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Supplemental information

Integrative systems biology characterizes

immune-mediated neurodevelopmental

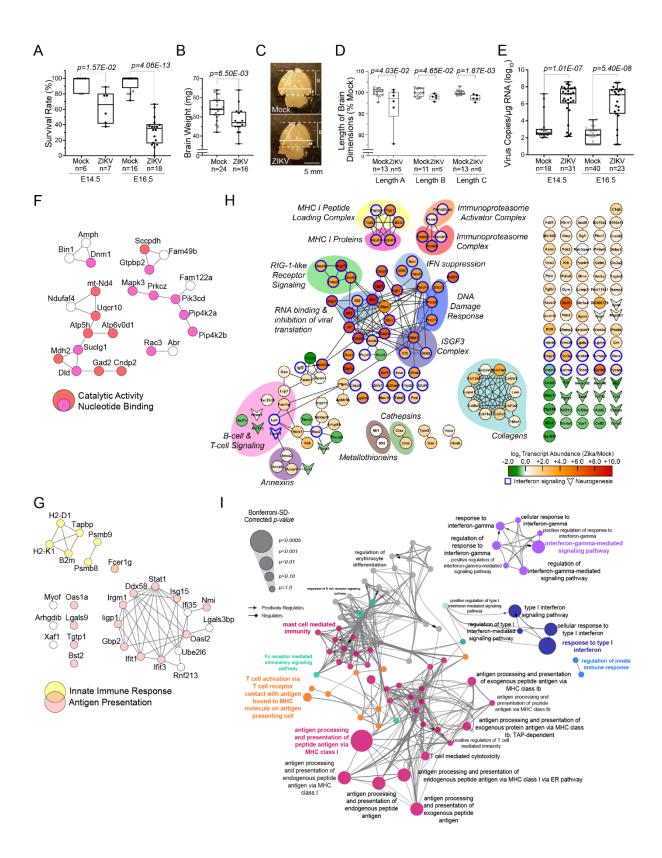
changes in murine Zika virus microcephaly

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Intraplacental zika virus injection leads to smaller brain size of the embryos and induces elevated innate immune response. (Related to Figure 1A-F, 2C-D, 3A, 3H)

