

1 Mapping human norovirus antigens during infection reveals the breadth of the humoral
2 immune response

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5 Timothy Palzkill

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8 **Supplementary Figure 1.** Schematic of HuNoV genomic phage library construction.

9 **Supplementary Figure 2.** Deep sequencing analysis of the inserts present in the NoV Jun-Fos
10 library before and after affinity selection versus day 14-30 post-infection sera.

11 **Supplementary Figure 3.** In-frame fraction analysis of NoV Jun-Fos library versus post-
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13 **Supplementary Figure 4.** Alignment of peptides with high coverage after two rounds of affinity
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20 Jun-Fos library before and after affinity selection versus GII.4 antisera.

21 **Supplementary Figure 8.** Dendrogram of GII.4 HOV epitope profiles.

22 **Supplementary Figure 9.** Alignment of epitope sequences of GI.1 Norwalk to that of GII.4
23 HOV.

24 **Supplementary Table 1.** List of epitopes from all the sera identified after two rounds of affinity
25 selections with the NoV Jun-Fos library.

26 **Supplementary Table 2.** List of epitopes from all the sera identified after two rounds of affinity
27 selections with the GII.4 HOV Jun-Fos library.

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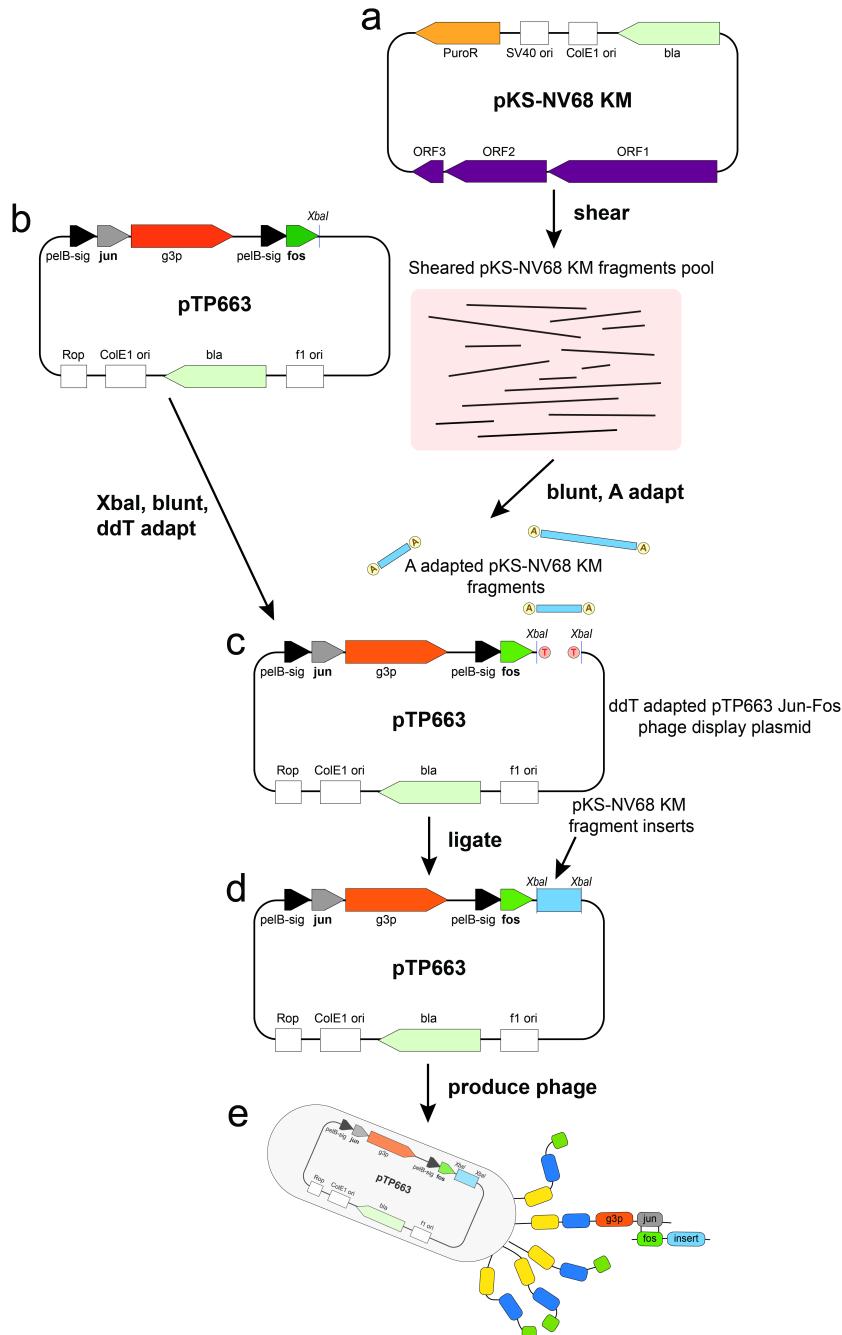
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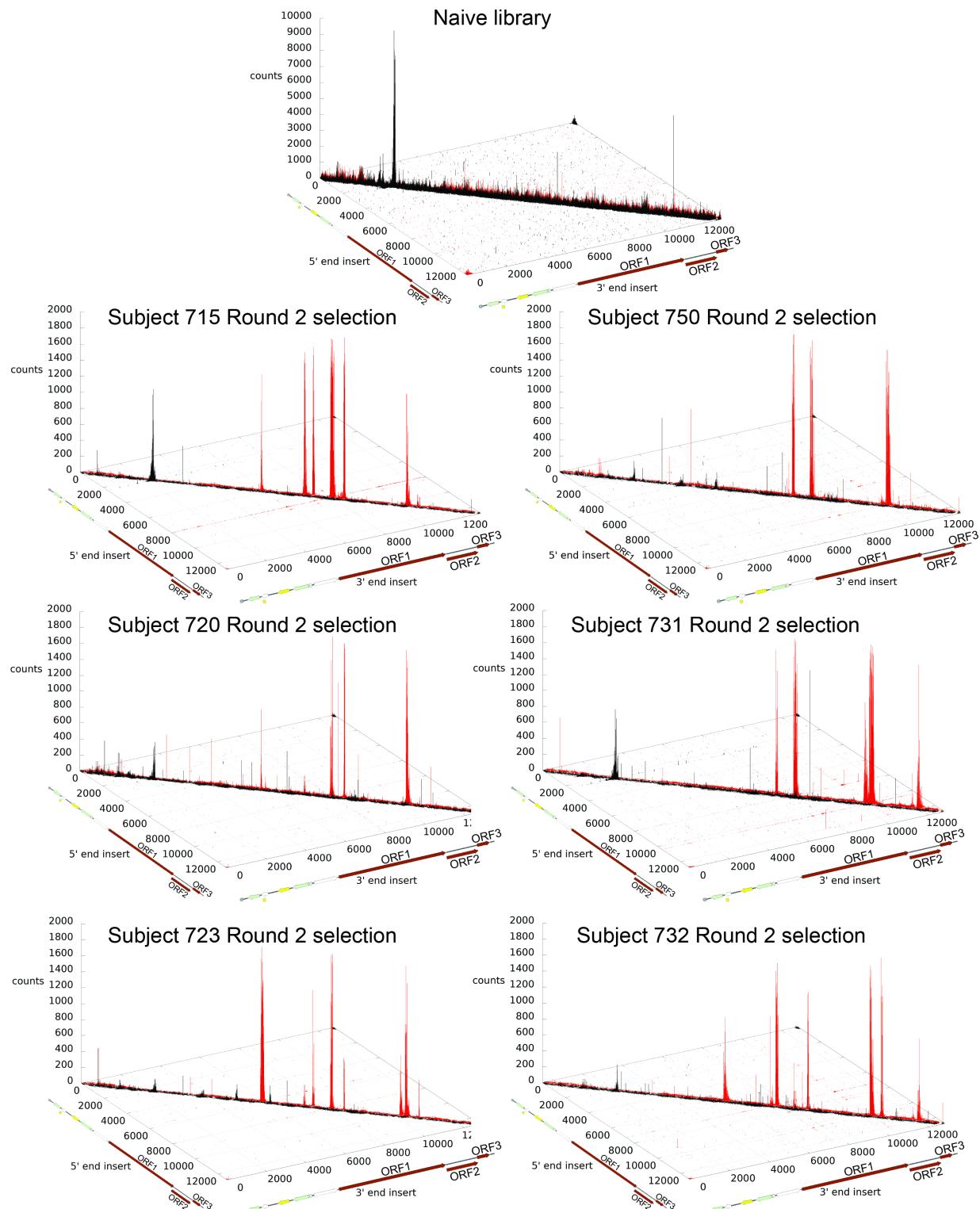
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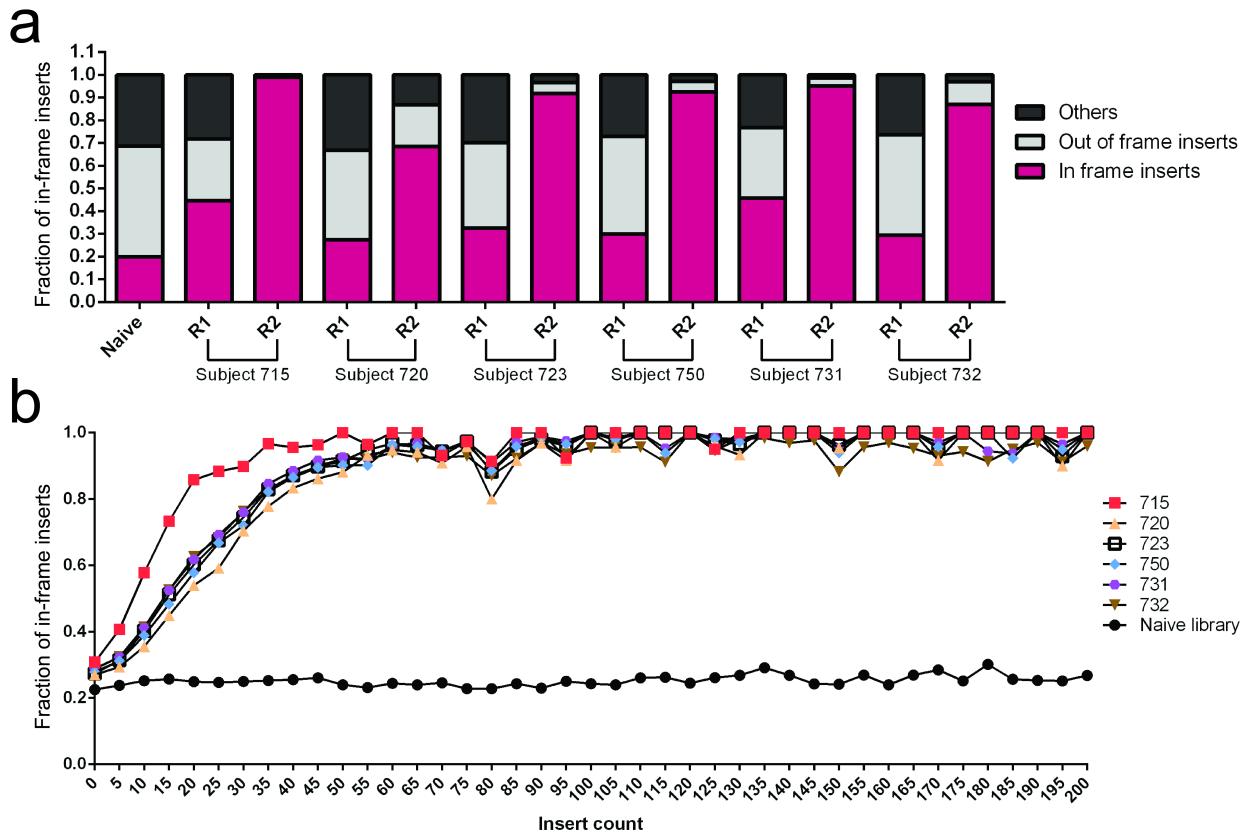
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36 **Supplementary Figure 1.** Schematic of HuNoV genomic phage library construction. **a**
 37 Schematic illustration of pKS-NV68 KM plasmid, which contains the GI.1 open reading frames
 38 (ORFs) as indicated in purple, that was sheared to construct the HuNoV phage library. Since
 39 the entire plasmid was sheared, the library contains inserts from all regions of the plasmid in
 40 addition to ORFs 1 to 3. **b** Schematic illustration of the pTP663 Jun-Fos phage display plasmid.
 41 **c** Sheared DNA of 100 to 500 bp was adapted with A. **d** The pTP663 plasmid was digested with
 42 XbaI, the ends were adapted with T, and the fragments were ligated with the A-adapted sheared
 43 DNA to form the library. **e** The library clones were transformed into *E. coli*, pooled, and helper
 44 phage was added to propagate phages for the phage library to be used for affinity selection.



