

Tubular aggregates: their association with myalgia

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SUMMARY Three thousand consecutive muscle biopsies were reviewed for the presence of tubular aggregates and their association with clinical symptomatology. Tubular aggregates were detected in 19 patients (0.6%). Twelve of these nineteen patients had severe myalgia, and the most abundant tubular aggregates were found in biopsies of patients with myalgia. Seven patients had only myalgia as their clinical symptomatology with normal physical examination. An additional five patients with tubular aggregates and myalgia had concomitant amyotrophic lateral sclerosis (2) or neuropathy (3). The high incidence of myalgia associated with tubular aggregates in our patients and the fact that tubular aggregates originate from sarcoplasmic reticulum suggest a role played by this structure in the pathogenesis of myalgia.

Tubular aggregates are visible by light microscopy within Type II muscle fibres as a mass in the subsarcolemmal region with strong NADH-tetrazolium reductase (NADH-TR) and negative succinic dehydrogenase reactions. Ultrastructurally they are closely packed, parallel, double-walled tubules which appear to be a proliferation of sarcoplasmic reticulum.¹ The diagnostic importance of tubular aggregates is uncertain because they are present in muscles of patients with a wide variety of disorders.¹⁻²¹ Recently tubular aggregates have been described in muscle biopsy specimens of patients with myalgia who are otherwise normal.²¹⁻²⁶ The fact that tubular aggregates originate from sarcoplasmic reticulum, which has an important role in regulation of muscle contracture and relaxation, suggests a relationship between tubular aggregates, sarcoplasmic reticulum and myalgia. To clarify this relationship, we reviewed muscle biopsy specimens for the presence of tubular aggregates and their association with clinical symptomatology.

Methods

We analysed all 3000 muscle biopsy specimens at our institution in the preceeding seven years. All were from the

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quadriceps or biceps muscles. For histochemical studies, tissue had been frozen with liquid nitrogen, sectioned at 10 μ in a cryostat, and stained with the following methods: Modified Gomori Trichrome (TRI), NADH-TR, adenosine triphosphatase (ATPase) at pH 9.4, succinic dehydrogenase and phosphorylase. For electron microscopy, tissue had been fixed in 1% glutaraldehyde buffered at pH 7.4 for 1½ h and then fixed in 1% osmium tetroxide for another hour. Some material was then embedded in Epon and sections 1 μ thick were cut for all blocks, stained with toluidine blue and examined under the light microscopy. Thin sections for electron microscopy were then stained with uranyl acetate and with lead citrate.

Results

Among 3000 patients 19 (0.6%) had histochemically typical tubular aggregates; 17 patients were male and two female. With the TRI stain, the tubular aggregates appeared as bright red staining subsarcolemmal material. Tubular aggregates were seen only in the Type II fibres, were darkly stained by the NADH-TR, and unstained by ATPase and succinate dehydrogenase (fig 1). Further studies including electron microscope and epoxy resin histology were done in some cases which showed closely packed, parallel, double-walled tubules (fig 2).

Five biopsies showed less than 1% fibres affected with tubular aggregates, six between 1-10%, and eight had more than 10% fibres with tubular aggregates. This was determined after obtaining a 100 \times photograph of each biopsy specimen and counting the number of tubular aggregates per 200 muscle fibres.

