Supporting Information

Nearest-neighbor parameters for the prediction of RNA duplex stability in diverse *in vitro* and cellular-like crowding conditions

Saptarshi Ghosh,¹ Shuntaro Takahashi,¹ Dipanwita Banerjee,¹ Tatsuya Ohyama,¹ Tamaki Endoh,¹ Hisae Tateishi-Karimata,¹ and Naoki Sugimoto^{1,2,*}

¹Frontier Institute for Biomolecular Engineering Research (FIBER), ²Graduate School of Frontiers of Innovative Research in Science and Technology (FIRST), Konan University, 7-1-20 Minatojimaminamimachi, Kobe 650-0047, Japan

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Supporting Method

Energetic estimation of nearest-neighbor (NN) parameters for G-U base pair and loop contribution

For G-U wobble base pairs, we estimated the stability of NN parameters including G-U pair according to the rules of the changes in NN pairs containing A-U pair because both base pairs have two hydrogen bonds, that are approximated to have same energy contribution of terminal A-U pair (Table S19) (1). For example, $\Delta G^{\circ}_{37, NN, no cosolute}$ value for r(GA/UU) in 100 mM NaCl was determined to be -0.96 kcal mol⁻¹ from its value of -1.27 kcal mol⁻¹ in 1 M NaCl, applying the same extent of reduction in the stability observed for r(AA/UU) in 100 mM NaCl from its value in 1 M NaCl. The m_{cs} values for NN pairs containing G-U base pairs in different cosolutes were considered to be same as determined for NN pairs having A-U pair (Table S19). Empirical relationships were proposed for the stability of different types of loop structures in varying concentrations of Na⁺ and Mg²⁺, although negligible cation dependency was observed for shorter loops (2). A previous study suggested that the destabilization effect of the hairpin tetraloop was reduced by 58% in 40 wt% PEG200 (Table S20) (3). Based on this information and as the stability of the loop changes with changing the water activity of the solution, we formulated an empirical relation for loop stability in the cosolute solutions ($\Delta G^{\circ}_{37, loop, cosolute}$) from their stability in the absence of cosolute ($\Delta G^{\circ}_{37, loop, no cosolute}$) and change in the water activity of cosolute solution (Δa_{w}) as follows:

 $\Delta G^{\circ}_{37, \text{ loop, cosolute}} = \Delta G^{\circ}_{37, \text{ loop, no cosolute}} \cdot (1 - 11.8 \cdot \Delta a_{w})$

 Δa_w is 0.049 for 40 wt% PEG200. The factor 11.8 was determined to match the extent of the reduction in the measured stability of a tetraloop in 40 wt% PEG200 (Table S20).

		Nearest-neighbor combinations present in duplex									
No.	Sequence ^b	AA	AU	UA	CU	CA	GU	GA	CG	GC	GG
		UU	UA	AU	GA	GU	CA	CU	GC	CG	CC
NS1	CAUGCC		1			2				1	1
NS2	CUAGGC			1	2					1	1
NS3	CAGCGG				1	1			1	1	1
NS4	CUACGC			1	1		1		1	1	
NS5	GAACUCC	1			1		1	2			1
NS6	GGCUGUCC										
NS7a	GGCUGUUC	1			1	1	1	1		1	1
NS7b	GGUUCUGC	1			1	1	1	1		1	1
NS8a	AGCUGUCU				3	1	1	1		1	
NS8b	AGUCUGCU				3	1	1	1		1	
NS9a	GGCAGUUC	1			1	1	1	1		1	1
NS9b	GGUUCAGC	1			1	1	1	1		1	1
NS10a	CGCUGUCG				1	1	1	1	2	1	
NS10b	CGUCUGCG				1	1	1	1	2	1	
NS11a	UGCUGUCA				1	3	1	1		1	
NS11b	UGUCUGCA				1	3	1	1		1	
NS12a	UAUGAGGA		1	1	1	1		2			1
NS12b	UAGAUGGA		1	1	1	1		2			1
NS13a	GUGCCGAG				1	1	1	1	1	1	1
NS13b	GCCGAGUG				1	1	1	1	1	1	1
NS14a	CGCUGUAG			1	2	1	1		1	1	
NS14b	CGUGCUAG			1	2	1	1		1	1	
NS15a	AUUGGAUACAAA	3	2	1		2	1	1			1
NS15b	AUACAUUGGAAA	3	2	1		2	1	1			1
NS16	GGCUCAAUUGAC	2	1		1	2	1	2		1	1
S1	UACAUGUA		1	2		2	2				
S2	GGUAUACC		1	2			2				2
S3	GGUUAACC	2		1			2				2
S4	CGAAUUCG	2	1					2	2		
S5	GCUUAAGC	2	1		2					2	
S6	CGUAUACG		1	2			2		2		
S7	CGCAUGCG		1			2			2	2	
S8	CGGUACCG			1			2		2		2
S9a	CAAGCUUG	2			2	2				1	
S9b	CUUGCAAG	2			2	2				1	
S10a	GAACGUUC	2					2	2	1		
S10b	GUUCGAAC	2					2	2	1		
S11a	GAUCCGGAUC		2					4	1		2
S11b	GGAUCGAUCC		2					4	1		2
S12a	AUGAGCUCAU		2		2		2	2		1	

Table S1. RNA sequences with nearest-neighbor frequencies^a

S12b	AUCAGCUGAU		2		2	2	2		1	
S13	UUACGCGUAA	2		2	2	2		2	1	
S14	AUCGCUAGCGAU		2	1			2	2	2	
S15a	CAUAGGCCUAUG		2	2	2	2			1	2
S15b	CUAUGGCCAUAG		2	2	2	2			1	2

^aTotal frequencies of the NN parameters are as follows: r(AA/UU) = 29, r(AU/UA) = 27, r(UA/AU) = 24, r(CA/GU) = 46, r(GU/CA) = 36, r(CU/GA) = 45, r(GA/CU) = 43, r(CG/GC) = 26, r(GC/CG) = 31, and r(GG/CC) = 33. ^bRNA duplex consists of denoted RNA strand (5' \rightarrow 3') and complementary RNA strand.

