

Supplementary Information

Solid acid-catalyzed one-step synthesis of oleacein from oleuropein

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1. General Information

1.1 Materials

All reagents were of research grade and used without further purification. TLC was performed on silica gel (60 F-254, 0.25 mm Plates). Column chromatography was carried out on Silica gel 60N (spherical, neutral, particle size 100-210 μm , Kanto Chemical Co., Inc.)

Oleuropein was purchased from Toronto Research Chemicals. Na-montmorillonite (Kunipia F) was purchased from Kunimine Industry in Japan. Sulfuric acid/Zirconium, Zirconium dioxide, Silica Alumina, γ -Aluminum oxide was purchased from FUJIFILM Wako Pure Chemical industry. Y-exchanged Zeolite (HSZ-320HOA) was purchased from TOSO industry. Amberlyst® was purchased from Organo corporation. Silica gel and dimethyl sulfoxide- d_6 were purchased from Sigma-Aldrich. Hydrochloric acid, *p*-toluenesulfonic acid and dimethyl sulfoxide were purchased from Kishida Chemical. Olive leaves was purchased from Hinata Food/SINSEI KOSAN company.

Olive leaves were from Shodoshima, Japan, and were purchased commercial from SHINSEI Co. Ltd. All local, national or international guidelines and legislation were adhered to in this study.

1.2 NMR spectroscopy

^1H NMR spectra were recorded in DMSO- d_6 and CDCl_3 on JEOL LA-400 spectrometer. Chemical shifts are expressed in ppm relative to tetramethylsilane (0 ppm) or CHCl_3 (7.28 ppm). The coupling constants are given in Hz. ^{13}C NMR spectra were recorded on the same spectrometers at 100 MHz, using the central resonance of CDCl_3 (δ_{C} 77.0 ppm) as the internal reference unless otherwise stated.

1.3 Mass spectroscopy

Low-resolution mass spectra (LRMS) were obtained on a Waters ZQ-2000 (ESI). The needle and cone voltage were +4.0 kV and 50 V, respectively. The sample solution was directly introduced into the apparatus at a flow rate of 20 $\mu\text{L}/\text{min}$.

1.4 Other analyses

Optical rotations were determined with a JASCO DIP-1000. The amount of water in DMSO was measured by Karl Fischer titration (Metrohm, 899 coulometer). The purity of test compounds was determined by analytical HPLC. For analytical HPLC, unless otherwise noted, a Discovery® HS C18 HPLC column (250 \times 4.6 mm I.D, 5 μm , Sigma-Aldrich.) was employed with a linear gradient of water: acetonitrile with gradient from 100:0 to 0:100 in 40 minutes at a flow rate of 1.0 mL/min on a Shimadzu Prominence system (UV, 254 nm).^{1,2}

2. Catalyst preparation and synthesis

2.1 Preparation of proton-exchanged montmorillonite

Na⁺-montmorillonite was protonated with hydrochloric acid in accordance with the procedure reported by Kaneda et al.³ A mixture of the Na⁺-montmorillonite (9.0 g) and 600 mL of HCl (0.22 wt%) was stirred at 90 °C for 24 hours. The obtained slurry was filtered and washed with deionized water and dried at 110 °C in air, followed by breaking the solid in a mortar to afford proton-exchanged montmorillonite as a gray powder (7.6 g). We also prepared the proton-exchanged montmorillonite catalysts with changing the concentration of hydrochloric acid (1.1, 0.55, 0.11, 0.055 wt%) and used them to investigate the effects of acid amounts in the catalyst.

2.2 Synthesis of oleacein and oleocanthal

2.2.1 Synthetic procedure of oleacein with hydrochloric acid and *p*-toluenesulfonic acid

Oleuropein (10 mg, purity >75%, 0.0138 mmol) was dissolved in 0.5 mL of DMSO-d₆ in an NMR tube. Its H₂O content was determined to be 1.36 mg (0.076 mmol) by Karl Fischer titration. Subsequently, tetramethyl benzene (0.5 mg, 3.7 μmol) was added to the tube as internal standard, followed by adding hydrochloric acid (10~0.1 mol%, 10 μL) or *p*-toluenesulfonic acid (10~0.1 mol%). This mixture was treated at 150 °C for 12 hours in an oil bath without stirring. After the reaction, NMR spectrum of this mixture was measured to calculate the NMR yield of oleacein relative to oleuropein.

2.2.2 Synthetic procedure of oleacein with solid acid catalysts

Oleuropein (10 mg, purity >75%, 0.0138 mmol) was dissolved in 0.5 mL of DMSO-d₆, to which tetramethyl benzene (0.5 mg, 3.7 μmol) was added as an internal standard in an NMR tube. Subsequently, a certain amount of solid acid catalysts was added and the NMR tube was left to stand in oil bath at 150 °C for 12 hours without stirring. After the reaction, NMR spectrum of this mixture was measured to calculate the NMR yield of oleacein relative to oleuropein.

