

Supplemental information

microRNA-132 regulates gene expression

programs involved in microglial homeostasis

Hannah Walgrave, Amber Penning, Giorgia Tosoni, Sarah Snoeck, Kristofer Davie, Emma Davis, Leen Wolfs, Annerieke Sierksma, Mayte Mars, Taofeng Bu, Nicola Thrupp, Lujia Zhou, Diederik Moechars, Renzo Mancuso, Mark Fiers, Andrew J.M. Howden, Bart De Strooper, and Evgenia Salta

Supplemental figures

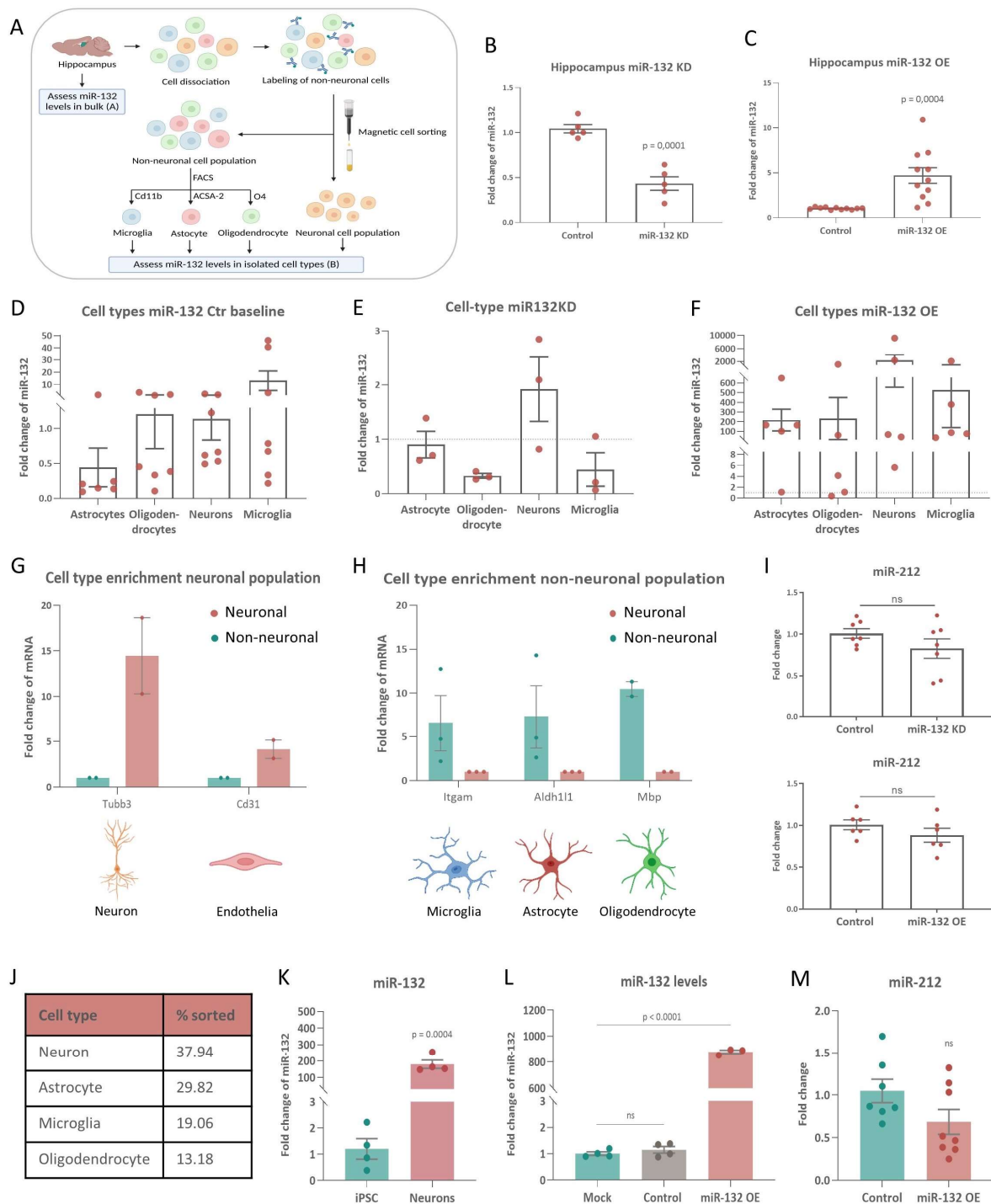


Figure S1. Hippocampal miR-132 expression profile. Related to STAR Methods and Figures 2, 3 and 6. **A.** Schematic representation of isolation protocol of four major brain cellular populations using MACS and FACS. **B.** Semi-quantitative real-time PCR of miR-132 levels in total hippocampus after intracerebroventricular injection with antagomiR (miR-132 KD) or control oligonucleotide. N = 5 mice per group. **C.** Semi-quantitative real-time PCR of miR-132 levels in total hippocampus after injection

with mimic (miR-132 OE) or control oligonucleotide. N = 11 mice per group. **D.** Semi-quantitative real-time PCR of miR-132 levels in isolated cell types of control-injected hippocampi. miR-132 levels in astrocytes, oligodendrocytes and microglia were normalized against neuronal levels. N = 6-7 independent experiments (each experiment contains 4-5 pooled hippocampi). **E.** Semi-quantitative real-time PCR of miR-132 levels in isolated cell types after injection with antagomiR (miR-132 KD) compared each time to corresponding control-injected samples. N = 3 independent experiments (each experiment contains 4-5 pooled hippocampi). **F.