		Measured	in the presence	e of 40 wt% PEG	200 ^b	Predicted in absence cosolute	n the of e ^c	Difference be with and wi cosolut	etween ithout :e
No.	Sequence ^a	ΔH°	T∆S°	ΔG°_{37}	T _m d	ΔG°_{37}	T_m^d	$\Delta\Delta G^{\circ}_{37}$	ΔT_m
		(kcal mol ⁻¹)	(kcal mol ⁻¹)	(kcal mol ⁻¹)	()	(kcal mol ⁻¹)	()	(kcal mol ⁻¹)	(C)
NS1	CAUGCC	-56.5 ± 1.9	-50.9 ± 1.7	-5.6 ± 0.3	31.0	-6.4	35.2	0.8	-4.2
NS2	CUAGGC	-57.5 ± 1.9	-51.4 ± 1.7	-6.1 ± 0.3	34.3	-7.2	39.8	1.1	-5.5
NS3	CAGCGG	-65.6 ± 3.0	-59.2 ± 2.7	-6.4 ± 0.4	36.4	-8.1	44.4	1.7	-8.0
NS4	CUACGC	-62.8 ± 4.7	-58.1 ± 4.4	-4.7 ± 0.6	27.8	-6.6	37.2	1.9	-9.4
NS5	GAACUCC	-71.8 ± 2.3	-65.9 ± 2.1	-5.9 ± 0.3	34.1	-8.1	43.3	2.2	-9.2
NS6	GGCUGUCC	-92.0 ± 2.0	-80.6 ± 3.0	-11.4 ± 0.6	54.3	-13.4	63.2	2.0	-8.9
NS7a	GGCUGUUC	-79.8 ± 1.9	-71.1 ± 1.7	-8.7 ± 0.7	45.6	-11.0	54.0	2.3	-8.4
NS7b	GGUUCUGC	-76.9 ± 2.1	-68.5 ± 1.9	-8.4 ± 0.8	44.9	-11.0	54.0	2.6	-9.1
NS8a	AGCUGUCU	-72.6 ± 2.1	-64.8 ± 1.9	-7.8 ± 0.9	42.4	-9.9	50.8	2.1	-8.4
NS8b	AGUCUGCU	-61.6 ± 0.6	-53.9 ± 0.6	-7.7 ± 0.3	42.6	-9.9	50.8	2.2	-8.2
NS9a	GGCAGUUC	-75.2 ± 2.3	-66.5 ± 2.0	-8.7 ± 0.9	46.2	-11.0	54.0	2.3	-7.8
NS9b	GGUUCAGC	-74.9 ± 2.2	-66.4 ± 2.0	-8.5 ± 0.9	45.1	-11.0	54.0	2.5	-8.9
NS10a	CGCUGUCG	-78.4 ± 2.4	-68.6 ± 2.1	-9.8 ± 1.0	49.8	-11.5	56.2	1.7	-6.4
NS10b	CGUCUGCG	-75.4 ± 2.4	-65.7 ± 2.1	-9.7 ± 1.0	50.3	-11.5	56.2	1.8	-5.9
NS11a	UGCUGUCA	-66.2 ± 1.8	-58.4 ± 1.6	-7.8 ± 0.7	43.0	-9.8	49.6	2.0	-6.6
NS11b	UGUCUGCA	-65.3 ± 0.9	-57.5 ± 0.8	-7.8 ± 0.4	42.9	-9.8	49.6	2.0	-6.7
NS12a	UAUGAGGA	-71.4 ± 1.4	-65.4 ± 1.3	-6.0 ± 0.6	34.7	-8.0	41.4	2.0	-6.7
NS12b	UAGAUGGA	-62.7 ± 0.8	-56.6 ± 0.7	-6.1 ± 0.3	35.2	-8.0	41.4	1.9	-6.2
NS13a	GUGCCGAG	-81.3 ± 1.3	-70.5 ± 1.1	-10.8 ± 0.5	54.3	-12.4	59.3	1.6	-5.0
NS13b	GCCGAGUG	-71.6 ± 2.1	-61.2 ± 1.8	-10.4 ± 0.8	54.1	-12.4	59.3	2.0	-5.2
NS14a	CGCUGUAG	-72.2 ± 0.9	-64.0 ± 0.8	-8.2 ± 0.4	44.1	-10.2	51.7	2.0	-7.6
NS14b	CGUGCUAG	-72.7 ± 1.9	-64.2 ± 1.7	-8.5 ± 0.8	45.8	-10.2	51.7	1.7	-5.9
NS15a	AUUGGAUACAAA	-105.8 ± 1.4	-97.4 ± 1.3	-8.4 ± 0.5	42.3	-10.5	48.4	2.1	-6.1
NS15b	AUACAUUGGAAA	-107.5 ± 1.2	-99.4 ± 1.1	-8.1 ± 0.5	41.6	-10.5	48.4	2.4	-6.8
NS16	GGCUCAAUUGAC	-118.5 ± 1.5	-102.8 ± 1.2	-15.7 ± 0.5	56.3	-16.2	61.7	0.5	-5.4
S1	UACAUGUA	-62.7 ± 2.9	-59.1 ± 2.8	-3.6 ± 0.3	28.2	-5.6	35.9	2.0	-7.7
S2	GGUAUACC	-71.9 ± 3.9	-65.7 ± 3.6	-6.2 ± 0.5	39.2	-9.1	51.8	2.9	-12.6
S3	GGUUAACC	-73.4 ± 2.8	-67.7 ± 2.6	-5.7 ±0.4	36.5	-8.7	50.3	3.0	-13.8
S4	CGAAUUCG	-70.3 ± 2.9	-65.8 ± 2.7	-4.5 ± 0.4	31.4	-7.0	38.5	2.5	-7.1
S5	GCUUAAGC	-70.4 ± 3.2	-64.3 ±2.9	-6.1 ± 0.4	38.6	-8.4	48.1	2.3	-9.5

Table S2. Thermodynamic parameters for RNA duplexes measured in the crowding condition andpredicted in the absence of cosolute

S6	CGUAUACG	-73.8 ± 3.5	-68.3 ± 3.3	-5.5 ± 0.5	35.7	-7.3	43.9	1.8	-8.2
S7	CGCAUGCG	-79.3 ±3.7	-69.7 ± 3.2	-9.6 ± 0.6	52.7	-10.6	55.5	1.0	-2.8
S8	CGGUACCG	-73.4 ±6.2	-64.4 ± 5.5	-9.0 ± 1.0	51.0	-11.7	62.3	2.7	-11.3
S9a	CAAGCUUG	-78.3 ± 2.4	-73.0 ± 2.3	-5.3 ± 1.0	35.8	-7.4	45.3	2.1	-9.5
S9b	CUUGCAAG	-77.5 ± 2.6	-71.5 ± 2.4	-6.0 ± 1.1	38.3	-7.4	45.3	1.4	-7.0
S10a	GAACGUUC	-73.9 ± 1.2	-68.2 ± 1.1	-5.7 ± 0.5	36.8	-7.8	45.4	2.1	-8.6
S10b	GUUCGAAC	-83.1 ± 3.4	-77.9 ± 3.2	-5.2 ± 1.3	35.4	-7.8	45.4	2.6	-10.0
S11a	GAUCCGGAUC	-103.9 ± 1.7	-91.6 ± 1.5	-12.3 ± 0.7	58.4	-13.8	60.6	1.5	-2.2
\$11b	GGAUCGAUCC	-100.1 ± 1.8	-88.7 ± 1.6	-11.4 ± 0.7	56.0	-13.8	60.6	2.4	-4.6
S12a	AUGAGCUCAU	-80.7 ± 2.3	-72.1 ± 2.0	-8.6 ± 0.3	48.8	-11.3	56.7	2.7	-7.9
S12b	AUCAGCUGAU	-83.3 ± 1.3	-75.1 ± 1.1	-8.2 ± 0.5	46.7	-11.3	56.7	3.1	-10.0
S13	UUACGCGUAA	-78.9 ±4.7	-71.7 ± 4.2	-7.2 ±0.6	42.2	-10.5	54.6	3.3	-12.4
S14	AUCGCUAGCGAU	-105.3 ± 4.7	-92.3 ± 4.8	-13.0 ± 0.8	60.0	-16.0	65.5	3.0	-5.5
\$15a	CAUAGGCCUAUG	-130.9 ± 2.1	-116.5 ± 1.9	-14.4 ± 0.7	59.1	-16.5	68.2	2.1	-9.1
\$15b	CUAUGGCCAUAG	-131.7 + 2.2	-117.2 + 2.2	-14.5 + 0.7	59.4	-16.5	68.2	2.0	-8.8

^aRNA duplex consists of a denoted RNA strand (5' \rightarrow 3') and complementary RNA strand. The pair of RNA oligonucleotides with identical nearest neighbors are shown by the numbers a and b. ^bAll experiments were performed in a buffer containing 100 mM NaCl, 10 mM Na₂HPO₄ (pH 7.0), and 1 mM Na₂EDTA. ^cThe values in the absence of cosolute were calculated from the parameters of 121 mM Na⁺ reported by Weber *et al.*(4) (Table S3) ^dMelting temperatures were calculated for a total strand concentration of 100 μ M.

Sequence	ΔH° _{NN}	$\Delta S^{\circ}_{\rm NN}$	ΔG° 37, NN, no cosolute
	(kcal mol ⁻¹)	(cal mol ⁻¹ K ⁻¹)	(kcal mol ⁻¹)
r(AA/UU)	-10.7	-32.2	-0.70
r(AU/UA)	-7.6	-22.8	-0.52
r(UA/AU)	-11.5	-32.9	-1.30
r(CA/GU)	-11.7	-32.0	-1.78
r(GU/CA)	-11.2	-29.2	-2.14
r(CU/GA)	-9.6	-25.1	-1.80
r(GA/CU)	-14.8	-40.6	-2.22
r(CG/GC)	-10.9	-28.1	-2.18
r(GC/CG)	-15.1	-38.0	-3.30
r(GG/CC)	-12.9	-31.6	-3.10
initiation	3.6	-1.5	4.09
per terminal AU	3.7	10.5	0.45
self-complementary ^b	0	-1.4	0.43
non-self-complementary	0	0	0

Table S3. NN parameters for RNA duplex formation in the absence of cosolute in 100 mM NaCl at 37 $^\circ C^a$

^aParameters are collected from the report by Weber et al.(4) ^bSymmetry factors for self-complementary sequences are independent of the solution conditions.

Table S4. Stability contribution of 40 wt% PEG200 ($\Delta\Delta G^{\circ}_{37, [40 wt\% PEG200]} = \Delta G^{\circ}_{37, [40 wt\% PEG200, 100 mM NaCl]} - \Delta G^{\circ}_{37, [no cosolute, 100 mM NaCl]}$) on the individual nearest-neighbor base pairs for RNA duplex formation at 100 mM NaCl

Sequence	
Sequence	220 37, [40 wt% PEG200]
	(kcal mol ⁻¹)
r(AA/UU)	0.13
r(AU/UA)	-0.03
r(UA/AU)	-0.03
r(CA/GU)	-0.36
r(GU/CA)	0.34
r(CU/GA)	0.03
r(GA/CU)	-0.02
r(CG/GC)	0.02
r(GC/CG)	0.23
r(GG/CC)	0.21
initiation	1.41
per terminal AU	0.40

 $^{a}\Delta G^{\circ}_{37, [40 wt\% PEG200, 100 mM NaCI]}$ were taken from Table 1 and $\Delta G^{\circ}_{37, [no cosolute, 100 mM NaCI]}$ were collected from Table S3.

