Appendix: Mo et al

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Appendix Table S1: Sequences of primers used in this study

Primers for cDNA cloning

Target	**Sequence (5' - 3')**
mRpL4 Forward	ATGTTGAACAATATTTTAAA
mRpL4 Reverse	CTAGACTTGATCCAGCTTAA
wap Forward (HA tag)	ATGtacccttacgttcctgattacgctagcctcTCCTCGAC CGCCGGAAA
wap Reverse	TTAGACCCGCAGGATCTC
human-mRpL4 Forward	ATGCTGCAGTTCGTCCGGGC
human-mRpL4 Reverse	CTAACAGCGGAGCCTGCACA
zebrafish-mRpL4 Forward	CGCTATCACGACCTTCCTGAC
zebrafish-mRpL4 Reverse	CCACTGAAGTCCATGCTGTTC
zebrafish-hey1 Forward	GGATACGCGCTGCTAAACTGTC
zebrafish-hey1 Reverse	TGGGACAAGCACAGTCGTTC

Primers for ChIP qPCR

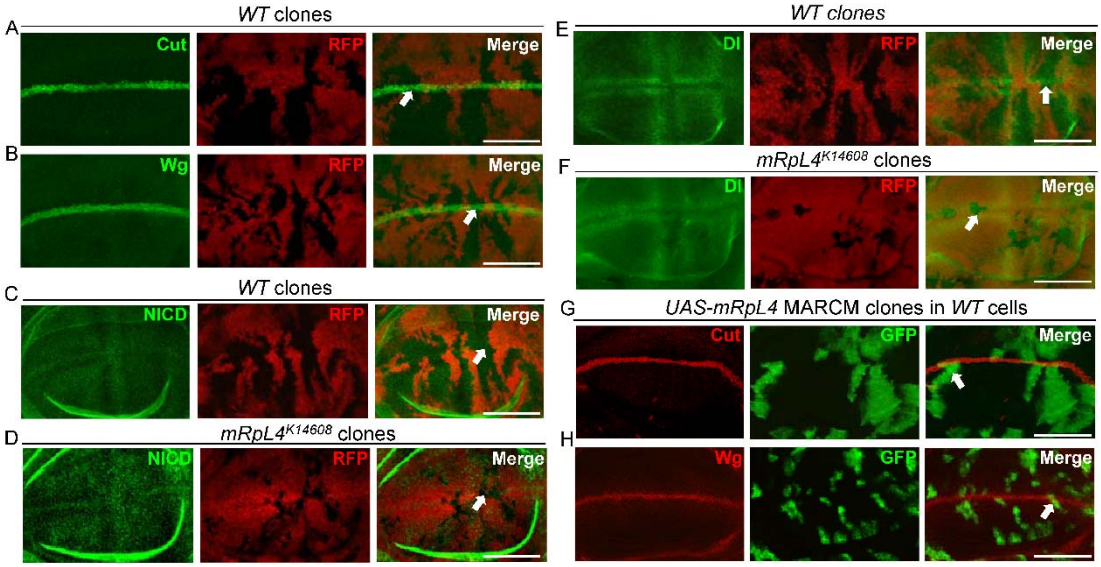
Target	**Sequence (5' - 3')**
mγ-mβ igr Forward	GGAGTTGAGGAGTTGGTCG
mγ-mβ igr Reverse	ATAAGTGTGGTTGGGTGCCT
mβ-tr Forward	AGAAGTGAGCAGCAGCCATC
mβ-tr Reverse	GCTGGACTTGAAACCGCACC
mβ-enh Forward	AGAGGTCTGTGCGACTTGG
mβ-enh Reverse	GGATGGAAGGCATGTGCT
mβ-mα igr Forward	AAGCCAGTGGACTCTGCTCT
mβ-mα igr Reverse	TGATCTCCAAGCGGAGTATG
mα-tr Forward	GCAGGAGGACGAGGAGGATG
mα-tr Reverse	GATCCTGGAATTGCATGGAG
m2-m3 igr Forward	GCGCGTATTTCCCAAATAAA
m2-m3 igr Reverse	GATTGTACGTGCATGGGAAA
m3-enh Forward	ACACACACAAACACCCATCC
m3-enh Reverse	CGAGGCAGTAGCCTATGTGA
m3-tr Forward	CGTCTGCAGCTCAATTAGTC
m3-tr Reverse	AGCCCACCCACCTCAACCAG
Cut enh Forward1	CCGATAAATCGGGGTTTTGGA
Cut enh Reverse1	ACGCCTGTTACCATAGTCGC
Cut enh Forward2	TTTGCCGACGTGAGAAACAC
Cut enh Reverse2	TCCTCCTTTTTCATACTTCATTCAC
Wg enh Forward1	CCGTACTTTTTCCGGACCAC
Wg enh Reverse1	CGCTAAGCCCCTGTTTGGT
Wg enh Forward2	TGCTCCTCTGACCACGATCC