** Semi-quantitative real-time PCR of miR-132 levels in isolated cell types after injection with mimic (miR-132 OE) compared each time to corresponding control-injected samples. N = 5 independent experiments (each experiment contains 4-5 pooled hippocampi). Dotted lines in E, F represent control levels. **G, H.** Semi-quantitative real-time PCR of cell type-specific marker genes comparing neuronal and non-neuronal isolated fractions in mouse brain: *Tubb3* (neurons), *Cd31* (endothelia), *Itgam* (microglia), *Aldh1l1* (astrocytes) and *Mbp* (oligodendrocytes). N = 2-3. **I.** Semi-quantitative real-time PCR of miR-212 levels in total hippocampus after intracerebroventricular injection with antagomiR (miR-132 KD, top panel)/mimic (miR-132 OE, bottom panel) or control oligonucleotide. N = 6-7 mice per group. **J.** Average percentages (%) of neurons, astrocytes, microglia and oligodendrocytes sorted for miR-132 level quantification depicted in Figure S1D-F. N = 5 independent experiments. **K.** Semi-quantitative real-time PCR of miR-132 levels in human iPSCs and iPSC-derived neurons. N = 4 biological replicates. **L.** Semi-quantitative real-time PCR of miR-132 levels in iPSC-derived microglia after treatment with mimic (miR-132 OE), control oligonucleotide or medium (mock). N = 3-4 biological replicates. **M.** Semi-quantitative real-time PCR of miR-212 levels in iPSC-derived microglia after treatment with mimic (miR-132 OE) or control oligonucleotide. N = 7-8 biological replicates. Values are presented as mean \pm SEM. In B, C, I, K and M Student's t-test was applied. In D-F, one-way ANOVA with Bonferroni correction was employed. In L, one-way ANOVA with Bonferroni correction was applied.