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46 **Supplementary Figure 2.** Deep sequencing analysis of the inserts present in the NoV Jun-Fos
 47 library before and after affinity selection versus day 14-30 post-infection sera. Distribution of
 48 inserts after two rounds of affinity selection with sera of subjects 715, 720, 723, 750, 731, and
 49 732.



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51 **Supplementary Figure 3.** In-frame fraction analysis of NoV Jun-Fos library versus post-
52 infection sera before and after affinity selection. **a** The fraction of all the in-frame inserts in the
53 naïve library and each round of affinity selection with subjects 715, 720, 723, 750, 731, and 732
54 sera. The fraction of the in-frame inserts is shown on the y-axis while the naïve library and the
55 individual sera names are shown on the x-axis. The fraction of the inserts that are in-frame to
56 the HuNoV ORFs are shown in magenta while the fraction of the out-of-frame inserts and the
57 inserts that are not within HuNoV ORFs are shown in gray and black, respectively. The fraction
58 of the in-frame inserts was determined by dividing the number of in-frame inserts by the total
59 number of inserts in that experiment. **b** The fraction of in-frame reads for each insert count
60 group in the naïve library and the libraries after the second round of affinity selection. The
61 fraction of in-frame inserts is shown on the y-axis while the insert count groups are shown on
62 the x-axis. The fraction of the in-frame inserts per insert count group is determined by dividing
63 the number of in-frame inserts by the total number of inserts in that insert count group.

