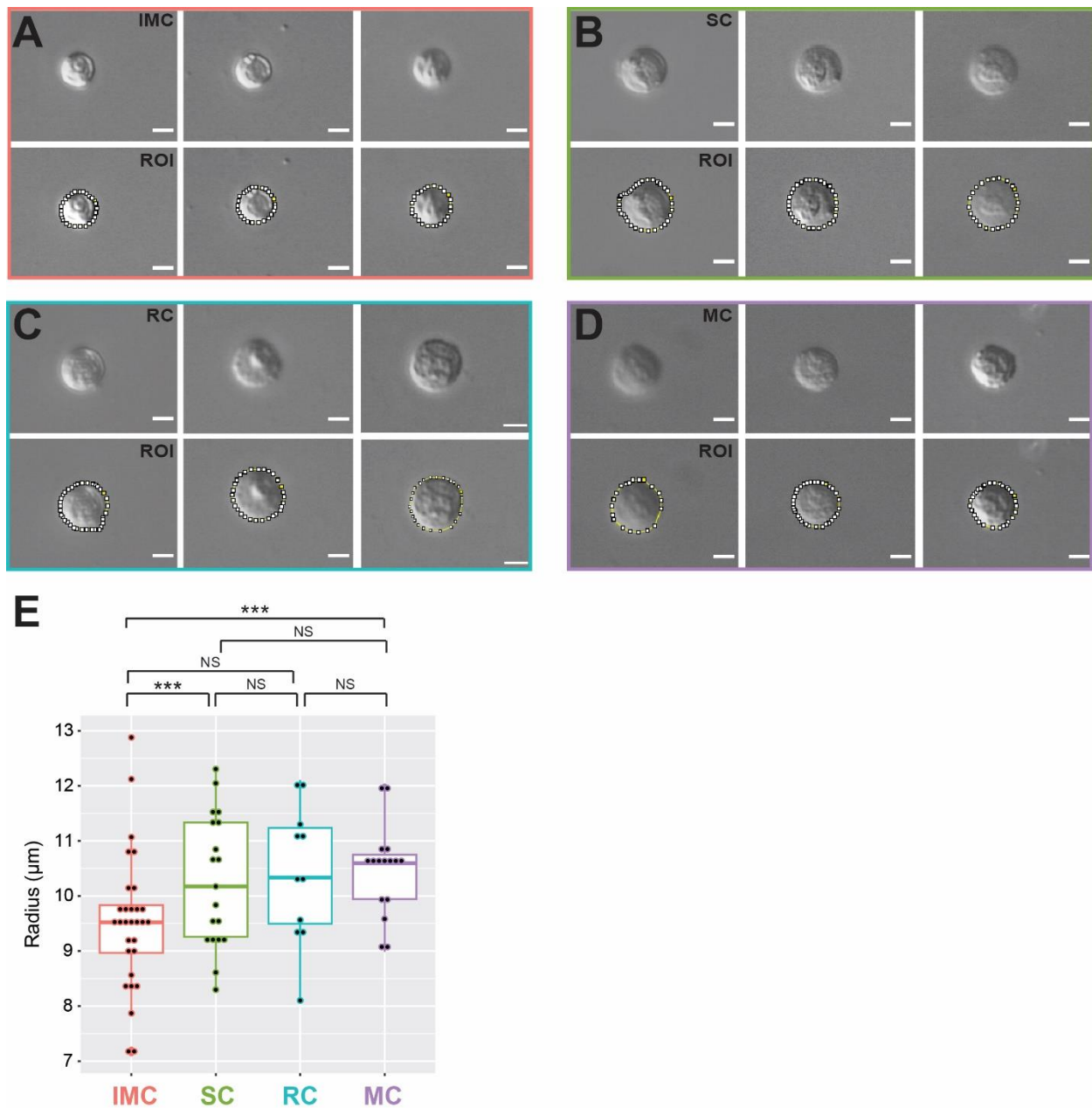


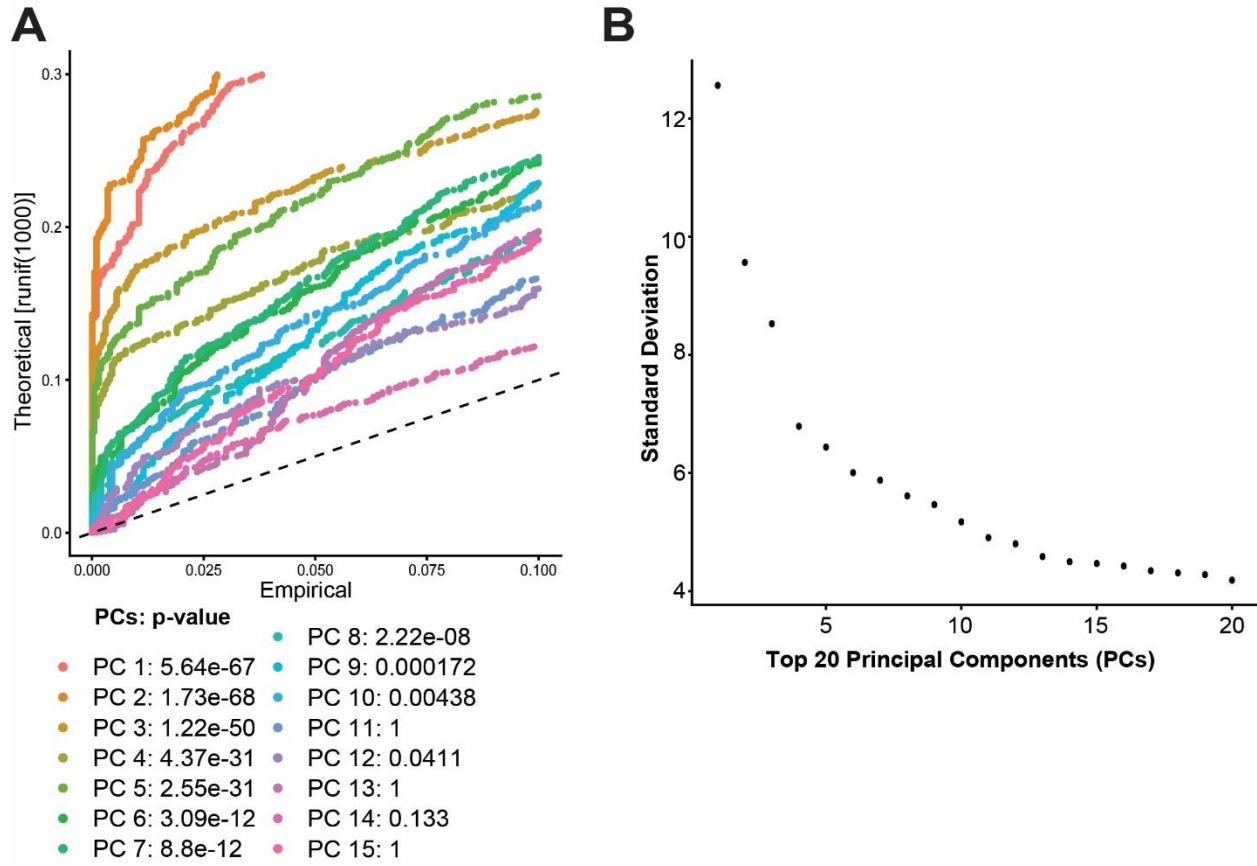
Additional file 1: Fig. S1. Workflow. Single cells from the stria vascularis (SV) were harvested manually from P30 *Slc26a4*^{+/+}, *Slc26a4*^{+/-}, and *Slc26a4*^{-/-} mice as a source of full-length RNA, which was used to prepare a library for single-cell RNA sequencing. Data analysis using uniform manifold approximation and projection (UMAP) identified four types of cochlear SV cells. Cluster-defining genes from unbiased clustering analysis was used to examine gene enrichment. In addition, we used clustering-defining genes to filter each cell type. Finally, we defined pH-

dependent differential expressed genes in *Slc26a4*^{-/-} as compared to *Slc26a4*^{+/+} in a cell-type-specific manner.