For the catalyst recycle experiments, the used solid catalyst was washed with methanol (5 mL × 3) and acetone (5 mL × 3), followed by drying at 110 °C and was used for the next run without further treatment. If the calcination was required, the catalyst was treated at 600 °C for 6 hours in the electric furnace before using for the next run.

The time course measurement was carried out by removing the reaction tube from oil bath at each time, followed by NMR measurement to calculate the yield of oleacein.

The effects of H₂O were evaluated by the H₂O content in DMSO-d₆ being defined. After the remained H₂O in DMSO-d₆ was removed with molecular sieve, a certain amount of H₂O was added to adjust the content by checking the H₂O content with Karl Fischer titration.

To investigate the effects of solvents, oleuropein (10 mg, purity >75%, 0.0138 mmol) was dissolved in each solvent in an NMR tube. The H₂O content of each solvent was measured by Karl Fischer

titration and a certain amount of H₂O was added to adjust the H₂O content to the same amount in DMSO (1.36 mg, 0.076 mmol). Subsequently, H-mont (20 mg) was added and the NMR tube was left to stand in oil bath at 150 °C without stirring. After 12 hours, the reaction solution was filtered to remove the catalyst. The organic layer was washed with water and extracted with AcOEt, dried over Na₂SO₄, filtered and concentrated. The residue was purified by silica gel chromatograph (hexane/AcOEt = 10:1 to 1:1) to get oleacein.

2.2.3 Synthetic procedure of oleacein using isolated oleuropein from olive leaves (Eq. S1)

Olive leaves (10 g) were soaked in methanol and water (40 mL, 4:1 v/v) for 12 hours at room temperature in accordance with the previous report.⁴ The green exuded solution was filtered and evaporated. The residue was roughly purified by silica gel column chromatography (CH₂Cl₂/MeOH = 10:1) to give (1.53 g, purity 88%, determined at 254 nm wavelength of HPLC) as a green powder.

The powder (1.53 g, containing 2.49 mmol of oleuropein, calculated from 88% purity) was dissolved in DMSO (10.0 mL, containing 7.06 mmol of H₂O, measured by Karl Fischer titration) in a 50 mL round-bottom flask. H-mont (3.06 g) was introduced in the mixture and the flask was left to stand in oil bath at 150 °C. After 3 hours, the complete consumption of oleuropein was confirmed by TLC and the flask was cooled to room temperature. The reaction solution was filtered to remove the catalyst. The organic layer was washed with water and extracted with AcOEt, dried over Na₂SO₄, filtered and concentrated. The residue was purified by silica gel chromatography (hexane/AcOEt = 10:1 to 1:1) to obtain oleacein (598mg, 75%).



Figure S1. Oleacein synthesis from olive leaves.

(-)-Oleacein; (3*S*,4*E*)-4-Formyl-3-(2-oxoethyl)-4-hexenoic acid 2-(3,4-dihydroxyphenyl) ethyl ester
 White solid. $[\alpha]_D^{26}$ -0.78 (c 0.32, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ = 9.65 (s, 1H), 9.19 (d, *J* = 2.0 Hz, 1H), 6.78 (d, *J* = 8 Hz, 1H), 6.71 (d, *J* = 2.0 Hz, 1H), 6.65 (q, *J* = 7.2 Hz, 1H), 6.60 (dd, *J* = 2.0, 8.0 Hz, 1H), 4.26-4.12 (m, 2H), 3.67-3.59 (m, 1H), 2.94 (dd, *J* = 18.0, 6.0 Hz, 1H), 2.83-2.80 (m, 3H), 2.78-2.71 (m, 1H), 2.63 (dd, *J* = 16.0, 6.4 Hz, 1H), 2.06 (d, *J* = 7.6 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ = 200.9, 195.7, 171.8, 155.2, 143.3, 143.1, 142.7, 130.4, 121.3, 116.3, 115.2, 65.2, 46.3, 37.0, 34.2, 27.2, 15.3; LRMS (ESI) calcd for C₁₇H₂₀NaO₆ [M+Na]⁺: 343.11. Found: 343.1.

