

TITLE

Hydra-elastin-like polypeptides increase rapamycin potency when targeted to cell-surface GRP78

AUTHORS

Hugo Avila¹, Jingmei Yu¹, Geetha Boddu¹, Alvin Phan¹, Anh Truong¹, Santosh Peddi¹, Hao Guo¹, Shin-Jae Lee^{1,2}, Mario Alba¹, Ethan Canfield⁴, Vicky Yamamoto⁵, James C. Paton⁶, Adrienne W Paton⁶, Amy S. Lee⁵, J. Andrew MacKay^{1,2,3}

AFFILIATIONS

1. USC School of Pharmacy, Department of Pharmacology and Pharmaceutical Sciences
2. USC Viterbi School of Engineering, Department of Biomedical Engineering
3. USC Keck School of Medicine, Department of Ophthalmology
4. USC School of Pharmacy, Mass Spectrometry Core
5. USC Keck School of Medicine, Department of Biochemistry and Molecular Medicine
6. University of Adelaide, Research Centre for Infectious Diseases, Department of Molecular and Biomedical Science

CORRESPONDING AUTHOR INFORMATION

MacKay, J. Andrew

1985 Zonal Ave., PSC 306A, Los Angeles, CA, 90089

Telephone: (323) 442-4118. Fax: (323) 442-4118. jamackay@usc.edu

L-5FA:

MRLLDTNRP LLPYVPGAGMGVQVETISPGDGRTFPKRGQTCVVHYTG MLEDGKKFDSSRDRNKPFKMLGKQEVIR
GWEEGVAQMSVGQRAKLTISPDYAYGATGHPGIIPPHATLVFDVELLKLEGVPGAGVPGAGVPGAGVPGAGVPGAG
VPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGV
GAGVPGAGVPGAGVPGAGVPGAGVQVETISPGDGRTFPKRGQTCVVHYTG MLEDGKKFDSSRDRNKPFKMLGKQ
EVIRGWEEGVAQMSVGQRAKLTISPDYAYGATGHPGIIPPHATLVFDVELLKLEGVPGAGVPGAGVPGAGVPGAGVPGAGV
GAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGV
GAGVPGAGVPGAGVPGAGVPGAGVPGAGVQVETISPGDGRTFPKRGQTCVVHYTG MLEDGKKFDSSRDRNKPFKML
GKQEVIRGWEEGVAQMSVGQRAKLTISPDYAYGATGHPGIIPPHATLVFDVELLKLEGVPGAGVPGAGVPGAGVPGAGV
GVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGV
PGAGVPGAGVPGAGVPGAGVPGAGVPGAGVQVETISPGDGRTFPKRGQTCVVHYTG MLEDGKKFDSSRDRNKPFK
MLGKQEVIRGWEEGVAQMSVGQRAKLTISPDYAYGATGHPGIIPPHATLVFDVELLKLEGVPGAGVPGAGVPGAGVPGAGV
GAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGV
GVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVQVETISPGDGRTFPKRGQTCVVHYTG MLEDGKKFDSSRDRNK
PFKMLGKQEVIRGWEEGVAQMSVGQRAKLTISP DYAYGATGHPGIIPPHATLVFDVELLKLE

P6-5FA:

MRLLDTNRP FLRYVPGAGMGVQVETISPGDGRTFPKRGQTCVVHYTG MLEDGKKFDSSRDRNKPFKMLGKQEVIR
GWEEGVAQMSVGQRAKLTISPDYAYGATGHPGIIPPHATLVFDVELLKLEGVPGAGVPGAGVPGAGVPGAGVPGAG
VPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGV
GAGVPGAGVPGAGVPGAGVPGAGVQVETISPGDGRTFPKRGQTCVVHYTG MLEDGKKFDSSRDRNKPFKMLGKQ
EVIRGWEEGVAQMSVGQRAKLTISPDYAYGATGHPGIIPPHATLVFDVELLKLEGVPGAGVPGAGVPGAGVPGAGVPGAGV
GAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGV
GAGVPGAGVPGAGVPGAGVPGAGVPGAGVQVETISPGDGRTFPKRGQTCVVHYTG MLEDGKKFDSSRDRNKPFKML
GKQEVIRGWEEGVAQMSVGQRAKLTISPDYAYGATGHPGIIPPHATLVFDVELLKLEGVPGAGVPGAGVPGAGVPGAGV
GVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGV
PGAGVPGAGVPGAGVPGAGVPGAGVPGAGVQVETISPGDGRTFPKRGQTCVVHYTG MLEDGKKFDSSRDRNKPFK
MLGKQEVIRGWEEGVAQMSVGQRAKLTISPDYAYGATGHPGIIPPHATLVFDVELLKLEGVPGAGVPGAGVPGAGVPGAGV
GAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGV
GVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVQVETISPGDGRTFPKRGQTCVVHYTG MLEDGKKFDSSRDRNK
PFKMLGKQEVIRGWEEGVAQMSVGQRAKLTISP DYAYGATGHPGIIPPHATLVFDVELLKLE

