Vimentin is Required for Tumor Progression and Metastasis in a Mouse Model of Non-Small Cell Lung Cancer

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Supplemental Figure Legends

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Supplemental Figure 1. (A) Experimental design. LSL-Kras^{G12D/+}Tp53^{flox/flox} (KPV^{+/+}) mice were 2 crossed with vimentin-knockout (Vim^{-/-}) mice. KPV^{+/+} and KPV^{-/-} mice were administered 3 4 adenoviral Cre recombinase (Ad-Cre) which resulted in gene recombination at LoxP sites. As a 5 control, null adenovirus (Ad-null) was administered to an independent cohort of mice. (B) Lungs were isolated from Ad-null-treated KPV^{++} and KPV^{--} mice, fixed, sectioned, and subjected to 6 7 H&E staining and vimentin immunohistochemical staining. Positive vimentin staining is brown, 8 and nuclei are blue. Scale bar: 200 µm. (C) Rosa26-LSL-LacZ reporter mice were administered 9 Ad-Null or Ad-Cre, and β-galactosidase staining was performed on whole lung samples; positive staining appears blue. (**D**) $KPV^{+/+}$ and $KPV^{-/-}$ mice were administered Ad-Null or Ad-Cre. Lunas 10 11 were harvested at 2, 8, and 12 weeks following adenoviral infection. DNA was isolated from the 12 tissue and PCR was performed to evaluate the presence of the wild-type (WT) and mutant (G12D) 13 Kras transcript. (E) Tumor cells were isolated from Ad-Cre-infected mice at 6 weeks post-14 infection. A Western blot was performed on KPV^{+/+} and KPV^{-/-} whole cell lysates to detect WT 15 and G12D-mutant KRAS. P53 mRNA (F) and protein levels (G) were detected through gRT-PCR and Western blot, respectively; MLE-12 cells were used as a positive control. (H) $KPV^{+/+}$ and 16 KPV^{-/-} mice were infected with null or Cre recombinase adenovirus (Ad-Null and Ad-Cre, 17 18 respectively) and were imaged at 2, 6, and 10 weeks post-infection (w.p.i.). Representative MRI 19 coronal (left) and transverse (right) images are shown. Hearts (H) are outlined in white. (I) Positive 20 staining was quantified from lung sections stained with either TTF-1 or Ki67 (see Figure 1D for 21 representative images) and normalized to either total lung area (TTF-1) or total tumor cell count (Ki67) (n=2-5). (J) $KPV^{+/+}$ and $KPV^{-/-}$ cells were plated overnight and were then treated with BrdU 22 for 4 hours. BrdU incorporation was detected and normalized to the KPV^{+/+} condition for each 23 independent trial (n=4). Data were compared using an unpaired, two-tailed t-test (**p<0.01). (K) 24 25 H&E-stained lung sections from 8 or 12 w.p.i. were evaluated for tumor grade by a pathologist. (J) Human lung tissue sections (or lymph node tissue where indicated) were stained with
antibodies against vimentin. LUAD=lung adenocarcinoma. Positive vimentin staining is brown,
and nuclei are blue. Scale bar: 200 µm. Data are presented as the mean ± standard deviation.

Supplemental Figure 2. (A) KPV^{+/+} and KPV^{-/-} cell lysates were subjected to a Western blot to 30 31 detect E-cadherin, N-cadherin, vimentin, and actin. Band signal was guantified and normalized to 32 actin loading controls and average KPV^{+/+} signal. Data were compared using an unpaired, twotailed t-test (*p<0.01; ***p<0.001; ****p<0.0001). (**B**) KPV^{+/+} and KPV^{-/-} cells were stained for 33 34 vimentin (green), for keratin 8 (red), and with Hoechst nuclear dve (blue). Scale bar: 10 µm, (C-**E**) Messenger RNA from $KPV^{+/+}$ and $KPV^{-/-}$ cell lysates was quantified via RNA sequencing. (**C**) 35 Principal component analysis (PCA) plot with each point representing one replicate (*black*, *KPV*^{+/+}; 36 37 grey, $KPV^{-/-}$). (**D**) Pearson's correlation plot. The correlation coefficient for each comparison is shown. (E) MA plot. Genes of interest are indicated. (F) Lungs were harvested from KPV^{++} and 38 KPV^{-/-} mice at 7 w.p.i., fixed, sectioned, and subjected to immunohistochemistry with antibodies 39 40 against E-cadherin and N-cadherin. Scale bar: 1 mm (whole lung), 100 µm (insets). 41 Representative images and quantification of positive staining are shown (n=2-5).