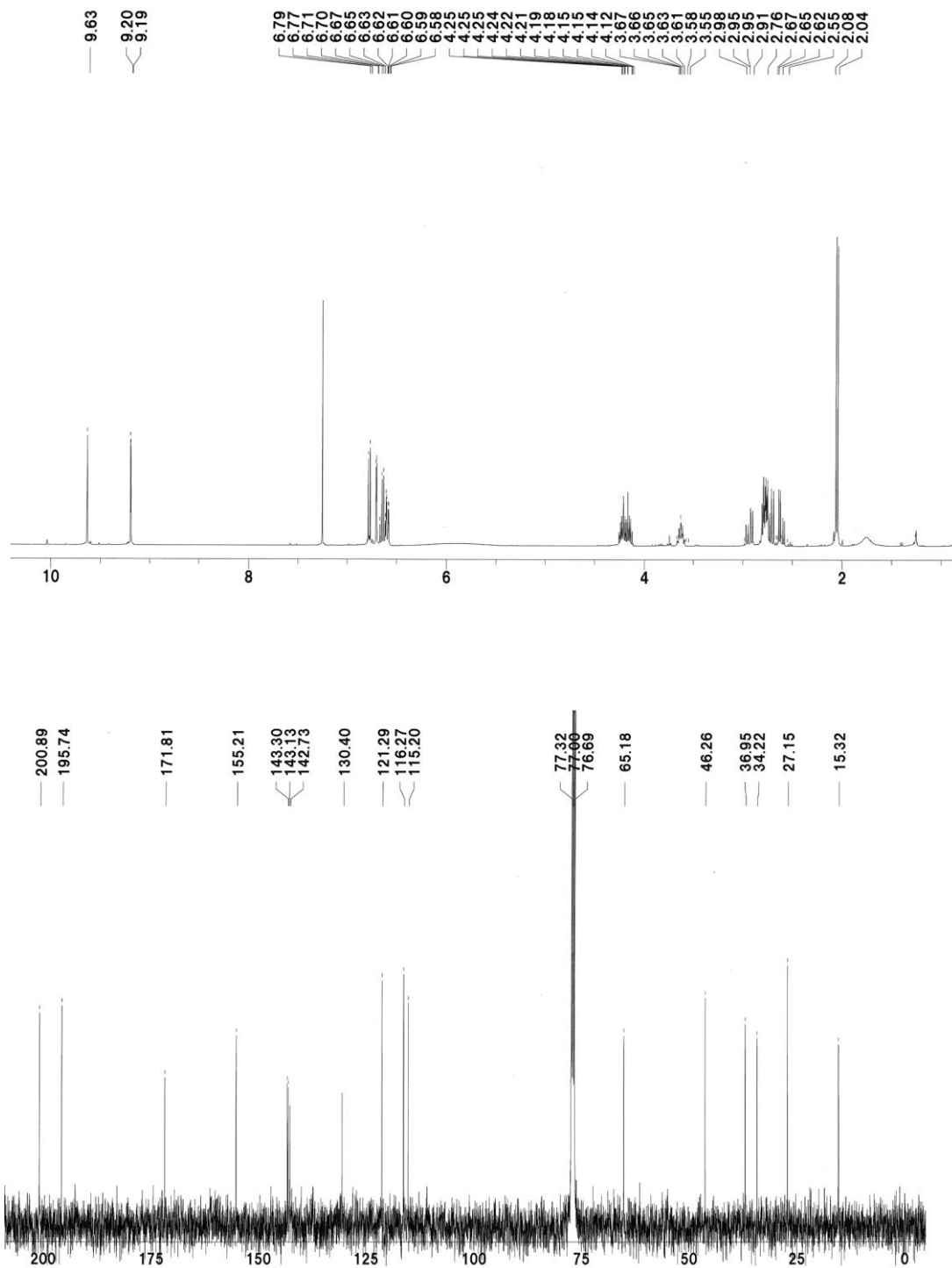


Figure S2. ¹H and ¹³C NMR chart of oleacein.

2.2.4 Synthesis of oleocanthal with proton-exchanged montmorillonite from ligstroside (Eq. S2)

Ligstroside (11 mg, 0.021 mmol) was synthesized according to the reported paper.⁵ It was dissolved in 10 mL of DMSO-d₆ containing 127 mg (7.06 mmol) of H₂O in an NMR tube. Subsequently, proton-exchanged montmorillonite (22 mg) was added to the tube, which was left to stand in oil bath at 150 °C for 12 hours. After the reaction, the reaction solution was filtered to remove the catalyst. The organic layer was washed with water and extracted with AcOEt, dried over Na₂SO₄, filtered and concentrated. The residue was purified by silica gel chromatography (hexane/AcOEt = 10:1 to 1:1) to afford oleocanthal (4.1 mg, 63%).

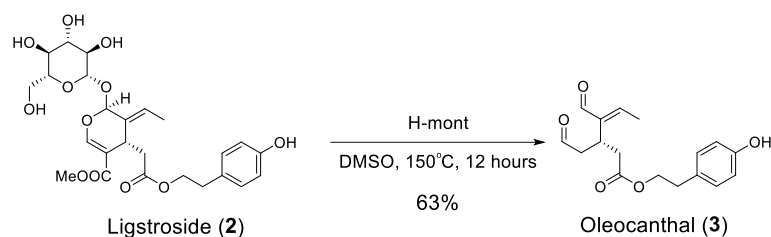


Figure S3. Oleocanthal synthesis from ligstroside.

