Real-life Experience and Diagnostic Utility of the BioFire Joint Infection PCR Panel in Bone and Joint Infections: Analysis of a Prospective Validation Study

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Table S1. Patient characteristics

	Total (n=57) Infected		Uninfected	P value	
		(n=39)	(n=18)		
Gender, male (%)	35 (61.4)	21 (53.8)	14 (77.8)	0.142	
Age, years (mean)	64.5	65.9	61.6	0.473	
Diabetes (%)	17 (29.8)	13 (33.3)	4 (22.2)	0.537	
Cardio- or cerebro- vascular disease (%)	20 (35)	14 (35.9)	6 (33.3)	1	
Chronic kidney disease (%)	8 (14)	6 (15.4)	2 (11.1)	1	
Inflammatory arthritis (%)	1 (1.8)	1 (2.6)	0 (0)	-	
Cancer (%)	5 (8.8)	4 (10.3)	1 (5.6)	1	
Immunocompromised (%)	3 (5.3)	2 (5.1)	1 (5.6)	1	
Sample type (%)					
Tissue sample	23 (40.4)	20 (51.3)	3 (16.7)	0.02	
Fluid sample	34 (59.6)	19 (48.7)	15 (83.3)		
Sample obtainment method (%)					
Arthrocentesis	30 (52.6)	16 (41)	14 (77.8)	0.012	
Intra-operative	27 (47.4)	23 (59)	4 (22.2)		
Suspected infection site ¹ (%)					
Bone	13 (22.8)	11 (28.2)	2 (11.1)	0.124	
Joint	44 (77.2)	28 (71.8)	16 (88.8)	0.191	
Native	22 (38.6)	14 (35.9)	8 (44.4)	1	
Prosthetic	22 (38.6)	14 (35.9)	8 (44.4)		
Soft tissue	Soft tissue 2 (3.5)		0 (0)	1	

Suspected post-	ed post-				
operative infection	33 (57.9)	24 (61.5)	9 (50)	0.565	
(%)					
Multiple prior	8 (24.2)	6 (25)	2 (22.2)	1	
revision surgeries (%)					
Antibiotics in					
preceding 14 days	20 (35.1)	14 (35.9)	6 (33.3)	1	
(%)					
Blood leukocyte	10.2	10.9	8.5	0.141	
count, 109/L (mean)					
Blood C-reactive	138	162	74	0.007	
protein, mg/L (mean)					
Arthrocentesis	34 (59.6)	20 (51.3)	14 (77.8)	0.083	
performed (%)					
Synovial fluid					
leukocyte count,	54.9	78	24.1	0.003	
109/L (mean)					
Synovial fluid	7 0.0	04.6	71.5	0.170	
neutrophil,	/8.8	84.6	71.5	0.172	
percentage (mean)					
Surgery performed	40 (70.1)	35 (89.7)	5 (27.7)		
(%)					
No infection found	1 (1.7)	0 (0)	1 (5.5)		
Debridement	32 (56 .2)	29 (74.3)	3 (16.7)		
One-stage joint	1 (1.7)	1 (2.6)	0 (0)	<0.001	
exchange					
Two-stage joint	5 (8.8)	5 (12.8)	0 (0)		
exchange					
Intramedullary	1 (1.7)	0 (0)	1 (5.5)		
nail exchange					
Empirical antibiotics	45 (78.9)	37 (94.9)	8 (44.4)	<0.001	
administered (%)					
Final diagnosis ¹				n/a	
Native joint septic	14	14	0	11/ a	

arthritis				
Prosthetic joint	14	14	0	
infection				
Spinal infection	8	8	0	
Osteomyelitis	1	1	0	
Device-associated	2	2	0	
osteomyelitis				
Soft tissue	9	9	0	
infection				
Uninfected	18	0	18	
Hospital length of	15	20	5	<0.001
stay, days (mean)				
Re-operation	7 (12.3)	7 (17.9)	0 (0)	0.565
performed (%)				
Length of follow-up,	44	52	27	0.015
days (mean)				
Outcome at last				
follow-up (%)				
No evidence of	46 (80.7)	29 (74.3)	17 (94.4)	
infection				0.180
Persistent or	7 (12.3)	6 (15.4)	$1 (5.6)^2$	
recurrent infection				
Death	4 (7)	4 (10.3)	0 (0)	
Ongoing antibiotics	17 (29.8)	16 (41)	1 (5.6)	0.011
at last follow-up (%)	· · · ·			

¹ More than one suspected infected site or diagnosis may apply to each patient

² A single patient who was uninfected at index event later developed persistent septic arthritis