			Measure	d ^b		Predicted				
No.	Sequence ^a	∆ <i>H</i> ° (kcal mol⁻¹)	ΔS° (cal mol ⁻¹ K ⁻¹)	ΔG°_{37} (kcal mol ⁻¹)	Т _m с (°С)	∆ <i>H</i> ° (kcal mol⁻¹)	∆S° (cal mol ⁻¹ K ⁻¹)	ΔG°_{37} (kcal mol ⁻¹)	Т _m с (°С)	
NS1	CAUGCC	-56.5	-164.1	-5.6	31.0	-58.3	-170.9	-5.3	30.6	
NS2	CUAGGC	-57.5	-165.7	-6.1	34.3	-57.5	-168.2	-5.3	30.7	
NS3	CAGCGG	-65.6	-190.9	-6.4	36.4	-58.4	-167.2	-6.5	37.1	
NS4	CUACGC	-62.8	-187.3	-4.7	27.8	-53.3	-156.9	-4.6	26.4	
NS5	GAACUCC	-71.8	-212.5	-5.9	34.1	-65.5	-191.8	-6.0	34.6	
NS6	GGCUGUCC	-92.0	-259.9	-11.4	54.3	-84.5	-236.0	-11.3	55.6	
NS7a	GGCUGUUC	-79.8	-229.2	-8.7	45.6	-79.7	-228.0	-9.0	46.9	
NS7b	GGUUCUGC	-76.9	-220.9	-8.4	44.9	-79.7	-228.0	-9.0	46.9	
NS8a	AGCUGUCU	-72.6	-208.9	-7.8	42.4	-64.3	-183.6	-7.4	41.0	
NS8b	AGUCUGCU	-61.6	-173.8	-7.7	42.6	-64.3	-183.6	-7.4	41.0	
NS9a	GGCAGUUC	-75.2	-214.4	-8.7	46.2	-79.7	-228.0	-9.0	46.9	
NS9b	GGUUCAGC	-74.9	-214.1	-8.5	45.1	-79.7	-228.0	-9.0	46.9	
NS10a	CGCUGUCG	-78.4	-221.2	-9.8	49.8	-77.1	-216.8	-9.8	51.0	
NS10b	CGUCUGCG	-75.4	-211.8	-9.7	50.3	-77.1	-216.8	-9.8	51.0	
NS11a	UGCUGUCA	-66.2	-188.3	-7.8	43.0	-66.1	-187.0	-8.1	44.6	
NS11b	UGUCUGCA	-65.3	-185.4	-7.8	42.9	-66.1	-187.0	-8.1	44.6	
NS12a	UAUGAGGA	-71.4	-210.9	-6.0	34.7	-65.1	-190.7	-6.0	34.3	
NS12b	UAGAUGGA	-62.7	-182.5	-6.1	35.2	-65.1	-190.7	-6.0	34.3	
NS13a	GUGCCGAG	-81.3	-227.3	-10.8	54.3	-80.8	-226.4	-10.6	53.4	
NS13b	GCCGAGUG	-71.6	-197.3	-10.4	54.1	-80.8	-226.4	-10.6	53.4	
NS14a	CGCUGUAG	-72.2	-206.4	-8.2	44.1	-76.6	-219.4	-8.5	45.4	
NS14b	CGUGCUAG	-72.7	-207.0	-8.5	45.8	-76.6	-219.4	-8.5	45.4	
NS15a	AUUGGAUACAAA	-105.8	-314.0	-8.4	42.3	-105.1	-312.6	-8.2	41.8	
NS15b	AUACAUUGGAAA	-107.5	-320.5	-8.1	41.6	-105.1	-312.6	-8.2	41.8	
NS16	GGCUCAAUUGAC	-118.5	-331.5	-15.7	56.3	-123.6	-351.8	-14.5	58.3	
S1	UACAUGUA	-62.7	-190.6	-3.6	28.2	-60.3	-183.3	-3.5	26.0	
S2	GGUAUACC	-71.9	-211.8	-6.2	39.2	-78.7	-232.3	-6.7	40.9	
S3	GGUUAACC	-73.4	-218.3	-5.7	36.5	-77.5	-230.8	-5.9	38.0	
S4	CGAAUUCG	-70.3	-212.2	-4.5	31.4	-71.1	-214.5	-4.6	32.3	
S5	GCUUAAGC	-70.4	-207.3	-6.1	38.6	-76.5	-226.6	-6.2	39.2	
S6	CGUAUACG	-73.8	-220.2	-5.5	35.7	-71.3	-213.1	-5.2	35.0	
S7	CGCAUGCG	-79.3	-224.7	-9.6	52.7	-79.5	-226.1	-9.4	52.1	

Table S5. Measured and predicted thermodynamic parameters for RNA duplex formation with 40 wt% PEG200 and 100 mM NaCl in 10 mM phosphate buffer (pH 7.0)

S8	CGGUACCG	-73.4	-207.6	-9.0	51.0	-79.7	-227.6	-9.1	51.0
S9a	CAAGCUUG	-78.3	-235.4	-5.3	35.8	-75.8	-224.7	-6.1	38.8
S9b	CUUGCAAG	-77.5	-230.5	-6.0	38.3	-75.8	-224.7	-6.1	38.8
S10a	GAACGUUC	-73.9	-219.9	-5.7	36.8	-71.3	-212.3	-5.5	36.0
S10b	GUUCGAAC	-83.1	-251.2	-5.2	35.4	-71.3	-212.3	-5.5	36.0
S11a	GAUCCGGAUC	-103.9	-295.3	-12.3	58.4	-103.1	-293.5	-12.1	57.5
S11b	GGAUCGAUCC	-100.1	-286.0	-11.4	56.0	-103.1	-293.5	-12.1	57.5
S12a	AUGAGCUCAU	-80.7	-232.5	-8.6	48.8	-86.4	-250.1	-8.8	48.8
S12b	AUCAGCUGAU	-83.3	-242.1	-8.2	46.7	-86.4	-250.1	-8.8	48.8
S13	UUACGCGUAA	-78.9	-231.2	-7.2	42.2	-82.0	-241.3	-7.2	42.7
S14	AUCGCUAGCGAU	-105.3	-297.6	-13.0	60.0	-109.3	-309.6	-13.3	60.2
S15a	CAUAGGCCUAUG	-130.9	-375.6	-14.4	59.1	-127.8	-365.3	-14.5	60.0
S15b	CUAUGGCCAUAG	-131.7	-377.9	-14.5	59.4	-127.8	-365.3	-14.5	60.0

^aRNA duplex consists of denoted RNA strand (5' \rightarrow 3') and complementary RNA strand. ^bErrors associated with the measured values were mentioned in the Table S2. ^cMelting temperatures were calculated for total strand concentration of 100 μ M.

Sequence	R a	L ^b	Р ^с	<i>C</i> (<i>p</i>) ^d	k e	V/N ^f
	(Å)	(Å)			(Å)	(L mol ⁻¹)
6-bp duplex	10.0	20.4	2.04	1.20	8.88	0.76
6-nt strand	5.0	20.4	4.08	1.76	6.09	0.52
12-bp duplex	10.0	40.8	4.08	1.76	12.17	1.04
12-nt strand	5.0	40.8	8.16	2.37	9.01	0.77
24-bp duplex	10.0	81.6	8.16	2.37	18.03	1.54
24-nt strand	5.0	81.6	16.32	3.03	14.12	1.20
36-bp duplex	10.0	122.4	12.24	2.75	23.29	1.99
36-nt strand	5.0	122.4	24.48	3.42	18.75	1.60
34-nt hairpin	10.0	57.8	5.78	2.06	14.71	1.25
34-nt strand	5.0	115.6	23.12	3.36	18.01	1.54

Table S6. Parameters for RNA-PEG excluded volume calculations

^a Radius of equivalent cylinder. ^bLength of equivalent cylinder. ^{a, b} Values are collected from the report of Knowles et al. (5) following both DNA and RNA oligomeric duplexes has almost same radius and length.(6) ^cp = L/R. ^d $C(p) = (1+p^2)^{3/2}/3p - p^2/3 + 2/3p - (1+p^2)^{1/2}/p + \ln [p+(1+p^2)^{1/2}]$. ^e $k = \pi pR/6C(p)$. ^fObtained by using equation 10 and Kuhn length (*l*) of PEG was taken as 11.9 Å.(5)

Sequence	∆V/N ª	$\Delta G^{\circ}_{37, ev, dx} / C \bullet N^{b}$	
Sequence	(L mol⁻¹)	(kcal mol ⁻² kg)	
6-bp duplex	-0.28	-0.17	
12-bp duplex	-0.50	-0.31	
24-bp duplex	-0.86	-0.52	
36-bp duplex	-1.21	-0.74	

Table S7. Excluded volume and energy contribution of excluded volume for different lengths ofRNA duplexes with PEG

 $^{\circ}$ Calculated from the values in Table S6 using equation 9. b Calculated by using equation 8 and taking density of water 0.99 kg L⁻¹ at 37 $^{\circ}$ C.(5)