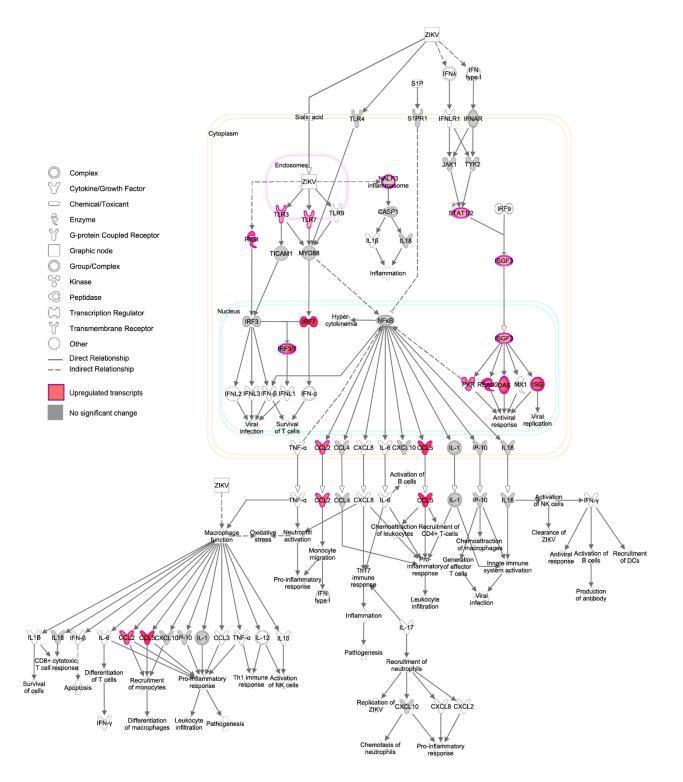
- (A) Survival rates of embryos at E14.5 and E16.5. Box plot shows median, interquartile range, and maximum/minimum values.
- (B) Brain weight in milligrams of Mock- or ZIKV-infected embryos at E16.5.
- (C) Morphometric analysis of a representative E16.5 brain in 3 dimensions. Representative measurements of Mock-infected brain (lengths A-C) are overlaid on a representative ZIKV-infected brain.
- (D) Measurements of lengths A-C expressed as percentage of lengths measured in Mock-infected brains.
- (E) Virus copy number at E14.5 and E16.5 following intraplacental ZIKV or Mock infection at E10.5.
- (F) Interaction network of proteins with strong positive correlation (Pearson correlation r > 0.90) with ZIKV polyprotein abundance. Node color indicates functional classification (red, catalytic activity; purple, catalytic and nucleotide-binding activities); edges represent high confidence interactions (confidence score = 0.70) between proteins in the network based on protein-protein interaction information curated by the STRING database; connected nodes are shown.
- (G) Interaction network of proteins included in cluster 1-1 of Figure 2C. Node color indicates functional classification (yellow, innate immune response; light pink, antigen presentation, orange, both); edges represent high confidence interactions (confidence score = 0.70) between proteins based on in the network based on STRING database.
- (H) Interaction network for the union of significantly increased-in-abundance and decreased-inabundance RNA-proteins pairs. Node color indicates RNA-level fold change (log2(ZIKV/Mock)); edges represent high confidence interactions between nodes in the network based on protein-protein interaction information curated by the STRING database. Nodes outlined in blue indicate are associated with interferon signaling and innate immune response. Arrow-head node shape indicates association with neurogenesis and neurodevelopment.
- (I) ClueGO network analysis of significantly increased-in-abundance RNA-protein pairs highlighting enriched immune-associated pathways (GO immune System Functional Pathway Database, EBI) network. Nodes indicate individual functional pathways. Node colors indicate interconnected pathways. Node size indicates significance of enrichment (p-value, Bonferroni step-down method) in the network. Edge width and transparency indicate kappa scores reflecting the similarity of genes associated with connected nodes.

Error bars indicate standard deviation.



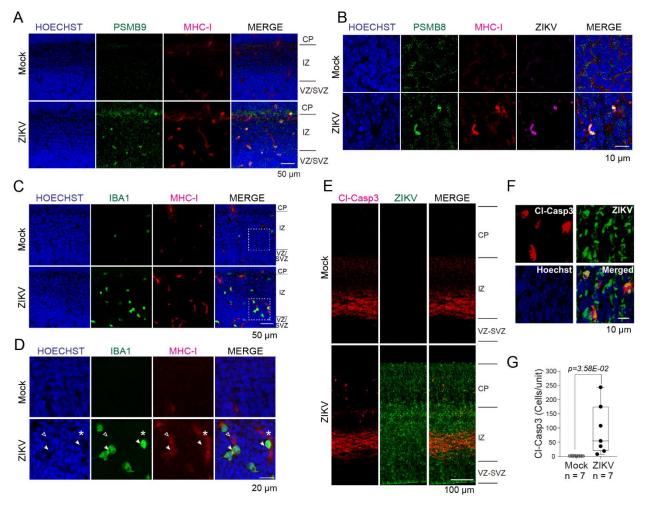
Congenital ZIKV infection activates the interferon-induced pathways in the embryonic brain. (Related to Figure 3A-F)

Significant transcript expression changes (q < 0.05) mapped to interferon-induced pathways based on pathway analysis conducted by QIAGEN Ingenuity Pathway Analysis. Node shape represents the function of the edges; solid line, direct interaction; dashed line, indirect interaction; red, upregulated transcripts; grey, no significant changes.



