Supplementary information

Structural basis for FGF hormone signalling

In the format provided by the authors and unedited

Supplementary Fig.1. Raw uncropped western blot images



Uncropped blotting for Figure 3



Uncropped blotting for Figure 4





panel b

Uncropped blotting for Figure 5



α-tubulin 💳 🖛

-70

-55



panel f for FGF1

FGFR1c^{∆PLBS}+FGFR1c^{∆SLBS}







panel f for FGF4









panel h



Uncropped blotting for Extended Data Figure 2



Uncropped blotting for Extended Data Figure 3









Uncropped Blotting for Extended Data Figure 6b





Uncropped Blotting For Extended Data Figure 7b





Uncropped Blotting For Extended Data Figure 7c





Uncropped Blotting For Extended Data Figure 7d







$FGFR2b^{\text{APLBS}} + FGFR2b^{\text{ASLBS}}$

Uncropped Blotting For Extended Data Figure 7e





Uncropped Blotting For Extended Data Figure 8b







FGFR2b^{APLBS}+FGFR1b^{ASLBS}

Uncropped Blotting For Extended Data Figure 8c





FGFR2b^{ΔPLBS}+FGFR1b^{ΔSLBS}

Uncropped Blotting For Extended Data Figure 8e





Uncropped Blotting For Supplementary Figure3



Supplementary Fig.2. Purification of all three physiologically possible human FGF23-FGFR- α Klotho-HS quaternary complexes.

Representative size exclusion chromatograms of FGF23-FGFR1c- α Klotho-HS (a), FGF23-FGFR3c- α Klotho-HS (b), and FGF23-FGFR4- α Klotho-HS (c) quaternary complexes obtained over Superdex 200 column. Under the chromatograms, Coomassie blue-stained SDS-PAGE gels show the contents of the indicated main peak fractions within the dashed lines. Results are from at least three replicates. Molecular weight markers are labelled. Bands corresponding to FGF23, FGFRs and α Klotho are indicated. **d.** Purified samples of FGF23^{WT} and its mutants (FGF23^{ΔHBS}, FGF23^{ΔNT}, and FGF23^{ΔSRBS}) were analyzed by SDS-PAGE confirming similar purity and quantity. e. Size exclusion chromatography-multi-angle light scattering (SEC-MALS) analysis of 1:1:1 mixture of FGF23, FGFR1c and α Klotho in the absence (green line) or presence of HS (blue line). Horizontal dashed line indicates molecular mass of the main peak.



Supplementary Fig.3. Hypothetical 1:2:2:1 or 2:4:2:2 FGF23-FGFR- α Klotho-

HS quaternary complexes.

a, Cryo-EM maps of FGF23-FGFR1c-αKlotho-HS (left), FGF23-FGFR3c- α Klotho-HS (center) and FGF23-FGFR4- α Klotho-HS (right) quaternary complexes shown at low threshold levels of 0.4, 0.3 and 0.3, respectively. Note the presence of an extra weak density (in black dashed box) adjacent to the 1:2:1:1 quaternary complex. **b**, Top: Surface rendering in two views showing that the extra density represents an additional α Klotho molecule (α Klotho^s in wheat color) that packs via its KL2 domain against the KL2 domain of α Klotho component (α Klotho^P) of 1:2:1:1 guaternary complexes (colored as in Fig. 2a). Beneath the surface representation, the 1:2:1:1 FGF23-FGFR- α Klotho-HS guaternary complex and α Klotho^S are pulled apart and rotated along the vertical axis to expose the α Klotho^P- α Klotho^S contact sites (in purple). Cartoon representation of the boxed region highlighting residues (rendered in purple sticks) at the α Klotho- α Klotho interface. **c**. Based on this observation, we contemplated the possibility that FGF23 signaling may entail higher order assemblies featuring a 1:2:2:1 (left) or a 2:4:2:2 stoichiometry (right), the latter resulting from symmetric apposition of two sets of 1:2:1:1 FGF23-FGFR1caKlotho-HS quaternary complexes. Such higher order complexes seemed plausible particularly on cell membrane where reduced dimensionality would promote weak interactions between transmembrane α Klotho molecules. **d-f**. FGF23 signaling does not require 1:2:2:1 or 2:4:2:2 FGF23-FGFR1c-αKlotho-HS complexes. Wild-type transmembrane α Klotho (α KlothoTM) or its mutated versions harboring either a N782H/F784S/Y788R or D776A/N779A/Q780S triple mutation - introduced to disrupt α Klotho- α Klotho contacts - were coexpressed with FGFR1c^{WT} in L6 cell lines. Expression levels of FGFR1c and α KlothoTM were confirmed by western blotting (**d**). These cell lines were exposed to a fixed concentration of FGF23 for increasing time intervals (e) or varying concentrations of FGF23 for a fixed time (f). FGFR1c activation/signaling was monitored by western blotting analysis of total cell lysates with antibodies against phosphorylated FGFR and its downstream signal transducers. Note that in both sets of experiments, α Klotho mutants elicited comparable capacity as wild type α KlothoTM in promoting FGF23 signaling as evident by similar levels of FGFR1c/PLC γ 1/FRS2 α /ERK phosphorylation. These cell-based mutagenesis data strongly argued against a physiological role for higher order 1:2:2:1 or 2:4:2:2 FGF23-FGFR1c-aKlotho-HS assemblies in FGF23 signaling and accordingly, we did not consider them Experiments were performed in biological triplicates with similar further. results.