Clinically, 12 of 19 patients had severe myalgia,

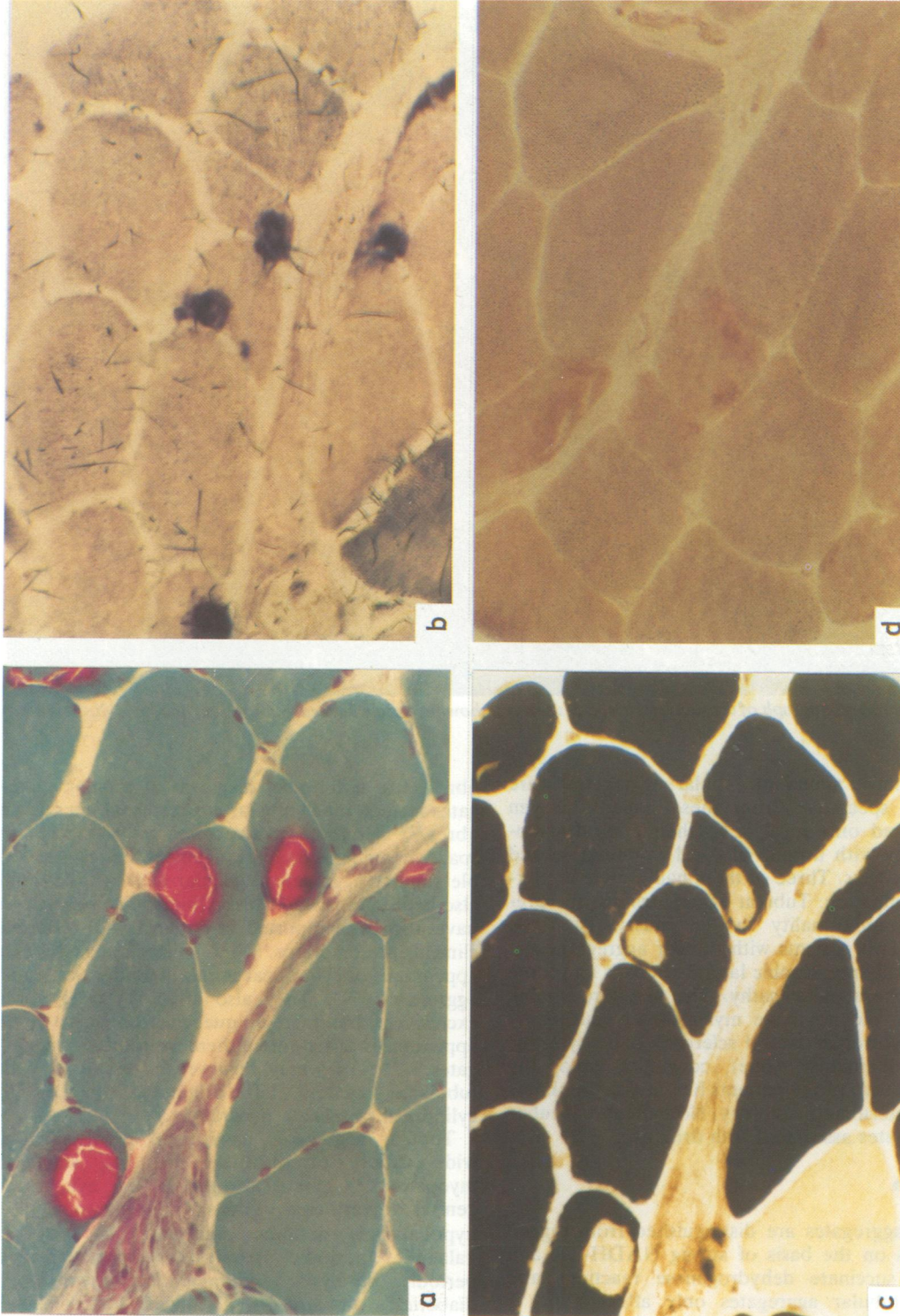


Fig 1 Histochemical demonstration of the tubular aggregates in cross section of the quadriceps muscle. The tubular aggregates are seen as bright red staining subsarcolemmal mass with the TRJ (A), darkly stained with the NADH-TR (B) and unstained with ATPase and succinic dehydrogenase (C and D). Type II muscle fibres are light in (B) and dark in (C). ($\times 250$)