(-)-Oleocanthal; (3*S*,4*E*)-4-Formyl-3-(2-oxoethyl)-4-hexenoic acid 2-(4-hydroxyphenyl) ethyl ester, Colorless oil. $[\alpha]_D^{26} -0.76$ (c 0.25, CHCl₃) ¹H NMR (400MHz, CDCl₃) δ 9.64 (s, 1H), 9.24 (d, *J* = 2.0 Hz, 1H), 7.04 (d, *J* = 8.0 Hz, 1H), 6.76 (d, *J* = 8.8 Hz, 1H), 6.64 (q, *J* = 7.1 Hz, 1H), 4.25-4.18 (m, 2H), 3.68-3.58 (m, 1H), 2.99 (m, 1H), 2.82 (t, *J* = 7.0 Hz, 2H), 2.79-2.58 (m, 3H), 2.07 (d, 6.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃)δ = 200.4, 195.1, 171.9, 154.3, 154.2, 143.2, 130.0, 129.8, 115.3, 65.1, 46.2, 36.9, 34.2, 27.2, 15.3; LRMS (ESI) calcd for C₁₇H₁₈O₅ [M+H]⁺: 305.14. Found: 305.1.

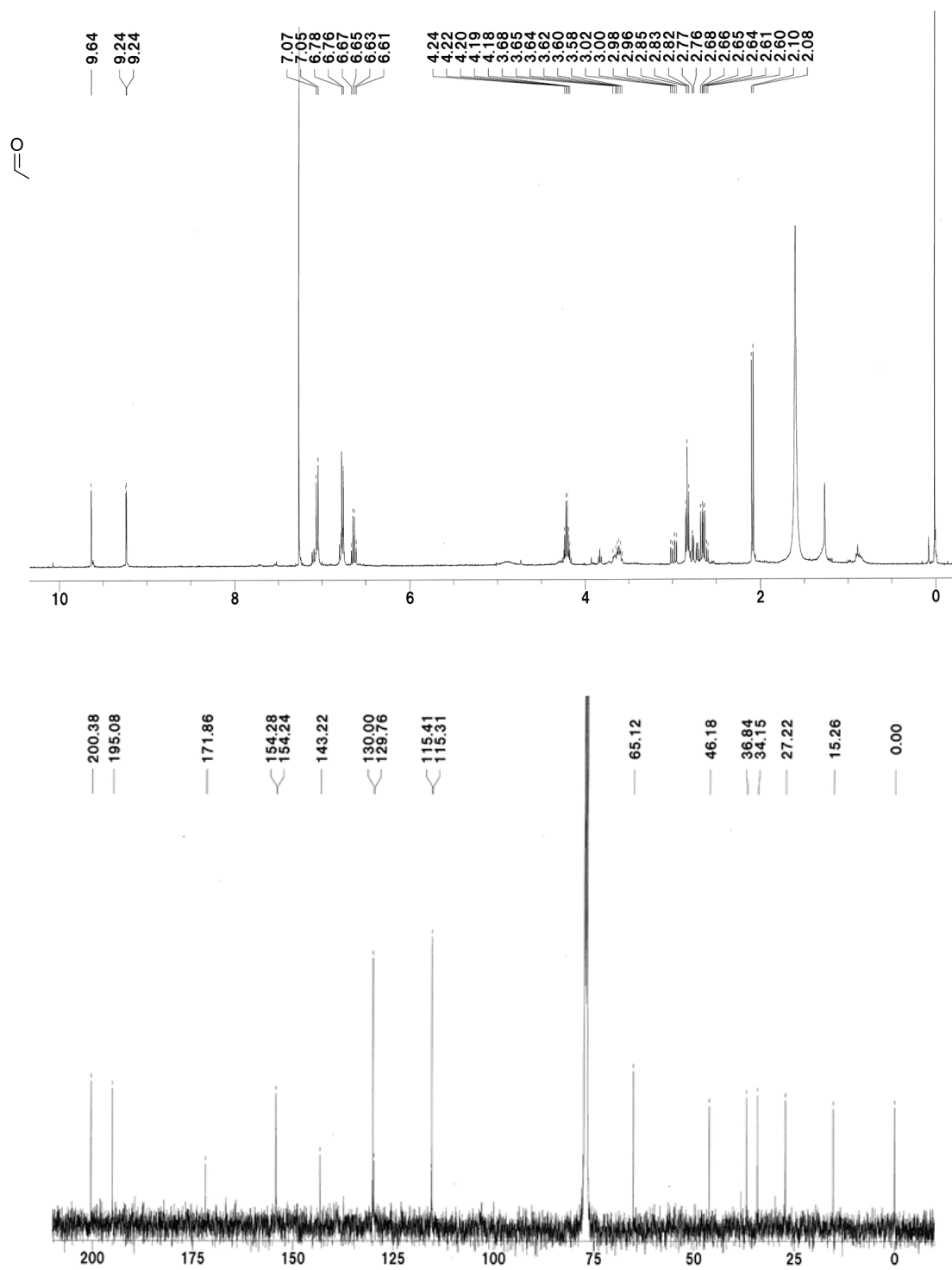


Figure S4. ¹H and ¹³C NMR chart of oleocanthal.