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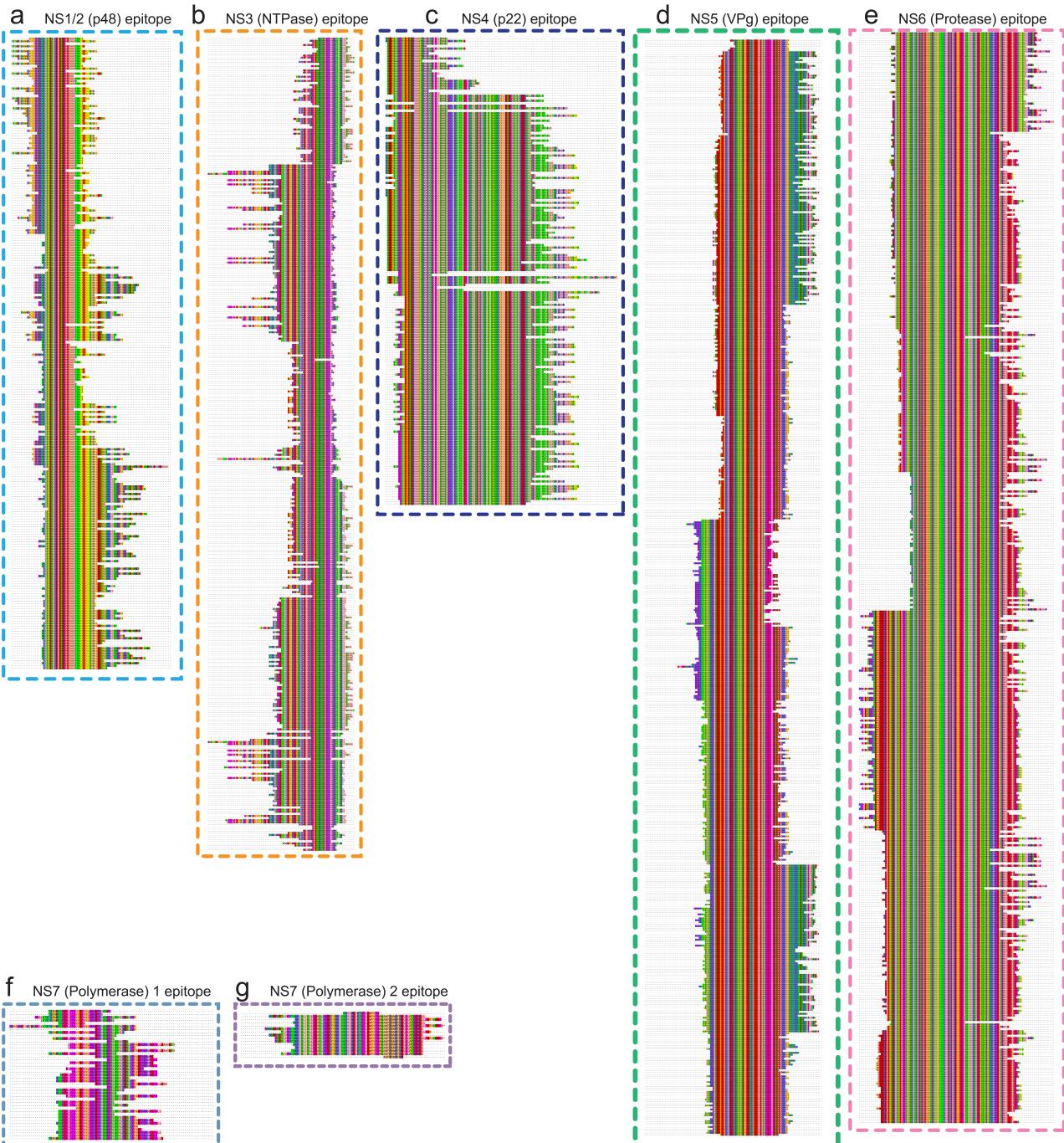
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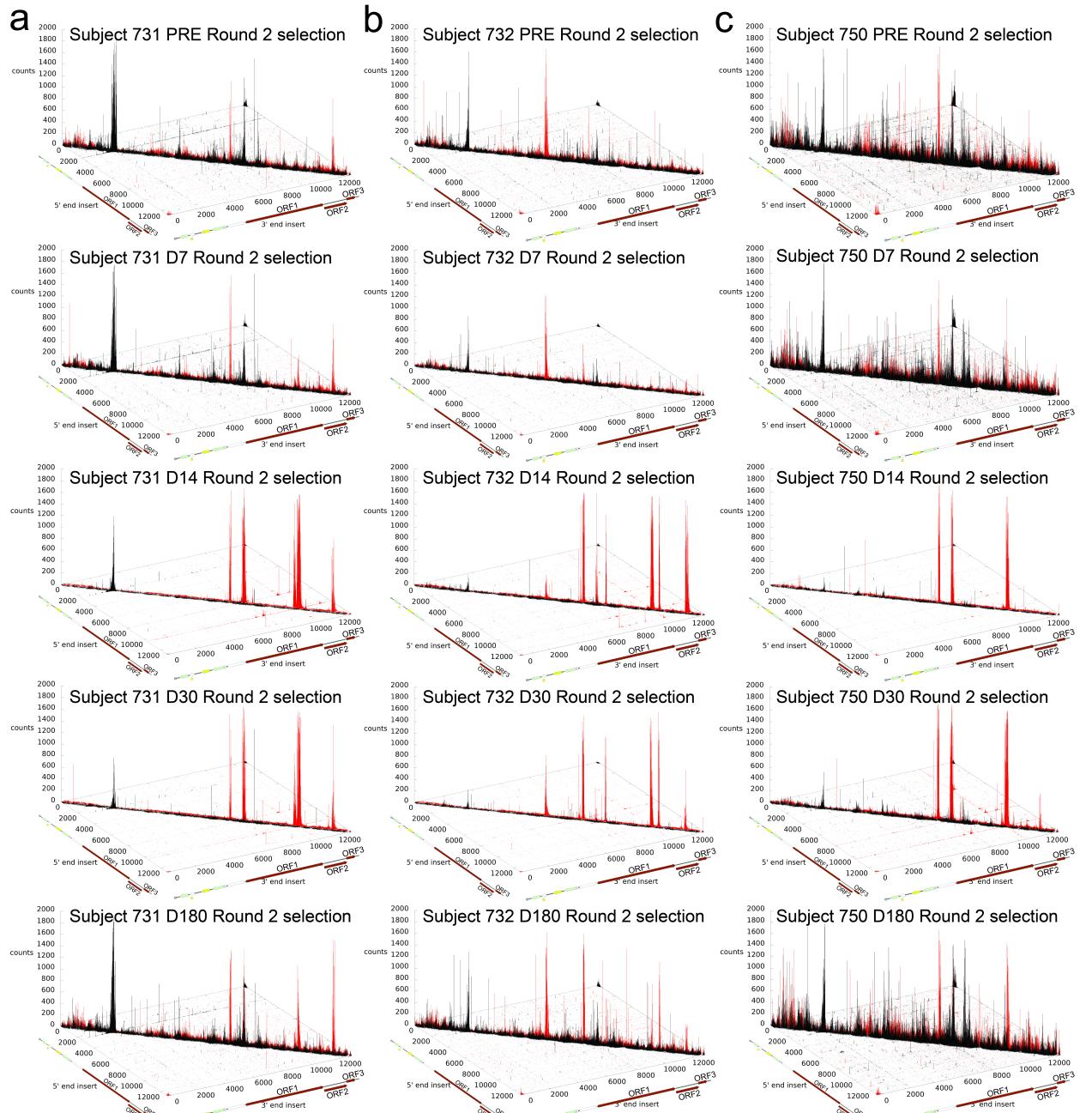
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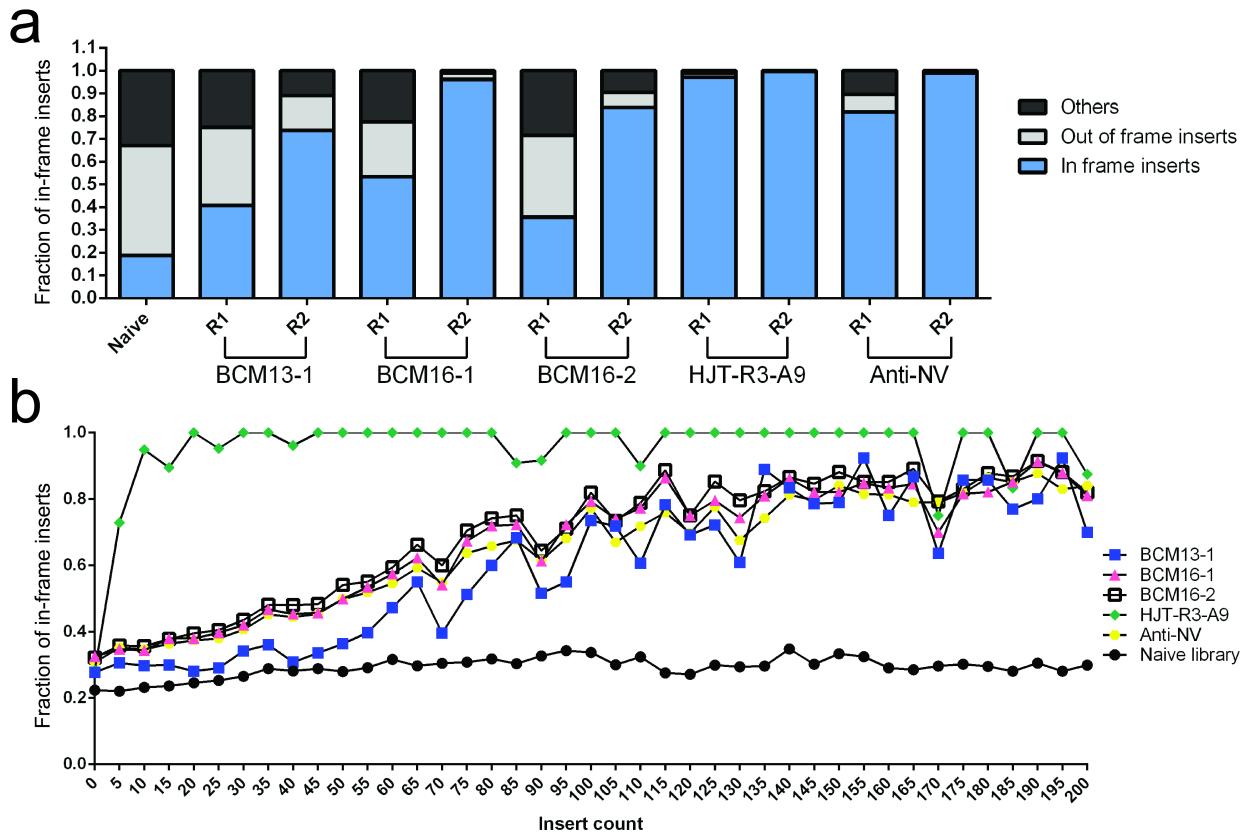


70 **Supplementary Figure 4.** Alignment of peptides with high coverage after two rounds of affinity
 71 selection with sera of subject 715. The numbering at the top indicates the amino acid residue
 72 positions in the nonstructural proteins. The Jalview Zappo color scheme is used, in which
 73 aliphatic/hydrophobic residues (I, L, V, A, and M) are peach, aromatic residues (F, W, and Y)
 74 are gold, positively charged residues (K, R, and H) are blue, negatively charged residues (D and
 75 E) are red, hydrophilic residues (S, T, N, and Q) are green, conformationally special residues (G
 76 and P) are magenta, and cysteine is yellow. The peptide alignments define the anti-GI.1
 77 nonstructural protein epitopes in **a** NS1/2 (p48), **b** NS3 (NTPase), **c** NS4 (p22), **d** NS5 (VPg), **e**
 78 NS6 (protease), and **(f, g)** NS7 (RdRp).