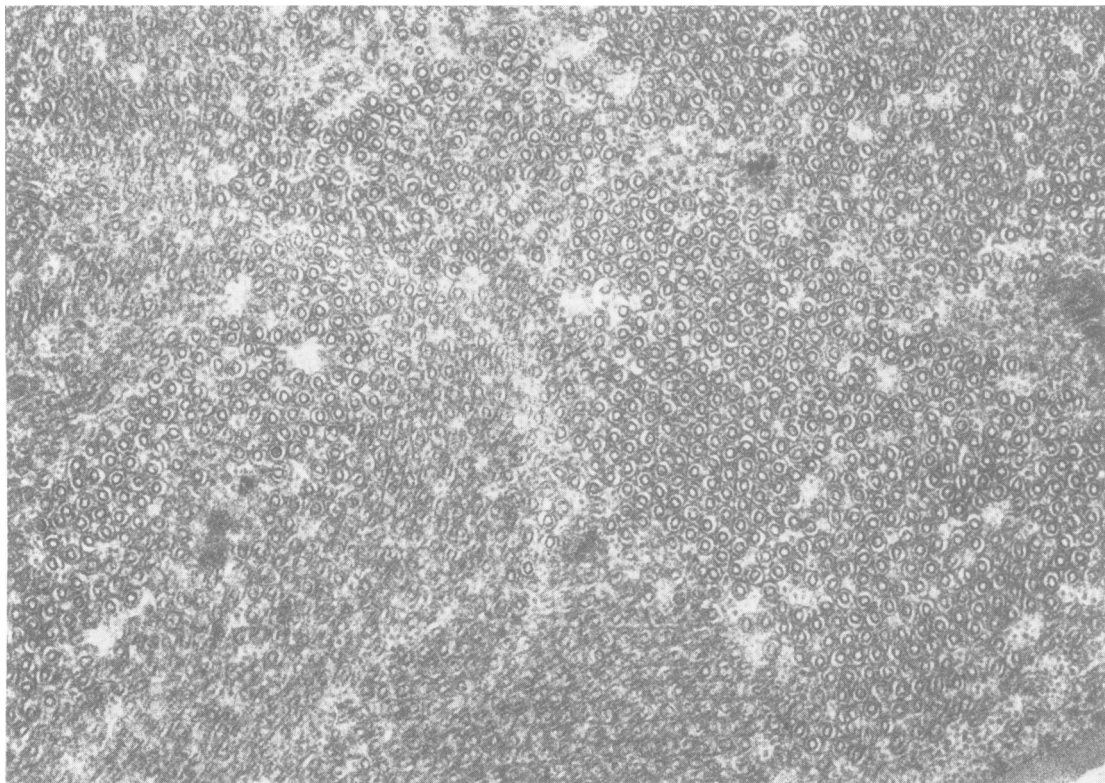


Fig 2 Electron micrograph of the same muscle, cross section, showing closely packed, parallel and double-walled tubules. ($\times 50,000$)

and the most abundant tubular aggregates were found in specimens from these patients. Seven patients had only myalgia as their clinical symptomatology with normal physical examination and laboratory tests. Three of them had muscle tenderness on palpation. Tubular aggregates was the predominant abnormality in the muscle of these patients. Two patients with tubular aggregates and myalgia had amyotrophic lateral sclerosis and three had peripheral neuropathy. The seven remaining cases who did not have myalgia had neuropathy (two) amyotrophic lateral sclerosis (one), systemic lupus erythematosus (one), congenital myopathy (one), and two of the patients had muscle weakness with undetermined aetiology. No patient had tubular aggregates associated with drug use.

Discussion

Tubular aggregates are distinguished from ragged red fibres on the basis of strong NADH-TR and negative succinate dehydrogenase reactions and because tubular aggregates only affect Type II

fibres. The differential diagnosis of tubular aggregates include other tubular structures within muscle fibres.¹²⁻¹⁶ Honeycomb structures are round empty spaces in an hexagonal arrangement that can resemble tubular aggregates. Tubular aggregates should also be differentiated from cylindrical spirals, which have also been reported in patients with different clinical presentations.¹¹⁻²⁷⁻²⁹ Histochemically, the appearance of cylindrical spirals is similar to tubular aggregates.¹¹⁻²⁷⁻²⁹ They also affect Type II fibres exclusively, but their unique electron microscopic appearances differentiate them from tubular aggregates.¹¹⁻²⁷⁻²⁹ They may, however, be associated with tubular aggregates.¹¹⁻²⁷ The origin and significance of cylindrical spirals are uncertain.

Tubular aggregates have been reported in a wide variety of disorders including alcoholic myopathy,¹⁶⁻¹⁷ gyrate atrophy of the eye,² congenital myasthenia gravis,⁴⁻⁶⁻¹⁴ myotonia,⁷⁻¹⁵⁻²¹ hypokalaemic periodic paralysis,¹⁸⁻⁹⁻¹⁸⁻²¹ hyperkalaemic periodic paralysis,¹ normokalaemic periodic paralysis,¹³ inflammatory myopathy,²¹ diabetic amyotrophy,³ hyperaldosteronism,¹²

prophyria cutanea tarda¹ and chronic drug exposure.¹ A similar abnormality has been found in muscles of mice after anoxia³⁰ and after local injection of botulinum,³¹ tetanus toxin,³¹ and perhexiline maleate³³ and in murine dystrophic heterozygotes.³⁴ Tubular aggregates have also been described in association with myalgia in otherwise normal individuals.²¹⁻²⁶ Twelve of 19 of our patients with tubular aggregates had severe myalgia. Seven patients had myalgia as their sole clinical symptomatology and despite extensive investigation, no definite diagnosis could be made. Like patients reported by Rosenberg *et al.*,²¹ myalgia was not necessarily precipitated by exercise. Three of these cases had severe muscle tenderness on palpation. Tubular aggregates was the prominent abnormality in the muscle of these patients. Five other patients with tubular aggregates and myalgia had concomitant amyotrophic lateral sclerosis or neuropathy. In these patients myalgia was one of their major complaints.