P13-5FA:

MRLLDTNRPFLFYVPGAGMGVQVETISPGDGRTFPPKRGQTCVVHYTGMLEDGKKFDSSRDRNKPFKFMLGKQEVIR
GWEEGVAQMSVGQRAKLTISPDYAYGATGHPGIIPPHATLVFDVELLKLEGVPGAGVPGAGVPGAGVPGAGVPGAG
VPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGV
GAGVPGAGVPGAGVPGAGVPGAGVQVETISPGDGRTFPPKRGQTCVVHYTGMLEDGKKFDSSRDRNKPFKFMLGKQ
EVIRGWEEGVAQMSVGQRAKLTISPDYAYGATGHPGIIPPHATLVFDVELLKLEGVPGAGVPGAGVPGAGVPGAGVPGAGV
GAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGA
GVPAGVPGAGVPGAGVPGAGVPGAGVQVETISPGDGRTFPPKRGQTCVVHYTGMLEDGKKFDSSRDRNKPFKFML
GKQEVIRGWEEGVAQMSVGQRAKLTISPDYAYGATGHPGIIPPHATLVFDVELLKLEGVPGAGVPGAGVPGAGVPGAGV
GVPAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGV
PGAGVPGAGVPGAGVPGAGVPGAGVQVETISPGDGRTFPPKRGQTCVVHYTGMLEDGKKFDSSRDRNKPFKFML
MLGKQEVIRGWEEGVAQMSVGQRAKLTISPDYAYGATGHPGIIPPHATLVFDVELLKLEGVPGAGVPGAGVPGAGVPGAGV
GAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGAGVPGA
GVPAGVPGAGVPGAGVPGAGVPGAGVQVETISPGDGRTFPPKRGQTCVVHYTGMLEDGKKFDSSRDRNKPFK
FKFMLGKQEVIRGWEEGVAQMSVGQRAKLTISPDYAYGATGHPGIIPPHATLVFDVELLKLE

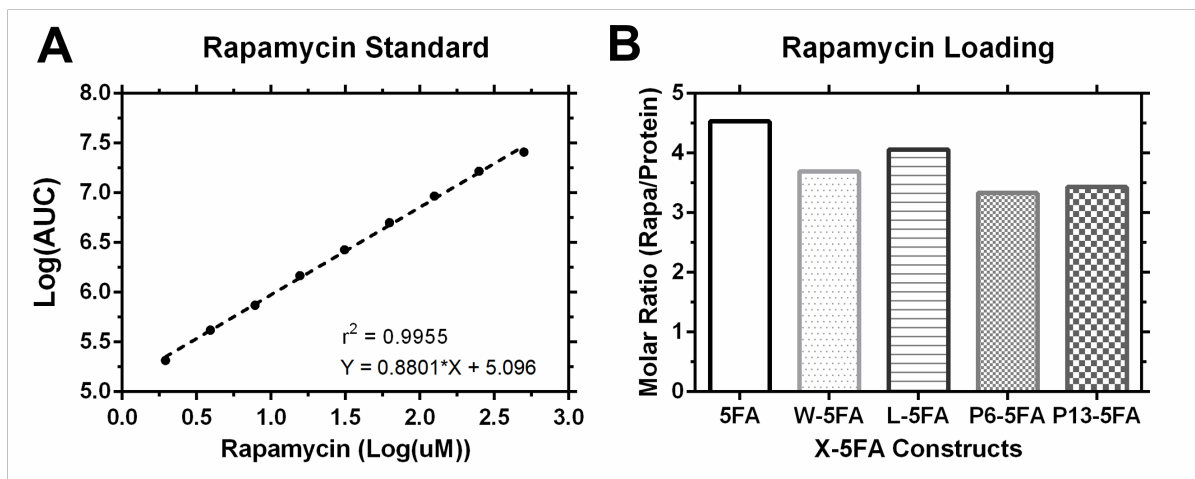
Supplemental Table S2: MALDI-TOF (M+XH)/X charge states

