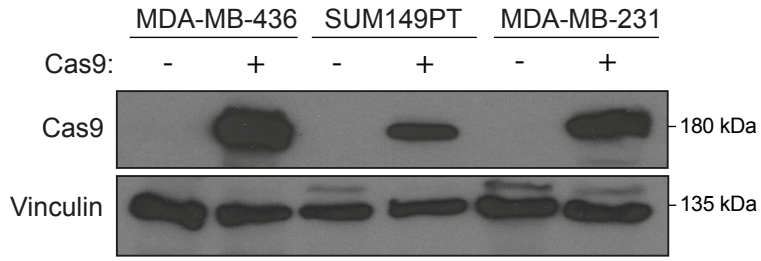
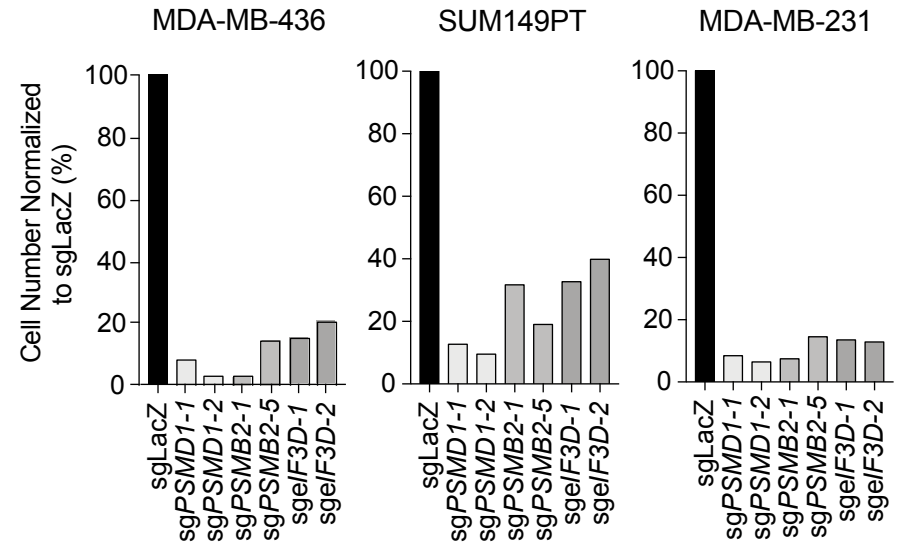


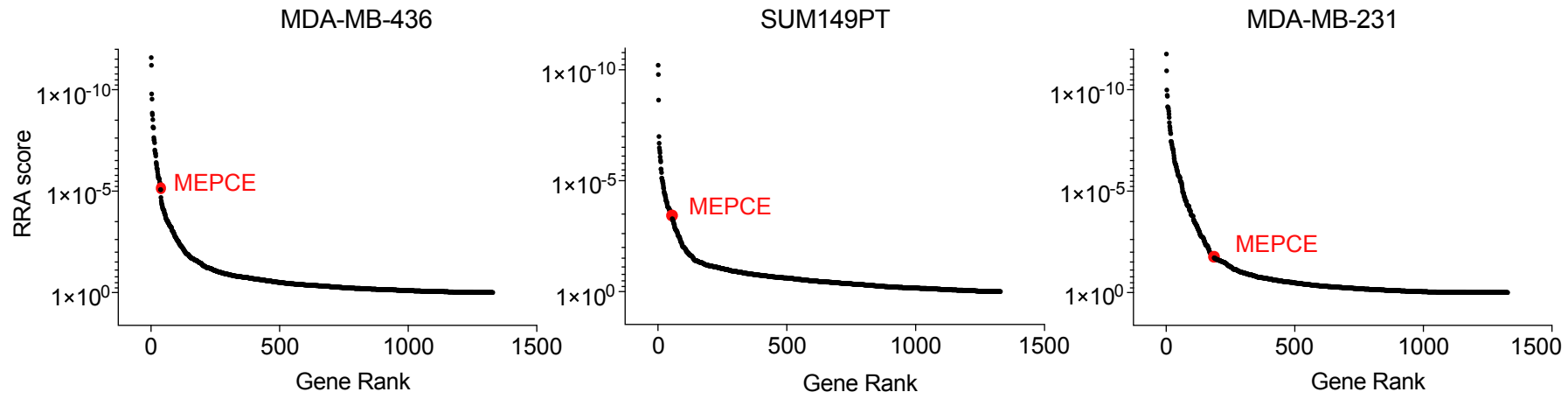
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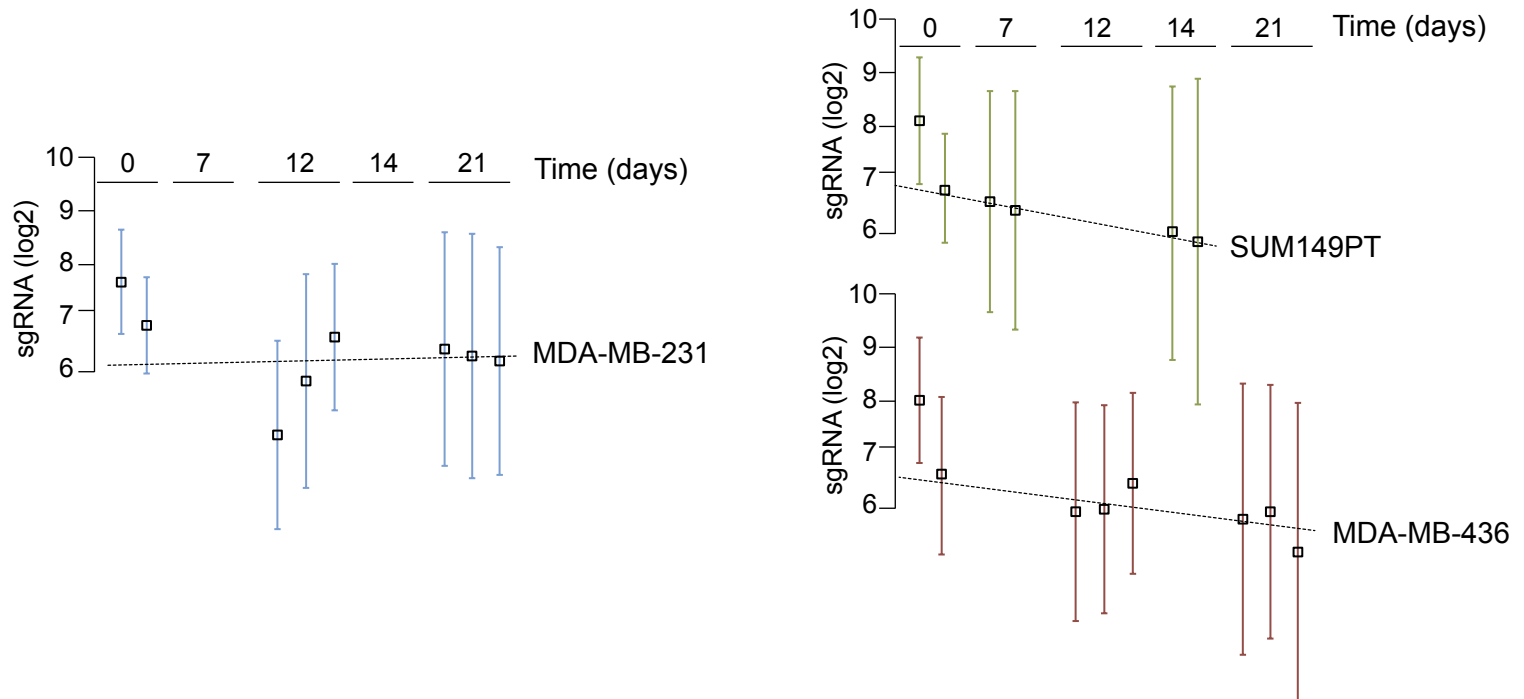


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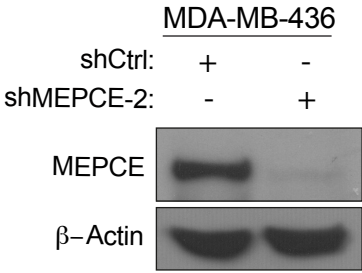
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MEPCE

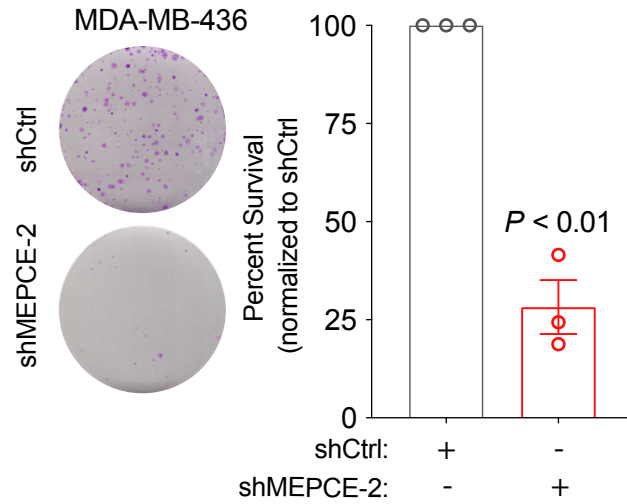


Supplementary Figure S1. Validation of Cas9 functionality in breast cancer cell lines used in CRISPR- Cas9 dropout screen and screen analysis. (A) Western blot showing the overexpression of Cas9 in MDA-MB-436, SUM149PT and MDA-MB-231 clones stably transduced with the lentiviral Lenti-Cas9-2A-Blast construct. Vinculin was used as a loading control. (B) Validation of CRISPR-Cas9 gene editing efficiency in Cas9-expressing MDA-MB-436, SUM149PT and MDA-MB-231 cells. Bar graph representing cell proliferation 9-10 days post- transduction with a control sgRNA construct (sgLacZ) or sgRNAs targeting essential genes *PSMD1*, *PSMB2* and *EIF3D*. Cell number is normalized to control sgRNA (sgLacZ). (C) Scatter plot representing gene rank vs RRA score calculated by the MAGeCK-VISPR pipeline for each cell line. *MEPCE* is highlighted. (D) Analysis of *MEPCE* gRNAs in indicated cell lines used for CRISPR screening. RRA: robust rank aggregation.

