gao, A. Schippers, N. Chen, Y. Yuan, M. Zhang, Q. Li, B. Liao, W. Shu, L. Huang, Depth-related variability in viral communities in highly stratified sulfidic mine tailings. *Microbiome* 8, 89 (2020).
44. Y. Hao, X. Zhao, H. Ai, S. Gao, W. Teng, J. Zheng, W. Shu, Microbial biogeography of acid mine drainage sediments at a regional scale across southern China. *FEMS Microbiol. Ecol.* 98, fiac002 (2022).
45. S.M. Abramov, D. Straub, J. Tejada, L. Grimm, F. Schadler, A. Bulaev, H. Thorwarth, R. Amils, A. Kappler, S. Kleindienst, Biogeochemical niches of Fe-cycling communities influencing heavy metal transport along the Rio Tinto, Spain. *Appl. Environ. Microbiol.* 88, e0229021 (2022).
46. T. Liu, A.N. Zhang, J. Wang, S. Liu, X. Jiang, C. Dang, T. Ma, S. Liu, Q. Chen, S. Xie, T. Zhang, J. Ni, Integrated biogeography of planktonic and sedimentary bacterial communities in the Yangtze River. *Microbiome* 6, 16 (2018).
47. D.V. Meier, W. Bach, P.R. Girguis, H.R. Gruber-Vodicka, E.P. Reeves, M. Richter, C. Vidoudez, R. Amann, A. Meyerdierks, Heterotrophic Proteobacteria in the vicinity of diffuse hydrothermal venting. *Environ. Microbiol.* 18, 4348–4368 (2016).
48. L.A. Zinke, B.K. Reese, J. McManus, C.G. Wheat, B.N. Orcutt, J.P. Amend, Sediment microbial communities influenced by cool hydrothermal fluid migration. *Front. Microbiol.* 9, 1249 (2018).
49. A. Teske, G. Wegener, J.P. Chanton, D. White, B. MacGregor, D. Hoer, D. de Beer, G. Zhuang, M.A. Saxton, S.B. Joye, D. Lizarralde, S.A. Soule, S.E. Ruff, Microbial communities under distinct thermal and geochemical regimes in axial and off-axis sediments of Guaymas Basin. *Front. Microbiol.* 12, 633649 (2021).
50. P. Cruaud, V. Adrien, P. Patricia, C. Jean-Claude, L. Françoise, T. Laurent, G. Anne, C. Marie-Anne, Microbial communities associated with benthic faunal assemblages at cold seep sediments of the Sonora Margin, Guaymas Basin. *Front. Marine Sci.* 2, 53 (2015).
51. S.E. Ruff, J.F. Biddle, A.P. Teske, K. Nittel, A. Boetius, A. Ramette, Global dispersion and local diversification of the methane seep microbiome. *Proc. Nat. Acad. Sci. USA* 112, 4015–4020 (2015).
52. F.H. Coutinho, F.A.B. von Meijenfheldt, J.M. Walter, J.M. Haro-Moreno, M. López-Pérez, M.C. van Verk, C.C. Thompson, C.A.N. Cosenza, L. Appolinario, R. Paranhos, A. Cabral, B.E. Dutilh, F.L. Thompson. Ecogenomics and metabolic potential of the South Atlantic Ocean microbiome. *Sci. Total Environ.* 765, 142758 (2021).
53. B.J. Tully, J.F. Heidelberg, Potential mechanisms for microbial energy acquisition in oxic deep-sea sediments. *Appl. Environ. Microbiol.* 82, 4232–4243 (2016).

54. S.A. Carr, F. Schubotz, R.B. Dunbar, C.T. Mills, R. Dias, R.E. Summons, K.W. Mandernack, Acetoclastic *Methanosaeta* are dominant methanogens in organic-rich Antarctic marine sediments. *ISME J.* 12, 330–342 (2018).
55. A.I. Garber, J.R. Zehnpfennig, C.S. Sheik, M.W. Henson, G.A. Ramírez, A.R. Mahon, K.M. Halanych, D.R. Learman, Metagenomics of antarctic marine sediment reveals potential for diverse chemolithoautotrophy. *mSphere* 6, e0077021 (2021).
56. X. Zhang, W. Xu, Y. Liu, M. Cai, Z. Luo, M. Li, Metagenomics reveals microbial diversity and metabolic potentials of seawater and surface sediment from a hadal biosphere at the Yap Trench. *Front. Microbiol.* 9, 2402 (2018).
57. K. Sivakumar, G. Scarascia, N. Zaouri, T. Wang, A.H. Kaksonen, P. Hong, Salinity-mediated increment in sulfate reduction, biofilm formation, and quorum sensing: a potential connection between quorum sensing and sulfate reduction? *Front. Microbiol.* 10, 188 (2019).
58. M. Clark, R. Edelman, M.L. Duley, J. Wall, M. Fields, Biofilm formation in *Desulfovibrio vulgaris* Hildenborough is dependent upon protein filaments. *Environ. Microbiol.* 9, 2844–2854 (2007).
59. B.K. Amos, Y. Sung, K.E. Fletcher, T.J. Gentry, W. Wu, C.S. Criddle, J. Zhou, F.E. Löffler, Detection and quantification of *Geobacter lovleyi* strain SZ: implications for bioremediation at tetrachloroethene- and uranium-impacted sites. *Appl. Environ. Microbiol.* 73, 6898-904 (2007).
60. A. Fite, G.T. Macfarlane, J.H. Cummings, Identification and quantitation of mucosal and faecal *Desulfovibrio* using real time polymerase chain reaction. *Gut* 53, 523-529 (2004).