Charge state	5FA (Da)	W (Da)	L (Da)	P6 (Da)	P13 (Da)
(M+1H)/1	95527.9	97486.5	97657.5	97549.2	97852.3
(M+2H)/2	47684.5	48845.3	48976	48899	48904.7
(M+3H)/3	32178.1	32558.2	32606.2	32617	32485.6

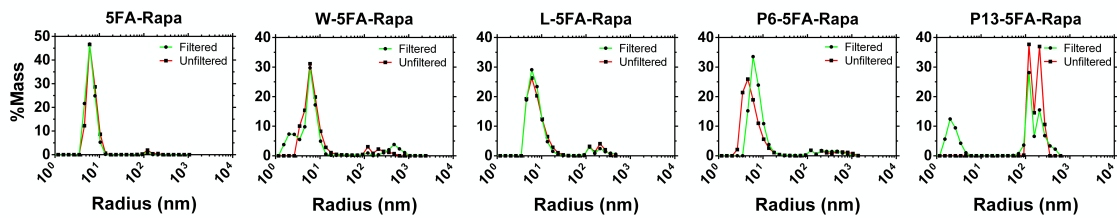
Supplemental Table S3: Molar Extinction Coefficients for ELPs as Evaluated by UV-Vis 280 nm

Sample	5FA	W-5FA	L-5FA	P6-5FA	P13-5FA
Extinction coefficient ($A_{280\text{ nm}} M^{-1}cm^{-1}$) ¹	50,100	61,100	51,590	51,590	51,590

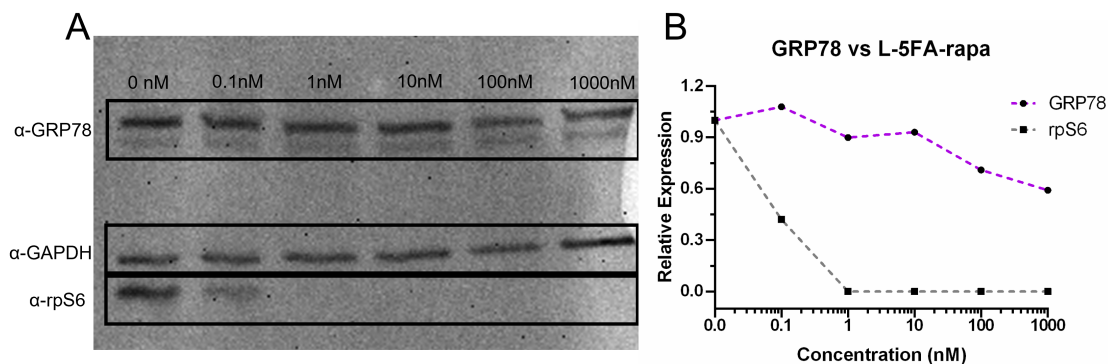
¹ Estimated using Pace et al. (1995)



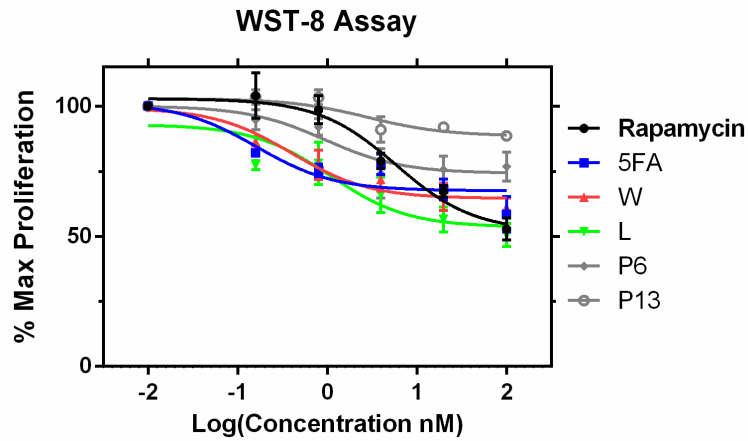
Supplemental Figures S1. Analytical Reverse Phase High Performance Liquid Chromatography was used to quantify rapamycin. RP-HPLC was performed as described in the methods. **A)** Using the AUCs observed after injection of standards, a linear calibration curve was developed. This curve was used to estimate the rapamycin concentrations within unknown samples, which were 725, 58, 211, 150, 266 uM for 5FA, W-5FA, L-5FA, P6-5FA, and P16-5FA respectively. The protein concentrations were estimated independently using UV-Vis spectroscopy at 280 nm using the molar extinction coefficients for each Hydra-ELP (**Supplemental Fig S3**), which were 160, 16, 52, 45, and 78 uM for 5FA, W-5FA, L-5FA, P6-5FA, and P16-5FA respectively. **B)** Using these, the loading ratio of Rapamycin per molecule was estimated at 4.5, 3.7, 4.1, 3.3, and 3.4 for 5FA, W-5FA, L-5FA, P6-5FA, and P16-5FA, respectively.