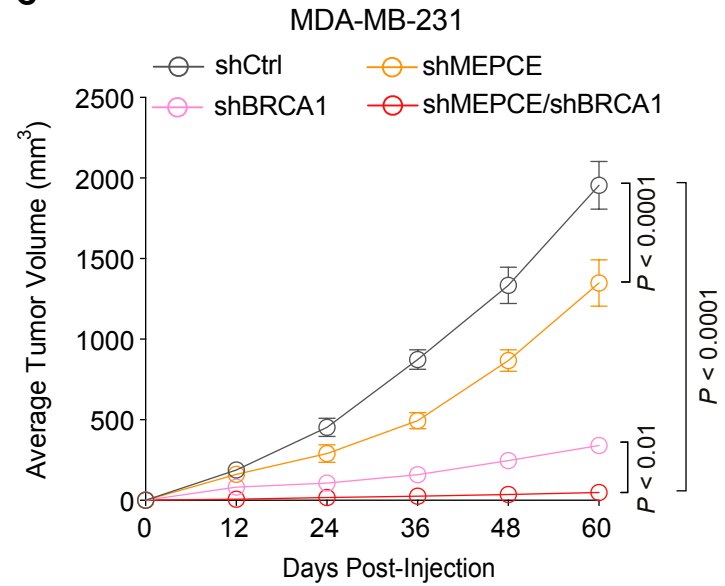
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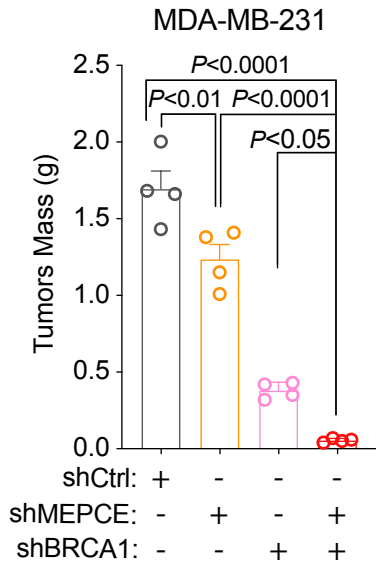
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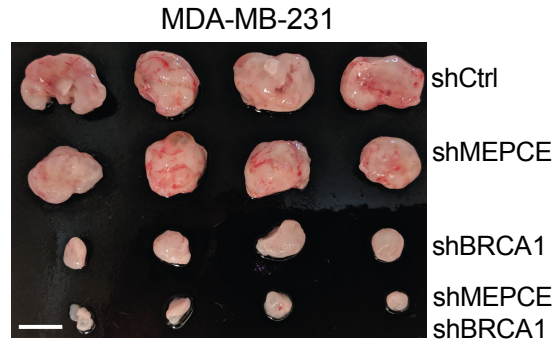
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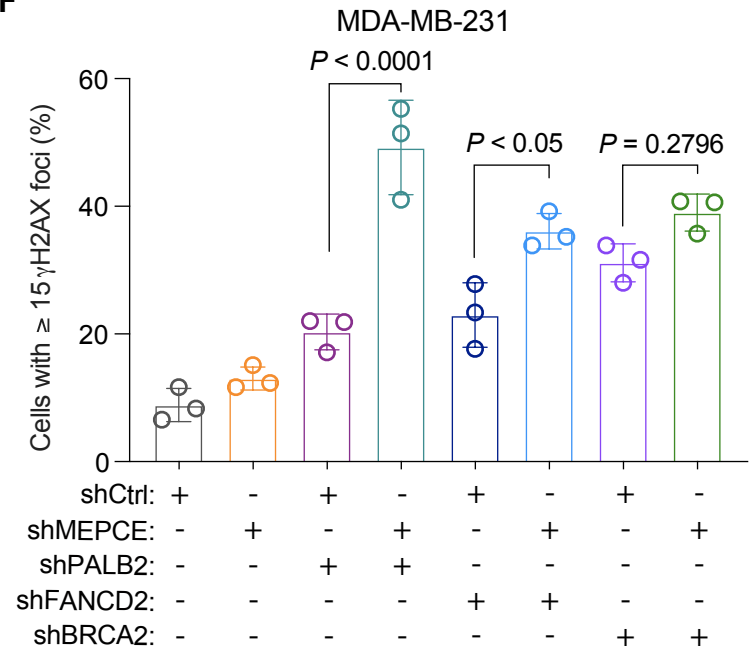
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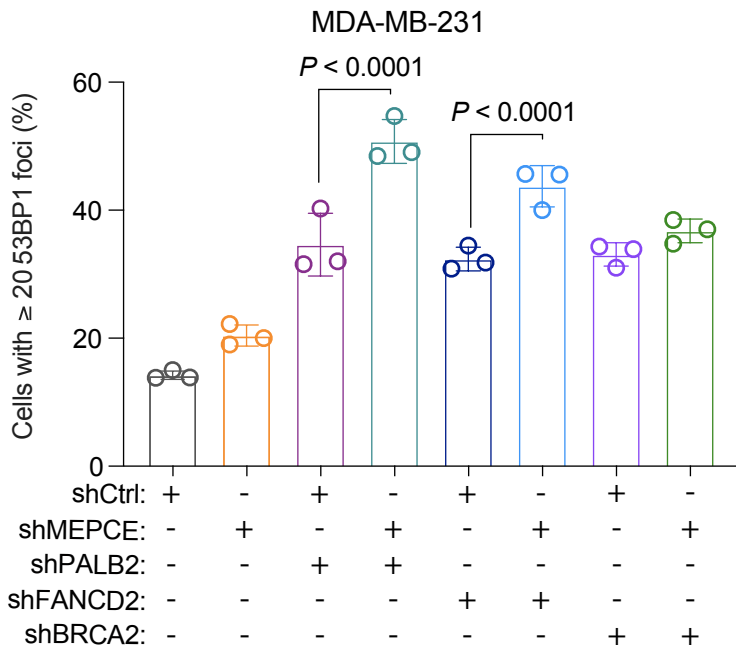
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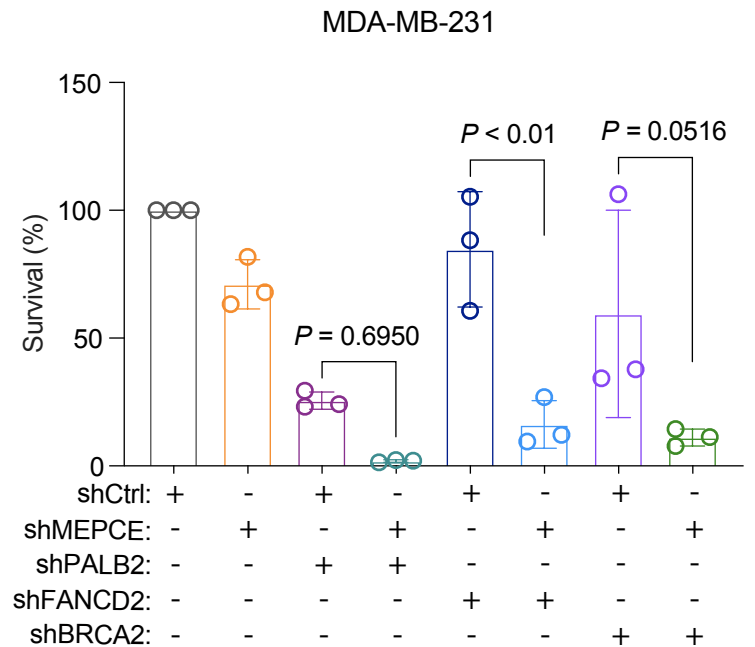
