

Supplemental Online Content

Yu AF, Moore ZR, Moskowitz CS, et al. Association of circulating cardiomyocyte cell-free DNA with cancer therapy–related cardiac dysfunction in patients undergoing treatment for *ERBB2*-positive breast cancer. *JAMA Cardiol*. Published online May 31, 2023. doi:10.1001/jamacardio.2023.1229

eMethods

eFigure. Levels of cardiomyocyte cfDNA and high-sensitivity troponin I

This supplemental material has been provided by the authors to give readers additional information about their work.

eMethods

cfDNA extraction (12278, 12278_C, 12649, 12773, 12828)

cfDNA was extracted from 0.6-1 mL plasma using the MagMAX Cell-Free DNA Isolation Kit (ThermoFisher catalog # A29319) on the KingFisher Flex Purification System (ThermoFisher) according to the manufacturer's protocol. Samples were eluted in 45-58 µL elution solution.

Detection of unmethylated FAM101A cfDNA fragments by digital droplet PCR (12278_B, 12528, 12649_B, 12808, 12828_C)

cfDNA underwent bisulfite conversion using the DNA Methylation-Gold kit (Zymo Research) following the manufacturer's instructions. Assays specific for the detection of unmethylated CpGs after bisulfite conversion at the *FAM101A* fragments were ordered through Bio-Rad.

Assay name	Forward sequence	Reverse sequence	Probe sequence	Fluorophore
FAM101A_MethyFam	TATGGTTTGGTAATTTATTTAGAG	AAATACAAATCCCACAAATAAA	AATGTATGGTCAAATGTAGTGTGGG	Fam
FAM101A_MethyHex			AAAAATACTCAACTTCCATCTACAATT	Hex

Cycling conditions were tested to ensure optimal annealing/extension temperature as well as optimal separation of positive from empty droplets. Optimization was done with a known positive control, an ultramer spiked into gDNA.

After PicoGreen quantification, <0.1-9.0 ng bisulfite treated cfDNA were combined with locus-specific primers, FAM- and HEX-labeled probes, MseI, and digital PCR Supermix for probes (no dUTP). All reactions were performed on a QX200 ddPCR system (Bio-Rad catalog # 1864001) and each sample was evaluated in technical duplicates. Reactions were partitioned into a median of ~18,500 droplets per well using the QX200 droplet generator. Emulsified PCRs were run on a 96-well thermal cycler using cycling conditions identified during the optimization step (95°C 10'; 40 cycles of 94°C 30' and 55°C 1'; 98°C 10'; 4°C hold). Plates were read and analyzed with the QuantaSoft software to assess the number of droplets positive for FAM-tagged probes, HEX-tagged probes, both, or neither.

