## Human umbilical cord mesenchymal stem cell-derived exosomes ameliorate liver steatosis by promoting fatty acid oxidation and reducing fatty acid synthesis

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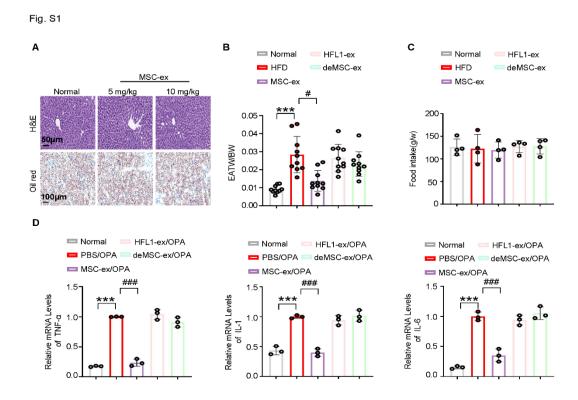


Fig. S1 MSC-ex inhibits weight gain of inguinal adipose tissue and inflammation in livers of HFD mice

(A) C57BL/6 mice were placed on high-fat diet (HFD, 40%) and administered 5 mg/kg and 10 mg/kg of MSC-ex *i.v.* From the 10th week to the 14th week of HFD feeding. Representative Images of Haematoxylin and eosin (H&E; upper; Scale bars, 50  $\mu$ m) and oil red O (bottom; Scale bars, 20  $\mu$ m) staining of liver sections. (B) Changes in the EATW/BW of mice (EATW/BW = relative weight of epididymal adipose tissue to body weight). (C) The food consumption was measured by weighing the food used. (D) Inflammatory transcription factor expression at mrna level (n=3 in each group). Data are represented as the mean  $\pm$  s.e.m. Statistical analyses was performed by a one-way ANOVA (panels B-D). \*\*\* P < 0.001 versus HFD group.



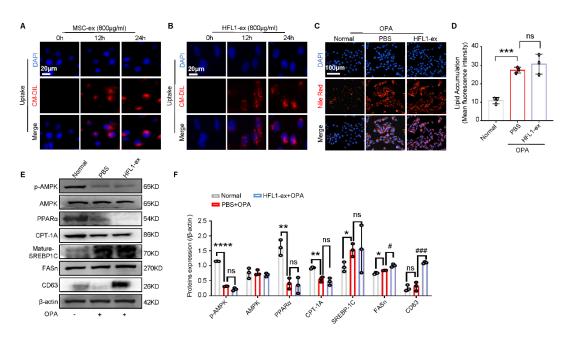


Fig. S2 HFL1-ex inactivates the AMPK signaling pathway and increases lipid accumulation (A-B) Representative fluorescent images of CM-DIL labeled MSC-ex (A) and HFL1-ex (B) in L02 cells at 0, 12, and 24 h. Scale bars, 100μm. (C-D) The intracellular lipid droplets in L02 cells subjected to oleate and palmitate (OPA) stimulation (2.0 mM, 2:1 ratio) in combination with HFL1-ex (800 μg/ml) or PBS treatment for 24 h were visualized by Nile red staining (C) and quantified by Image J for three random areas (D). Scale bars, 100 μm. (E-F) Immunoblotting of AMPK pathway proteins in L02 cells subjected to OPA stimulation (2.0 mM, 2:1 ratio) combined with HFL1-ex (800 μg/ml) or PBS treatment for 24 h (E) and quantification of the results (F). Data are represented as the mean ± s.e.m. Statistical analyses was performed by a one-way ANOVA (D and F). \* P <0.05, \*\*\* P <0.01, \*\*\*\* P <0.001, \*\*\*\*\* P <0.0001 versus normal group; # P <0.05, ### P <0.001 versus PBS group.