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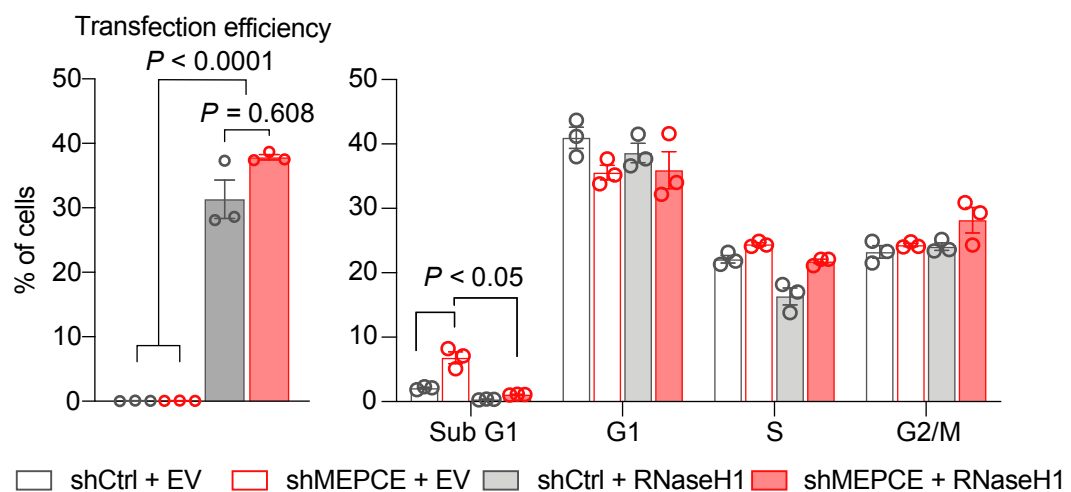
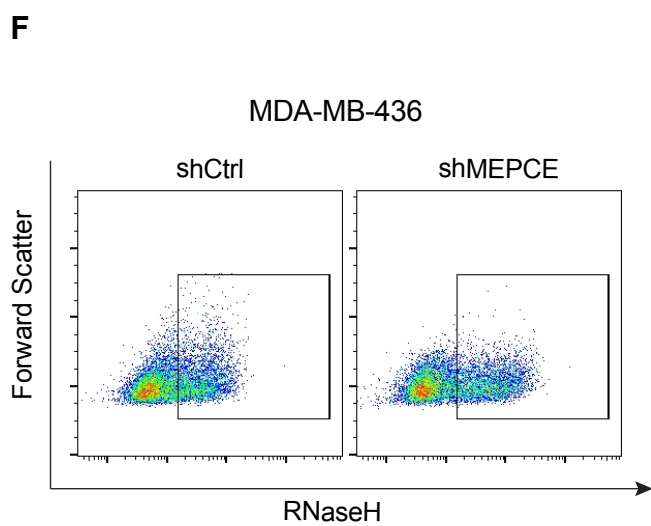
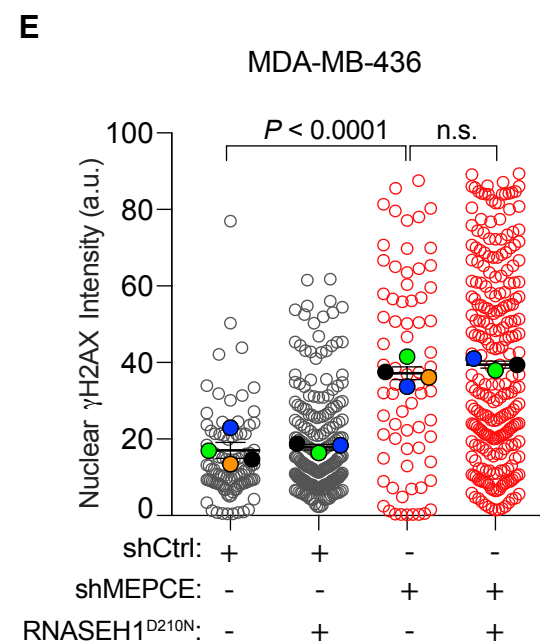
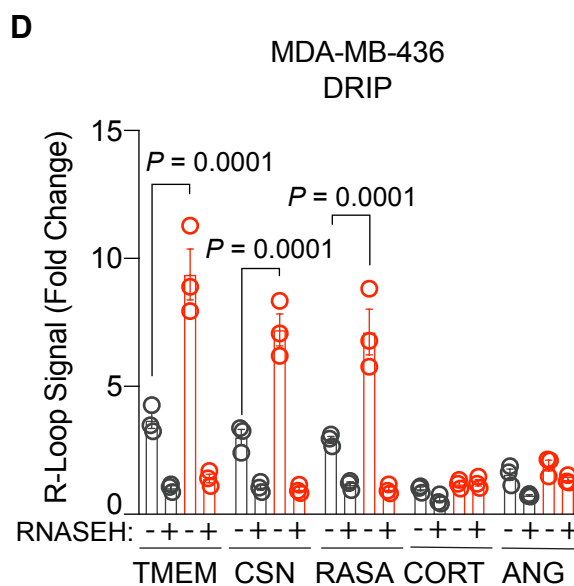
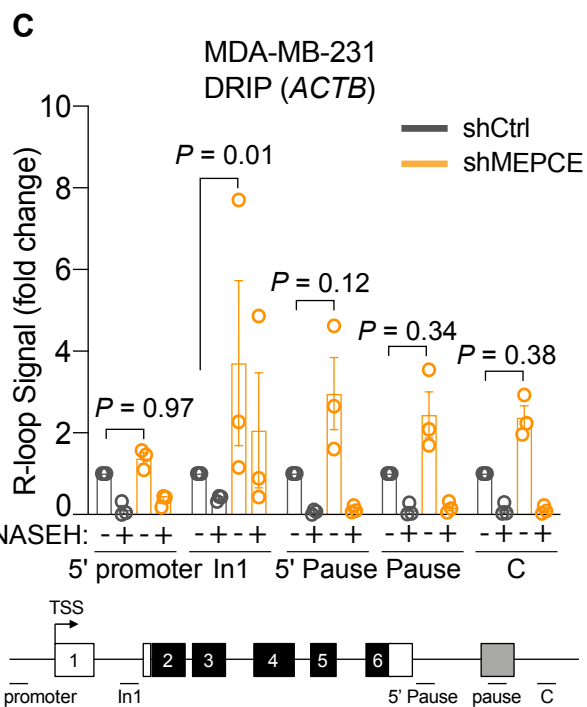
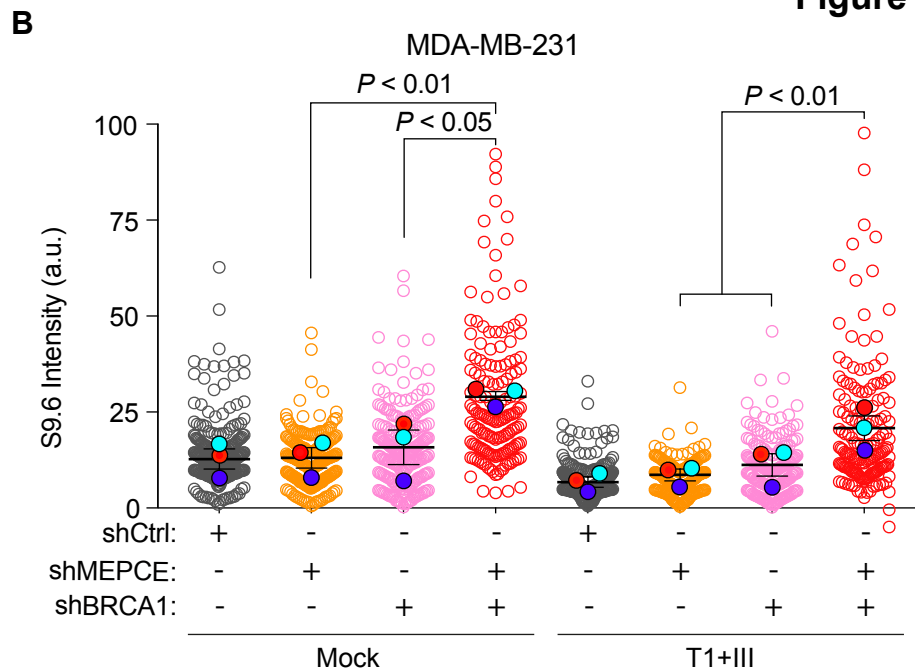
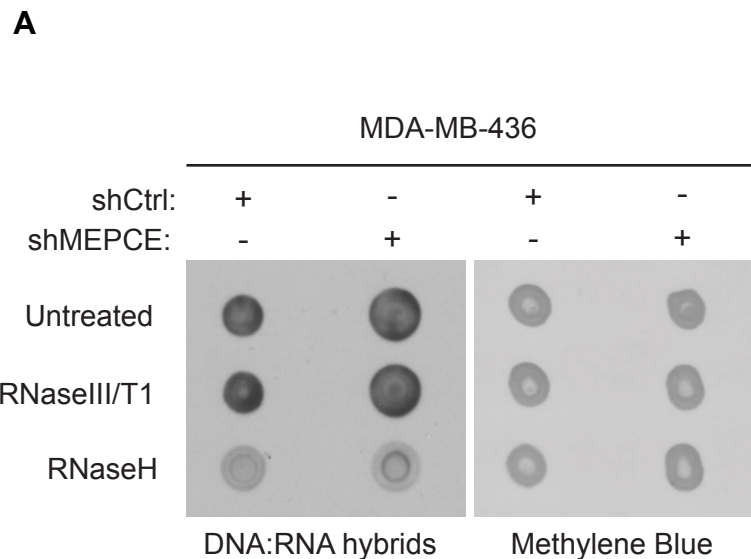
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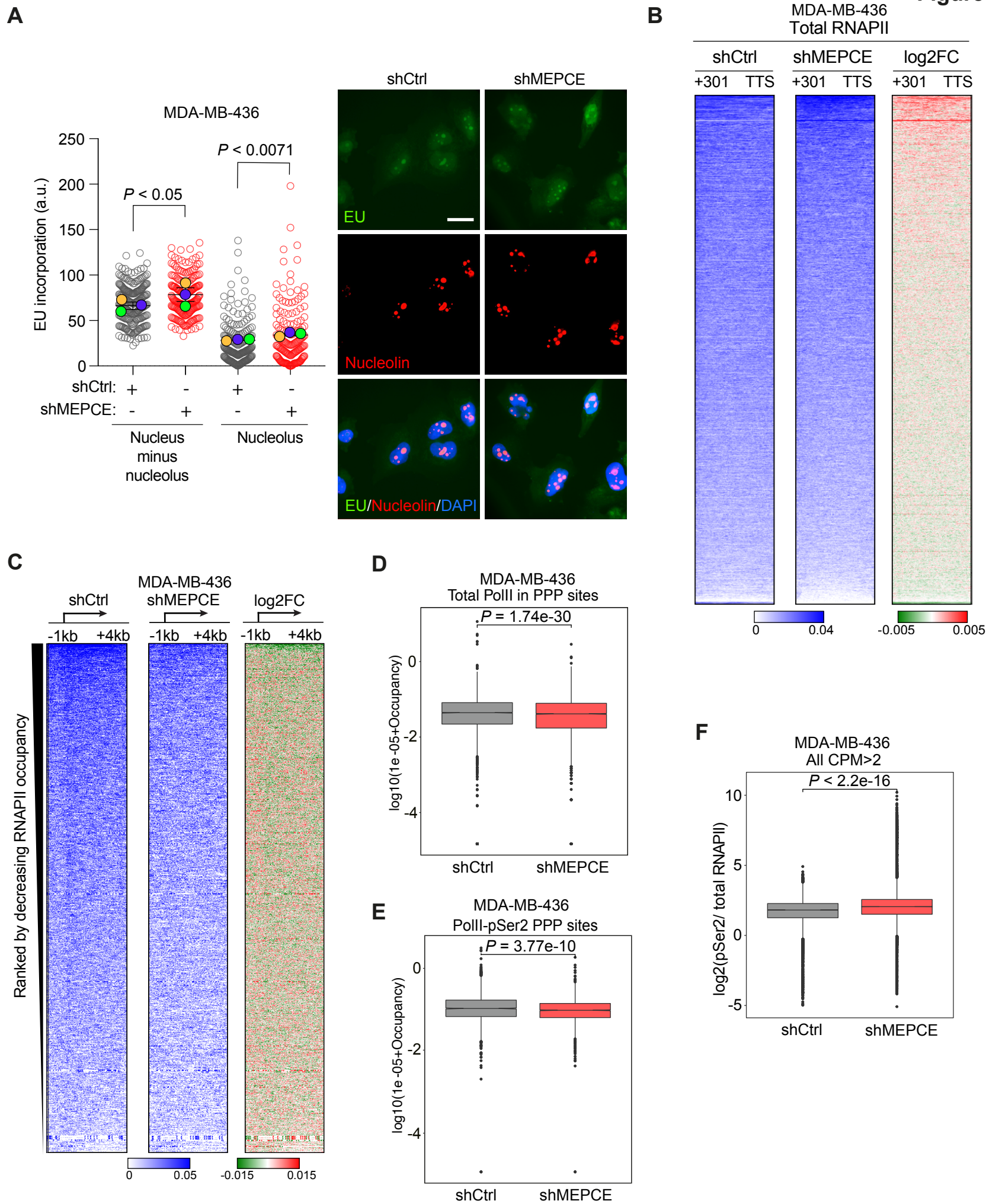
H