	ΔH°	T∆S°	∆G° 37	ΔG° 37, ev, dup ^a	ΔG° 37, wa, dup ^b	Water activity ^c
Solution	(kcal mol ⁻¹)	(cal mol ⁻¹ K ⁻¹)	(kcal mol ⁻¹)	(kcal mol ⁻¹)	(kcal mol ⁻¹)	
In 100 mM NaCl						
No cosolute	-97.8 ± 1.6	-83.7 ± 1.5	-14.1 ± 0.1			0.996
10 wt% EG	-97.9 ± 4.0	-84.3 ± 3.8	-13.6 ± 0.3	-0.45	0.95	0.970
15 wt% EG	-101.1 ± 1.0	-87.7 ± 0.9	-13.4 ± 0.1	-0.68	1.38	0.959
5 wt% Gly	-95.4 ± 2.6	-81.5 ± 2.3	-13.9 ± 0.3	-0.15	0.35	0.987
10 wt% Gly	-98.4 ± 1.6	-84.4 ± 1.5	-13.6 ± 0.1	-0.30	0.80	0.975
15 wt% Gly	-98.6 ± 1.0	-85.2 ± 0.8	-13.4 ± 0.1	-0.45	1.15	0.965
20 wt% Gly	-97.3 ± 1.7	-84.2 ± 1.6	-13.1 ± 0.1	-0.61	1.61	0.951
5 wt% 1,3 PDO	-100.1 ± 1.2	-86.2 ± 1.1	-13.9 ± 0.1	-0.18	0.38	0.985
10 wt% 1,3 PDO	-100.4 ± 3.1	-86.7 ± 2.9	-13.7 ± 0.3	-0.37	0.77	0.971
20 wt% 1,3 PDO	-103.5 ± 0.5	-90.0 ± 0.5	-13.5 ± 0.1	-0.73	1.33	0.947
40 wt% 1,3 PDO	-105.8 ± 2.2	-92.9 ± 2.1	-12.9 ± 0.1	-1.47	2.67	0.907
10 wt% 2-ME	-100.8 ± 1.6	-87.7 ± 1.6	-13.1 ± 0.1	-0.37	1.37	0.975
40 wt% 1,2-DME	-94.7 ± 5.5	-81.5 ± 5.1	-13.2 ± 0.3	-1.24	2.14	0.965
10 wt% PEG200	-102.2 ± 1.8	-88.3 ± 1.6	-13.9 ± 0.2	-0.56	0.76	0.985
20 wt% PEG200	-100.7 ± 1.3	-87.3 ± 1.2	-13.4 ± 0.1	-1.12	1.82	0.971
30 wt% PEG200	-101.4 ± 1.7	-88.4 ± 1.6	-13.0 ± 0.2	-1.67	2.77	0.956
40 wt% PEG200	-96.2 ± 1.9	-84.2 ± 1.8	-12.0 ± 0.2	-2.23	4.33	0.947
50 wt% PEG200	-90.6 ± 1.0	-79.6 ± 1.0	-11.0 ± 0.1	-2.79	5.89	0.931
20 wt% PEG400	-96.0 ± 5.4	-82.5 ± 5.1	-13.5 ± 0.3	-1.12	1.72	0.980
20 wt% PEG600	-103.5 ± 2.9	-88.9 ± 2.7	-14.6 ± 0.2	-1.21	0.71	0.983
In 1 M NaCl						
No cosolute	-98.4 ± 1.2	-81.9 ± 1.2	-16.5 ± 0.1			0.967
20 wt% EG	-99.8 ± 1.9	-84.8 ± 1.8	-15.0 ± 0.1	-0.90	2.40	0.907
15 wt% 1,3 PDO	-103.8 ± 2.1	-88.0 ± 2.0	-15.8 ± 0.2	-0.55	1.25	0.933
15 wt% 2-ME	-99.5 ± 0.6	-84.9 ± 0.5	-14.6 ± 0.1	-0.55	2.45	0.927
20 wt% 1,2-DME	-99.0 ± 2.7	-83.8 ± 2.7	-15.2 ± 0.2	-0.62	1.92	0.934
10 wt% PEG200	-98.5 ± 1.8	-82.4 ± 1.6	-16.1 ± 0.1	-0.56	0.96	0.953
20 wt% PEG200	-100.5 ± 2.4	-85.1 ± 2.2	-15.4 ± 0.2	-1.12	2.22	0.939
40 wt% PEG200	-92.3 ± 2.3	-78.9 ± 2.7	-13.4 ± 0.7	-2.23	5.33	0.911

Table S8. Thermodynamic parameters of GAUUACGCCUG in different cosolute solutions and the water activity values of the solutions

^aCalculated by using equation 11. Excluded volume for Gly, 1,3 PDO, 2-ME and 1,2-DME were considered to be same as EG due to similar molecular weights. ^b $\Delta G^{\circ}_{37, wa, dup} = \Delta G^{\circ}_{37} - \Delta G^{\circ}_{37, no cosolute} - \Delta G^{\circ}_{37, ev, dup}$. ^cWater activities were calculated at 37 °C from corresponding osmolality values of respective solutions calculated as described in Methods section. Error limit was ± 0.002.

			NaCl ^b	NaCl ^b KCl ^c					
No.	Sequence ^a	∆ <i>H</i> ° (kcal mol⁻¹)	$T\Delta S^{\circ}$ (cal mol ⁻¹ K ⁻¹)	ΔG°_{37} (kcal mol ⁻¹)	T _m ^d (°C)	∆ <i>H</i> ° (kcal mol⁻¹)	<i>T∆S</i> ° (cal mol ⁻¹ K ⁻¹)	ΔG°_{37} (kcal mol ⁻¹)	T _m d (°C)
NS3	CAGCGG	-65.6 ± 3.0	-59.2 ± 2.7	-6.4 ± 0.4	36.4	-55.2 ± 2.7	-49.1 ± 2.4	-6.1 ± 0.4	34.3
NS5	GAACUCC	-71.8 ± 2.3	-65.9 ± 2.1	-5.9 ± 0.3	34.1	-63.2 ± 1.9	-57.8 ± 1.8	-5.4 ± 0.3	31.2
NS10a	CGCUGUCG	-78.4 ± 2.4	-68.6 ± 2.1	-9.8 ± 1.0	49.8	-71.9 ± 0.7	-63.1 ± 0.6	-8.8 ± 0.1	47.1
NS11a	UGCUGUCA	-66.2 ± 1.8	-58.4 ± 1.6	-7.8 ± 0.7	43.0	-62.6 ± 1.6	-55.6 ± 1.4	-7.0 ± 0.2	39.6
NS13a	GUGCCGAG	-81.3 ± 1.3	-70.5 ± 1.1	-10.8 ± 0.5	54.3	-72.5 ± 3.7	-62.8 ± 3.2	-9.7 ± 0.6	51.1
NS13b	GCCGAGUG	-71.6 ± 2.1	-61.2 ± 1.8	-10.4 ± 0.8	54.1	-71.8 ± 5.4	-62.0 ± 4.7	-9.8 ± 0.9	51.6
S2	GGUAUACC	-71.9 ± 3.9	-65.7 ± 3.6	-6.2 ± 0.5	39.2	-72.3 ± 2.0	-66.8 ± 1.8	-5.5 ± 0.3	36.1
S3	GGUUAACC	-73.4 ± 2.8	-67.7 ± 2.6	-5.7 ± 0.4	36.5	-66.7 ± 3.3	-61.7 ± 3.1	-5.0 ± 0.5	33.6
S5	GCUUAAGC	-70.4 ± 3.2	-64.3 ± 2.9	-6.1 ± 0.4	38.6	-64.2 ± 1.1	-59.1 ± 1.0	-5.1 ± 0.1	34.1
S7	CGCAUGCG	-79.3 ± 3.7	-69.7 ± 3.2	-9.6 ± 0.6	52.7	-75.1 ± 2.9	-66.6 ± 2.6	-8.5 ± 0.5	48.6
S8	CGGUACCG	-73.4 ± 6.2	-64.4 ± 5.5	-9.0 ± 1.0	52.7	-69.9 ± 3.5	-61.0 ± 3.1	-8.6 ± 0.6	50.4
S11a	GAUCCGGAUC	-103.9 ± 1.7	-91.6 ± 1.5	-12.3 ± 0.7	58.4	-105.3 ± 4.1	-93.9 ± 3.7	-11.4 ± 0.6	55.0
S12a	AUGAGCUCAU	-80.7 ± 2.3	-72.1 ± 2.0	-8.6 ± 0.3	48.8	-79.6 ± 3.5	-72.2 ± 3.2	-7.4 ± 0.5	44.1
S12b	AUCAGCUGAU	-83.3 ± 1.3	-75.1 ± 1.1	-8.2 ± 0.5	46.7	-79.8 ± 2.7	-72.8 ± 2.4	-7.0 ± 0.4	42.1
S13	UUACGCGUAA	-78.9 ± 4.7	-71.7 ± 4.2	-7.2 ± 0.6	42.2	-77.7 ± 1.7	-71.5 ± 1.5	-6.2 ± 0.2	38.8
S14	AUCGCUAGCGAU	-105.3 ± 4.7	-92.3 ± 4.8	-13.0 ± 0.8	60.0	-100.7 ± 1.9	-89.4 ± 1.6	-11.3 ± 0.3	55.4

Table S9. Thermodynamic parameters for RNA duplexes measured in the presence of NaCl and KCl with 40 wt% PEG200

^aRNA duplex consists of denoted RNA strand (5' \rightarrow 3') and complementary RNA strand. ^bExperiments were performed in a buffer containing 100 mM NaCl, 10 mM Na₂HPO₄, and 1 mM Na₂EDTA in 40 wt% PEG200 at pH 7.0. Values were collected from Table S2. ^cExperiments were performed in a buffer containing 100 mM KCl, 10 mM K₂HPO₄, and 1 mM K₂EDTA in 40 wt% PEG200 at pH 7.0. ^dMelting temperatures were calculated for total strand concentration of 100 μ M.