Table S2. Pathogens identified in infected cases

Staphylococcus aureus	6
Cutibacterium acnes	3
Staphylococcus epidermidis	2
Streptococcus pyogenes	2
Streptococcus agalactiae	2
Streptococcus dysgalactiae	2
Escherichia coli	2
Enterobacter cloacae complex	2
Pseudomonas aeruginosa	2
Klebsiella pneumoniae	1
Klebsiella aerogenes	1
Aggregatibacter aphrophilus	1
Staphylococcus hominis	1
Streptococcus intermedius	1
Streptococcus gallolyticus	1
Finegoldia magna	1
Bacteroides fragilis	1
Anaerococcus spp.	1
Ureaplasma urealyticum	1
Mixed flora	1

Case	Description
1	Staphylococcus epidermidis in culture and 16s rRNA PCR
2	Cutibacterium acnes in culture
3	Staphylococcus epidermidis in culture and 16s rRNA PCR
4	Cutibacterium acnes in culture
5	Staphylococcus aureus ¹ in culture
6	Enterobacter cloacae ¹ in culture and 16s rRNA PCR
7	Cutibacterium acnes in culture
8	Aggregatibacter aphrophilus in blood culture and tissue 16s rRNA PCR
9	Ureaplasma urealyticum in 16s rRNA PCR
10	Staphylococcus hominis in 16s rRNA PCR
11	BioFire, culture and 16s rRNA PCR were negative
12	BioFire, culture and 16s rRNA PCR were negative
13	BioFire, culture and 16s rRNA PCR were negative
14	BioFire, culture and 16s rRNA PCR were negative
15	BioFire, culture and 16s rRNA PCR were negative
16	BioFire, culture and 16s rRNA PCR were negative
17	BioFire, culture and 16s rRNA PCR were negative

Table S3. Description of BioFire-negative infected cases (n=17)

¹ Organism is included in the BioFire panel

Case	Infection status	BioFire result	Culture result	Description
1	Infected	Negative	Positive	Staphylococcus epidermidis in culture and 16s
				rRNA PCR
2	Infected	Negative	Positive	Cutibacterium acnes in culture
3	Infected	Negative	Positive	Staphylococcus epidermidis in culture and 16s
		-		rRNA PCR
4	Infected	Negative	Positive	Cutibacterium acnes in culture
5	Infected	Negative	Positive	Staphylococcus aureus in culture
6	Infected	Negative	Positive	Enterobacter cloacae in culture and 16s rRNA
		C		PCR
7	Infected	Negative	Positive	Cutibacterium acnes in culture
				Streptococcus spp. in BioFire, Streptococcus
8	Infected	Positive	Negative	dysgalactiae in 16s rRNA PCR, recent antibiotic
				treatment
				Streptococcus spp. in BioFire, Streptococcus
9	Infected	Positive	Negative	dysgalactiae in 16s rRNA PCR, recent antibiotic
				treatment
10	Uninfected	Negative	Positive	Cutibacterium acnes in single specimen culture
11	Uninfected	Negative	Positive	Bacillus spp. in single specimen culture
12	Uninfected	Negative	Positive	Cutibacterium acnes in single specimen culture

Table S5. Diagnostic modalities sensitivity rates according to specimen type, obtainment

Modality	Fluid	Tissue	Arthro-	Intra-	Native	Prosthetic	Bone	Soft	Recent	No recent
	sample	sample	centesis	operative	joint	joint		tissue	antibiotics	antibiotics
BioFire	58%	55%	63%	52%	43%	71%	44%	100%	62%	54%
Gram stain	26%	40%	13%	39%	29%	36%	22%	100%	31%	35%
Culture	68%	70%	63%	74%	43%	86%	67%	100%	62%	73%
16s rRNA PCR	64%	81%	64%	79%	45%	80%	100%	100%	82%	68%

method, infection site and recent antibiotic exposure

Supplementary S6. 16s rRNA procedure

DNA Extraction

DNA Extraction was performed using Seegene's STARMAG (Seegene Inc., S. Korea) on a robotic Hamilton liquid handler. Homogenized ground solid tissue or synovial fluid were used according to availability in each case.

Initial PCR

5ul of DNA was used as template for initial PCR.

PCR reaction was as follows:

Reagent	ul		
Roche Sybr ready-			
mix		15	
16s F primer 10um		1	
16s R primer 10um		1	
DNA template		5	
h20		8	
total		30	

PCR Thermocycler Program was as follows:

Temperature		Time	
	95	5	min
	95	15	sec
	58	30	sec
	72	30	sec
repeat		x29	
	72	3	min

Primers used for 16s (~750bp):

16SAF CCAGACTCCTACGGGAGGCAG

16SAR ACATTTCACAACACGAGCTGACGA