

#### Supplemental Figure 1. Additional characterization of acute survival and fat mass loss in

**CLP/DCS mice**. Combined experiments (A) acute Kaplan-Meier survival for control and CLP/DCS. Control and CLP/DCS data subset is depicted in Figure 1A. (B-C) Control and CLP/DCS percent change from baseline in body weight (B) and fat mass (C). Data from mouse MRI on day 3 or 4 is represented as day 3. (D) Harvested wet weight, as change in percent of control, of fat pad normalized to baseline body weight in control and CLP/DCS day 4, 8, and 28. Data from harvested fat mass on days 27 or 28 is represented on graph as day 28. A = log-rank test. B-C = Two-way ANOVA with Bonferroni multiple comparisons (BMC). D = One-way ANOVA with BMC. Significance: \*p<0.05, \*\*/&p<0.01, \*\*\*/#p<0.001. A, n = 16-45. B, n = 16-45. C, n =12-14. Each n = one mouse.



**Supplemental Figure 2.** Role of CLP versus CLP/DCS in survival, body, lean, and fat mass. (A) Kaplan-Meier survival curve of control, cecal ligation and puncture (CLP), and CLP and daily chronic stress (CLP/DCS) mice to experiment endpoint. (B-D) Control, CLP, and CLP/DCS percent change from baseline in body mass (B), fat mass (C), and lean mass (D). (E) Harvested wet weight, as change in percent of control, of tibialis anterior (TA), gastrocnemius (GR), and fat pad normalized to baseline body weight, respectively, in control, CLP, and CLP/DCS groups. Data from harvested TA, GR, and fat mass on days 27 or 28 is represented on graph as day 28. Control and CLP/DCS data subset is also depicted in Figure 1A and C-D and Supplemental Figure 1B-D. A = log-rank test. B - D = Two-way ANOVA with BMC. E = One-way ANOVA with BMC. B, significance: \*p<0.05, &p<0.01, #p<0.001 versus control. C - D, significance: \*CLP or CLP/DCS versus control, %CLP versus CLP/CS; see supplemental tables 2 and 3 for all p-value comparisons in panel C and D, respectively. N = 12-16.









### Supplemental Figure 3. Raw muscle and fat masses from control and CLP/DCS mice across

**time.** (A) Baseline body mass between control and all CLP/DCS mice from combined experiments where data is represented in Figure 1, Supplemental Figure 1, and 2. (B-D) Control and CLP/DCS raw mass of harvested TA (B), GR (C), and fat (D). A = Student's unpaired t-test. B – D = One-way ANOVA with BMC. Significance is \*p<0.05, \*\*p<0.01, \*\*\*p<0.001. A, n = 16-45. B – D, n = 11-14. Each n = one mouse.



#### Supplemental Figure 4. Satellite cell reduction during sepsis does not affect mouse fat mass

or body weight. (A-F) Pax7-DTA vehicle or tamoxifen treated control or CLP/DCS mice (A) Kaplan-Meier survival curve, (B) percent change in weight from baseline, (C) percent change in fat mass from baseline, and percent change of controls of harvested wet weights normalized to initial body weight of (D) TA, (E) GR, or (F) fat pad. A = log-rank test. B - C = Two-way ANOVA with BMC where: \*significant vs. Control Vehicle and Control Tamoxifen and ^significant vs. Control Tamoxifen; n = 4-7. D - F = One-way ANOVA with BMC where significance is \*p<0.05, \*\*p<0.01, \*\*\*p<0.001; n = 3-5. Each n = one mouse.



### Supplemental Figure 5. Control and CLP/DCS reduction of satellite cell experiment raw

**muscle and fat masses.** (A) Baseline body mass between control and CLP/DCS mice treated with vehicle or tamoxifen. (B-D) Control and CLP/DCS vehicle or tamoxifen treated raw mass of harvested TA (B), GR (C), and fat (D). A - D = One-way ANOVA with BMC where significance is p<0.05, p<0.01, p<0.001. N = 4-7. Each n = one mouse.





Supplemental Figure 6. Satellite cells isolated from post sepsis mice do not have drastic alterations in their differentiation capacity. (A) Muscle stem cell (MuSC) differentiation in control and 3 days after CLP surgery (day 4). Quantification over time of MuSC (top) confluence, (middle) object count, and (bottom) object area fold changes. (B) Control and day 26 CLP/DCS proliferation measurements of MuSCs, with quantification of (top) confluence, (middle) object count, and (bottom) object area fold changes. (C-D) Myotube width and length from control or CLP/DCS differentiated MuSCs at day 4 or day 26. A - B = Two-way ANOVA with BMC. C - D = Student's unpaired t-test. Significance is p<0.05, p<0.01, p<0.01. N = 2-5. Each n = satellite cells isolated from a single mouse.