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43 **Supplemental Figure 3.** (A) A549 cells were treated with 2 µM withaferin A (WFA) for 1 hour. 44 Cells were fixed and stained for vimentin (white). A phase contrast image was used to identify cell 45 borders (dashed line). Scale bar: 10 µm, 5 µm (inset). (B) A549 cells were treated with WFA for 46 the indicated dose and time. A Western blot is shown; vimentin and GAPDH antibodies were used 47 to probe for these proteins. (C) A549 cells were treated with DMSO control or 1 or 2 µM WFA and 48 subjected to a scratch wound assay. After 24 hours, wound closure was assessed. 49 Representative images (left) and quantitation (right) are shown. Data were compared to the 50 vehicle control using an unpaired, two-tailed t-test (*p<0.05; ***p<0.001).

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Supplemental Figure 4. (**A**) Lungs isolated from vehicle- or WFA-treated $KPV^{+/+}$ mice at 6 weeks after adenoviral Cre infection were fixed, sectioned, and subjected to immunohistochemistry for the indicated markers; slides were scanned and signal was quantified using Histoquest. TTF-1 and Ki67 were normalized to total lung area. Data were subject to an unpaired, two-tailed t-test (**p<0.01; ***p<0.001). (**B**) Lungs isolated from vehicle- and WFA-treated $KPV^{+/+}$ mice were sectioned and stained for phospho-Serine55 vimentin (green) and DAPI. Scale bar: 50 µm, 5 µm (*insets*).

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60 Supplemental Figure 5. (A) Select gene expression values from RNA sequencing. All gene comparisons shown ($KPV^{+/+}$ vs. $KPV^{-/-}$) have FDR<0.05 after adjusting for multiple comparisons; 61 therefore, all gene differences shown between $KPV^{+/+}$ vs. $KPV^{-/-}$ cells are statistically significant. 62 63 (B) SLC7A11 levels were measured by Western blot and normalized to an actin loading control and the average KPV^{++} control. (C) Metabolite data were normalized to the total ion count per 64 65 sample, log-transformed, and subjected to an unpaired, two-tailed t-test. P-values were corrected for multiple comparisons (*adjusted p-value<0.05). (**D**) Lungs were harvested from $KPV^{+/+}$ mice 66 67 treated with WFA or vehicle control at 6 weeks after adenoviral Cre infection, fixed, sectioned, 68 and subjected to anti-GPX antibody staining. Scale bar: 1 mm. (E) KPV^{+/+} and KPV^{-/-} cells were 69 treated with the ML162 (1 µM), DFO (100 µM), and/or Fer-1 (10 µM) for 48 hours; cell death was 70 quantified with an LDH assay. Groups were compared using a 2-way ANOVA with multiple comparisons. All data are presented as the mean ± standard deviation. (****p<0.0001). (F) A 71 72 schematic model representing the ferroptosis-related mechanisms that are affected by loss of 73 vimentin in KRAS-mutant, p53-null lung adenocarcinoma cells. Created with BioRender.com

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Supplemental Figure 6. (A) Luciferase-tagged $KPV^{+/+}$ (*Luc-KPV*^{+/+}) cells were treated with CRISPR-Cas9 to knock out vimentin (*Luc-KPV*^{-/-}). Cells were subjected to a Western blot and probed for vimentin and actin. (**B**) Representative IVIS images for each week after flank injection 78 (n=9-11 mice per group). At 3 weeks post-injection, primary tumors were removed and lung 79 metastases were tracked for an additional 1 week. Intensity overlays show the accumulation of 80 luciferase-labeled cells. Luciferin signal was quantified from primary flank tumors (C) and the 81 lungs (D). (F) Flank tumor volume was measured with calipers each week. Volume was calculated 82 using the formula Volume=(length² x width)/2. For C-E, data were subjected to a two-way ANOVA 83 with multiple comparisons. (G) At week 3, tumors were removed and weighed. For G, data were 84 subjected to a one-way ANOVA with multiple comparisons. (*p<0.05; **p<0.01). Data are 85 presented as the mean ± standard deviation.

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Supplemental Table 1. List of antibodies used. For proteins with multiple antibodies used, the figures in which they are used are indicated. If no figure is indicated, the antibody was used for all instances in which that protein was detected. WB=Western blot; IHC=immunohistochemistry; IF=immunofluorescence.

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