Table S10. Dielectric constants (ε_r) of the NaCl and KCl solutions in the presence and absence of cosolutes at 37 °C

Solutions	ε _r					
In 100 mM NaCl with 10 mM Na ₂ HPO ₄ , and 1 mM Na ₂ EDTA						
absence of cosolute	76.8 ± 0.6					
40 wt% PEG200	39.4 ± 0.4					
40 wt% EG	62.0 ± 0.5					
In 100 mM KCl with 10 mM $K_2 HPO_4$, and 1 mM $K_2 EDTA$						
absence of cosolute	74.5 ± 0.6					
40 wt% PEG200	42.5 ± 0.4					
40 wt% EG	63.2 ± 0.5					

			NaCl ^b				KClc		
No.	Sequence ^a	∆ <i>H</i> ° (kcal mol ⁻¹)	TΔS° (cal mol ⁻¹ K ⁻¹)	ΔG°_{37} (kcal mol ⁻¹)	T _m ^d (°C)	∆ <i>H</i> ° (kcal mol ⁻¹)	TΔS° (cal mol ⁻¹ K ⁻¹)	ΔG°_{37} (kcal mol ⁻¹)	T _m ^d (°C)
NS1	CAUGCC	-62.0 ± 3.2	-55.0 ± 2.9	-7.0 ± 0.5	39.2	-57.6 ± 5.4	-50.7 ± 4.8	-6.9 ± 0.8	38.5
NS2	CUAGGC	-54.7 ± 5.5	-47.1 ± 4.9	-7.6 ± 0.9	42.5	-69.4 ± 7.3	-62.0 ± 6.6	-7.4 ± 1.0	40.7
NS5	GAACUCC	-67.3 ± 3.5	-59.4 ± 3.2	-7.9 ± 0.5	43.1	-71.1 ± 4.1	-63.5 ± 3.7	-7.6 ± 0.6	41.6
NS11a	UGCUGUCA	-71.8 ± 1.7	-61.6 ± 1.5	-10.2 ± 0.3	54.1	-69.4 ± 1.8	-59.5 ± 1.5	-9.9 ± 0.3	53.1
S6	CGUAUACG	-74.2 ± 0.4	-67.0 ± 0.3	-7.2 ± 0.1	43.3	-77.3 ± 3.0	-70.4 ± 2.8	-6.9 ± 0.4	41.6
S8	CGGUACCG	-85.3 ± 1.9	-73.8 ± 1.6	-11.5 ± 0.3	59.9	-82.0 ± 2.6	-70.6 ± 2.3	-11.4 ± 0.4	60.0
S12a	AUGAGCUCAU	-83.2 ± 8.0	-71.6 ± 6.9	-11.6 ± 1.3	60.9	-77.0 ± 3.8	-66.0 ± 3.3	-11.0 ± 0.7	59.8
S13	UUACGCGUAA	-85.2 ± 3.1	-75.1 ± 2.8	-10.1 ± 0.5	53.6	-84.8 ± 3.7	-75.1 ± 3.3	-9.7 ± 0.6	52.1

Table S11. Thermodynamic parameters for RNA duplexes measured in the presence of NaCl and KCl without any cosolute

^aRNA duplex consists of denoted RNA strand (5' \rightarrow 3') and complementary RNA strand. ^bExperiments were performed in a buffer containing 100 mM NaCl, 10 mM Na₂HPO₄, and 1 mM Na₂EDTA at pH 7.0. ^cExperiments were performed in a buffer containing 100 mM KCl, 10 mM K₂HPO₄, and 1 mM K₂EDTA at pH 7.0. ^dMelting temperatures were calculated for total strand concentration of 100 μ M.

			NaCl ^b				KClc		
No.	Sequence ^a	Δ H° (kcal mol⁻¹)	<i>T∆S</i> ° (cal mol ⁻¹ K ⁻¹)	ΔG°_{37} (kcal mol ⁻¹)	<i>T</i> _m ^d (°C)	Δ H° (kcal mol⁻¹)	<i>T</i> ΔS° (cal mol⁻¹K⁻¹)	ΔG°_{37} (kcal mol ⁻¹)	T _m d (°C)
NS2	CUAGGC	-58.4 ± 6.1	-52.3 ± 5.8	-6.1 ± 0.8	33.8	-55.5 ± 3.2	-49.7 ± 2.9	-5.8 ± 0.4	33.2
NS5	GAACUCC	-65.3 ± 1.4	-59.2 ± 1.2	-6.1 ± 0.2	35.2	-65.5 ± 5.6	-59.7 ± 5.2	-5.8 ± 0.7	33.9
S2	GGUAUACC	-69.3 ± 4.7	-63.5 ± 4.3	-5.8 ± 0.7	36.0	-72.0 ± 3.2	-66.4 ± 3.0	-5.6 ± 0.4	36.5
S3	GGUUAACC	-71.4 ± 2.3	-66.1 ± 2.1	-5.3 ± 0.3	35.1	-67.2 ± 2.2	-62.0 ± 2.0	-5.2 ± 0.3	34.7
S5	GCUUAAGC	-68.3 ± 2.7	-62.6 ± 2.5	-5.7 ± 0.4	36.5	-64.1 ± 1.2	-58.8 ± 1.1	-5.3 ± 0.2	35.0
S8	CGGUACCG	-77.2 ± 4.2	-68.3 ± 3.7	-8.9 ± 0.7	50.1	-74.8 ± 0.9	-66.0 ± 0.8	-8.8 ± 0.1	50.2
S12a	AUGAGCUCAU	-80.4 ± 2.0	-71.9 ± 1.8	-8.5 ± 0.3	48.1	-75.1 ± 6.4	-67.1 ± 5.7	-8.0 ± 0.9	46.8
S13	UUACGCGUAA	-88.0 ± 2.8	-81.3 ± 2.6	-6.7 ± 0.4	40.8	-89.4 ± 5.8	-83.0 ± 5.4	-6.4 ± 0.7	39.8

Table S12. Thermodynamic parameters for RNA duplexes measured in the presence of NaCl and KCl with 40 wt% EG

^aRNA duplex consists of denoted RNA strand (5' \rightarrow 3') and complementary RNA strand. ^bExperiments were performed in a buffer containing 100 mM NaCl, 10 mM Na₂HPO₄, and 1 mM Na₂EDTA in 40 wt% EG at pH 7.0. ^cExperiments were performed in a buffer containing 100 mM KCl, 10 mM K₂HPO₄, and 1 mM K₂EDTA in 40 wt% EG at pH 7.0. ^dMelting temperatures were calculated for total strand concentration of 100 μ M.

No.	Sequence ^a	Measured ΔG°_{37} ^b (kcal mol ⁻¹)	Calculated ΔG° ₃₇ ^c (kcal mol⁻¹)
NS3	CAGCGG	-6.1	-5.6
NS5	GAACUCC	-5.4	-5.1
NS10a	CGCUGUCG	-8.8	-8.7
NS11a	UGCUGUCA	-7.0	-7.1
NS13a	GUGCCGAG	-9.7	-9.4
NS13b	GCCGAGUG	-9.8	-9.4
S2	GGUAUACC	-5.5	-5.8
S3	GGUUAACC	-5.0	-5.0
S5	GCUUAAGC	-5.1	-5.3
S7	CGCAUGCG	-8.5	-8.3
S8	CGGUACCG	-8.6	-8.0
S11a	GAUCCGGAUC	-11.4	-10.8
S12a	AUGAGCUCAU	-7.4	-7.7
S12b	AUCAGCUGAU	-7.0	-7.7
S13	UUACGCGUAA	-6.2	-6.4
S14	AUCGCUAGCGAU	-11.3	-11.9

Table S13. Measured and calculated ΔG°_{37} for RNA duplexes under the molecular crowding environment of 40 wt% PEG200 with 100 mM KCl

^aRNA duplex consists of denoted RNA strand (5' \rightarrow 3') and complementary RNA strand. ^bErrors associated with the measured values were mentioned in the Table S9. ^cCalculated using equation 15 and the parameters of Table 1.