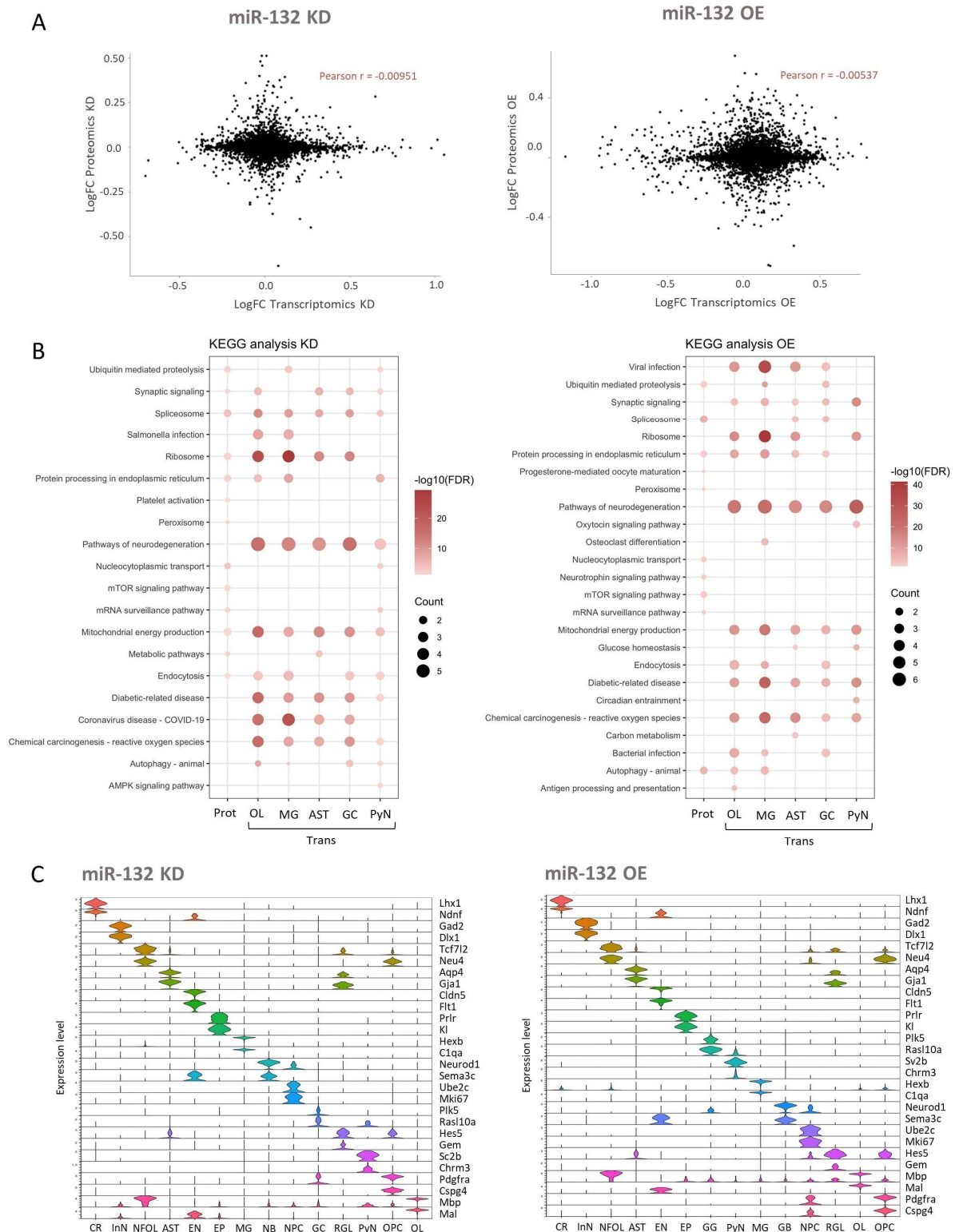


Figure S2. Correlation between proteomic and transcriptional miR-132 response. Related to Figures 2 and 3. **A.** Correlation analysis between the common protein and pseudo-bulked gene pairs. Logarithmic fold change (logFC) comparing miR-132 KD/OE with corresponding controls is plotted. Pearson correlation: $R = -0.00951$ (miR-132 KD) and -0.00537 (miR-132 OE). **B.** Significantly enriched KEGG pathways of differentially expressed proteins (Prot) or genes (Trans) in distinct cell types. Count

