Supporting Information

N-acylated ciprofloxacin derivatives: synthesis and in vitro biological evaluation as antibacterial and anticancer agents

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Figure S1. Growth curve analysis of *S. aureus* ATCC 6538 at absorbance of 600 nm (OD600) with or without different concentrations of A. compound 5, B. compound 10, C. compound 11, D. CP for 18h. The growth curve data were plotted as average values with standard deviations of n = 3.

Table S1. MIC [ug/ml] values of compounds 5, 10, 11 and ciprofloxacin, used in biofilm eradication assay.

	Concentration [µg/ml]															
		4 MIC 2 MIC					MIC				1/2 MIC					
Bacteria strain	Compound															
	5	10	11	СР	5	10	11	СР	5	10	11	СР	5	10	11	СР
E. coli 25988	0.1	0.1	0.1	0.1	0.05	0.05	0.05	0.03	0.025	0.025	0.025	0.015	0.013	0.0125	0.0125	0.0075
S. aureus 6538	0.8	0.8	0.4	0.5	0.4	0.4	0.2	0.25	0.200	0.200	0.100	0.125	0.1	0.1	0.05	0.0625
P. aeruginosa 15442	0.8	0.4	0.4	0.2	0.4	0.2	0.2	0.12	0.200	0.100	0.100	0.060	0.1	0.05	0.05	0.03

Table S2. DNA gyrase (PDB ID: 5BTC [1]) and DNA topoisomerase IV (PDB ID: 3RAD [2]) binding data based on docking results for compounds 1-13 and CP.

Compound	DNA	gyrase	DNA topoisomerase IV			
	CS	BE	CS	BE		
		(kcal/mol)		(kcal/mol)		
1	433	-7.86	660	-6.7200		
2	360	-8.65	597	-7.3400		
3	454	-7.64	714	-6.7100		
4	409	-8.46	701	-6.8700		
5	343	-9.12	662	-7.9100		
6	340	-9.93	754	-8.2300		
7	350	-10.79	647	-9.0800		
8	408	-7.99	645	-7.4900		
9	399	-8.48	672	-7.6300		
10	418	-9.49	833	-8.5300		
11	381	-10.29	830	-9.2100		
12	392	-10.74	854	-10.0900		
13	171	-12.46	219	-12.2000		
СР	753	-7.26	698	-6.1700		

CS = number of members of the largest cluster calculated for 1000 docking runs using RMSD cutoff tolerance = 3 Å.

BE = binding free energy values estimated using AutoDock4 energy function for the representative ligand structure of the largest cluster.

Table S3. Trypan blue assay. The effect of compounds **3**, **15** and **21** on live cell number and viability in PC3 and HaCaT cells. Cells were incubated for 72 h with tested compounds used in their IC₅₀ concentrations, then cells were harvested, stained with trypan blue, and analyzed using cell counter. Data are expressed as the mean \pm SD. "-" control without compound, "Human metastatic prostate cancer (PC3), "Human immortal keratinocyte cell line from adult human skin (HaCaT).

		Compound	Cell number x 10 ⁶	Cell number	Viability (%)
				(% of control)	
		-	2.4 ± 0.90	100	98 ± 1.01
	PC3 ^a	3	0.04 ± 0.01	1.97	52 ± 2.40
Cancer cell line		15	0.3 ± 0.08	12.04	82 ± 2.64
	_	21	0.2 ± 0.03	6.22	78 ± 1.50
		-	1.5 ± 0.45	100	97 ± 2.01
Normal cell line	HaCaT ^b	3	0.2 ± 0.01	12.15	75 ± 2.23
		15	0.8 ± 0.10	48.38	90 ± 3.04
		21	0.3 ± 0.01	19.35	94 ± 4.01



Figure S2. Trypan blue assay. The effect of compounds **3**, **15** and **21** on live cell number and viability in HaCaT cells. Compounds were used in their IC₅₀ for PC3 cells, 2.02 μ M, 15.7 μ M and 4.8 μ M, respectively.

Table S4. The effect of compounds **3**, **15** and **21** on early and late apoptosis or necrosis in PC3 and HaCaT cells detected with Annexin V-FITC/PI by flow cytometry. Cells which were Annexin V:FITC positive and PI negative were identified as early apoptosis, and Annexin V:FITC and PI positive as late apoptosis or necrosis. Data are expressed as the mean \pm SD from 3 independent experiments.

		PC3 cells		
Compound	Live cells	Early apoptosis	Late apoptosis	Dead cells
control	92.03 ± 1.04	7.61 ± 1.05	0.21 ± 0.06	0.15 ± 0.04
3	20.27 ± 0.45	1.51 ± 0.04	12.12 ± 2.18	66.10 ± 6.31
15	61.21 ± 2.06	1.15 ± 0.10	16.49 ± 2.65	21.15 ± 3.29
21	2.59 ± 0.20	28.22 ± 0.98	16.28 ± 2.80	52.91 ± 3.99
		HaCaT cells		
Compound	Live cells	Early apoptosis	Late apoptosis	Dead cells
control	95.78 ± 0.77	2.30 ± 0.65	0.97 ± 0.22	0.95 ± 0.17
3	42.70 ± 2.81	10.07 ± 1.22	37.24 ± 2.27	9.99 ± 1.17
15	87.32 ± 2.20	11.63 ± 2.93	0.77 ± 0.05	0.3 ± 0.01
21	54.53 ± 3.34	10.52 ± 1.07	20.71 ± 4.34	14.24 ± 3.78



Figure S3. The effect of CP conjugates 3, 15, 21 on ROS production in PC3 and HaCaT cells. Cells were incubated with tested compounds at their IC₅₀ concentration for 2 and 12 h. Fluorescence intensity (FI) of the probe was measured by rhodamine (5 μ M). The results are expressed as mean ± SD of three experiments, each of them performed in triplicate. ***p \leq 0.0001, ** p \leq 0.001, *p \leq 0.01, as compared to the control.

¹H and ¹³C NMR spectra of synthesized Ciprofloxacin derivatives **1-21**.



























































S30



















S39