Continued



Supplementary Fig.4. FGFR^P-FGFR^S contacts deprive FGFR^S from FGF23 binding by inducing a D3 rotation and by occluding ligand binding sites in FGFR^S.

a, Left: ribbon diagram of 1:2:1 FGF23-FGFR1c-HS component of the quaternary complex. α Klotho is omitted for simplicity. Center: superimposition of FGFR^s (cyan) onto FGFR^P (pale green) via D2 domain alignment reveals a large rotation of D3 domain of FGFR^s pivoted around Cterminal pole of D2 domain. This rotation is caused by contacts with FGFR^P and FGF23 and renders the conformation of FGFR^S incompatible with FGF23 binding. Right: zoomed-in view of the boxed region showing that if another FGF23 were to bind FGFR^S, its core region would sterically clash with the FGFR^P is omitted for clarity. FGFR^s D3 domain. **b**, Center: Cartoon representation of the quaternary complex with boxes demarcating the contacts involving D2-D3 linker and top (N-terminal) end of D3 domain of FGFR^P and FGFR^s chains. Left: expanded view of the boxed region showing that the D2-D3 linker and the β F- β G loop in D3 of FGFR^P play critical roles in ligand binding. Specifically, Arg-250 from the D2-D3 linker and Ser-346 from the β F- β G loop make hydrogen bonds with Asp-125 and His-117 of FGF23, respectively. Right: expanded view of the boxed region showing that the D2-D3 linker and β F- β G loop of FGFR1^s are trapped in the FGFR^P-FGFR^s dimer interface and hence are inaccessible for FGF23.



Supplementary Table 1. List of contacts at sites 1 and 2 of the asymmetric $FGFR^{P}$ -FGFR^S interface within the FGF23-FGFR1c- α Klotho-HS quaternary complex.

| Contact sites | Residues of FGFR ^P | Residues of FGFR ^s |
|----------------------------------|-------------------------------|-------------------------------|
| site 1 (hydrophobic interaction) | Ala-170, Ala-171, | lle-347 |
| | Val-221, Pro-222 | |
| site 1 (hydrogen bonds) | Ala-171, Ser-219 | Arg-250 |
| site 1 (hydrogen bonds) | Asp-218, Ser-219 | Lys-172 |
| site 2 (hydrophobic interaction) | lle-256 | lle-256, Tyr-280 |
| site 2 (hydrogen bonds) | Glu-249 | Arg-254 |
| site 2 (hydrogen bonds) | Arg-250 | Pro-252 |
| site 2 (hydrogen bonds) | Leu-257 | Gln-258 |
| site 2 (hydrogen bonds) | Ala-259 | Ala-259 |

Supplementary Table 2. List of interactions at the FGF23-FGFR^S interface within the FGF23-FGFR1c- α Klotho-HS quaternary complex.

| Residues of FGF23 | Residues of FGFR ^s |
|---------------------------------|-------------------------------|
| Pro-151, Leu-120 (core) | lle-203 |
| Glu-121 (core) | Ser-223 |
| Asn-150 (core) | Asp-200, Arg-202 |
| Try-25, Pro-26 (N-terminus) | Leu-290, Leu-342, Leu-349 |
| Ser-29 (N-terminus) | Arg-254, Ser-350 |
| Leu-31, and Leu-32 (N-terminus) | lle-256, Ala-259 |