Supplementary Figure S2. Loss of MEPCE in *BRCA1*-null and some HR-defective cells causes severe growth defect *in vivo*, leads to elevated DNA damage, and R-loop accumulation. (A) Western blot of MEPCE expression in indicated cells. (B) Representative images of colony forming assay and quantification in indicated MDA-MB-436 cells. (C) *In vivo* growth curve of MDA-MB-231 control (shCtrl), shMEPCE, shBRCA1 and shMEPCE/shBRCA1 xenografts depicting changes in tumor volume over the duration of the experiment measured by an external caliper. (D) Quantification of tumor mass post-resection. (E) Representative images of tumors at day 60 post-injection (n = 4 per condition; bar = 1 cm). (F) Quantification of γ H2AX foci in indicated MDA-MB-231 cells. (n = 3). (G) Quantification of 53BP1 foci in indicated MDA-MB-231 cells. (n = 3). (H) Quantification of survival in indicated MDA-MB-231 cells. B, Student's *t*-test. C, two-way ANOVA (Tukey's multiple comparisons test). D, F, G, and H, one-way ANOVA (Tukey's multiple comparisons test). For B, D, F, G, and H, mean \pm SEM is shown.

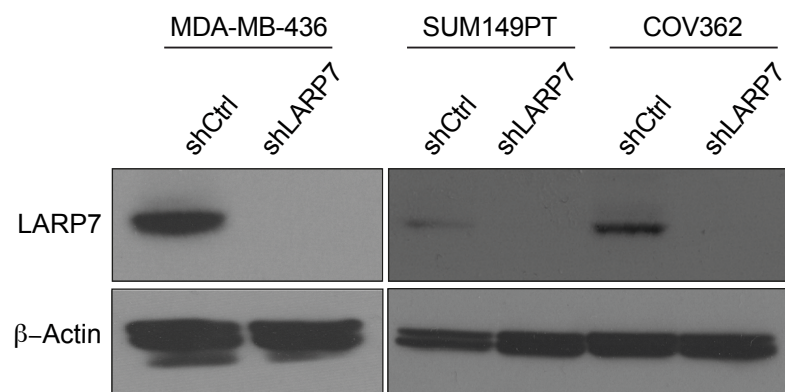


Supplementary Figure S3. MEPCE depletion increases R-loops in BRCA1-deficient cancer cells and short-term RNASEH1 overexpression does not significantly alter cell cycle profile in MEPCE-depleted or control cells. (A) Dot blot depicting S9.6 (DNA:RNA hybrid) levels in MDA-MB-436 cells. Methylene blue is used as loading control. **(B)** Quantification of anti-DNA-RNA hybrid (S9.6) immunofluorescence and quantification of nuclear and nucleolar hybrid levels in response to indicated treatment conditions (Mock, RNASET1 (T1), RNASEIII (III), and/or RNASEH (H)). **(C)** DRIP-qPCR using S9.6 antibody in indicated MDA-MB-231 cells in *ACTB* gene at the indicated loci. Enrichment is compared to negative control (IgG) followed by normalization to shCtrl cells (mean \pm SEM). **(D)** DRIP-qPCR using S9.6 antibody in indicated MDA-MB-436 cells at the indicated loci. Enrichment is compared to negative control (IgG) followed by normalization to shCtrl cells (mean \pm SEM). **(E)** Quantification nuclear γ H2AX intensity in MEPCE-depleted MDA-MB-436 cells and control cells transiently transfected with RNASEH1-D210N **(F)** RNASEH1 transfection efficiency (FACS profile) and cell cycle profiles of MEPCE-depleted and control MDA-MB-436 cells as indicated. (n = 3). B-F, each dot represents a biological triplicate. B and E, one-way ANOVA (Tukey's multiple comparisons test), C and D, two-way ANOVA (Tukey's multiple comparisons test). F, one-way ANOVA (Tukey's multiple comparisons test).

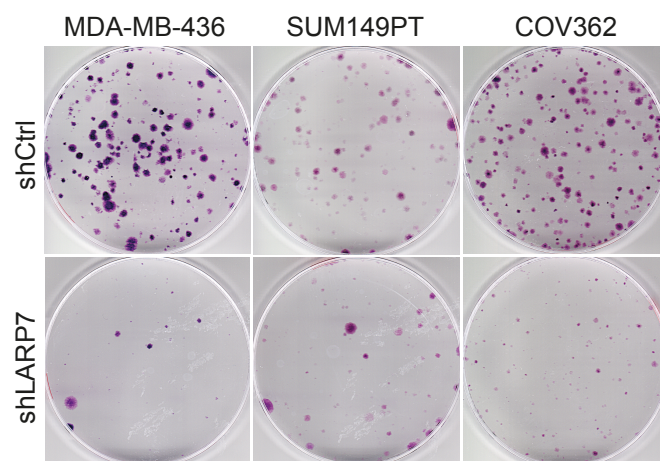


Supplementary Figure S4. Loss of MEPCE in *BRCA1*-mutant cells alters transcription and occupancy of total RNAPII and pSer2 RNAPII. (A) Quantification and representative images of EU incorporation in indicated nuclear compartments in MEPCE-depleted MDA-MB-436 and control cells. (n = 3). Bar = 20 μ m. (B) Heatmaps of total RNAPII occupancy in the +301bp to transcription termination site (TTS) in indicated MDA-MB-436 cells. Log₂ fold change is shown. (C) PolII occupancy in -1kb and +4kb surrounding the transcription start site in genes with PRR>1 in shCtrl and shMEPCE MDA-MB-436 cells. Log₂ fold change is shown. (D) Total PolII occupancy in the promoter region (-100bp to +300bp surrounding the transcription start site) which includes promoter proximal pause site. Log₁₀ values are shown. (E) PolII-pSer2 occupancy in the promoter region (-100bp to +300bp surrounding the transcription start site) which includes promoter proximal pause site. Log₁₀ values are shown. (F) Boxplots showing the ratio of pSer2 levels compared to total RNAPII enrichment in the promoter region in indicated MDA-MB-436 cells for all regions. A, D-F, Student's *t*-test.