represents the % of included proteins/genes that are part of each process. Color represents significance and top 20 GO terms are displayed (Fisher's Exact test, p-values corrected with false discovery rate, adjusted p-value < 0.05 considered significant). **C.** Violin plots of normalized expression levels of cell type-specific marker genes in distinct cell clusters used for cluster annotation. OL, Oligodendrocyte; MG, microglia; AST, astrocyte; GC, granule cell; PyN, hippocampal pyramidal neuron; CR, Cajal-Retzius neuron; InN, inhibitory/GABAergic neuron; NFOL, newly-formed oligodendrocyte; EN, endothelial cell; EP, epithelial cell; NB, neuroblast; NPC, neuronal precursor cell; RGL, radial-glia like cell; OPC, oligodendrocyte precursor cell.

Figure S3. Characterization of microglial cellular states. Related to Figure 5. **A, E.** UMAP visualizing subset microglial population from miR-132 KD (A) or miR-132 OE (E) and corresponding control cells. Cells are colored according to identified clusters. UMAPs show integration of samples and different experimental runs, or amount of identified genes, percent of mitochondrial genes and amount of unique molecular identifiers (UMIs) per cell. **B, F.** UMAP visualizing *ex vivo* activated microglial populations from miR-132 KD (B) or miR-132 OE (F) and corresponding control cells. **C, G.** scProportion test comparing miR-132 KD (C) or OE (G) to control cells present in different cellular states. p-value < 0.05 is considered significant. **D, H.** Expression of top 10 enriched genes per cellular state in miR-132 KD (D) or miR-132 OE (H). **I.** Volcano plot showing differentially expressed genes (DEGs) comparing miR-132 OE cells to their corresponding controls in the DAM state. Significant DEGs indicated in red (Wilcoxon rank-sum test using Bonferroni for p-value correction, adjusted p-value < 0.05 considered significant). **J.** GO biological processes significantly enriched among the most differentially expressed genes, with color indicating significance (Fisher's Exact test, p-values corrected with FDR, adjusted p-value < 0.05 considered significant). Homeostatic, homeostatic microglia; DAM, disease-associated microglia; IRM, interferon-response microglia; CRM, cytokine-response microglia; Cycling, cycling microglia. **K.** UMAP of microglial cells ordered along pseudotime. Color indicates pseudotime ordering. **L.** Integrated UMAP of all microglial cells from the miR-132 KD and miR-132 OE experiments. Color indicates the annotated subpopulations (left), sample (middle) or experiment (right).