2.2.5 HPLC charts

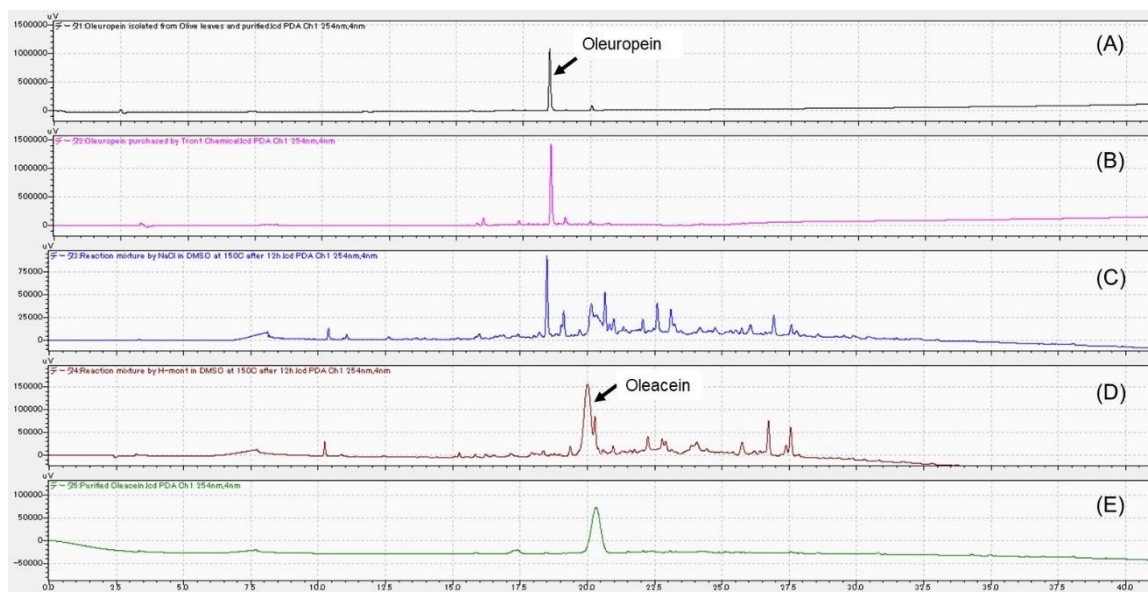
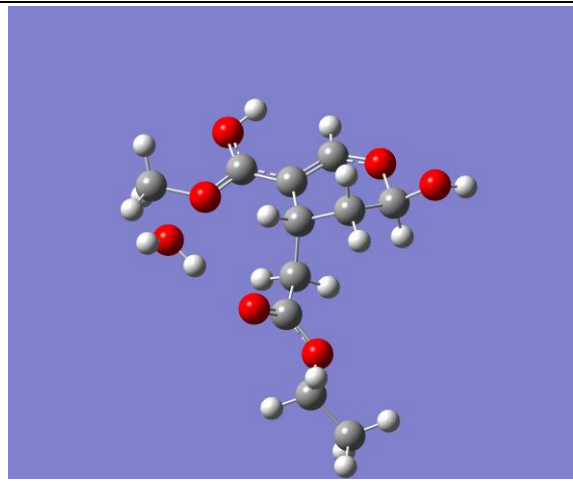
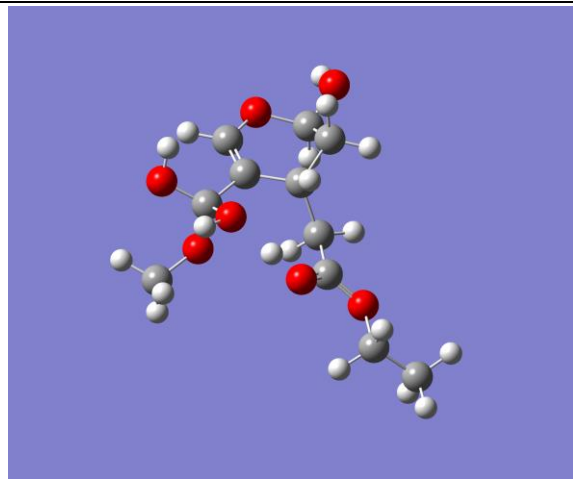
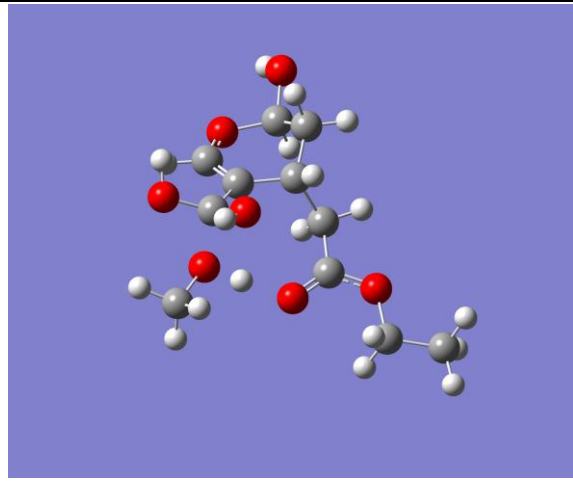
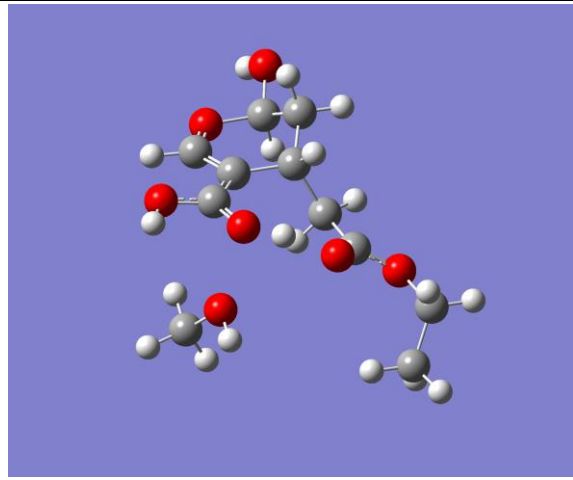