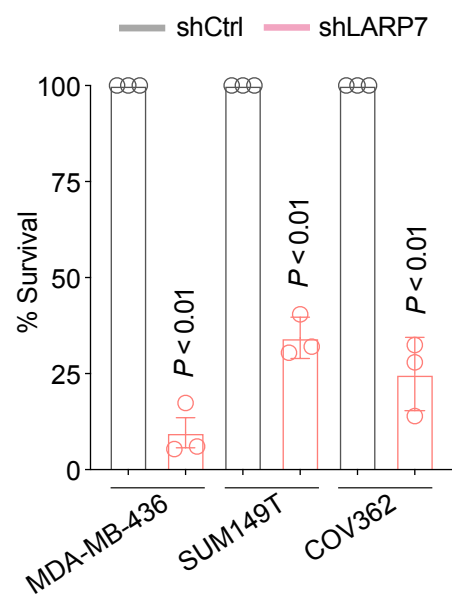
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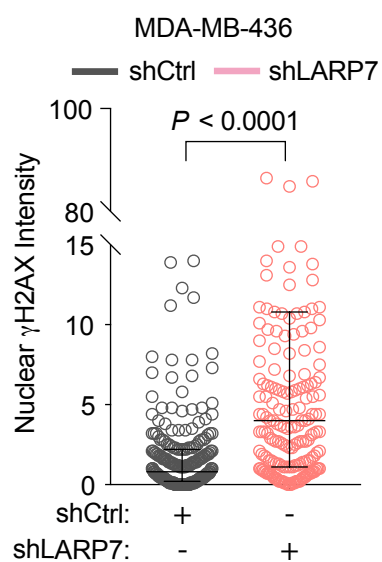
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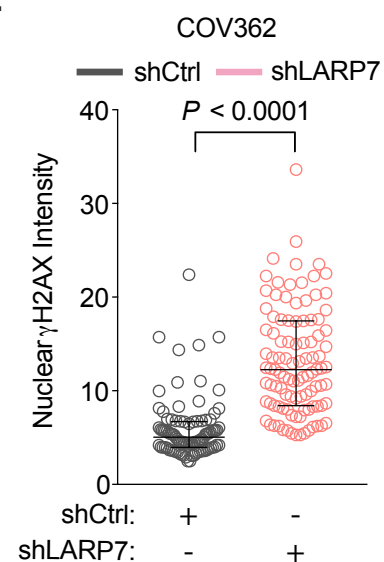
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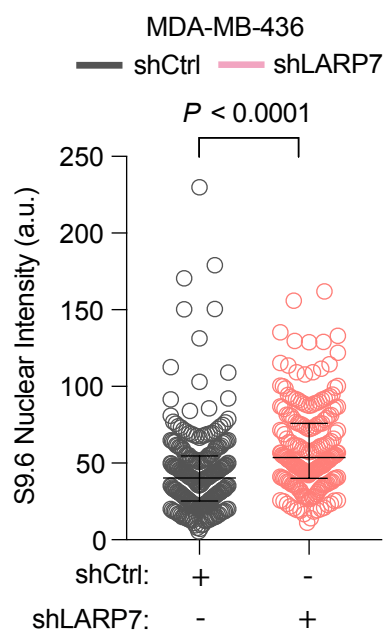
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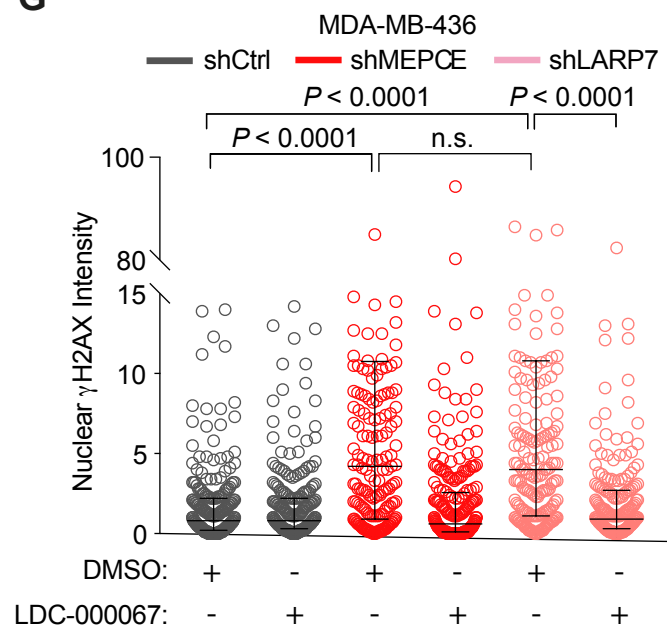
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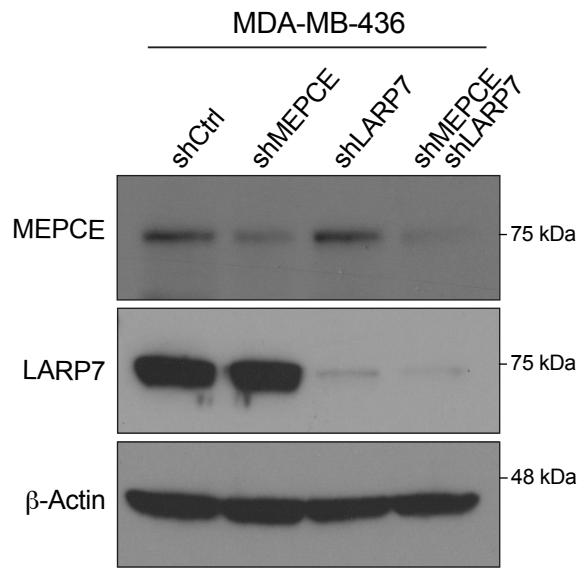
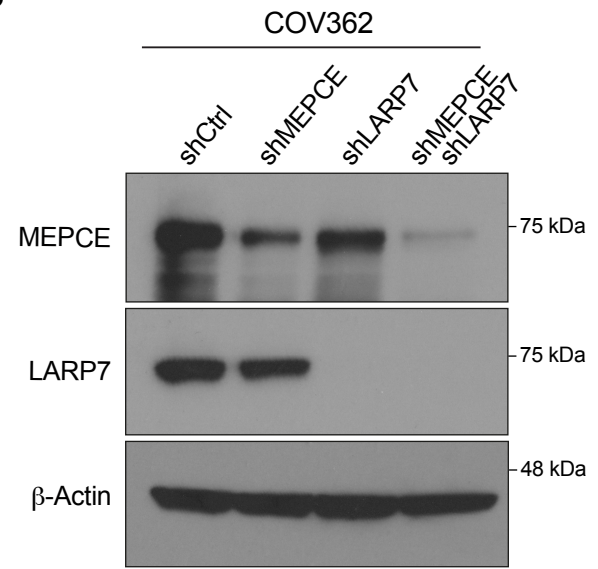
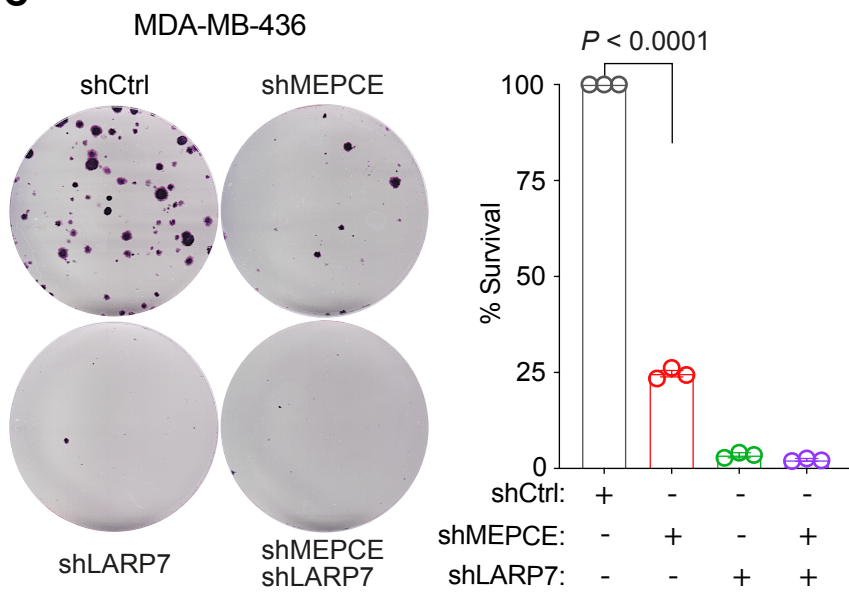
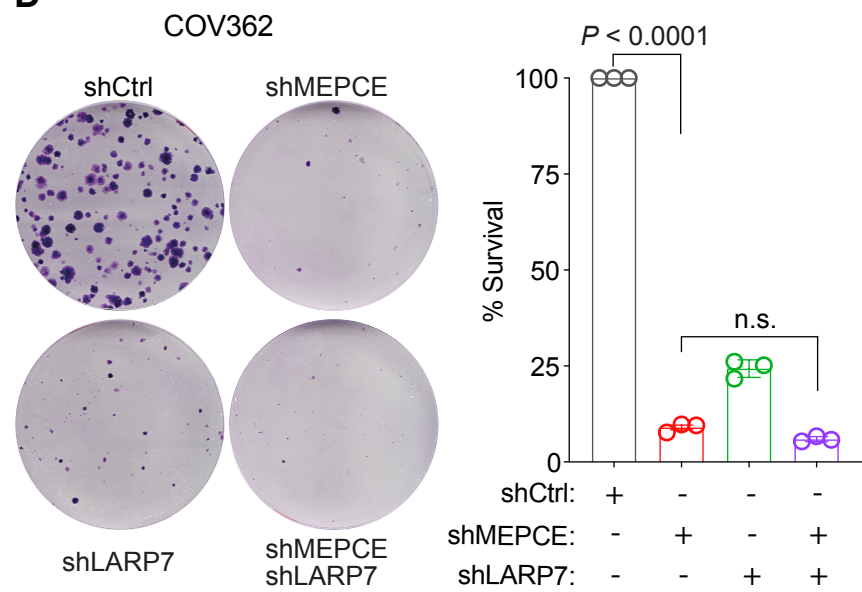
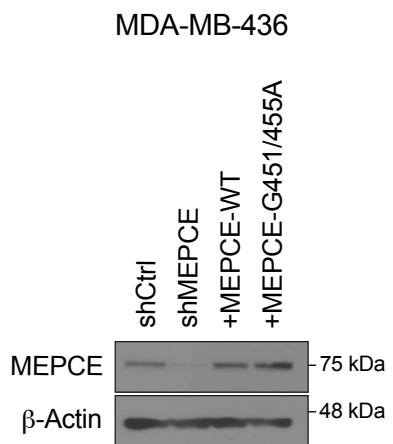
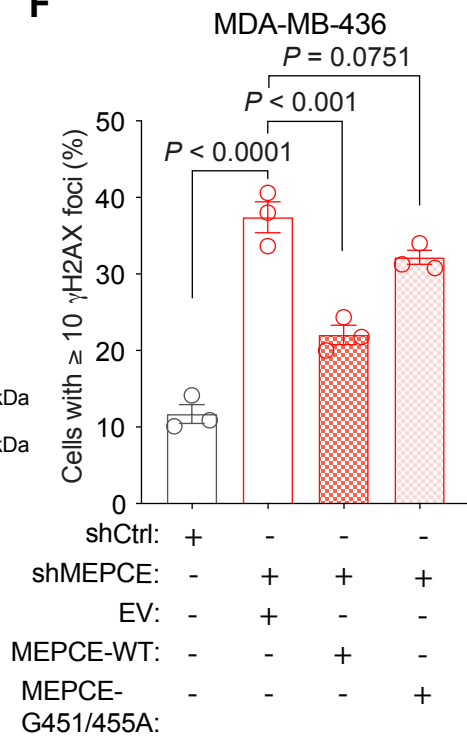
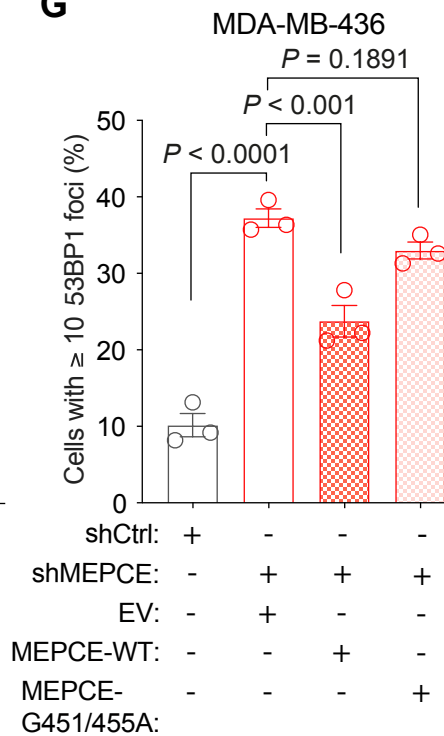
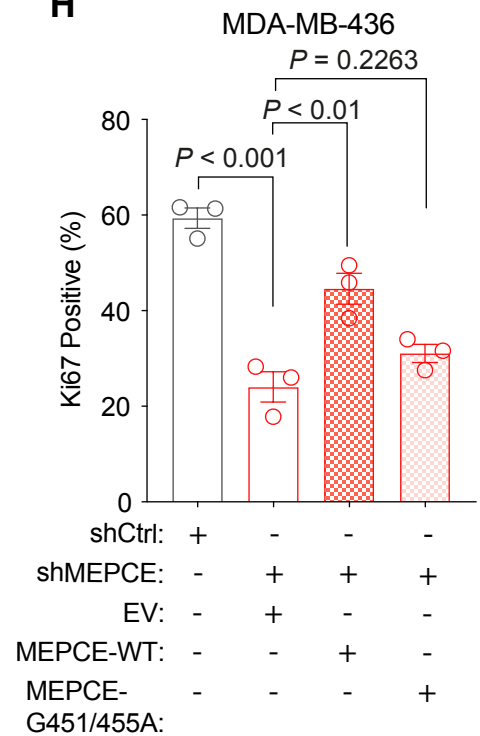
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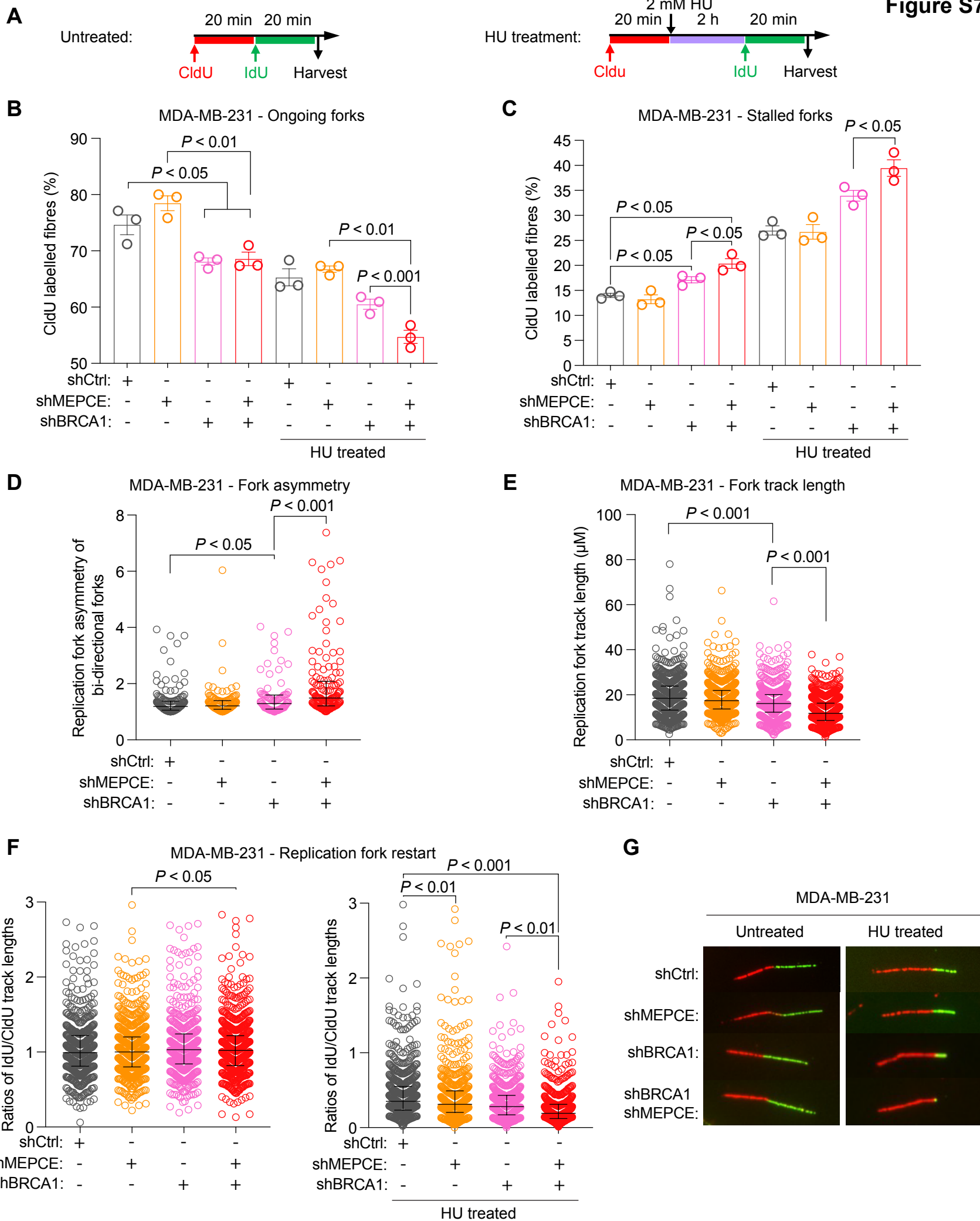
G