Wg enh Reverse2	CCTTAAGCCGCCTCGACTG
Wg enh Forward3	CGCCGAAACATTTCGAGAAACA
Wg enh Reverse3	CGTGACGCACAAGACCTTTA
Wg enh Forward4	GCATTGCGCAACGTTCGGT
Wg enh Reverse4	AAATGTTTATTGACAGGCAGCGG
Vg enh Forward1	CCTCTCCCGCTTTTTGCTAAC
Vg enh Reverse1	ACTGGACACTGGAAACCAGC

Primers for RT-qPCR assay

Target	**Sequence (5' - 3')**
zebrafish-mRpL4 Forward	TCAGATCTTCCAGTTGTGCG
zebrafish-mRpL4 Reverse	GTTTGAAGTTCCGTTGCCAG
zebrafish-notch1a Forward	CGACACCACACACACATGCT
zebrafish-notch1a Reverse	AGTGGCAGTTGTAGGTGTTG
zebrafish-hey1 Forward	GCCTTTGAGAAACAGGGCTCAG
zebrafish-hey1 Reverse	AGCGTGAGCATCAAAGTAACCT
zebrafish-her4.1 Forward	AGGAGAACTGAACACAAGACAC
zebrafish-her4.1 Reverse	TGCTGTTGATTCGCTCTCG
zebrafish-her6 Forward	GGCTTCGGAACACAGAAAG
zebrafish-her6 Reverse	TGACCCAAGCTTTCGTTGA
zebrafish-her15.1 Forward	TCGCTCTGCTCAGAGAAACA
zebrafish-her15.1 Reverse	ACCACTGGCTTTCGCAA
zebrafish-β-actin Forward	ATCTTCACTCCCCTTGTTCAC
zebrafish-β-actin Reverse	TCATCTCCAGCAAAACCGG

Appendix Figure S1. Expression pattern of Notch signaling components.



A Representative images of wing imaginal disc (n > 10 wing discs) bearing wild type clones stained for Cut.

B Representative images of wing imaginal disc (n > 10 wing discs) bearing wild type clones stained for Wg.

C Representative images of wing imaginal disc (n > 10 wing discs) bearing wild type clones stained for NICD.

D Representative images of wing imaginal disc (n > 15 wing discs) bearing *mRpL4*^{K14608} clones stained for NICD.

E Representative images of wing imaginal disc (n > 15 wing discs) bearing wild type clones stained for Dl.

F Representative images of wing imaginal disc (n > 15 wing discs) bearing *mRpL4*^{K14608} clones stained for Dl.