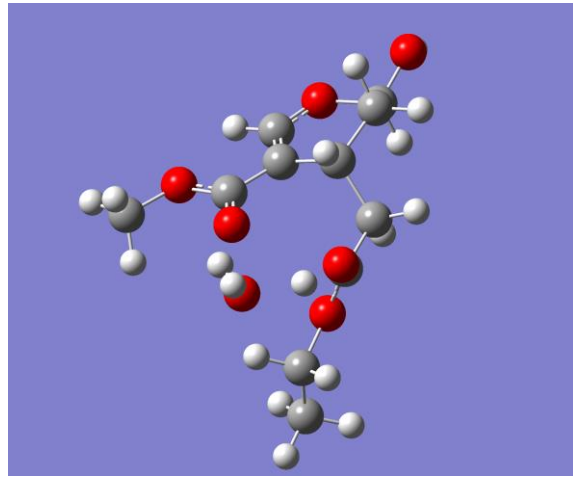
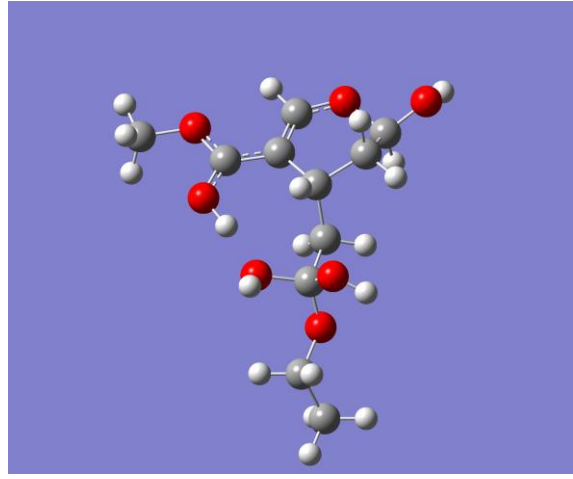
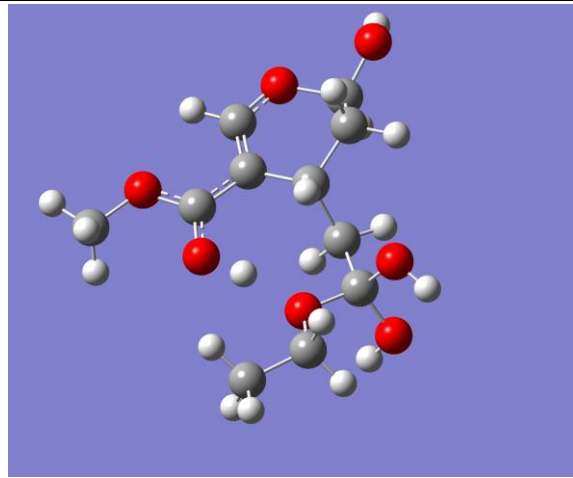
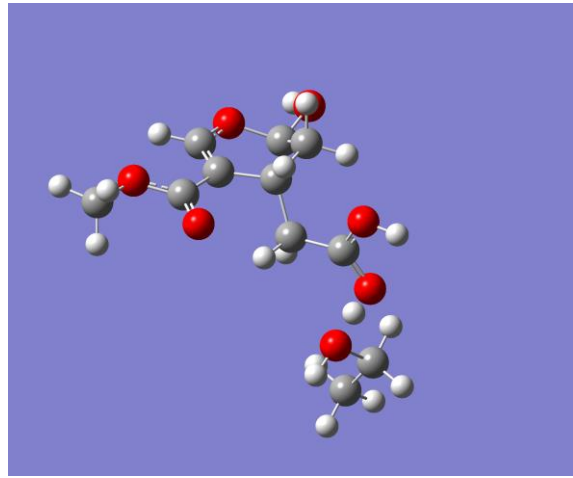
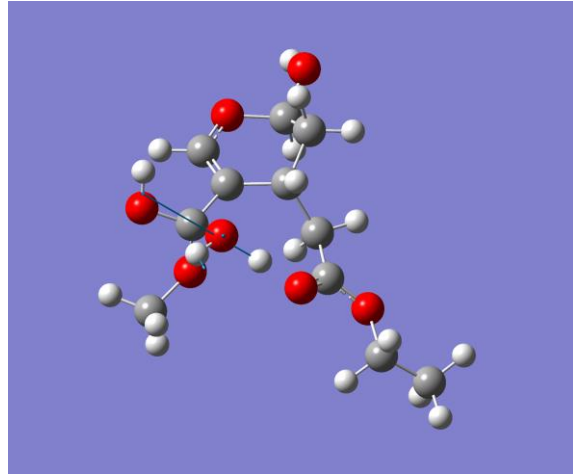