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81 **Supplementary Figure 5.** Deep sequencing analysis of the inserts present in the HuNoV Jun-
 82 Fos library before and after affinity selection versus pre- and post-infection sera. **a** Distribution
 83 of inserts after two rounds of affinity selection with subject 731 sera before challenge, 7 days
 84 after challenge, 14 days after challenge, 30 days after challenge, and 180 days after challenge.
 85 **b** The distribution of inserts after two rounds of affinity selection with subject 732 sera before
 86 challenge, 7 days after challenge, 14 days after challenge, 30 days after challenge, and 180
 87 days after challenge. **c** The distribution of inserts after round 2 of affinity selection with subject
 88 750 sera before challenge, 7 days after challenge, 14 days after challenge, 30 days after
 89 challenge, and 180 days after challenge.



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91 **Supplementary Figure 6.** In-frame fraction analysis of GII.4 HOV Jun-Fos library before and
92 after affinity selection versus GII.4 antisera. **a** The fraction of all the in-frame inserts in the naïve
93 library and each round of affinity selection with subjects BCM16-1, BCM13-1, and BCM16-2
94 sera, HJT-R3-A9 antibody, and Anti-NV. The fraction of the in-frame inserts is shown on the y-
95 axis while the naïve library and the individual sera names are shown on the x-axis. The fraction
96 of the inserts that are in-frame to the HuNoV ORFs are shown in blue while the fraction of the
97 out-of-frame inserts and the inserts that are not within HuNoV ORFs are shown in gray and
98 black, respectively. The fraction of the in-frame inserts is determined by dividing the number of
99 in-frame inserts by the total number of inserts in that experiment. **b** The fraction of in-frame
100 reads for each insert count group in the naïve library and the libraries after affinity selections.
101 The fraction of in-frame inserts is shown on the y-axis while the insert count groups are shown
102 on the x-axis. The fraction of the in-frame inserts per insert count group is determined by
103 dividing the number of in-frame inserts by the total number of inserts in that insert count group.

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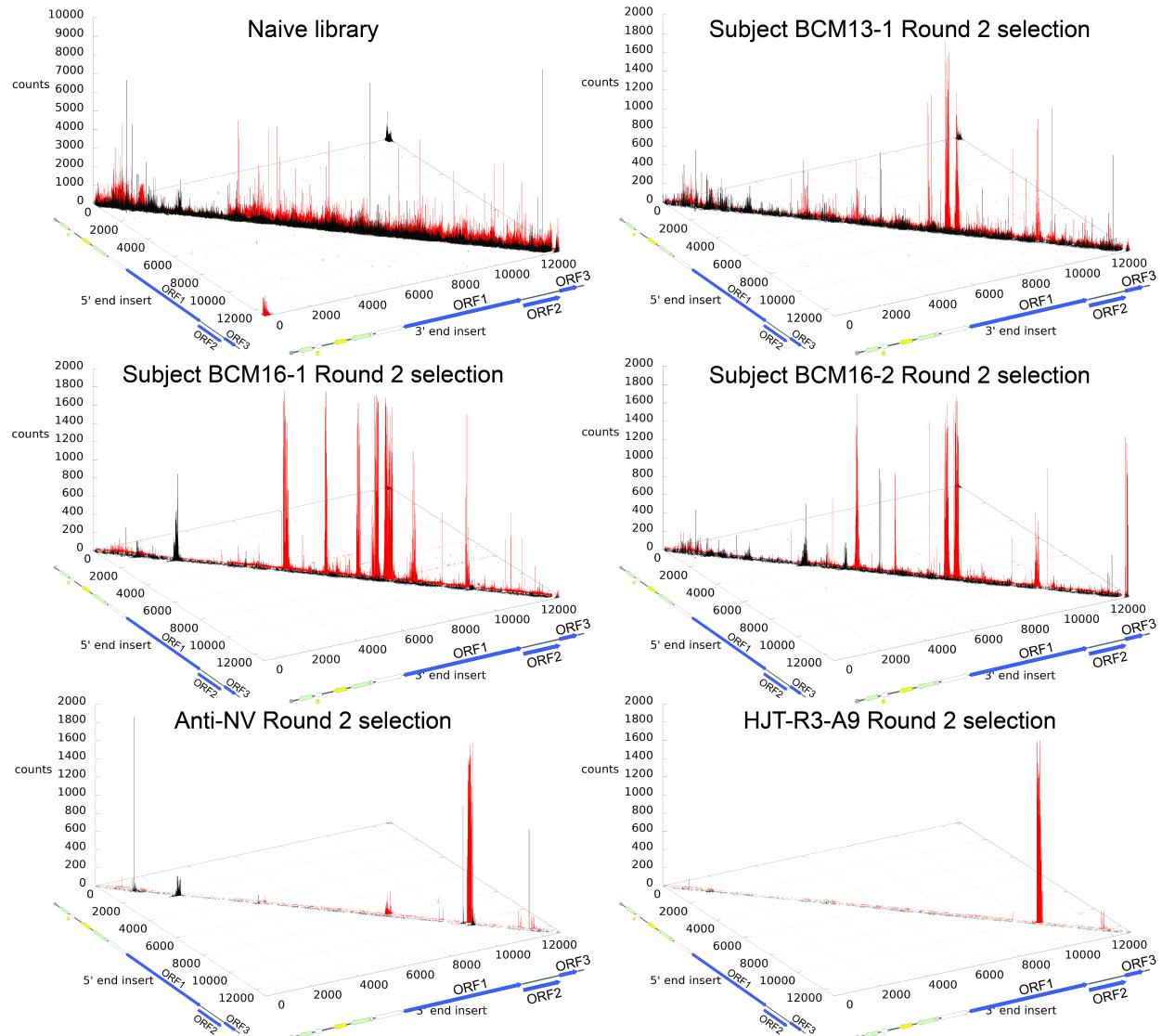
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111 **Supplementary Figure 7.** Deep sequencing analysis of the inserts present in the GII.4 HOV
 112 Jun-Fos library before and after affinity selection versus GII.4 antisera. Distribution of inserts
 113 after two rounds of affinity selection with BCM16-1 sera, BCM13-1 sera, BCM16-2 sera, anti-NV
 114 rabbit polyclonal antibodies, HJT-R3-A9 scFv antibody.