Supplemental Figure 7. The percentage of regenerating myofibers and number of centrally located nuclei are not altered post-second muscle injury in sepsis recovered mice. (A) The percentages of regenerating myofibers in vehicle and injury control or CLP/DCS mice. (B-C) Percentage of regenerating fibers containing 1, 2, or 3+ centrally located nuclei in control and CLP/DCS mice for (B) vehicle treated or (C) injury treated TA's. A - C = Student's unpaired t-test where all values are not significant (p>0.05). N = 3-6. Each n = one mouse.



#### Supplemental Figure 8. Differentially expressed genes in CLP/DCS day 4, 8, and 28 versus

**control.** Combined experimental (A) heatmaps depicting expression of the top ten upregulated and top ten downregulated differentially expressed genes (DEGs) for CLP/DCS day 4, 8, and 28 versus control. (B-C) Log2FC and negative log p-values for top upregulated (B) and downregulated (C) DEGs in CLP/DCS day 4, 8, and 28 versus control from A. (D-E) Venn diagram illustrating the overlapping upregulated (D) or downregulated (E) DEGs in CLP/DCS day 4, 8, and 28 versus control. Each n = pooled satellite cells isolated from 2-3 mice. Satellite cells harvested from mice on day 27 or 28 are labeled as day 28. Top DEGs were defined by: Day 4 = p-value < 0.05 and -1.75 > LOG2FC > 1.75; Day 8 = p-value < 0.05 and -2 > LOG2FC > 2; Day 28 = p-value < 0.05 and -1.25 > LOG2FC > 1.25.



### Supplemental Figure 9. Metabolite changes in gastrocnemius from CLP/DCS day 4, 8, and 28

**versus control.** Combined experimental (A-B) scaled intensity fold change and p-values for the top ten upregulated (A) and top ten downregulated (B) metabolites in whole GR muscle from CLP/DCS day 4, 8, and 28 versus control. (C-D) Venn diagram illustrating the overlapping upregulated (C) or downregulated (D) top altered metabolites from A in CLP/DCS day 4, 8, and 28 versus control. GRs that were harvested on day 27 or 28 are labeled as day 28. Top metabolites determined via Student's unpaired t-test with Welch's correction (Metabolon), p<0.05. N = 6 per group. Each n = one mouse.



3

4

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5

Day 28

#### Supplemental Figure 10. Longitudinal altered metabolic pathways in post-CLP/DCS mice

**gastrocnemius.** Combined experimental (A-C) top altered metabolic pathways enrichment ratios and negative log p-values from GR CLP/DCS day 4 (A), day 8 (B), and day 28 (C) versus control. (D) Venn diagram depicting overlapping top altered metabolic pathways. GRs that were harvested on day 27 or 28 are labeled as day 28. N = 6 per group. Each n = one mouse.

# Supplemental Tables

P-values	Control	Day 4	Day 8	Day 26- 28	
Control		0.4093	0.0021	0.4691	
Day 4	0.4093		0.0111	0.9023	
Day 8	0.0021	0.0111		0.0078	
Day 26-28	0.4691	0.9023	0.0078		
<b>Supplemental Table 1.</b> p-values from nonlinear fit pairwise comparisons of minimum feret diameter means from control, day 4, day 8, and day 28 myofibers from figure 1E.					

Time	Comparison	Adjusted p-value	
Day 4	Control vs. CLP	<0.0001	
	Control vs. CLP/DCS	<0.0001	
	CLP vs. CLP/DCS	>0.9999	
Day 7	Control vs. CLP	<0.0001	
	Control vs. CLP/DCS	<0.0001	
	CLP vs. CLP/DCS	>0.9999	
Day 10	Control vs. CLP	<0.0001	
	Control vs. CLP/DCS	<0.0001	
	CLP vs. CLP/DCS	>0.9999	
Day 13	Control vs. CLP	<0.0001	
	Control vs. CLP/DCS	<0.0001	
	CLP vs. CLP/DCS	0.7314	
Day 17	Control vs. CLP	<0.0001	
	Control vs. CLP/DCS	0.0002	
	CLP vs. CLP/DCS	0.0269	
Day 20	Control vs. CLP	<0.0001	
	Control vs. CLP/DCS	0.0005	
	CLP vs. CLP/DCS	0.1302	
Day 23	Control vs. CLP	<0.0001	
	Control vs. CLP/DCS	0.0065	
	CLP vs. CLP/DCS	0.0809	
Day 26	Control vs. CLP	0.0005	
	Control vs. CLP/DCS	0.0361	
	CLP vs. CLP/DCS	0.5489	
<b>Supplemental Table 2.</b> Longitudinal fat mass p-values from two- way ANOVA with BMC between control, CLP, and CLP/DCS from supplementary figure 2C.			