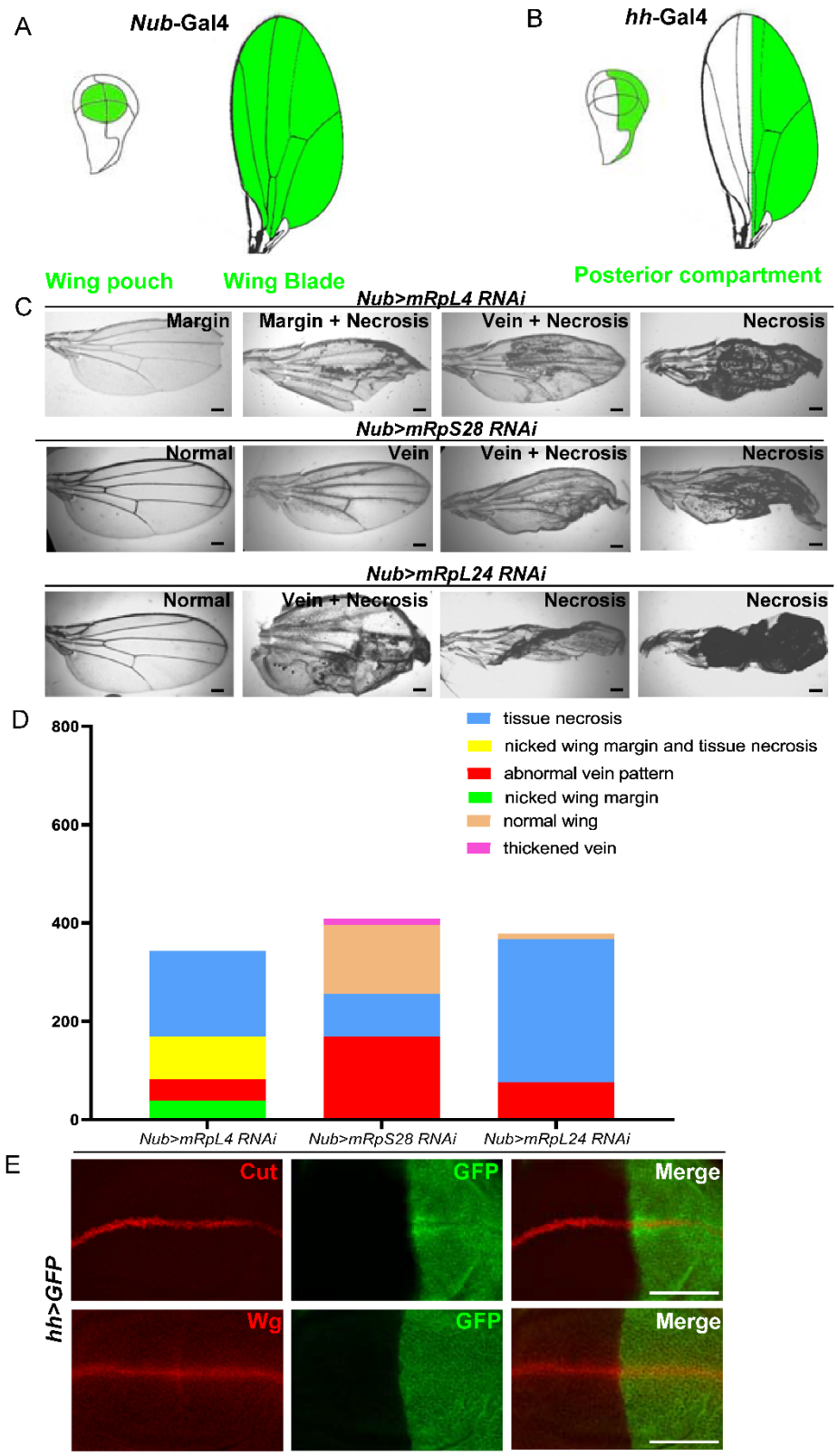
G Representative images of wild type wing imaginal disc (n > 10 wing discs) bearing *UAS-mRpL4* MARCM clones stained for Cut.

H Representative images of wild type wing imaginal disc (n > 10 wing discs) bearing

UAS-mRpL4 MARCM clones stained for Wg.

Data information: Clones are marked by absence of RFP (*A-F*), while the MARCM clones are marked by GFP (*G* and *H*). Representative clones are marked by white arrows. Scale bars = 100 μm.

Appendix Figure S2. Effects of MRPs knock-down.



A, B Cartoons showing the areas where *Nub*-Gal4 (*A*) and *hh*-Gal4 (*B*) are active.

C Representative images of adult wings (n > 15 wings) showing diverse defects upon

knock-down of *mRpL4*, *mRpS28* and *mRpL24* by *Nub*-Gal4 driven RNAi.

D Quantification of adult wing defects. mRpL4 RNAi led to wing margin nicks in 11.08% of the wings, and wing margin nicks along with tissue necrosis in 25.66% of the wings (n = 343). After knock-down mRpS28 (n = 408) or mRpL24 (n = 378) by RNAi, wing margin nicking defects were not observed.

E Representative images of wing imaginal disc (n > 15 wing discs) expressing GFP under the control of *hh*-Gal4 stained for Cut and Wg.

Data information: Scale bars = 100 μm.