Supplementary Figure S5. Depletion of LARP7 in *BRCA1*-mutant cells leads to growth defects and genomic instability. (A) Western blots indicating LARP7 expression in indicated *BRCA1*-mutated cells (MDA-MB-436, SUM149PT, COV362). (B) Representative images are shown for clonogenic growth in indicated *BRCA1*-mutated cells. 500 cells were seeded in 6-cm dishes. Cells were fixed and quantified after 21-28 days of growth. (C) Quantification from (B). (D) Quantification of γ H2AX staining in indicated cells. (E) Quantification of γ H2AX staining intensity in indicated COV362 cells. (F) Quantification of S9.6 staining in indicated cells. (G) Quantification of γ H2AX staining in indicated cells treated with vehicle control or 10 μ M LDC-000067 for 2 hrs (median with interquartile range, >100 nuclei were scored per condition). C, D, E, and F, Student's *t*-test. G, one-way ANOVA.

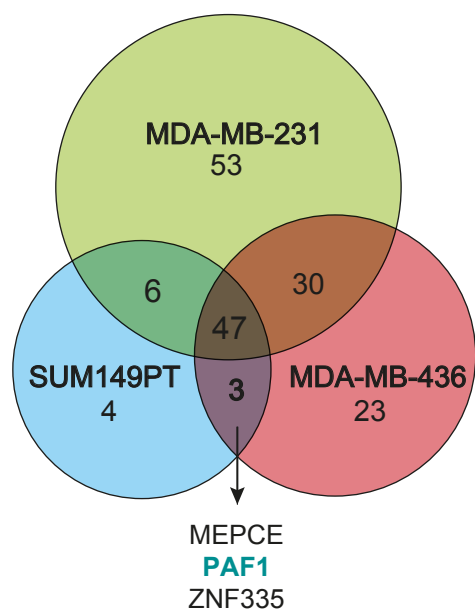
A**B****C****D****E****F****G****H**

Supplementary Figure S6. Concurrent loss of MEPCE and LARP7 in *BRCA1*-mutant cells does not leads to additive growth defects and MEPCE catalytic activity is important for suppressing DNA damage in *BRCA1*-null cells. (A, B) Western blots indicating MEPCE and LARP7 expression in MDA-MB-436 (A) and COV362 (B) cells transduced with shRNAs targeting a control, MEPCE, or LARP7 sequence as indicated. **(C, D)** Representative images and quantification of clonogenic growth of indicated MDA-MB-436 (C) and COV362 (D) cells. 500 cells were seeded in 6-cm dishes. Colonies were fixed and quantified after 21-28 days of growth (n = 3). **(E)** Western blot showing reconstitution of shMEPCE cells with MSCV-MEPCE-WT or MSCV-MEPCE-G451/455A. **(F)** Quantification of cells with equal to or greater than 10 γ H2AX foci in MDA-MB-436 shCtrl cells or shMEPCE MDA-MB-436 cells reconstituted with EV (empty vector), full-length MEPCE-WT or catalytically inactive MEPCE-G451/455A mutant. (n = 3). **(G)** Quantification of cells with equal to or greater than 10 foci of 53BP1 in MDA-MB-436 shCtrl cells or shMEPCE MDA-MB-436 cells reconstituted with EV, full-length MEPCE-WT or catalytically inactive MEPCE-G451/455A mutant. (n = 3). **(H)** Quantification of Ki67 positive shCtrl MDA-MB-436 cells and shMEPCE MDA-MB-436 cells reconstituted with EV, full-length MEPCE-WT or catalytically inactive MEPCE-G451/455A mutant. (n = 3). C, D, F, G, and H, mean \pm SEM are shown. Statistical significance was calculated using one-way ANOVA (Tukey's multiple comparisons test). n.s.: not significant.