Supplemental Figure S2. Distribution of Hydrodynamic Radii of Hydra-ELPs complexed with Rapamycin. Rapamycin loaded Hydra-ELPs were characterized by dynamic light scattering with and without filtration to analyze the distribution of particle diameters. Rapamycin does not appear to significantly affect the radius of loaded Hydra-ELPs except for P13-5FA-Rapamycin, which shows a significant distribution between 10^2 nm and 10^3 nm. Filtration through a $0.02 \mu\text{m}$ filter does not appear to affect distribution except for P13-5FA-Rapa, which is split into two populations, one of which overlaps with the unfiltered population ($X > 100$ nm). A slightly smaller proportion of the unfiltered P13-5FA-Rapa population has a radius similar to filtered P13-5FA without rapamycin (**Fig. 4B**). For the remaining, there is a slight increase in radius to about 10 nm for all Hydra-ELPs, except for P13-5FA-Rapa, which remains consistent with filtered P13-5FA without rapamycin.



Supplemental Figure S3: GRP78 expression is attenuated after treatment with L-5FA-rapa in a concentration dependent manner. After samples from a concentration-based assay were imaged for rpS6 and GAPDH (**Fig. 6A**), the membrane was blocked with 5% BSA and incubated with anti-BiP/GRP78 rabbit mAb (Cell Signaling Technologies Inc., Boston, MA, Part #3177) overnight at 4°C . **A**) Images were acquired after incubation with anti-rabbit horse radish peroxidase and imaged on an iBright system. The relative expression of GRP78 and rpS6 at each concentration were normalized to GAPDH and quantified as a ratio to 0 nM. While rpS6 phosphorylation appears to significantly decrease with increasing L-5FA-rapa concentration (~ 1 nM), GRP78 is marginally affected beginning at (~ 100 nM). **B**) GRP78 appears to slightly decrease relative to GAPDH, but not directly proportional to a decrease in rpS6 phosphorylation. The data confirms reports in literature that suggest exposure to rapamycin affects mTORC1-dependent expression of GRP78. This effect appears to be at a range beyond the relevant therapeutic concentration for rpS6 phosphorylation.



Supplemental Figure S4: BT474 cells were treated with Hydra-ELPs loaded with rapamycin and assayed using a cell-viability assay. After reaching 30% confluence, cells were treated with Hydra-ELPs (5FA, W-5FA, L-5FA, P6-5FA, or P13-5FA) loaded with rapamycin at the following concentrations: 100, 20, 4, 0.8, or 0.16 nM. Cells were incubated for 48-hrs and assayed using the WST-8 reagent (Abcam, #228554) following the manufacturer protocol and read for absorbance 450 nm. Data is displayed as Log(concentration) vs. the percent of maximum possible proliferation. The absorbance of each sample was normalized to the difference between the untreated cells and cell-free blank wells. After fitting to a dose-response curve in Graphpad Prism, IC50 values were calculated for Rapamycin, 5FA-Rapa, W-5FA-Rapa, L-5FA-Rapa, P6-Rapa, and P13-5FA-Rapa at 5.8 (± 13.8) nM, 0.17 (± 0.70) nM, 0.49 (± 1.15) nM, 0.80 (± 4.7) nM, 0.94 (± 3.9) nM, and 2.8 (± 22.6) nM, respectively. Values indicate mean (\pm SD) for triplicates.