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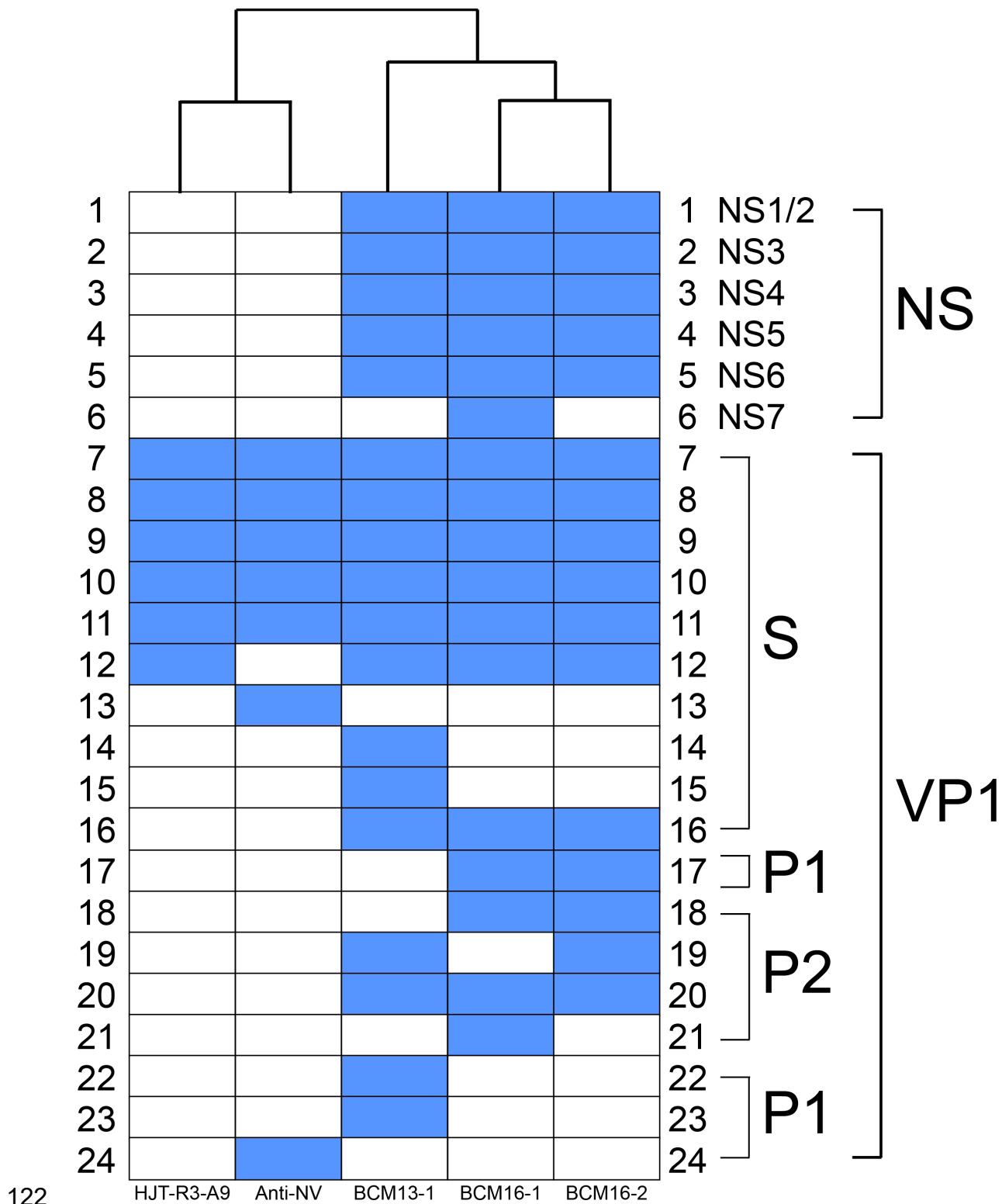
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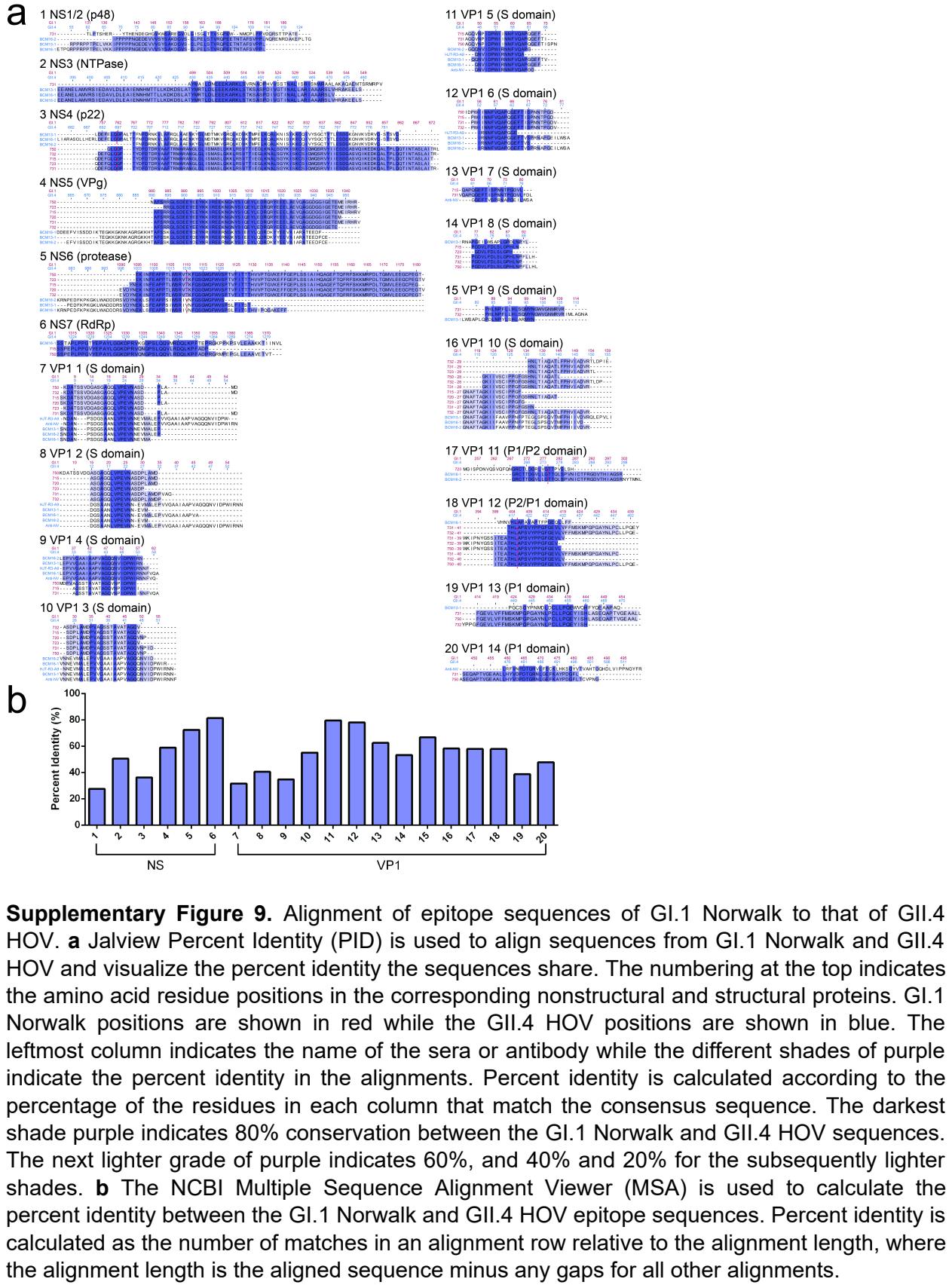
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122 **Supplementary Figure 8.** Dendrogram of GII.4 HOV epitope profiles. A dendrogram comparing
123 the epitope profiles shared among the four sera and HJT-R3-A9 scFv antibody. Blue blocks
124 indicate the epitope that an individual has in the specified nonstructural or structural protein
125 domain.
126