Table S14. Measured and predicted ΔG°_{37} of RNA duplexes in different cosolutes and NaCl concentrations

Solution/Sequences ^a	Measured ΔG°_{37} (kcal mol ⁻¹)	Predicted ΔG°_{3} (kcal mol ⁻¹)
GGCUCAAUUGAC in 100 mM NaCl		
10 wt% PEG200	-15.1 ± 0.8	-15.4
20 wt% PEG200	-14.8 ± 0.6	-14.7
30 wt% PEG200	-14.0 ± 0.7	-14.0
40 wt% PEG200	-13.8 ± 0.6	-13.7
In 20 wt% EG at 100 mM NaCl		
GGAUCGAUCC	-12.7 ± 0.7	-13.1
AUCAGCUGAU	-9.9 ± 0.6	-9.9
GGCUCAAUUGAC	-14.4 ± 0.6	-15.0
In 20 wt% 1,3 PDO at 100 mM NaCl		
GAUCCGGAUC	-14.5 ± 0.7	-12.9
GGCUCAAUUGAC	-14.7 ± 0.6	-14.7
In 20 wt% PEG2000 at 100 mM NaCl ^b		
GGCUCAAUUGAC	-16.9 ± 0.8	-16.3
GAUUACGCCUG	-15.5 ± 0.5	-15.4
In 20 wt% PEG8000 at 100 mM NaCl ^c		
GGCUCAAUUGAC	-16.7 ± 0.4	-16.5
GGCUCAAUUGAC in 1 M NaCl		
10 wt% PEG200	-18.9 ± 0.8	-17.9
20 wt% PEG200	-18.3 ± 0.6	-17.2
In 20 vol% PEG200 at 1 M NaCl ^d		
UCAUGA ^e	-3.0 ± 0.1	-3.8
ACUGCG ^e	-6.7 ± 0.0	-7.2
AUGGAC ^e	-5.9 ± 0.1	-6.1
GCGAUA ^e	-5.3 ± 0.2	-5.6
GCUAUG ^e	-5.2 ± 0.2	-5.2
ACCGGU ^e	-7.1 ± 0.1	-6.0
AGCGCU ^e	-6.8 ± 0.1	-6.4
CACGUG ^e	-5.8 ± 0.0	-5.7
CAGCUG ^e	-5.9 ± 0.0	-6.7
CCAUGG ^e	-6.5 ± 0.0	-6.7
CCUAGG ^e	-6.9 ± 0.0	-6.4
CUGCAG ^e	-6.2 ± 0.1	-6.7
GACGUC ^e	-6.6 ± 0.0	-5.8
GAGCUC ^e	-6.9 ± 0.1	-6.8
GCAUGC ^e	-6.9 ± 0.1	-7.0

CGCGCG ^e	-7.7 ± 0.2	-8.3
CGGCCG ^e	-8.5 ± 0.1	-8.9
GCCGGC ^e	-10.2 ± 0.5	-9.9
GCGCGC ^e	-9.0 ± 0.4	-9.2
UAUAUA ^e	-2.3 ± 0.2	-1.9
UAAUAUUA ^e	-1.9 ± 0.4	-1.2
AACUAGUU ^e	-6.0 ± 0.0	-4.5
ACUAUAGU ^e	-5.8 ± 0.0	-5.2
AGAUAUCU ^e	-5.7 ± 0.1	-5.6
GAUAUAUC ^e	-5.5 ± 0.0	-5.3
GAAUAUUC ^e	-4.7 ± 0.1	-4.6
AACCGGUU ^e	-9.0 ± 0.3	-7.6
ACUGCAGU ^e	-9.2 ± 0.1	-9.3
GCAAUUGC ^e	-8.1 ± 0.1	-8.6
GAACGUUC ^e	-7.8 ± 0.1	-7.4
AGCGCGCU ^e	-12.5 ± 0.5	-11.9
AGCCGGCU ^e	-13.9 ± 0.2	-12.5
GCGAUCGC ^e	-12.4 ± 0.4	-11.7
GACCGGUC ^e	-12.9 ± 0.2	-12.0
UUAUCGAUAA ^e	-6.9 ± 0.0	-6.8
UAUCGAUA ^e	-5.8 ± 0.0	-5.2
AGCGCU ^e	-6.6 ± 0.1	-6.4
CGCGCG ^e	-8.6 ± 0.2	-8.3

^aRNA duplex consists of denoted RNA strand (5' \rightarrow 3') and complementary RNA strand. ^{b,c}Water activities were measured to be 0.987 and 0.990 for PEG2000 and PEG8000, respectively, at their 20 wt% solution in 10 mM Na-phosphate buffer with 100 mM NaCl and 0.1 M Na₂EDTA. ^d20 (v/v)% PEG200 is equivalent to 21.7 (w/w)% PEG200 at 37 °C. Since the contribution of water activity and excluded volume effect will be negligibly different between 21.7 wt% and 20 wt%, we calculated ΔG°_{37} in 20 vol% PEG200 using the parameters obtained for 20 wt% PEG200. ^eMeasured values were collected from the report by Adams and Znosko.(7)

			122 mM N	la⁺ ^b		Int	racellular catio	on condition ^c	
No.	Sequence ^a	∆ <i>H</i> ° (kcal mol⁻¹)	$T\Delta S^{\circ}$ (cal mol ⁻¹ K ⁻¹)	ΔG°_{37} (kcal mol ⁻¹)	T _m ^d (°C)	∆ <i>H</i> ° (kcal mol⁻¹)	<i>TΔS</i> ° (cal mol⁻¹K⁻¹)	ΔG° ₃₇ (kcal mol ⁻¹)	<i>T</i> _m ^d (°C)
NS3	CAGCGG	-65.6 ± 3.0	-59.2 ± 2.7	-6.4 ± 0.4	36.4	-56.5 ± 1.4	-50.2 ± 1.3	-6.3 ± 0.2	36.0
NS5	GAACUCC	-71.8 ± 2.3	-65.9 ± 2.1	-5.9 ± 0.3	34.1	-76.9 ± 2.8	-70.9 ± 2.6	-6.0 ± 0.4	35.1
NS10a	CGCUGUCG	-78.4 ± 2.4	-68.6 ± 2.1	-9.8 ± 1.0	49.8	-74.7 ± 1.8	-65.4 ± 1.5	-9.3 ± 0.3	48.7
NS11a	UGCUGUCA	-66.2 ± 1.8	-58.4 ± 1.6	-7.8 ± 0.7	43.0	-61.0 ± 2.3	-53.6 ± 2.0	-7.4 ± 0.4	41.6
NS13a	GUGCCGAG	-81.3 ± 1.3	-70.5 ± 1.1	-10.8 ± 0.5	54.3	-73.1 ± 4.6	-62.9 ± 4.0	-10.2 ± 0.8	53.0
NS13b	GCCGAGUG	-71.6 ± 2.1	-61.2 ± 1.8	-10.4 ± 0.8	54.1	-74.1 ± 3.2	-63.8 ± 2.7	-10.3 ± 0.5	53.6
S2	GGUAUACC	-71.9 ± 3.9	-65.7 ± 3.6	-6.2 ± 0.5	39.2	-73.4 ± 1.4	-67.5 ± 1.3	-5.9 ± 0.2	37.9
S3	GGUUAACC	-73.4 ± 2.8	-67.7 ± 2.6	-5.7 ± 0.4	36.5	-68.5 ± 2.9	-63.1 ± 2.5	-5.4 ± 0.4	35.4
S5	GCUUAAGC	-70.4 ± 3.2	-64.3 ± 2.9	-6.1 ± 0.4	38.6	-74.0 ± 5.1	-68.7 ± 4.8	-5.3 ± 0.7	35.4
S7	CGCAUGCG	-79.3 ± 3.7	-69.7 ± 3.2	-9.6 ± 0.6	52.7	-78.1 ± 5.2	-69.3 ± 4.6	-8.8 ± 0.9	50.3
S8	CGGUACCG	-73.4 ± 6.2	-64.4 ± 5.5	-9.0 ± 1.0	52.7	-75.0 ± 1.7	-65.9 ± 1.5	-9.1 ± 0.3	52.0
S11a	GAUCCGGAUC	-103.9 ± 1.7	-91.6 ± 1.5	-12.3 ± 0.7	58.4	-94.8 ± 3.3	-83.3 ± 2.9	-11.5 ± 0.5	57.0
S12a	AUGAGCUCAU	-80.7 ± 2.3	-72.1 ± 2.0	-8.6 ± 0.3	48.8	-80.9 ± 5.8	-72.8 ± 5.2	-8.1 ± 0.7	46.8
S12b	AUCAGCUGAU	-83.3 ± 1.3	-75.1 ± 1.1	-8.2 ± 0.5	46.7	-81.9 ± 1.9	-74.2 ± 1.8	-7.7 ± 0.3	44.7
S13	UUACGCGUAA	-78.9 ± 4.7	-71.7 ± 4.2	-7.2 ± 0.6	42.2	-75.7 ± 2.2	-68.8 ± 2.0	-6.9 ± 0.3	41.7
S14	AUCGCUAGCGAU	-105.3 ± 4.7	-92.3 ± 4.8	-13.0 ± 0.8	60.0	-95.0 ± 8.3	-83.4 ± 7.3	-11.6 ± 1.5	57.1

Table S15. Thermodynamic parameters for RNA duplexes measured in the presence of NaCl and exact intracellular cation condition with 40 wt% PEG200

^aRNA duplex consists of denoted RNA strand (5' \rightarrow 3') and complementary RNA strand. ^bExperiments were performed in a buffer containing 100 mM NaCl, 10 mM Na₂HPO₄, and 1 mM Na₂EDTA in 40 wt% PEG200 at pH 7.0. Values were collected from Table S2. ^cExperiments were performed in a buffer containing 120 mM KCl, 10 mM NaCl, 0.5 mM MgCl₂, 0.0001 mM CaCl₂ and 10 mM K₂HPO₄ in 40 wt% PEG200 at pH 7.2. ^dMelting temperatures were calculated for total strand concentration of 100 μ M.