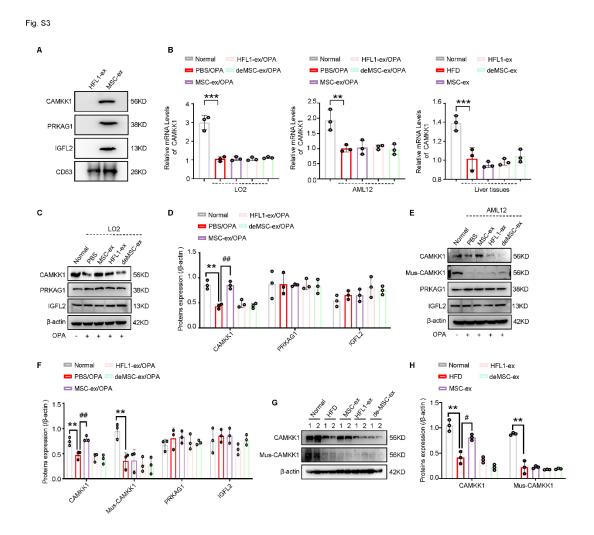


Fig. S3 Expression of CAMKK1 in HFL1-ex/MSC-ex, HFL1-ex/MSC-ex treated hepatocytes and livers

(A) CAMKK1, PRKAG1, and IGFL2 expression in MSC-ex and HFL1-ex were examined by immunoblotting. (B) QRT-PCR analyses of CAMKK1 mRNA expression (n=3 biological replicates per group). \*\* P < 0.01, \*\*\* P < 0.001 versus normal group. (C-D) The expression of CAMKK1, PRKAG1, and IGFL2 in L02 cells was detected by immunoblotting (C) and quantified (D). \*\* P < 0.01 versus normal group; ## P < 0.01 versus PBS group. (E-F) The expression of CAMKK1, PRKAG1, and IGFL2 in AML12 cells was detected by immunoblotting (E) and quantified (F). \*\* P < 0.01 versus normal group; ## P < 0.01 versus PBS group. (G-H) Immunoblotting analyses of CAMKK1 and

Mus-CAMKK1 proteins in mice placed on a HFD diet for 10 weeks followed by 10 mg/kg MSC-ex or 10 mg/kg HFL1-ex or MSC-ex-free conditional medium supernatant (deMSC-ex) and PBS treatment for 4 weeks ( $\bf G$ ) and quantification of the results ( $\bf H$ ).

\*\* P <0.01 versus normal chow diet group; # P <0.05 versus HFD group. Data are represented as the mean  $\pm$  s.e.m. Statistical analyses was performed by a one-way ANOVA ( $\bf B$ ,  $\bf D$ ,  $\bf F$ , and  $\bf H$ ).

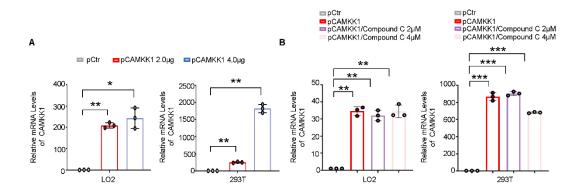


Fig. S4 CAMKK1 expression is not affected by Compound-C

(A) QRT-PCR analyses of CAMKK1 mRNA expression in L02 and 293T cells transfected with pCAMKK1 (2 μg and 4 μg) or empty plasmid (pCtr) and normalized to β-actin expression. n=3 biological replicates per group. (B) QRT-PCR analyses of CAMKK1 mRNA expression in L02 and 293T cells transfected with pCAMKK1 (4 μg) or pCtr with or without Compound-C (2 μM and 4 μM) treatment for 24 h. Expression was normalized to β-actin expression. n=3 biological replicates per group. Data are represented as the mean  $\pm$  s.e.m. Statistical analyses by unpaired two-tailed student's t-test. \* P <0.05, \*\* P <0.01, \*\*\* P <0.001 versus pCtr group.