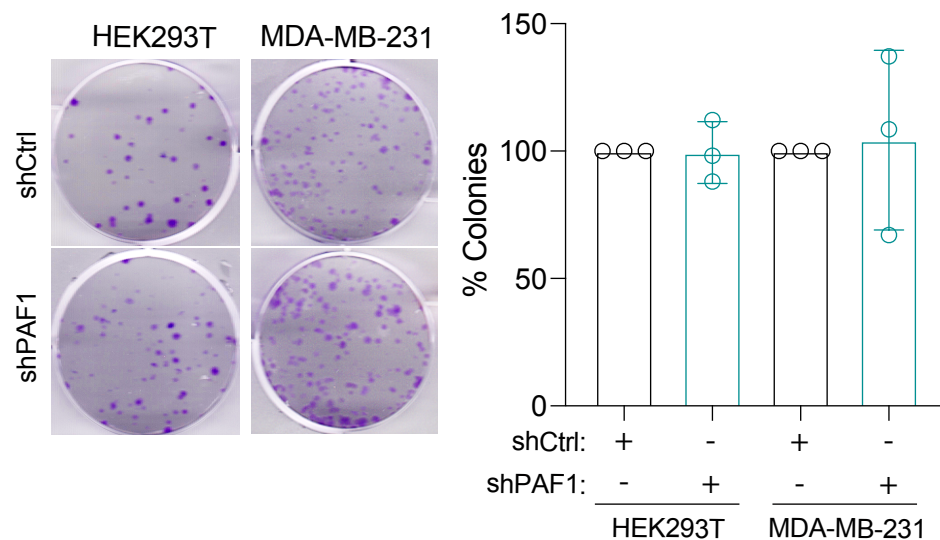


Supplementary Figure S7. MEPCE is dispensable for replication fork progression and stability in BRCA1-proficient cells. (A) Schematic of conditions for DNA fibre experiments in figure B-G. (B, C) Quantification of the percentage of ongoing forks (B), and stalled forks (C) in indicated MDA-MB-231 cells. (D) Quantification of fork asymmetry in indicated MDA-MB-231 cells. (E) Quantification of replication fork track length in indicated MDA-MB-231 cells. (F) Quantification of replication fork restart in indicated cells under indicated conditions in indicated MDA-MB-231 cells. (G) Representative images of ongoing replication forks from F. In all cases data is representative of the average of 3 independent experimental repeats. For A and B, mean \pm SEM are shown. At least 1200 CldU labelled replication fork structures were counted in total. For D, at least 90 bi-directional replication forks were analysed in total. For E and F, at least 580 ongoing replication fork structures were analysed in total. B and C, Students *t*-test was carried out. D, E, and F, Mann Whitney Rank Sum test.

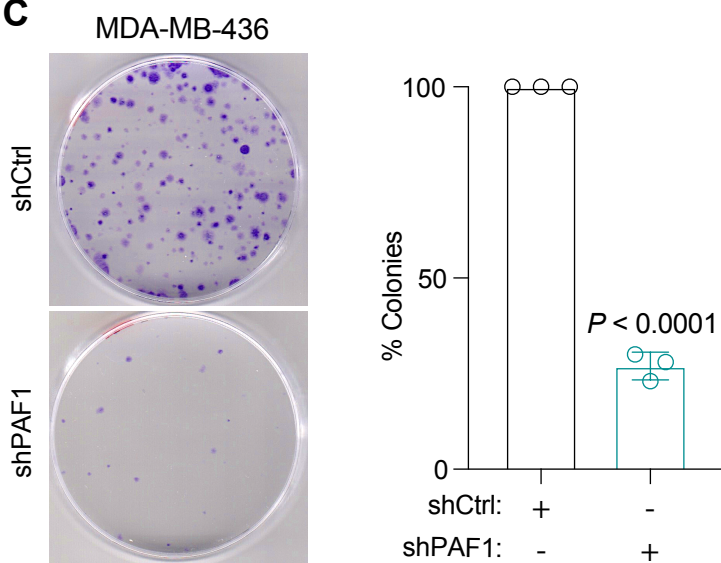
A MDA-MB-231 & SUM149PT Hits at FDR < 0.1
MDA-MB-436 Hits at FDR < 0.2