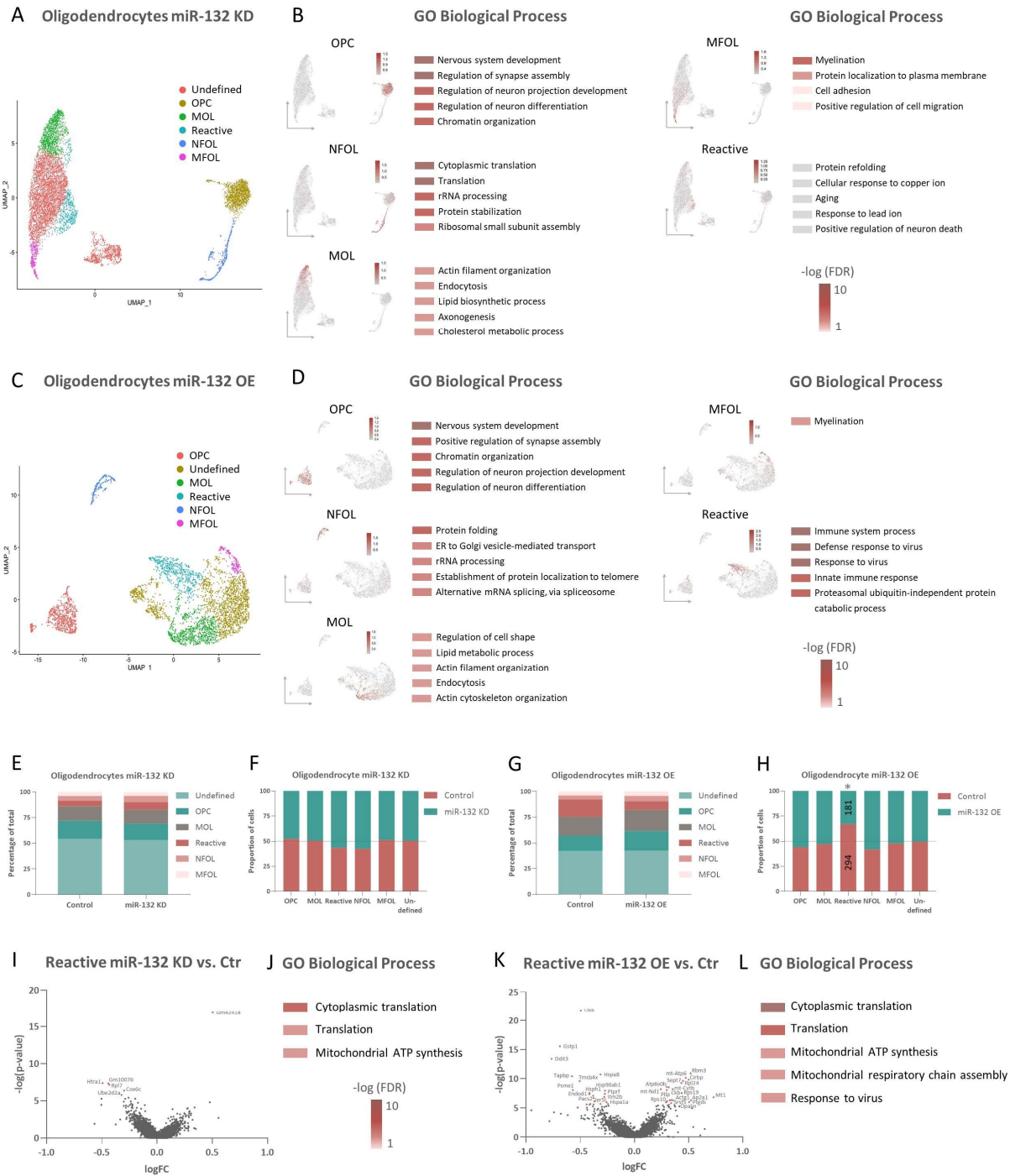


Figure S4. miR-132 overexpression decreases the proportion of reactive oligodendrocytes. Related to Figure S5. A, C. UMAP visualizes subset oligodendrocytic lineage from miR-132 KD (A) or miR-132 OE (C) and corresponding control cells, colored according to identified clusters. **B, D.** UMAP plots colored based on the signature score of the combined gene set that characterizes each individual oligodendrocytic subpopulation. Top 5 unique GO biological processes of significantly enriched genes are provided per subtype; color is indicated according to significance (Fisher's Exact test, p-values corrected with FDR, adjusted p-value < 0.05 considered significant). **E, G.** Normalized percentage of cells present in distinct cellular types/states per condition in miR-132 KD (E) and miR-132 OE (G). **F, H.**

Normalized percentage of cells present in distinct cellular types/states per cell type/state in miR-132 KD (F) and miR-132 OE (H). Significance indicated with an asterisk (Chi-squared test, p-value = 0.0007). The number of cells present in miR-132 OE or control condition is depicted on the graph. **I, K.** Volcano plot showing differentially expressed genes (DEGs) comparing miR-132 KD (I) or miR-132 OE (K) cells to their corresponding controls in reactive oligodendrocytes. Significant DEGs indicated in red (Wilcoxon rank-sum test using Bonferroni for p-value correction, adjusted p-value < 0.05 considered significant). **J, L.** GO biological processes significantly enriched among the top 500 differentially expressed genes (depicted in I, K), with color indicating significance (Fisher's Exact test, p-values corrected with FDR, adjusted p-value < 0.05 considered significant). OPC, oligodendrocyte precursor cell; MOL, mature oligodendrocyte; Reactive, reactive oligodendrocyte; NFOL, newly-formed oligodendrocyte; MFOL, myelin-forming oligodendrocyte.

oligodendrocyte precursor cell; MOL, mature oligodendrocyte; Reactive, reactive oligodendrocyte; NFOL, newly-formed oligodendrocyte; MFOL, myelin-forming oligodendrocyte.