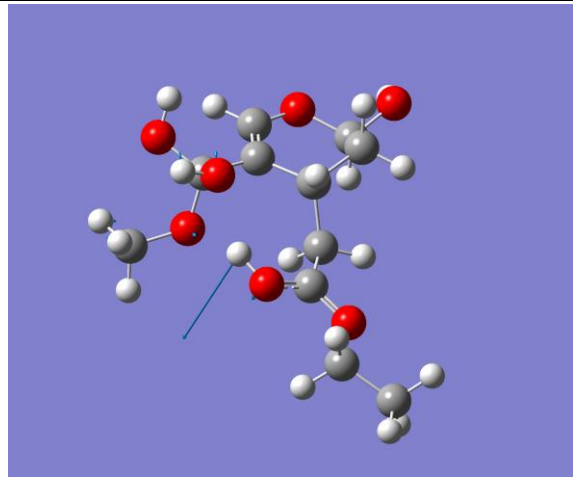
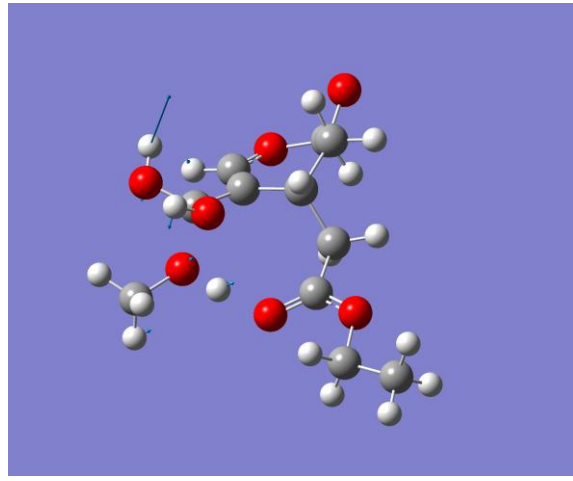
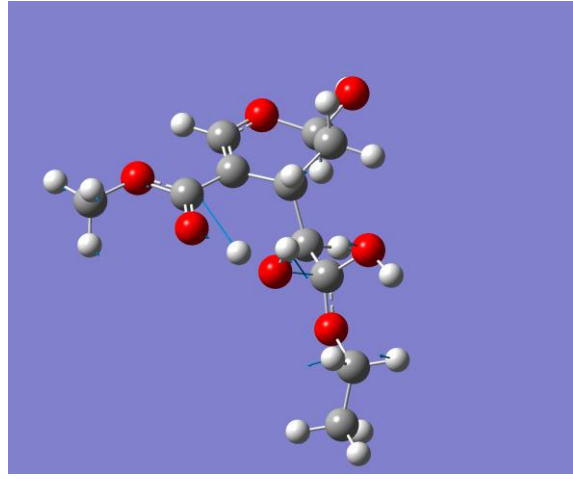
Figure S5. Comparison of HPLC charts. (A) oleuropein (isolated from olive leaves, purity:88%), (B) oleuropein (purchased from Toronto Research Chemicals, purity:75%), (C) reaction solution according to ref. 6 (oleuropein was treated with 2 eq. of NaCl and 10 eq. of H₂O in DMSO at 150 °C for 12 h), (D) reaction solution according to 2.2.3, and (E) purified oleacein from the reaction solution of (D).