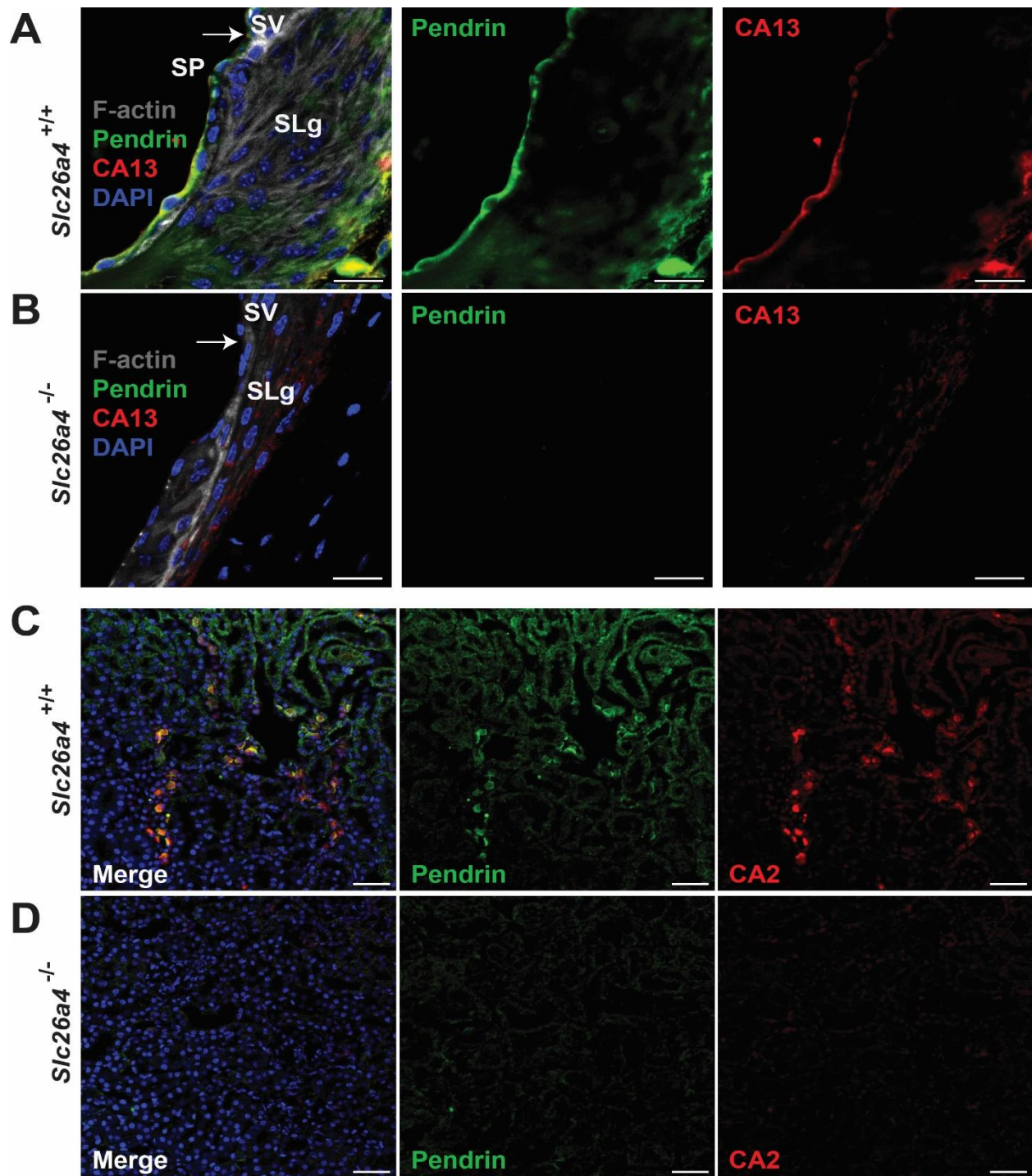


Additional file 1: Fig. S2. (Related to Figure 1): SV Cells and size measurement. (A-D) Representative images of isolated cells for the four-cell type cluster. Measure the cell's perimeter as a region of interest (ROI). Scale bar: 10 µm. (IMC, intermediate cell; MC, marginal cell; RC, root cell; SC, spindle cell). **(A)** IMCs of the cochlear SV. The median perimeter was 59.80 ± 1.42 µm (n=30). **(B)** SCs of the cochlear SV. The median perimeter was 64.49 ± 1.73 µm (n=19). **(C)** RCs of the cochlear SV. The median perimeter was 65.38 ± 2.34 µm (n=11). **(D)** MCs of the

cochlear SV. The median perimeter was $65.79 \pm 1.29 \mu\text{m}$ (n=16). **(E)** Box plots of the radius of each cell. Y-axis is the radius of cells *, $p < 0.05$. The radius of IMC ($9.52 \pm 0.23 \mu\text{m}$) is significantly smaller than the radius of SC ($10.26 \pm 0.28 \mu\text{m}$; $p = 0.043$) and MC ($10.47 \pm 0.21 \mu\text{m}$; $p = 0.0033$). There is no difference in the radius of IMC and RC ($p = 0.2$). There is no difference in the radius of SC and RC ($p = 0.76$). There is no difference in the radius of SC and MC ($p = 0.55$). There is no difference in the radius of RC and MC ($p = 0.58$). Each dot represents a single cell.



Additional file 1: Fig. S3. (Related to Figure 1): Quality control and principal components (PCs) of the SV cells. **(A)** PCs are shown as solid-colored curves. There is a sharp drop-off in p-values after the first 8 PCs. A dashed line is a uniform distribution. **(B)** A ranking of principal components (PCs) based on the percentage of variance. The black dots are 20 different PCs.



Additional file 1: Fig. S4. (Related to Figure 3): Representative confocal images showing the cellular localization of pendrin and carbonic anhydrases in the inner ear (A-B) and kidney (C-D). (A-B) Immunostaining of pendrin (green) and CA13 (red) nuclei (blue), and F-actin (grey). Scale bar: 20 μ m. Note: In *Slc26a4*^{-/-} mice, pendrin and CA13 staining are negative. (C-D) Cryosections

of the mouse kidney tissue are shown as positive and negative expression controls. Scale bar: 50 μm . Note: In *Slc26a4*^{+/+} mice (**C**), pendrin (green) and CA2 (red) co-localize in the intercalated cells. However, in *Slc26a4*^{-/-} mice (**D**), pendrin and CA2 staining are negative.