Immunohistochemistry staining results show activation of antigen presentation pathways, recruitment of macrophages/microglia, and induction of apoptosis after ZIKV infection. (Related to Figure 3D-E)

- (A) Immunohistochemistry (IHC) staining of cortices of Mock- or ZIKV-infected E14.5 brains with proteasome 20S Subunit Beta 9 (PSMB9) (green), MHC-I (red), and Hoechst 33342 (nucleus, blue). CP, cortical plate; IZ, intermediate zone; VZ, ventricular zone; SVZ, subventricular zone. Scale bar = 50 μm.
- (B) IHC staining of cells in the E14.5 CP of Mock- or ZIKV-infected brains stained for PSMB8 (green), MHC-I (red), ZIKV (purple), and Hoechst (nucleus, blue). Scale bar = 10 μm.
- (C) IHC staining of the VZ/SVZ, IZ, and CP of Mock- or ZIKV-infected E14.5 brains stained for allograft inflammatory factor 1 (IBA1) (green), MHC-I (red), and Hoechst (nucleus, blue).
- (D) Higher magnification of the sections indicated by the dotted square in 3F. Triangles, MHC-I-presenting neural cells; arrows, cells with overlapping signals of IBA1 and MHC-I, suggesting the engulfment of MHC-I-presenting cells by macrophage/microglia. Note that the IBA-positive cells are located adjacent to the endothelial cells of blood vessels, indicated by asterisks. Scale bar = 20 µm.
- (E) IHC staining of Mock-or ZIKV-infected E16.5 brains with Cleaved Caspase 3 (CI-Casp3) (red), ZIKV (green), and Hoechst (nucleus, blue), scale bar = 100 μm.
- (F) Higher magnification of **D** of the ZIKV-infected brain, scale bar = $10\mu m$.
- (G) Quantification of cleaved caspase 3 (CI-Casp3)-positive cells per unit area in Mock-or ZIKV-infected embryonic brains. Mock, n = 7 from 7 litters; zika, n = 7 from 6 litters. Error bars indicate standard deviation.



ZIKV infection downregulates neurodevelopmental genes/proteins leading to diminished numbers of neurons in the cortical plate. (Related to Figure 3G)

Interaction network for the protein-RNA pairs that did not match with respect to the changes in abundance (Quadrant II and IV of **Figure 3G**). Node color represents the function of the proteins (light blue, neurodevelopmental genes; purple, cell cycle; green, cell adhesion; peach, collagen and cell adhesion); edges represent high confidence interactions (confidence score = 0.70) between proteins in the network based on protein-protein interaction information curated by the STRING database.

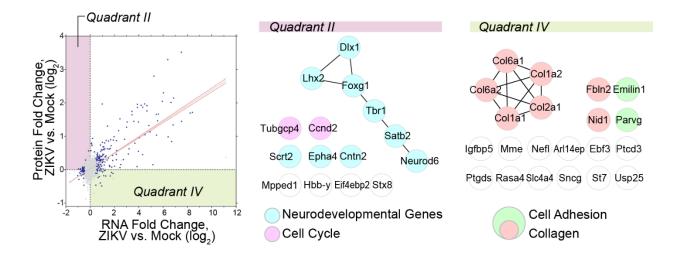


Table S7

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Downregulated protein-transcript pairs reported to have neurodevelopmental functions (*Related to Figure 4A*)

Mouse gene symbol	Neurodevelopmental functions
Dlx2	Terminal differentiation of interneurons; development of ventral forebrain ^{95, 96, 97, 98}
Ep400	Survival of the oligodendrocytes and myelination in the vertebrate central nervous system ⁹⁹
Kif26a	Repression of cell growth signaling pathway during enteric nervous system development ¹⁰⁰
Plxna4	Axon guidance and migration of sympathetic neurons ¹⁰¹
Ptprz1	Inhibition of proliferation and promotion of maturation of oligodendrocytes; establishment of contextual memory ^{102, 103}
Sez6	Dendrite elongation and branching and excitatory synapse formation of the cerebral cortical neurons ¹⁰⁴
Tiam2	Neurite growth and growth cone remodeling of the hippocampal neurons; migration of cerebral cortical neurons ^{105, 106}

Supplemental References (Related to Table S7)