Time	Comparison	Adjusted p-value	
Day 4	Control vs. CLP	<0.0001	
	Control vs. CLP/DCS	<0.0001	
	CLP vs. CLP/DCS	>0.9999	
Day 7	Control vs. CLP	0.0002	
	Control vs. CLP/DCS	<0.0001	
	CLP vs. CLP/DCS	0.0896	
Day 10	Control vs. CLP	0.4154	
	Control vs. CLP/DCS	0.0005	
	CLP vs. CLP/DCS	0.0499	
Day 13	Control vs. CLP	>0.9999	
	Control vs. CLP/DCS	0.1933	
	CLP vs. CLP/DCS	0.9850	
Day 17	Control vs. CLP	>0.9999	
	Control vs. CLP/DCS	>0.9999	
	CLP vs. CLP/DCS	>0.9999	
Day 20	Control vs. CLP	>0.9999	
	Control vs. CLP/DCS	>0.9999	
	CLP vs. CLP/DCS	0.5624	
Day 23	Control vs. CLP	>0.9999	
	Control vs. CLP/DCS	>0.9999	
	CLP vs. CLP/DCS	>0.9999	
Day 26	Control vs. CLP	0.5578	
	Control vs. CLP/DCS	>0.9999	
	CLP vs. CLP/DCS	>0.9999	
<b>Supplemental Table 3.</b> Longitudinal lean mass p-values from two-way ANOVA with BMC between control, CLP, and CLP/DCS from supplementary figure 2D.			

Time	Comparison	Adjusted p-value		
Day 4	Control Vehicle vs. Control Tamoxifen	>0.9999		
	Control Vehicle vs. CLP/DCS Vehicle	0.0034		
	Control Vehicle vs. CLP/DCS Tamoxifen	<0.0001		
	Control Tamoxifen vs. CLP/DCS Vehicle	0.0007		
	Control Tamoxifen vs. CLP/DCS Tamoxifen	<0.0001		
	CLP/DCS Vehicle vs. CLP/DCS Tamoxifen	0.6269		
Day 7	Control Vehicle vs. Control Tamoxifen	>0.9999		
	Control Vehicle vs. CLP/DCS Vehicle	0.0296		
	Control Vehicle vs. CLP/DCS Tamoxifen	0.0002		
	Control Tamoxifen vs. CLP/DCS Vehicle	0.0171		
	Control Tamoxifen vs. CLP/DCS Tamoxifen	0.0001		
	CLP/DCS Vehicle vs. CLP/DCS Tamoxifen	0.9821		
Day 10	Control Vehicle vs. Control Tamoxifen	>0.9999		
	Control Vehicle vs. CLP/DCS Vehicle	0.9318		
	Control Vehicle vs. CLP/DCS Tamoxifen	0.0011		
	Control Tamoxifen vs. CLP/DCS Vehicle	>0.9999		
	Control Tamoxifen vs. CLP/DCS Tamoxifen	0.0052		
	CLP/DCS Vehicle vs. CLP/DCS Tamoxifen	0.0629		
Day 14	Control Vehicle vs. Control Tamoxifen	>0.9999		
	Control Vehicle vs. CLP/DCS Vehicle	>0.9999		
	Control Vehicle vs. CLP/DCS Tamoxifen	0.0015		
	Control Tamoxifen vs. CLP/DCS Vehicle	>0.9999		
	Control Tamoxifen vs. CLP/DCS Tamoxifen	0.0080		
	CLP/DCS Vehicle vs. CLP/DCS Tamoxifen	0.0665		
Day 19	Control Vehicle vs. Control Tamoxifen	>0.9999		
	Control Vehicle vs. CLP/DCS Vehicle	0.4899		
	Control Vehicle vs. CLP/DCS Tamoxifen	0.0032		
	Control Tamoxifen vs. CLP/DCS Vehicle	>0.9999		
	Control Tamoxifen vs. CLP/DCS Tamoxifen	0.0231		
	CLP/DCS Vehicle vs. CLP/DCS Tamoxifen	0.3374		
Day 27	Control Vehicle vs. Control Tamoxifen	>0.9999		
	Control Vehicle vs. CLP/DCS Vehicle	>0.9999		
	Control Vehicle vs. CLP/DCS Tamoxifen	0.0277		
	Control Tamoxifen vs. CLP/DCS Vehicle	>0.9999		
	Control Tamoxifen vs. CLP/DCS Tamoxifen	0.0884		
	CLP/DCS Vehicle vs. CLP/DCS Tamoxifen	0.2441		
<b>Supplemental Table 4.</b> Longitudinal p-values from two-way ANOVA with BMC of lean mass between control vehicle, control tamoxifen, CLP/DCS vehicle, and CLP/DCS tamoxifen from figure 2B.				

