Correlating the differences in the receptor binding domain of SARS-CoV-2 spike variants on their interactions with human ACE2 receptor

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Figure S1: Cloning of hACE gene and characterization. Overview of molecular cloning of hACE2 gene into lentiviral plasmid by Gibbson assembly (A). The fragmentation of pLenti-hACE2-P2A-PuroR plasmid by restriction digestion (Nhel and BamHI enzyme) was predicted insilico by Snapgene software (B) and confirmed by in vitro digestion (C).



Figure S2: Gating strategy of Flow cytometric evaluation. P1 Shows Population of the 293ThACE2 cells. To discriminate the doublets, single cells were gated from P1 Population. PE+ve cells were gated from the singlet population. Unstained (A), Histograms for Isotype controls for Streptavidin-PE (B), Wild spike bound cells (C). The PE positive was gated against isotype as a control.



Spike variants binding affinity to hACE2 (MFI)

Figure S3. Histogram plots show the level of spike variants binding to dimeric hACE2 receptor present on surface of the cells at different concentrations, generated by FlowJo software. Unstained-cells only, Control-cells + streptavidin–PE conjugate.



Lane 1- 293T-ACE2 cells protein lysate-40 µg Lane 2- HEK-293T cells protein lysate-40 µg Lane 3- Prestained Protein Ladder Lane 4-HEK-293T cells protein lysate-20 µg Lane 5- 293T-ACE2 cells protein lysate-20 µg

Figure S4. Whole blot images of western blot, performed with 20 µg and 40 µg of protein lysate from HEK-293T and 293T-ACE2 cell lines, and prestained protein ladder (Bio-Helix, #PMB01-0500).

Table S1. Primer List

Primer name	Primer sequence 5'- 3' direction
Human ACE2 cloning	F-CCAGAACACAGGTGTCGTGACGCGGGAT
	CCGCCACCATGTCAAGCTCTTCCTGGCTCC
	R- GAAGTTGGTGGCGCCGCTGCCGCTAGCA
	AAGGAGGTCTGAACATCATCAGTGTTTTG
Human ACE2	F- GGGATCAGAGATCGGAAGAAGAAA
	R-AGGAGGTCTGAACATCATCAGTG
Human GAPDH	F- CTGGGCTACACTGAGCACC
	R- AAGTGGTCGTTGAGGGCAATG