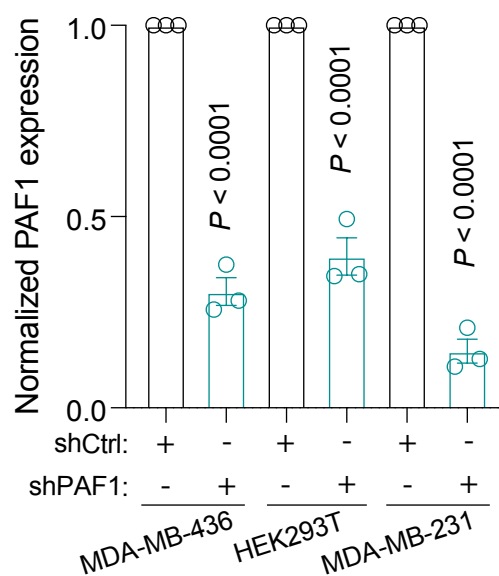
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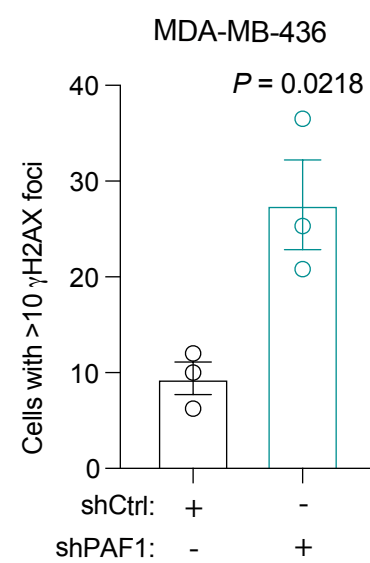
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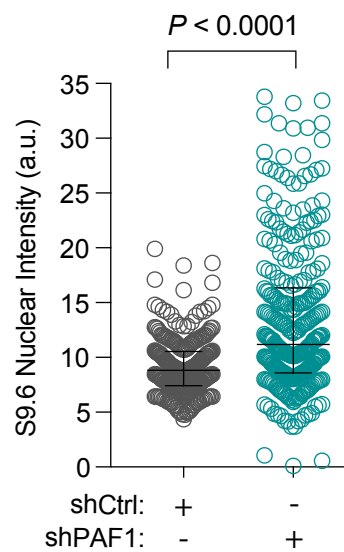
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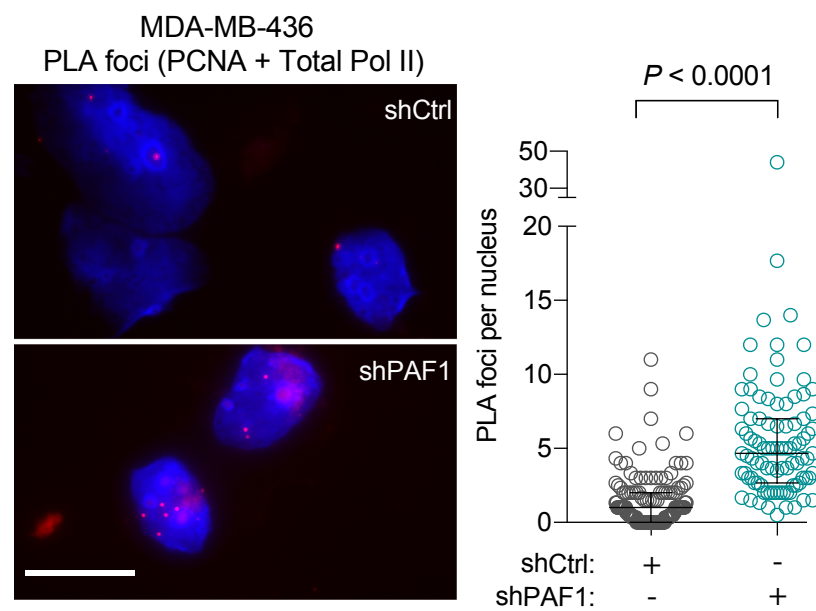
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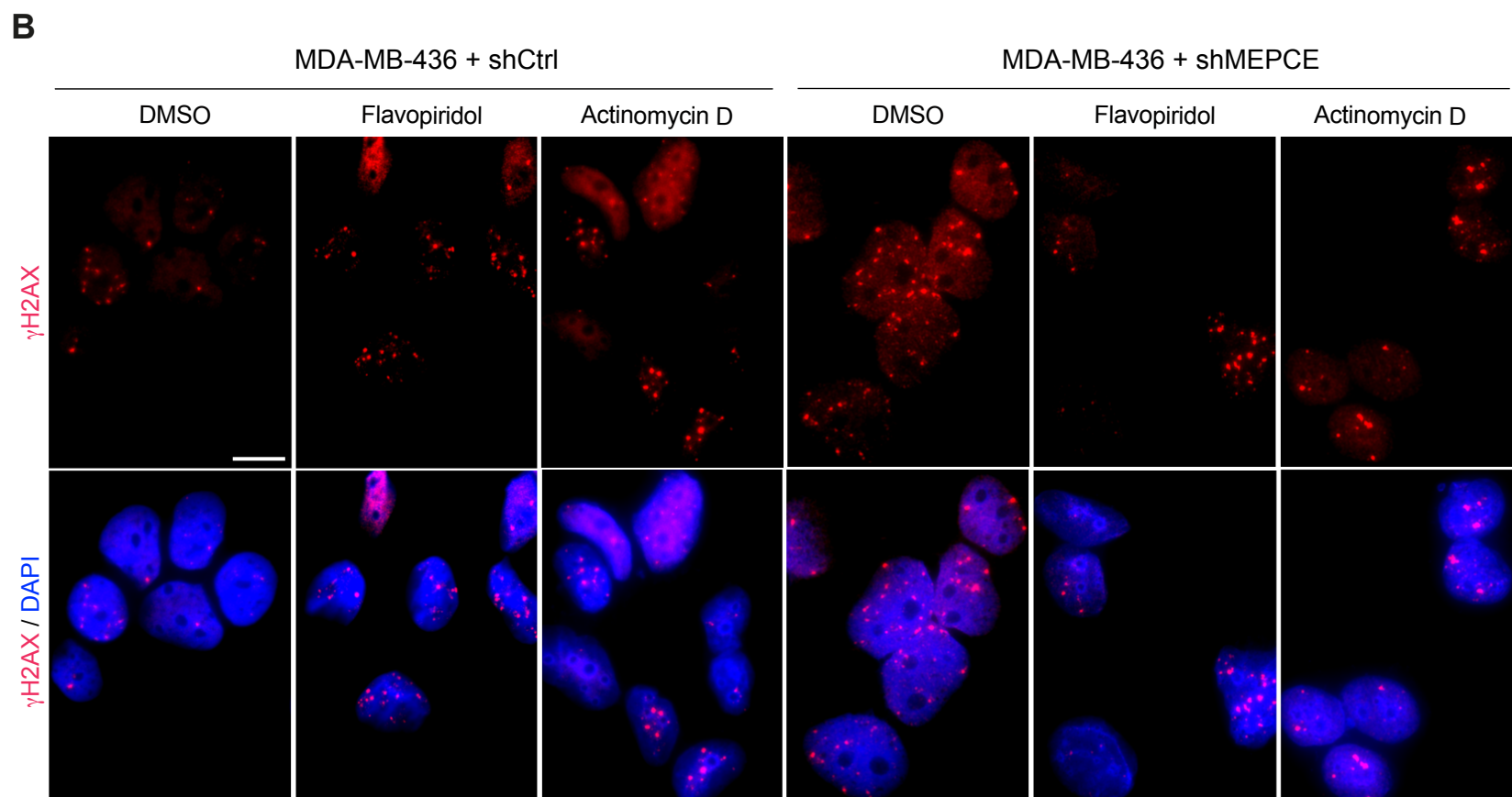
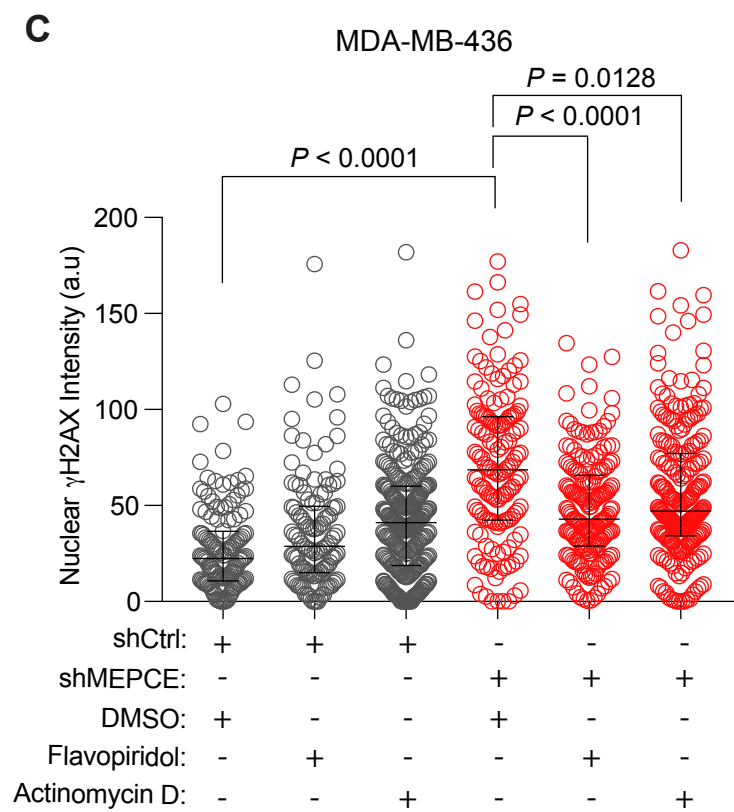
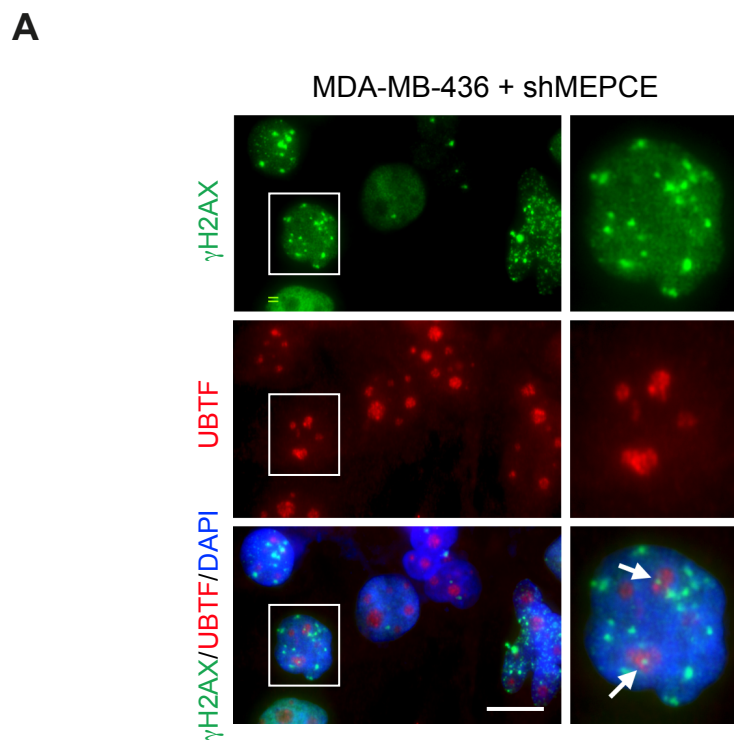
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Supplementary Figure S8. PAF1 deficiency in BRCA1-deficient cells leads to an increase in R-loop accumulation, genomic instability, and TRC. (A) Venn diagram representation of negatively selected genes in the indicated cell lines used for CRISPR-Cas9 screen. (B, C) Representative images and quantification of clonogenic growth of indicated cells. 500 cells were seeded and allowed to grow for 21-28 days. (D) *PAF1* mRNA expression normalized to 18S. (E) Quantification of cells with ≥ 10 spontaneous γ H2AX foci in indicated cells. (F) Quantification of S9.6 immunofluorescence in indicated cells. >100 nuclei were scored per condition. (G) Representative images and quantification of PLA (PCNA + Total Pol II) foci. Bar = 20 μ m. For B, C, D, and E, mean \pm SEM is shown, and Student's *t*-test was conducted. F and G, median with interquartile range is shown and statistical significance was calculated using Mann-Whitney Rank Sum test.



Supplementary Figure S9. MEPCE depletion causes non-nucleolar and nucleolar DNA damage.

(A) Representative images showing γ H2AX and UBTF immunofluorescence in MEPCE-depleted MDA-MB-436 cells. Bar = 25 μ M. (B) Representative images showing γ H2AX immunofluorescence in indicated MDA-MB-436 cells after treatment with DMSO, Flavopiridol (1 μ M), and Actinomycin D (25ng/ml) for 1 hr. Bar = 25 μ M. (C) Quantification of γ H2AX immunofluorescence in cells from (B). A minimum of 100 cells were quantified per replicate. Three biological experiments were completed. Statistical significance was calculated using one-way ANOVA (Tukey's multiple comparisons test).

Supplementary Table 1. Full list of RT-qPCR primers.

<i>ACTB</i> 5'Promoter	5'-CCACCTGGGTACACACAGTCT-3'	5'-TGCCTTGTCACCCTTTCTTG-3'
<i>ACTB</i> In1	5'-CGGGGTCTTTGTCTGAGC-3'	5'-CAGTTAGCGCCCAAAGGAC-3'
<i>ACTB</i> 5'Pause	5'-TTACCCAGAGTGCAGGTGTG-3'	5'-CCCCAATAAGCAGGAACAGA-3'
<i>ACTB</i> Pause	5'-GGGACTATTTGGGGGTGTCT-3'	5'-TCCCATAGGTGAAGGCAAAG-3'
<i>ACTB</i> C	5'-TGGGCCACTTAATCATTCAAC-3'	5'-CCTCACTTCCAGACTGACAGC-3'
<i>GAPDH</i> PPP	5'-CTCCTGTTCGACAGTCAGC-3'	5'-TTCAGGCCGTCCCTAGC-3'
<i>GAPDH</i> In5	5'-ATAGGCGAGATCCCTCAA-3'	5'-TGAAGACGCCAGTGGAC-3'
<i>GAPDH</i> TES	5'-CCCTGTGCTCAACCAGT -3'	5'-CTCACCTTGACACAAGCC-3'
<i>TEFM</i> PPP	5'-CTTGGAGATGAGCGGGTCTG-3'	5'-GACAGACGGGAAATCACCCC-3'
<i>TEFM</i> In2	5'-TGGCCAATGTGGTGAAAGCC-3'	5'-GGGACTACAGGCCACGCC-3'
<i>TEFM</i> TES	5'-ACCACATAGACTTTATGACAGAGAA-3'	5'-TCAATCCATGCTTGTGAAGCAA-3'
<i>18S</i>	5'-CAGCCACCCGAGATTGAGCA-3'	5'-TAGTAGCGACGGGCGGTGTG-3'
<i>PAF1</i>	5'-CCTGACACCTACCGCATCG-3'	5'-TGTA CTCTGTCTTTCGCATCCA-3'
<i>TMEM242</i>	5'-ATGCGCTATGTCTGTGGACC-3'	5'-GGCCCCAGTAAAGGTTAGCA-3'
<i>CSNK1D</i>	5'-GCGAGTTGAGTTCAGAGTCCA-3'	5'-CTGCCTACAGTTGGGACAGT-3'
<i>RASAI</i>	5'-AACCATGGGAAGCTCCCTCTA-3'	5'-TCCACAATGCAAGCACCTTC-3'
<i>CORT</i>	5'-TTCTCCAGAAGCAAGCGCAC-3'	5'-GAGCAATCAGTGCCACAAA-3'
<i>ANG</i>	5'-GGGTGAGTAGCGTCTCTTTCG-3'	5'-TATCAGCAAGCAGGACAGGGT-3'