Table S6: Sequences of primers used. Related to STAR Methods and Figures 6 and S1.

Primer	Sequence	Cat#
mmu-miR-132-3p	U AACAGUCUACAGCCAUGGUCG	204129, Exiqon, Qiagen
Primer	Forward	Reverse
<i>hAPOC1</i>	TGGTTCTGTCGATCGTCTTG	GAAAACCACTCCCGCATCT
<i>hCD9</i>	TTCCTCTTGGTGATATTCGCCA	AGTTCAACGCATAGTGGATGG
<i>hLGALS3</i>	GTGAAGCCCAATGCAAACAGA	AGCGTGGGTTAAAGTGAAGG
<i>hP2RY12</i>	GAAGACCACCAGGCCATTTA	TCATGCCAGACTAGACCGAAC
<i>hCX3CR1</i>	AGTGTCAACGACATTTACCTCC	AAGGCGGTAGTGAATTTGCAC
<i>hSSH2</i>	TTCAAGGATGGAGGAACTGG	CATGAAGATGTCTGGGCTGA
<i>hPDIA3</i>	AAGCTCAGCAAAGACCCAAA	CACTTAATTCACGGCCACCT
<i>hMAF</i>	AAGTCGACCACCTCAAGCAG	GCTTCCAAAATGTGGCGTAT
<i>hDDX5</i>	GTCAACAAGAGCGTGACTGG	GCGAGCAGTTCTTCCAATTC
<i>hSEC62</i>	AGTAATAGCGGCCACCCTCT	GGAGTCAATGAAGCCCAT
<i>hCD164</i>	AAGTGGGGAACACGACAGAC	TGAAACTGGCTGCATCAAAG
<i>hUBC</i>	GGGTCGCAGTTCTTGTTTGT	GGAGGGATGCCTTCCTTATC
<i>hPSMB4</i>	TGTCACCGAAAAAGGTGTTG	AAGAGTCTATCTTTGAAC TAGCCAAG
<i>hCASP1</i>	AGTGCAGGACAACCCAGCTA	AGATAATGAGAGCAAGACGTGTG
<i>hCASP3</i>	CATGGAAGCGAATCAATGGACT	CTGTACCAGACCGAGATGTCA
<i>hXIAP</i>	GAAGACCCTTGGGAACAACA	TGAAACTGAACCCCATTCGT
<i>hBCL2</i>	GGTATTGGTGAGTCGGATCG	AAGAGTGAGCCCAGCAGAAC
<i>hGAPDH</i>	TCAAGAAGGTGGTGAAGCAGG	ACCAGGAAATGAGCTTGACAAA
<i>mTubb3</i>	TGGAGCGCATCAGCGTATAC	GCCCTGGGCACATACTTGTG
<i>mCd31</i>	AAGCAGCACTTTCAGTCA	CATCTCCACGGGTTTCTGTT
<i>mltgam</i>	GGCTTTGGACAGAGTGTGGT	GCTGGGGGACAGTAGAAACA
<i>mAldh1l1</i>	CCATCAACTGGACCCCTCATT	CCTGAACCATCCCTTTGATG
<i>mMbp</i>	ACTACCCATTATGGCTCCCTG	GAGGTGGTGTTCGAGGTGTC
<i>mGapdh</i>	TTGATGGCAACAATCTCCAC	CGTCCCGTAGACAAAATGGT
<i>mActin</i>	AGCCATGTACGTAGCCATCC	CTCTCAGCTGTGGTGGTGAA