			Measure	d ^b			Predicted	c	
No.	Sequence ^a	∆ <i>H</i> ° (kcal mol⁻¹)	∆ <i>S</i> ° (cal mol ⁻¹ K ⁻¹)	∆ <i>G</i> ° ₃₇ (kcal mol⁻¹)	<i>T</i> ^d (°C)	∆ <i>H</i> ° (kcal mol ⁻¹)	∆ <i>S</i> ° (cal mol⁻¹K⁻¹)	∆G° ₃₇ (kcal mol⁻¹)	<i>T</i> _m ^d (°C)
NS3	CAGCGG	-56.5	-161.9	-6.3	36.0	-58.4	-167.2	-6.5	37.1
NS5	GAACUCC	-76.9	-228.6	-6.0	35.1	-65.5	-191.8	-6.0	34.6
NS10a	CGCUGUCG	-74.7	-210.9	-9.3	48.7	-77.1	-216.8	-9.8	51.0
NS11a	UGCUGUCA	-61.0	-172.8	-7.4	41.6	-66.1	-187.0	-8.1	44.6
NS13a	GUGCCGAG	-73.1	-202.8	-10.2	53.0	-80.8	-226.4	-10.6	53.4
NS13b	GCCGAGUG	-74.1	-205.7	-10.3	53.6	-80.8	-226.4	-10.6	53.4
S2	GGUAUACC	-73.4	-217.6	-5.9	37.9	-78.7	-232.3	-6.7	40.9
S3	GGUUAACC	-68.5	-203.4	-5.4	35.4	-77.5	-230.8	-5.9	38.0
S5	GCUUAAGC	-74.0	-221.5	-5.3	35.4	-76.5	-226.6	-6.2	39.2
S7	CGCAUGCG	-78.1	-233.4	-8.8	50.3	-79.5	-226.1	-9.4	52.1
S8	CGGUACCG	-75.0	-212.5	-9.1	52.0	-79.7	-227.6	-9.1	51.0
S11a	GAUCCGGAUC	-94.8	-268.6	-11.5	57.0	-103.1	-293.5	-12.1	57.5
S12a	AUGAGCUCAU	-80.9	-234.7	-8.1	46.8	-86.4	-250.1	-8.8	48.8
S12b	AUCAGCUGAU	-81.9	-239.2	-7.7	44.7	-86.4	-250.1	-8.8	48.8
S13	UUACGCGUAA	-75.7	-221.8	-6.9	41.7	-82.0	-241.3	-7.2	42.7
S14	AUCGCUAGCGAU	-95.0	-268.9	-11.6	57.1	-109.3	-309.6	-13.3	60.2

Table S16. Measured thermodynamic parameters for RNA duplexes in exact intracellular cation condition and the predicted values using parameters in 40 wt% PEG200 with 100 mM NaCl

^aRNA duplex consists of denoted RNA strand (5' \rightarrow 3') and complementary RNA strand. ^bErrors associated with the measured values were mentioned in the Table S15. ^cPredicted using the parameters of Table 1. ^dMelting temperatures were calculated for total strand concentration of 100 μ M.

Sequence	$\Delta G^{\circ}_{25, \text{ NN, [no cosolute, 122 mM Na}^+]^a}$	ΔG° 25, NN, ev [40 wt% PEG200] ^b	$\Delta G^{\circ}_{25, \text{ NN, Wa}}$ [40 wt% PEG200] ^c	$m_{\rm PEG}{}^{\rm d}$
	(kcal mol ⁻¹)	(kcal mol ⁻¹)	(kcal mol ⁻¹)	(kcal mol ⁻¹)
r(AA/UU)	-1.10	-0.21	0.37	7.6
r(AU/UA)	-0.81	-0.21	0.09	1.9
r(UA/AU)	-1.70	-0.21	0.19	3.9
r(CA/GU)	-2.16	-0.21	-0.16	-3.3
r(GU/CA)	-2.50	-0.21	0.56	11.4
r(CU/GA)	-2.12	-0.21	0.19	3.9
r(GA/CU)	-2.70	-0.21	0.30	6.1
r(CG/GC)	-2.53	-0.21	0.22	4.5
r(GC/CG)	-3.78	-0.21	0.50	10.1
r(GG/CC)	-3.48	-0.21	0.34	6.9
initiation	4.05	-0.21	1.63	33.2
per terminal AU	0.57	NA	0.51	10.4

Table S17. Parameters for $\Delta G^{\circ}_{NN, [no cosolute, 122 mM Na^+]}$, $\Delta G^{\circ}_{NN, ev}$, $\Delta G^{\circ}_{NN, wa}$ and prefactor of PEG (m_{PEG}) at 25 °C

^aCalculated from the ΔH° and ΔS° values in Table S3. ^b $\Delta G^{\circ}_{25, NN, ev}$ [40 wt% PEG200] were calculated similarly as done in Table 2. ^c $\Delta G^{\circ}_{25, NN, ev}$ [40 wt% PEG200] = $\Delta G^{\circ}_{25, NN, [40 wt% PEG200, 122 mM Na^{+}]} - \Delta G^{\circ}_{25, NN, [no cosolute, 122 mM Na^{+}]} - \Delta G^{\circ}_{25, NN, ev}$ [40 wt% PEG200]. $\Delta G^{\circ}_{25, NN, [40 wt% PEG200, 122 mM Na^{+}]} - \Delta G^{\circ}_{25, NN, ev}$ [40 wt% PEG200]. $\Delta G^{\circ}_{25, NN, [40 wt% PEG200, 122 mM Na^{+}]}$ were calculated from the ΔH° and ΔS° values in Table 1. ^dDetermined by using equation 13.

Solution		(ACUG)₃			(ACUG)₀	
	ΔG° _{25, wa} a (kcal mol ⁻¹)	ΔG° _{25, ev} b (kcal mol ⁻¹)	ΔG° _{25, [crowder]} c (kcal mol ⁻¹)	ΔG° _{25, wa} a (kcal mol ⁻¹)	ΔG° _{25, ev} b (kcal mol ⁻¹)	$\Delta G^{\circ}_{25, [crowder]}^{c}$ (kcal mol ⁻¹)
10 wt% PEG200	1.0	-0.6	0.4	1.6	-1.0	0.6
20 wt% PEG200	2.3	-1.2	1.1	3.7	-2.0	1.7
30 wt% PEG200	3.7	-1.7	2.0	5.8	-3.0	2.8
40 wt% PEG200	4.5	-2.3	2.2	7.2	-4.0	3.2
50 wt% PEG200	6.0	-2.9	3.1	9.5	-5.1	4.4

Table S18. Calculated $\Delta G^{\circ}_{25, [crowder]}$ for (ACUG)₃ and (ACUG)₆ in various PEG200 concentrations at 25 °C in 100 mM NaCl

 $^{a}\Delta G^{\circ}_{25, wa}$ were calculated from the m_{PEG} values in Table S17 and Δa_{w} values using equation 13 at 298 K. $^{b}\Delta G^{\circ}_{25, ev}$ were calculated using equation 11 at 298 K. $^{c}\Delta G^{\circ}_{25, [crowder]} = \Delta G^{\circ}_{25, wa} + \Delta G^{\circ}_{25, ev}$.

