Supplementary information

SOSIP.664	Fab	Affinity	$T(k_{on})^b$	$T (k_{off})^b$	χ^{2b}	k_t^{b}	$T(k_t)^c$
trimer		purification				(RU/Ms)	
BG505	PGT145	2G12	$9.4 \cdot 10^2$	$1.2 \cdot 10^{3}$	$4.4 \cdot 10^{-2}$	$1.0 \cdot 10^{15}$	1.5
		(n=3)	± 33	± 67	$\pm 5.9 \cdot 10^{-3}$	$\pm 7.0 \cdot 10^{14}$	± 0
	PGT151	2G12	$3.0 \cdot 10^{2}$	$1.3 \cdot 10^{2}$	0.62	$2.0 \cdot 10^{22}$	1.5
		(n=2)	± 5.0	± 0	\pm 1.8 \cdot 10 ⁻²	\pm 1.3 \cdot 10 ²²	± 0
B41	PGT145	2G12	$1.6 \cdot 10^2$	51	0.12	$2.5 \cdot 10^{16}$	0.14
		(n=3)	± 3.3	± 2.9	± 0	\pm 1.4 \cdot 10 ¹⁶	$\pm 7.3 \cdot 10^{-2}$
		PGT145	$2.1 \cdot 10^{2}$	3.2	0.19	$4.8 \cdot 10^{9}$	1.5
		(n=3)	± 18	± 3.1	$\pm 2.2 \cdot 10^{-2}$	$\pm 2.4 \cdot 10^9$	± 0.46
	DCT151	2G12	$1.2 \cdot 10^{2}$	$1.4 \cdot 10^2$	0.13	$1.2 \cdot 10^{22}$	6.3 · 10 ⁻⁴
	FUIIJI	(n=3)	±16	± 18	$\pm 3.5 . 10^{-2}$	\pm 1.2 \cdot 10 ²²	$\pm 3.1 \cdot 10^{-4}$

SI Table 1. Validation of Langmuir modeling^{*a*}

^{*a*} Tabulated values are means \pm S.E.M of *n* independent replicates.

^b Kinetic parameters were validated by global fitting. T value > 10 confirms the statistical significance of the parameter value. χ^2 values, the average squared residuals between experimental and fitted data, were determined as measures the goodness of the fit.

^{*c*} The mass transfer constant = $k_t > 10^9$ and low value (<10) of T(*kt*) indicate lack of significant mass-transfer limitation.

SOSIP.664 trimer	Fab	Affinity purification	T (kon1) ^b	$T (k_{off1})^b$	T (kon2) ^b	T (k _{off2}) ^b	χ^{2b}
BG505	PGT145	2G12 (n-3)	$9.8 \cdot 10^2$	0.47	$1.3 \cdot 10^2$	$3.6 \cdot 10^2$	$1.5 \cdot 10^{-2}$ + 1.6 \cdot 10^{-3}
	PGT151	2G12 (n=2)	$2.5 \cdot 10^2$ ± 10	$ \begin{array}{r} \underline{0.10} \\ 0.15 \\ \pm 5.0 \cdot 10^{-3} \end{array} $		$\frac{1}{78}$ ± 1.5	$ \begin{array}{r} \underline{1.0} & 10 \\ 0.10 \\ \pm 2.0 \cdot 10^{-3} \end{array} $
B41	PGT145	2G12 (n=3)	83 ± 26	44 ± 11	$1.8 \cdot 10^2 \pm 22$	39 ± 8.0	$\begin{array}{c} 8.0 \cdot 10^{-2} \\ \pm 1.1 \cdot 10^{-2} \end{array}$
		PGT145 (n=3)	$1.1 \cdot 10^2 \pm 45$	27 ± 10	$1.9 \cdot 10^2 \pm 29$	25 ± 3.5	$0.13 \pm 2.7 \cdot 10^{-2}$
	PGT151	2G12 (n=2)	$1.1 \cdot 10^2 \pm 11$	$1.3 \cdot 10^2 \pm 47$	95 ± 8.0	$1.2 \cdot 10^2 \pm 25$	$0.10 \pm 1.2 \cdot 10^{-2}$

SI Table 2. Validation of heterogeneous-ligand modeling ^a

^{*a*} Tabulated values are means \pm S.E.M of *n* independent replicates.

^b Kinetic parameters were validated by global fitting. *T* value > 10 confirms the statistical significance of the parameter value. χ^2 values, the average squared residuals between experimental and fitted data, were determined as measures the goodness of the fit.





SI Figure 1. Analysis of bNAb binding to differentially bNAb-purified B41 SOSIP.664 by ELISA. Each diagram shows the binding of one bNAb to BG505 (top row) or B41 (bottom row) SOSIP.664 trimers purified in three ways (color-coded legend). The optical densities (OD₄₅₀) are plotted on the y axes as functions of the bNAb concentrations.



SI Figure 2. Extent of neutralization in vitro as a predictor of protection in vivo. Extent of neutralization is depicted on the y axis (%) as a function the x-fold excess of NAb concentration over IC₅₀ in the neutralization reaction (mixture of NAb and virus). A. The diagram elaborates on a meta-analysis of data from passive immunization of macaques with bNAbs followed by SHIV challenge [1]. Logistic modeling showed that protection of 95% of animals required an ID₅₀ of the animal sera at challenge of ~700 (green vertical line). The meta-analysis suggested that 50% protection in vivo corresponded to 93.7% neutralization in vitro, 75% protection to 99.8% neutralization, and 95% protection to > 99.9% neutralization. That was under the assumption of an asymptotic approach to 100% neutralization, but the values for instantaneous inhibitory potential were based on the empirical data. The red and blue curves illustrate possibilities for smaller discrepancies between protection in vivo and neutralization in vitro. The arbitrary coincidence of 95% protection and neutralization is chosen as an example. NAbs present at 700fold higher concentration than their IC₅₀ (*i.e.*, ID₅₀ of the solution is 700) would neutralize 95% of the virus input, given the Hill coefficient = 0.45 and PF = 0% or a Hill coefficient = 1.0 and PF = 5%. Under the assumption of similar ID₅₀ (a product of NAb affinity and concentration [2]) and efficacy, *in vivo* and *in vitro*, the example might be deemed to show realistic correspondences. The unknown is the degree of neutralization of the inoculum *in vivo* in the relevant compartment, *i.e.*,

perhaps exclusively at the portal of entry. In **B** the condition that the ID₅₀ is the same *in vitro* and *in vivo* is relaxed. Neutralization *in vitro* is mediated by NAbs in serum; protective neutralization *in vivo* is postulated to occur at the mucosal site of viral deposition. The concentration of the infused NAb is known to be lower in the mucosal interstitial fluid and lumen than in the sera, and more markedly so intrarectally than intravaginally [3], reducing ID₅₀ accordingly. Furthermore, virus and target cells differ in important respects between *in vivo* and *in vitro* conditions, which may also reduce the ID₅₀ [4-7]. The diagram shows a simulation of the degree of neutralization under the realistic assumption that ID₅₀ is 37-fold lower in the mucosa than in serum through the combined NAb concentration and potency reductions, *e.g.*, 3.7-fold of the former and 10-fold of the latter. The serum ID₅₀ value of 700 from [1], corresponding to protection of 95% of the animals, would, with asymptote 100% and Hill coefficient 1.0, give 99.86% neutralization, which might seem high considering the proportion of infected animals. But with the above assumptions about the mucosa, neutralization there would be 95%. We are not arguing that percentages of neutralization in the mucosa and protection of animals have to be identical, just that under realistic assumptions they do not necessarily differ much.

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