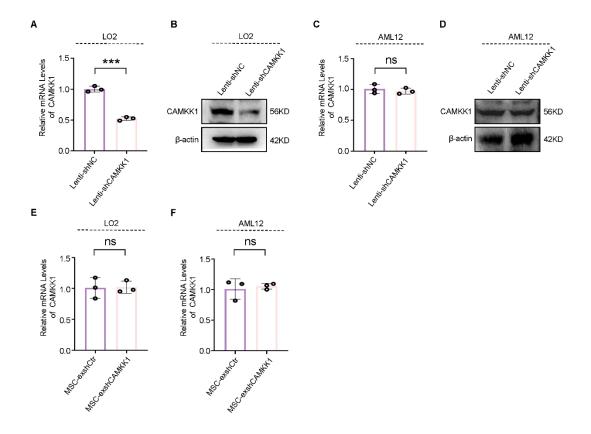


Fig. S5 Identification of CAMKK1 Knockdown in Lenti-shCAMKK1 transfected hepatocytes and MSC-ex<sup>shCAMKK1</sup> treated hepatocytes (A) QRT-PCR analyses of CAMKK1 mRNA expression in L02 cells treated with recombinant lentivirus (pLKO) (pLKO.1-GFP-Puro-shCAMKK1 or pLKO.1-Puro-shRNA, 15MOI) for 72 h. n=3 in each group; \*\*\* *P* <0.001 versus Lenti-shNC group. (B) The expression of CAMKK1 in L02 cells was detected by immunoblotting. (C) QRT-PCR analyses of CAMKK1 mRNA expression in AML12 cells treated with recombinant lentivirus (pLKO) (pLKO.1-GFP-Puro-shCAMKK1 or pLKO.1-Puro-shRNA, 15MOI) for 72 h. n=3 in each group. (D) The expression of CAMKK1 in AML12 cells was detected by immunoblotting. (E) QRT-PCR analyses of CAMKK1 mRNA expression in L02 cells treated with MSC-ex<sup>shCtr</sup> (800 μg/ml) or MSC-ex<sup>shCAMKK1</sup> (800 μg/ml) for 24 h. n=3 in each group. (F) QRT-PCR

analyses of CAMKK1 mRNA expression in AML12 cells treated with MSC-ex $^{shCtr}$  (800  $\mu g/ml$ ) or MSC-ex $^{shCAMKK1}$  (800  $\mu g/ml$ ) for 24 h. n=3 in each group. Data are represented as the mean  $\pm$  s.e.m. Statistical analyses by unpaired two-tailed student's t-test.

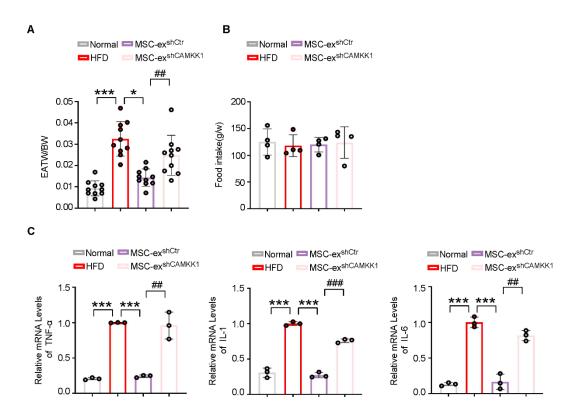


Fig. S6 MSC-ex<sup>shCAMKK1</sup> reverses MSC-ex<sup>shCtr</sup> decreased hepatic inflammation in the livers from HFD mice

(A) C57BL/6 mice were placed on a high-fat diet (HFD, 40%) and administered 10 mg/kg of MSC-ex<sup>shCtr</sup> or MSC-ex<sup>shCAMKK1</sup> *i.v.* from the 10th week to the 14th week of HFD feeding. As a control, the same volume of PBS was injected. Changes in the EATW/BW of mice (EATW/BW = relative weight of epididymal adipose tissue to body weight). (B) The food consumption was measured by weighing the food used. (C) Inflammatory transcription factor expression at mRNA level (n=3 in each group). Data are represented as the mean  $\pm$  s.e.m. Statistical analyses was performed by a one-way ANOVA (panels **A-C**). \* P < 0.05, \*\*\* P < 0.001 versus HFD group; ## P < 0.01, ### P < 0.001 versus MSC-ex<sup>shCtr</sup> (10 mg/kg) group.