Sequence ^a	$\Delta G^{\circ}_{37 \text{ NN}, [no, cosolute, 122 mM Na^+]}$	<i>т</i> _{РЕБ/2-МЕ/1,2 DME}	m _{EG/Gly/1,3 PDO}
	(kcal mol ⁻¹)	(kcal mol ⁻¹)	(kcal mol ⁻¹)
r(GA/UU)	-0.96	7.1	2.9
r(GG/UU)	-0.38	7.1	2.9
r(AG/UU)	-0.41	7.1	2.9
r(AU/UG)	-0.64	3.9	1.6
r(UG/AU)	-0.98	3.9	1.6

Table S19. Estimated NN parameters including G-U wobble base pairs present in Im-4U

 ${}^{a}\Delta G^{\circ}{}_{37}$ in 1 M NaCl solution were collected from collected from the report by Turner et al. (1)

Sequence ^a	In the absence of PEG200 ^b	In the presence of 40 wt% PEG200	
	(kcal mol ⁻¹)	(kcal mol ⁻¹)	
5'GGAG <u>GAAA</u> CUCC 3'	1.47 ± 0.06	0.63 ± 0.09	
5'GAAG <u>GAAA</u> CUUC 3'	1.36 ± 0.06	0.57 ± 0.09	

Table S20. ΔG°_{37} of GAAA tetraloop in the absence and presence of 40 wt% PEG200

^aUnderlined region is the loop sequence. ^bValues were collected from the report by Leonard and Blose.(3)

Solution	Δa_{w}	$\Delta G^{\circ}_{37^{a}}$
		(kcal mol ⁻¹)
10 wt% EG	0.026	-0.5
20 wt% EG	0.047	-1.3
30 wt% EG	0.072	-2.2
40 wt% EG	0.087	-2.8
10 wt% Gly	0.021	-0.2
20 wt% Gly	0.045	-1.0
30 wt% Gly	0.076	-2.0
40 wt% Gly	0.101	-2.8
10 wt% 1,3 PDO	0.025	-0.4
20 wt% 1,3 PDO	0.049	-1.2
30 wt% 1,3 PDO	0.066	-1.9
40 wt% 1,3 PDO	0.089	-2.7

Table S21. Predicted ΔG°_{37} of RNA hairpin Im-4U in different cosolute concentrations at 100 mM NaCl along with the Δa_w values in those cosolute solutions

 ${}^{a}\Delta G^{\circ}_{37} = \Delta G^{\circ}_{37, \text{ no cosolute, 100 mM NaCl}} + \Delta G^{\circ}_{37, \text{ wa}} + \Delta G^{\circ}_{37, \text{ ev}}$, as shown in Figure S6. $\Delta G^{\circ}_{37, \text{ no cosolute, 100 mM NaCl}}$ were calculated from the data in Table 2 and Table S19. $\Delta G^{\circ}_{37, \text{ wa}}$ were determined from the m_{cs} values in Table S19 and Δa_{w} values in this Table and equation 13. $\Delta G^{\circ}_{37, \text{ loop, [1M NaCl]}}$ values were collected from the report of Turner et al.(1) $\Delta G^{\circ}_{37, \text{ ev}}$ was determined from the data in Table S6 and equation 8. For hairpin, $\Delta V = V_{\text{hairpin}} - V_{\text{single strand}}$.



Figure S1. Representative CD spectra of 20 μ M RNA oligonucleotides at 4 °C in a buffer containing 0.1 M NaCl, 10 mM Na₂HPO₄ (pH 7.0) and 1 mM Na₂EDTA in the absence (black) and presence (red) of 40 wt% PEG200.



Figure S2. Renaturation and denaturation curves for some representative RNA oligonucleotides at 100 μ M concentration in a buffer containing 0.1 M NaCl, 10 mM Na₂HPO₄ (pH 7.0), 1 mM Na₂EDTA and 40 wt% PEG200.





Figure S3. Plot of individual $\Delta G^{\circ}_{37, NN}$ against the concentration of Na⁺. Values are collected from the report by Weber et al.(4)



Figure S4. UV melting profiles of 10 μ M RNA duplexes with the varying concentrations of NaCl. Samples were dialyzed overnight in Milli-Q water before setting the cation concentrations.



Figure S5. Relationship between measured ΔG°_{37} of RNA duplexes in **(A)** 122 mM K⁺ solution or **(B)** intracellular cation concentration and predicted ΔG°_{37} in 122 mM Na⁺ solution under the crowding condition of 40 wt% PEG200. Measured values are provided in Table S9 and Table S15, respectively and predicted values collected from Table S5.

5' GUUGA ^A CUUUUAAAU ^A G 11111 GAGGAUUUA G lm-4U $\Delta G^{\circ}_{37, \text{ hairpin}} = 3\Delta G^{\circ}_{37} (AA/UU) + \Delta G^{\circ}_{37} (CU/GA) + \Delta G^{\circ}_{37} (AU/UA) + \Delta G^{\circ}_{37} (UA/AU) + \Delta G^{\circ}_{37} ($ $2\Delta G_{37}^{\circ}(GA/UU) + \Delta G_{37}^{\circ}(GG/UU) + \Delta G_{37}^{\circ}(AG/UU) + \Delta G_{37}^{\circ}(AU/UG) +$ $\Delta G^{\circ}_{37} (\text{UG/AU}) + \Delta G^{\circ}_{37, \text{ initiation}} + \Delta G^{\circ}_{37, \text{ terminal GU}} + \Delta G^{\circ}_{37, \text{ hairpin loop}} + \Delta G^{\circ}_{37, \text{ internal loop}}$ ΔG°_{37} (pred.) = $\Delta G^{\circ}_{37, \text{ hairpin, no cosolute}} + \Delta G^{\circ}_{37, \text{ hairpin, wa}} + \Delta G^{\circ}_{37, \text{ hairpin, ev}}$ $\Delta G^{\circ}_{37} (AA/UU) = \{ -0.70 + (7.1 \times 0.049) \}$ ΔG°_{37} (CU/GA) = { -1.80 + (5.1 x 0.049)} $\Delta G^{\circ}_{37}(UA/AU) = \{ -1.30 + (3.9 \times 0.049) \}$ $\Delta G^{\circ}_{37} (AU/UA) = \{ -0.52 + (3.9 \times 0.049) \}$ $\Delta G^{\circ}_{37} (GA/UU) = \{ -0.96 + (7.1 \times 0.049) \}$ $\Delta G^{\circ}_{37} (GG/UU) = \{ -0.38 + (7.1 \times 0.049) \}$ $\Delta G^{\circ}_{37} (AG/UU) = \{ -0.41 + (7.1 \times 0.049) \}$ $\Delta G^{\circ}_{37} (AU/UG) = \{ -0.64 + (3.9 \times 0.049) \}$ $\Delta G^{\circ}_{37, \text{initiation}} = \{ 4.09 + (33.3 \times 0.049) \}$ $\Delta G^{\circ}_{37} (\text{UG/AU}) = \{ -0.98 + (3.9 \times 0.049) \}$ $\Delta G^{\circ}_{37, \text{ hairpin loop}} = \{4.95 - (11.8 \times 5.25 \times 0.049)\}$ $\Delta G^{\circ}_{37, \text{ terminal GU}} = \{ 0.45 + (8.2 \times 0.049) \}$ $\Delta G^{\circ}_{37, \text{ internal loop}} = \{1.05 - (11.8 \times 1.05 \times 0.049)\}$ $\Delta G^{\circ}_{37 \text{ hairpin ev}} = -1.42$

 ΔG°_{37} (pred.) = 1.1 kcal mol⁻¹ ΔG°_{37} (exp.) = 0.4 kcal mol⁻¹

Figure S6. Secondary structure of RNA hairpin Im-4U and prediction of its stability in 40 wt% PEG200 at 100 mM NaCl.

References

- 1. Mathews, D.H., Sabina, J., Zuker, M. and Turner, D.H. (1999) Expanded sequence dependence of thermodynamic parameters improves prediction of RNA secondary structure11Edited by I. Tinoco. J. Mol. Biol., **288**, 911-940.
- 2. Tan, Z.-J. and Chen, S.-J. (2008) Salt Dependence of Nucleic Acid Hairpin Stability. *Biophys. J.*, **95**, 738-752.
- 3. Leonard, K.N. and Blose, J.M. (2018) Effects of osmolytes and macromolecular crowders on stable GAAA tetraloops and their preference for a CG closing base pair. *PeerJ*, **6**, e4236-e4236.
- 4. Ferreira, I., Jolley, E.A., Znosko, B.M. and Weber, G. (2019) Replacing salt correction factors with optimized RNA nearest-neighbour enthalpy and entropy parameters. *Chem. Phys.*, **521**, 69-76.
- 5. Knowles, D.B., LaCroix, A.S., Deines, N.F., Shkel, I. and Record, M.T. (2011) Separation of preferential interaction and excluded volume effects on DNA duplex and hairpin stability. *Proc. Natl. Acad. Sci. U. S. A.*, **108**, 12699-12704.
- 6. Liu, J.-H., Xi, K., Zhang, X., Bao, L., Zhang, X. and Tan, Z.-J. (2019) Structural Flexibility of DNA-RNA Hybrid Duplex: Stretching and Twist-Stretch Coupling. *Biophys. J.*, **117**, 74-86.
- 7. Adams, M.S. and Znosko, B.M. (2019) Thermodynamic characterization and nearest neighbor parameters for RNA duplexes under molecular crowding conditions. *Nucleic Acids Res.*, **47**, 3658-3666.