P-values	Control Vehicle	Control Tamoxifen	CLP/DCS Vehicle	CLP/DCS Tamoxifen	
Control Vehicle		0.2448	0.3057	0.0066	
Control Tamoxifen	0.2448		0.0191	0.0558	
CLP/DCS Vehicle	0.3057	0.0191		0.0001	
CLP/DCS Tamoxifen	0.0066	0.0558	0.0001		
<b>Supplemental Table 5</b> . p-values from nonlinear fit pairwise comparisons of minimum feret diameter means from control vehicle, control tamoxifen, CLP/DCS vehicle, and CLP/DCS tamoxifen myofibers from figure 2C.					

P-values	Control Vehicle	Control Injured	CLP/DCS Vehicle	CLP/DCS Injured
Control Vehicle		0.3072	0.7802	0.0003
Control Injured	0.3072		0.5016	0.0029
CLP/DCS Vehicle	0.7802	0.5016		0.0015
CLP/DCS Injured	0.0003	0.0029	0.0015	
<b>Supplemental Table 6</b> . p-values from nonlinear fit pairwise comparisons of minimum feret diameter means from control vehicle, control injured, CLP/DCS vehicle, and CLP/DCS injured myofibers from figure 4B.				

Ingenuity Canonical Pathways	-log(p-value)	Ratio	z-score	
Polyamine Regulation in Colon Cancer	23	0.403	#NUM!	
FAT10 Signaling Pathway	22.8	0.429	#NUM!	
BAG2 Signaling Pathway	19.2	0.298	#NUM!	
Protein Ubiquitination Pathway	17.1	0.142	#NUM!	
Inhibition of ARE-Mediated mRNA Degradation Pathway	12.9	0.161	1	
Huntington's Disease Signaling	11.4	0.114	#NUM!	
Hepatic Fibrosis / Hepatic Stellate Cell Activation	10.2	0.129	#NUM!	
p38 MAPK Signaling	6.32	0.127	1.941	
Atherosclerosis Signaling	5.77	0.115	#NUM!	
Actin Cytoskeleton Signaling	5.65	0.0857	1.069	
Calcium Signaling	5.33	0.088	0.707	
ILK Signaling	5.28	0.0909	0.775	
Dilated Cardiomyopathy Signaling Pathway	5.14	0.103	0.277	
Glycogen Biosynthesis II (from UDP-D-Glucose)	4.81	0.571	1	
Agranulocyte Adhesion and Diapedesis	4.27	0.0794	#NUM!	
PAK Signaling	4.19	0.102	0.333	
Neuroprotective Role of THOP1 in Alzheimer's Disease	4.19	0.102	-1.667	
Glucocorticoid Receptor Signaling	4.13	0.0551	#NUM!	
Tumor Microenvironment Pathway	4.1	0.0838	-1.069	
ERK5 Signaling	3.97	0.125	1.414	
ERK/MAPK Signaling	3.75	0.0748	1.387	
Cholecystokinin/Gastrin-mediated Signaling	3.52	0.0924	0.333	
Sertoli Cell-Sertoli Cell Junction Signaling	3.43	0.0728	#NUM!	
HIF1α Signaling	3.38	0.0721	0	
Cardiac Hypertrophy Signaling	3.3	0.0659	0.535	
<b>Supplemental Table 7.</b> Top 25 altered pathways from IPA analysis of RNA sequencing data from CLP/DCS day 4 post-septic satellite cells versus control (DEGs: p-value < 0.05 and -1.75 > LOG2FC > 1.75).				