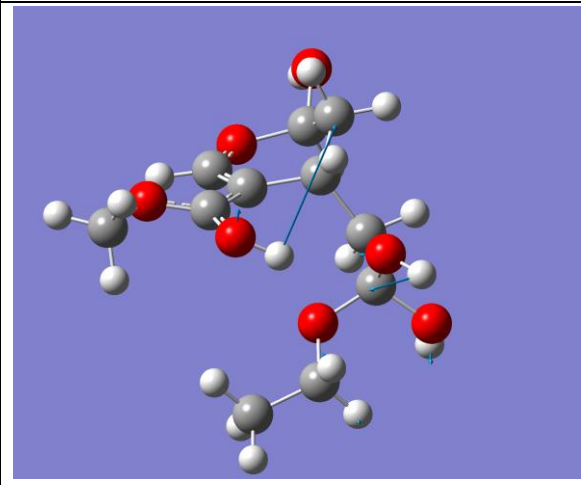
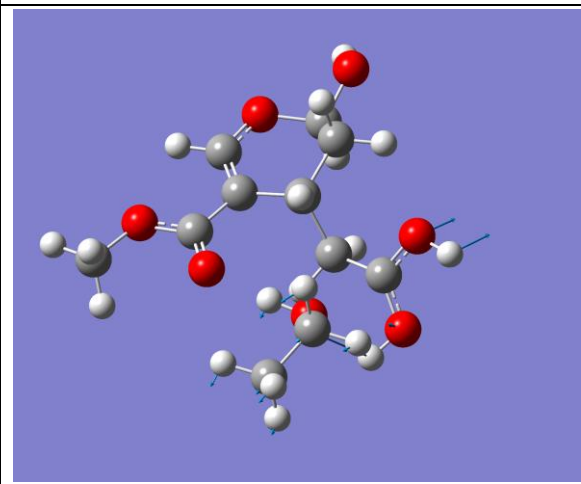
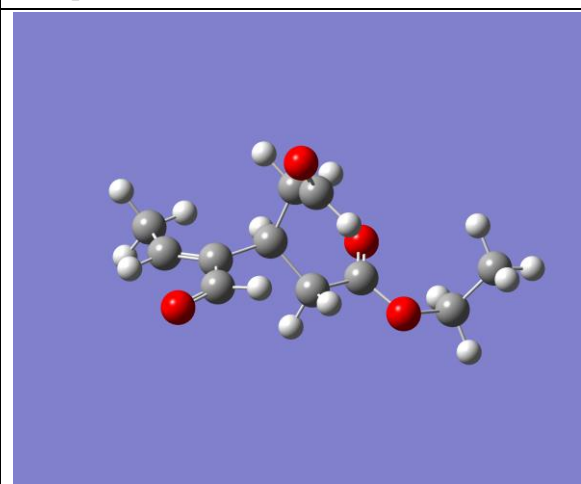
3. DFT calculation results (8 - 16, TS1 - TS6, 16+DMSO, 16+2DMSO)

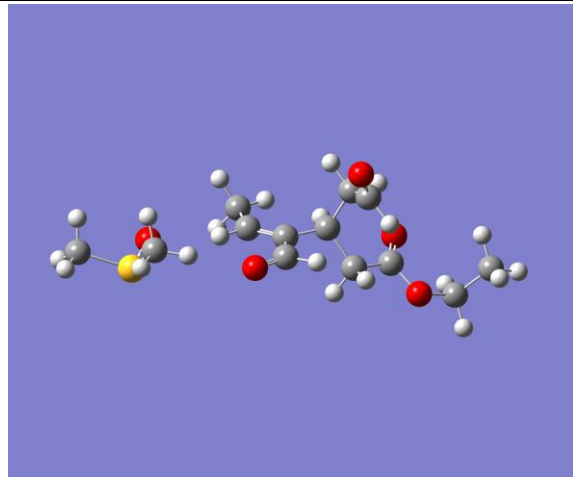
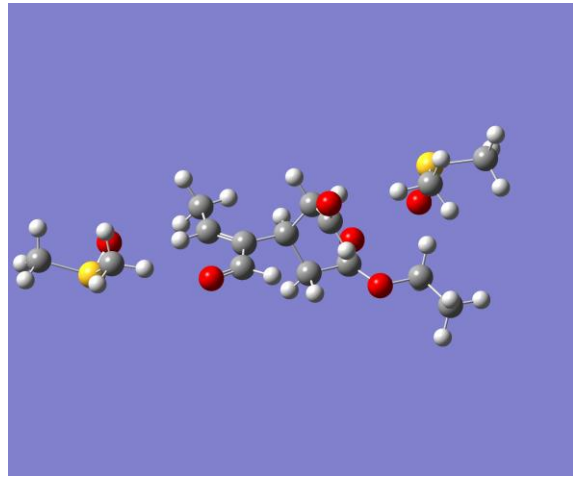
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	Basis Set	6-31+G(d)
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	RMS Gradient Norm	0.00000081
	Imaginary Freq	3.5221
	Dipole Moment	C1
	Point Group	FOPT

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