Supplementary Information for End-to-end protein-ligand complex structure generation with diffusion-based generative model

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The physical accuracy of the generated structures

To validate the physical accuracy of the generated structures, we calculated the distances between the C_{α} atoms of adjacent residues and compared their distribution with the expected distribution calculated from the experimental structures (Figure S1). All structures generated by DPL on the PDBbind test set (64 structures for each of the 207 complexes) were used for the analysis. Figure S1 shows that the distribution of distances was concentrated around the peak of the expected distribution within a range of ± 0.5 Å. This indicates that the structures generated by DPL are physically plausible, suggesting that a reasonable result could be obtained by building an all-atom model and optimizing its structure using physics-based methods such as Rosetta [1].



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Dependence of protein structure reproducibility on training data

Figure S2 illustrates how the amount of training data with similar protein structures (TM-score > 0.5) affects the reproducibility of protein structures. This indicates that the reproducibility increases with the number of related training data, suggesting some overfitting to the protein structures used in the training. However, it also shows that DPL can generalize to proteins with little or no related training data. Extensive use of PDB-registered structures, including apo protein structures, for training would improve the reproducibility of protein structures.



Enhancing training data through random cropping

Randomly cropping partial structures from a large complex is a common method used to enhance the training data in protein structure prediction [2, 3]. For example, the contiguous cropping algorithm described in AlphaFold-Multimer [3] sequentially and randomly crops contiguous residues from each protein chain to fit within a given budget. Although this algorithm is designed for proteins and does not consider the ligand, it can be adapted to protein-ligand complexes by preferentially keeping protein structures in contact with the ligand. An example of such a procedure is shown in Algorithm S1. For this adaptation, contiguous residues are chosen to maximize contact with the ligand, rather than being chosen uniformly at random. One way to define the contact for each chain, $c_k \in \{0,1\}^{n_k}$, is to check if any atom of the residue is within 4 Å of any atom of the ligand. In addition, while the original algorithm randomly shuffles the chains before cropping to avoid bias, it is also possible to reorder them based on contacts with the ligand.

Algorithm S1 Random cropping of a complex structure. Adapted from the contiguous cropping algorithm described in AlphaFold-Multimer [3].

procedure CROP_COMPLEX(Set of chain lengths $\{n_k\}$, Set of chain contacts $\{c_k\}$, The total protein residue budget $N_{\rm res}$) Initialize $n_{\text{added}} = 0$ Initialize $n_{\text{remaining}} = N_{\text{res}}$ for k in $0, 1, \ldots N_{\text{chains}} - 1$ do Compute $n_{\text{remaining}} = n_{\text{remaining}} - n_k$ Compute crop_size_max = minimum $(N_{res} - n_{added}, n_k)$ Compute crop_size_min = minimum $(n_k, maximum(0, N_{res} - (n_{added} + n_{remaining})))$ Sample crop_size ~ uniform(crop_size_min, crop_size_max + 1) Compute $n_{\text{added}} = n_{\text{added}} + \text{crop}_{\text{size}}$ Compute crop_start = $\operatorname{argmax}_{s} \sum_{i=0}^{\operatorname{crop}_size} c_{ks+i}$ Initialize $\boldsymbol{m}_{\boldsymbol{k}} = 0$ Compute keep = $[crop_start, ..., crop_start + crop_size]$ Compute $m_{k \text{keep}} = 1$ end for return $\{m_k\}$ end procedure

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