Ingenuity Canonical Pathways	-log(p-value)	Ratio	z-score	
Oxidative Phosphorylation	49.7	0.536	7.421	
Mitochondrial Dysfunction	48.6	0.409	#NUM!	
Sirtuin Signaling Pathway	22.3	0.202	-3.162	
Estrogen Receptor Signaling	17.7	0.156	2.263	
Glucocorticoid Receptor Signaling	16.2	0.129	#NUM!	
Calcium Signaling	9.93	0.157	3.441	
Dilated Cardiomyopathy Signaling Pathway	8.92	0.178	-1.606	
TCA Cycle II (Eukaryotic)	8.76	0.458	3.317	
Glycolysis I	7.31	0.4	3.162	
Gluconeogenesis I	7.31	0.4	3.162	
Protein Ubiquitination Pathway	7.18	0.124	#NUM!	
ILK Signaling	6.69	0.136	1.877	
Hepatic Fibrosis / Hepatic Stellate Cell Activation	6.32	0.134	#NUM!	
Huntington's Disease Signaling	6	0.114	1.134	
Cellular Effects of Sildenafil (Viagra)	5.58	0.141	#NUM!	
Actin Cytoskeleton Signaling	5.36	0.114	3.273	
HIF1α Signaling	5.24	0.12	2.041	
Agranulocyte Adhesion and Diapedesis	4.55	0.112	#NUM!	
Role of NFAT in Cardiac Hypertrophy	4.35	0.109	3.273	
Amyotrophic Lateral Sclerosis Signaling	4.33	0.139	0.905	
Netrin Signaling	4.16	0.167	2.309	
Cardiac Hypertrophy Signaling	4.08	0.101	3.13	
Hypoxia Signaling in the Cardiovascular System	4.04	0.162	0.447	
Aspartate Degradation II	3.92	0.571	2	
Integrin Signaling	3.69	0.103	3.441	
<b>Supplemental Table 8.</b> Top 25 altered pathways from IPA analysis of RNA sequencing data from CLP/DCS day 8 post-septic satellite cells versus control (DEGs: p-value < 0.05 and -2 > LOG2FC > 2).				

Ingenuity Canonical Pathways	-log(p-value)	Ratio	z-score	
Oxidative Phosphorylation	59.1	0.527	7.353	
Mitochondrial Dysfunction	58.7	0.398	#NUM!	
Sirtuin Signaling Pathway	27.3	0.185	-4.11	
Estrogen Receptor Signaling	20.4	0.134	2.6	
Calcium Signaling	12.7	0.144	3.71	
Glucocorticoid Receptor Signaling	11.2	0.0861	#NUM!	
Glycolysis I	10.5	0.44	3.317	
Gluconeogenesis I	10.5	0.44	3.317	
TCA Cycle II (Eukaryotic)	9.3	0.417	3.162	
Dilated Cardiomyopathy Signaling Pathway	8.86	0.144	-0.535	
Role of NFAT in Cardiac Hypertrophy	8.24	0.114	4.264	
Dopamine-DARPP32 Feedback in cAMP Signaling	7.18	0.116	2.138	
Cellular Effects of Sildenafil (Viagra)	5.83	0.114	#NUM!	
Cardiac β-adrenergic Signaling	5.51	0.103	-0.577	
Synaptic Long Term Potentiation	5.32	0.116	2.138	
Cardiac Hypertrophy Signaling	5.19	0.0853	3.638	
nNOS Signaling in Skeletal Muscle Cells	5.1	0.188	#NUM!	
Senescence Pathway	4.72	0.0774	3.3	
Glycogen Biosynthesis II (from UDP-D-Glucose)	4.65	0.571	#NUM!	
3-phosphoinositide Degradation	4.47	0.0904	2.887	
HIF1α Signaling	4.45	0.0865	3.771	
White Adipose Tissue Browning Pathway	4.4	0.103	3.207	
D-myo-inositol-5-phosphate Metabolism	4.36	0.0885	2.887	
Protein Kinase A Signaling	4.33	0.0672	1.279	
D-myo-inositol (1,4,5,6)-Tetrakisphosphate Biosynthesis	4.27	0.0909	2.714	
<b>Supplemental Table 9.</b> Top 25 altered pathways from IPA analysis of RNA sequencing data from day 28 post-septic satellite cells versus control (DEGs: p-value < 0.05 and - 1.25 > LOG2FC > 1.25).				