143 **Supplementary Table 1.** List of epitopes from all the sera identified after two rounds of affinity
144 selections with the NoV Jun-Fos library. A total of 48 unique epitopes were identified. The
145 sequences and the positions in the GI.1 Norwalk ORF1 and ORF2 genome of each epitope are
146 included. The shared epitopes amongst the six individuals as well as the epitopes' sequences,
147 positions in the GI.1 Norwalk genome, length, and the number of sera that contain the epitopes
148 are also included.

		HUT-R3-A9 epitope sequence	HUT-R3-A9 epitope position	BCM131 epitope sequence	BCM131 epitope position	BCM161 epitope sequence	BCM161 epitope position	BCM162 epitope sequence	BCM162 epitope position	Anti-NV epitope sequence	Anti-NV epitope position	shared epitope sequence	shared epitope position	epitope length	shared among		
11-p48		RPPRPPPTPELVKQ PPPPNGEDEVVV SYSAKDGSVGLP LSTVQRQEETNTAF DVSQVPL	55 131	ETPQRPRPPPTPE LVKQIPPPNGED EVVVSYSAKOGVS GLPLETEVROPEE TNTAFAVPL	51 113	IPPPPNNGEDEVVV SYSAKDGSVGLP LSTVQRQEETNTAF DVSQVPL	68 127					IPPPPNNGEDEVVV SYSAKDGSVGLP LSTVQRQEETNTAF DVSQVPL	68 113	46	3		
22-NTPass		EEANELAMRSIED AVLDLEAEENNHT TLLDKSLSATYM RTLDLEEKARQLS TKASASPDIVGTTINA LLARIAAARSLLVR AKEELS	395 483	EEANELAMRSIED AVLDLEAEENNHT TLLDKSLSATYM RTLDLEEKARQLS TKASASPDIVGTTINA LLARIAAARSLLVR AKEELS	395 475	EEANELAMRSIED AVLDLEAEENNHT TLLDKSLSATYM RTLDLEEKARQLS TKASASPDIVGTTINA LLARIAAARSLLVR AKEELS	395 483					EEANELAMRSIED AVLDLEAEENNHT TLLDKSLSATYM RTLDLEEKARQLS TKASASPDIVGTTINA LLARIAAARSLLVR AKEELS	395 475	81	3		
33-p22		LEDFELLOGPALTIF NFRNKVLAFLRQ AAENKYGMLDTMK IWRKQVYKPE LKGALKNSKXCG VYSGCTTYLESQG KGNGKVNDRVQSTS VG	690 785	LEDFELLOGPALTIF EFELLOGPALTIF NFRNKVLAFLRQ AAENKYGMLDTMK IWRKQVYKPE LKGALKNSKXCG VYSGCTTYLESQG KGNGKVNDRVQSTS VG	678 785	TNFNDRNKVLAFLR QLAERKQYLMOT IWRKQVYKPE LKGALKNSKXCG VYSGCTTYLESQG KGNGKVNDRVQSTS VG	702 780					TNFNDRNKVLAFLR QLAERKQYLMOT IWRKQVYKPE LKGALKNSKXCG VYSGCTTYLESQG KGNGKVNDRVQSTS VG	702 779	78	3		
44-VPg		TEGKGKGNKAGR GKHKHTAFSSKGLS DEEYDEYKRIREE NOVYSEEEYLDR DKYKEEVAVARAT EEDFCE	874 944	TEGKGKGNKAGR GKHKHTAFSSKGLS DEEYDEYKRIREE NOVYSEEEYLDR DKYKEEVAVARAT EEDFCE	861 942	TEGKGKGNKAGR GKHKHTAFSSKGLS DEEYDEYKRIREE NOVYSEEEYLDR DKYKEEVAVARAT EEDFCE	864 944					TEGKGKGNKAGR GKHKHTAFSSKGLS DEEYDEYKRIREE NOVYSEEEYLDR DKYKEEVAVARAT EEDFCE	872 942	69	3		
55-protease		PEDPKPKGLWAD DORSVDYNEKLSE EAPPSSWSRNVFG SGWGFWSWPSLFSI TST	882 1037	KRNPEDPKPKGL WADDQVYDYN KLSFEPAPSVRSI VNFGSGWGFWS PSLFTSHMWPQG SVLEAKKTIILNL	979 1048	KRNPEDPKPKGL WADDQVYDYN KLSFEPAPSVRSI VNFGSGWGFWS PSLFTSHMWPQG SVLEAKKTIILNL	979 1029					PEDPKPKGLWAD DORSVDYNEKLSE EAPPSSWSRNVFG SGWGFWSI	982 1029	48	3		
66-RdRp				SSTALPLPGTYP AYLGKQDPRVKG GPSLQQVMRDQL KPFTEPRGKPKP SVLEAKKTIILNL	1220 1283							SSTALPLPGTYP AYLGKQDPRVKG GPSLQQVMRDQL KPFTEPRGKPKP SVLEAKKTIILNL	1220 1283	64	1		
71-VP1 (S domain)		NDANPSDGSAAAL VPEVNNNEEMALEP VVGAAIAAP/VAGQ AVPAGQON/DIPW F	6 51	NDANPSDGSAAAL VPEVNNNEEMALEP VVGAAIAAP/VAGQ AVPAGQON/DIPW F	5 31	NDANPSDGSAAAL VPEVNNNEEMALEP VVGAAIAAP/VAGQ AVPAGQON/DIPW F	5 28					NDANPSDGSAAAL VPEVNNNEEMALEP VVGAAIAAP/VAGQ AVPAGQON/DIPW F	6 54	6 31	26	5	
82-VP1 (S domain)		DGSAANLPV EVMALEPV VGAIA AVPAGQON/DIPW F	12 55	DGSAANLPV EVMALEPV VGAIA AVPAGQON/DIPW F	12 27	DGSAANLPV EVMALEPV VGAIA AVPAGQON/DIPW F	12 42					DGSAANLPV EVMALEPV VGAIA AVPAGQON/DIPW F	12 27	12 55	12 27	5	
93-VP1 (S domain)		NNEVMALEPV/VG AIAAPVAGQON/D PWIRNN	23 55	VNNEVMALEPV/VG AAAAPVAGQON/D PWIRNN	22 55	VNNEVMALEPV/VG AAAAPVAGQON/D PWIRNN	22 53					VNNEVMALEPV/VG AAAAPVAGQON/D PWIRNN	22 56	23 49	27	5	
104-VP1 (S domain)		LEPV/GAIAAPVA GQON/DIPWIRNN F	39 56	LEPV/GAIAAPVA GQON/DIPWIRNN F	29 54	LEPV/GAIAAPVA GQON/DIPWIRNN F	29 59					LEPV/GAIAAPVA GQON/DIPWIRNN F	30 58	30 54	25	5	
115-VP1 (S domain)	QA	QNIDPWRNNF QAPGGFET	45 59	QNIDPWRNNF QAPGGFET	44 66	QNIDPWRNNF QAPGGFET	45 64					QNIDPWRNNF QAPGGFET	45 62	45 69	15	5	
126-VP1 (S domain)		IRNNFVQAPGGE TVSPNQAPGGE SA	52 63	IRNNFVQAPGGE TVSPNQAPGGE SA	52 79	IRNNFVQAPGGE TVSPNQAPGGE SA	52 67					IRNNFVQAPGGE TVSPNQAPGGE SA	52 79	52 63	12	4	
137-VP1 (S domain)		RNAPELW GQDNLW										GGEFTVSPRNAPG BILWSA	61 79	61 79	19	1	
148-VP1 (S domain)		LWSAPLQGDLPN PY										RNAPELW GQDNLW	69 89	69 89	21	2	
159-VP1 (S domain)		GNATAFKII PPNPFTEGLPSQ VTMPFHII EPVL	76 97	GNATAFKII PPNPFTEGLPSQ VTMPFHII EPVL	111 156	GNATAFKII PPNPFTEGLPSQ VTMPFHII EPVL	111 147					GNATAFKII PPNPFTEGLPSQ VTMPFHII EPVL	111 149	111 147	37	4	
1610-VP1 (S domain)				GRCTTDGVLLGT QLSPVNCTTRGD THAGSR		GRCTTDGVLLGT QLSPVNCTTRGD THAGSR						GRCTTDGVLLGT QLSPVNCTTRGD THAGSR	264 297		34	2	
1711-VP1 (P1 domain)				NNYDPEEIPAPLG TPDFVGHQGLLT QTTKGDGSTRH KAT		NNYDPEEIPAPLG TPDFVGHQGLLT QTTKGDGSTRH KAT						NNYDPEEIPAPLG TPDFVGHQGLLT QTTKGDGSTRH KAT	309 348		40	2	
1812-VP1 (P2 domain)				TOTTKGDGSTRH KATVYGSAP VYTGSAFTPKLG S	335 357	TOTTKGDGSTRH KATVYGSAP VYTGSAFTPKLG S	338 364					TOTTKGDGSTRH KATVYGSAP VYTGSAFTPKLG S	335 357		23	2	
1913-VP1 (P2 domain)		TKGGDSTRH KATVYGSAP VYTGSAFTPKLG S	338 364	TKGGDSTRH KATVYGSAP VYTGSAFTPKLG S	338 364	TKGGDSTRH KATVYGSAP VYTGSAFTPKLG S	338 364					TKGGDSTRH KATVYGSAP VYTGSAFTPKLG S	338 364		27	3	
2014-VP1 (P2domain)				VHNHLAP VAPTF PGEOLI	413 434	VHNHLAP VAPTF PGEOLI						VHNHLAP VAPTF PGEOLI	413 434		22	1	
2115-VP1 (P2 domain)				PGCSGP NMOLD CLPQEW VHFY DPAQ PAMOL CLPQEW VHFY QEA PAQS DVALL	439 469							PGCSGP NMOLD CLPQEW VHFY DPAQ PAMOL CLPQEW VHFY QEA PAQS DVALL	439 469		31	1	
2216-VP1 (P1 domain)															445 476	32	1
2317-VP1 (P1 domain)												UFBPNP DGTGRULF ECKLH KSQVY TVTA HTGCHD LVPING YFR	475 516		42	1	
2418-VP1 (P1 domain)															475 516		

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150 **Supplementary Table 2.** List of epitopes from all the sera identified after two rounds of affinity
 151 selections with the GII.4 HOV Jun-Fos library. A total of 24 unique epitopes were identified. The
 152 sequences and the positions in the GII.4 HOV ORF1 and ORF2 genome of each epitope are
 153 included. The shared epitopes amongst the six individuals as well as the epitopes' sequences,
 154 positions in the GII.4 HOV genome, length, and the number of sera that contain the epitopes are
 155 also included.

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