- 95. Porteus, M.H., Bulfone, A., Ciaranello, R.D., and Rubenstein, J.L. (1991). Isolation and characterization of a novel cDNA clone encoding a homeodomain that is developmentally regulated in the ventral forebrain. Neuron *7*, 221-229. 10.1016/0896-6273(91)90260-7.
- 96. Anderson, S.A., Eisenstat, D.D., Shi, L., and Rubenstein, J.L. (1997). Interneuron migration from basal forebrain to neocortex: dependence on DIx genes. Science *278*, 474-476.
- 97. Colasante, G., Collombat, P., Raimondi, V., Bonanomi, D., Ferrai, C., Maira, M., Yoshikawa, K., Mansouri, A., Valtorta, F., Rubenstein, J.L., and Broccoli, V. (2008). Arx is a direct target of Dlx2 and thereby contributes to the tangential migration of GABAergic interneurons. J Neurosci 28, 10674-10686. 10.1523/JNEUROSCI.1283-08.2008.
- 98. Feng, L., Eisenstat, D.D., Chiba, S., Ishizaki, Y., Gan, L., and Shibasaki, K. (2011). Brn-3b inhibits generation of amacrine cells by binding to and negatively regulating DLX1/2 in developing retina. Neuroscience *195*, 9-20. 10.1016/j.neuroscience.2011.08.015.
- 99. Elsesser, O., Frob, F., Kuspert, M., Tamm, E.R., Fujii, T., Fukunaga, R., and Wegner, M. (2019). Chromatin remodeler Ep400 ensures oligodendrocyte survival and is required for myelination in the vertebrate central nervous system. Nucleic Acids Res *47*, 6208-6224. 10.1093/nar/gkz376.
- 100. Zhou, R., Niwa, S., Homma, N., Takei, Y., and Hirokawa, N. (2009). KIF26A is an unconventional kinesin and regulates GDNF-Ret signaling in enteric neuronal development. Cell *139*, 802-813. 10.1016/j.cell.2009.10.023.
- 101. Waimey, K.E., Huang, P.H., Chen, M., and Cheng, H.J. (2008). Plexin-A3 and plexin-A4 restrict the migration of sympathetic neurons but not their neural crest precursors. Dev Biol *315*, 448-458. 10.1016/j.ydbio.2008.01.002.
- 102. Tamura, H., Fukada, M., Fujikawa, A., and Noda, M. (2006). Protein tyrosine phosphatase receptor type Z is involved in hippocampus-dependent memory formation through dephosphorylation at Y1105 on p190 RhoGAP. Neurosci Lett *399*, 33-38. 10.1016/j.neulet.2006.01.045.
- 103. Lamprianou, S., Chatzopoulou, E., Thomas, J.L., Bouyain, S., and Harroch, S. (2011). A complex between contactin-1 and the protein tyrosine phosphatase PTPRZ controls the development of oligodendrocyte precursor cells. Proc Natl Acad Sci U S A *108*, 17498-17503. 10.1073/pnas.1108774108.
- 104. Gunnersen, J.M., Kim, M.H., Fuller, S.J., De Silva, M., Britto, J.M., Hammond, V.E., Davies, P.J., Petrou, S., Faber, E.S., Sah, P., and Tan, S.S. (2007). Sez-6 proteins affect dendritic arborization patterns and excitability of cortical pyramidal neurons. Neuron *56*, 621-639. 10.1016/j.neuron.2007.09.018.
- 105. Kawauchi, T., Chihama, K., Nabeshima, Y., and Hoshino, M. (2003). The in vivo roles of STEF/Tiam1, Rac1 and JNK in cortical neuronal migration. EMBO J *22*, 4190-4201. 10.1093/emboj/cdg413.
- 106. Matsuo, N., Terao, M., Nabeshima, Y., and Hoshino, M. (2003). Roles of STEF/Tiam1, guanine nucleotide exchange factors for Rac1, in regulation of growth cone morphology. Mol Cell Neurosci *24*, 69-81. 10.1016/s1044-7431(03)00122-2.