It is generally accepted that tubular aggregates originate in the sarcoplasmic reticulum, but their pathogenesis is unknown. Based on the finding of tubular aggregates in muscle biopsies of patients chronically using large amounts of analgesic-narcotic drugs and alcohol,^{1, 16} it has been postulated that they may form in response to exogenous or endogenous toxin. However, these reports failed to indicate why these medications were taken or why the muscle biopsies were performed,^{1, 16} and it may be that the reason for chronic analgesic use was for relief of myalgia.¹⁶ It has also been suggested that tubular aggregates may represent adaptive hyperplastic sarcoplasmic reticulum derivatives to improve calcium uptake in alcoholic myopathy.¹⁷ However, there was no history of chronic drug use or alcoholic abuse in our cases. We also doubt that this is a virus-induced lesion as postulated by Dunkle *et al.*¹⁸ and Bergman,^{8, 9} since no evidence of viral infection of muscle was seen. There is thus no satisfactory explanation for the formation of tubular aggregates, nor for their occurrence in a wide variety of diseases, but there may be an explanation of the high incidence of myalgia associated with them.

Sarcoplasmic reticulum has an important role in contraction and relaxation of muscle. The function of the sarcoplasmic reticulum is regulation of the calcium concentration in the vicinity of actin and myosin interaction. This regulation of calcium concentration is the control factor for turning on and off the basic contractile unit, the sarcomere. It is well known that malfunction of sarcoplasmic reticulum interferes with normal contraction and relaxation, and it may cause myalgia. The fact that tubular aggregates originates from sarcoplasmic reticulum and the high incidence of myalgia associated with

tubular aggregates in our patients and previous reports of similar cases,²¹⁻²⁶ suggest a role played by this structure in the pathogenesis of myalgia. The relationship between tubular aggregates, sarcoplasmic reticulum and calcium metabolism may also explain why tubular aggregates are seen only in the Type II fibres. Type II fibres have considerably more sarcoplasmic reticulum than the Type I fibres.³⁵ In Type II fibres, the sarcoplasmic reticulum is possibly qualitatively different from that of the Type I fibre.¹ Also it has been shown in rabbits that white muscle fibres (Type II) have more active calcium-concentrating microsoms than red muscle fibres (Type I).³⁶ These structural differences may predispose Type II fibres to tubular aggregates formation.

In spite of relationships between tubular aggregates, sarcoplasmic reticulum and myalgia, tubular aggregates are not always associated with myalgia. There may be different types of tubular aggregates,^{12, 16, 17, 37, 38} but there are classic forms of tubular aggregates in patients who never complain of myalgia. Also, in our patients who had electron microscopic studies, the pattern of tubular aggregates was similar in cases with or without myalgia. Degree of muscle fibre involvement with tubular aggregates may be an important factor for appearance of myalgia since the most abundant tubular aggregates were found in our patients with myalgia.

Based on our observation and review of literatures, we suggest the association of myalgia and tubular aggregates may be important. Although myalgia is the most prominent feature in the symptomatology of a wide variety of conditions, in many patients no firm diagnosis can be made despite extensive investigation.³⁹ Therefore, the presence of tubular aggregates in patients complaining of myalgia may be a hallmark and should not be regarded as a nonspecific finding. Recognition of these combinations in time may lead to a better understanding of the significance of tubular aggregates. This relationship between tubular aggregates, sarcoplasmic reticulum and myalgia may also have important therapeutic implications, since drugs which are known to affect these structures may prove beneficial in alleviating symptoms of myalgia.

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References

- 1 Engel WK, Bishop DW, Cunningham GG. Tubular aggregates in Type II muscle fibers: Ultrastructural and histochemical correlation. *Ultrastructure Research* 1970;31:507-25.

- ² Sipila I, Simell O, Rapola J, *et al.* Gyrate atrophy of the choroid and retina with hyperornithinemia: Tubular aggregates and Type II fibre atrophy in muscle. *Neurology (Minneapolis)* 1979; **29**:996-1005.
- ³ Chokroverty S, Reyes MG, Rubino FA, Tonaki H. The syndrome of diabetic amyotrophy. *Ann Neurol* 1977; **2**:181-4.
- ⁴ Morgan-Hughes JA, Lecky BR, London DN, Murray NMF. Alteration in the number and affinity of junctional acetylcholine receptors in a myopathy with tubular aggregates. *Brain* 1981; **104**:279-95.
- ⁵ Engel AG, Lambert EH, Mulder DM, *et al.* A newly recognized congenital myasthenic syndrome attributed to a prolonged open time of the acetylcholine-induced ion channel. *Ann Neurol* 1982; **11**:553-69.
- ⁶ Dobkin BH, Verity MA. Familial neuromuscular disease with Type I fibre hypoplasia, tubular aggregates, cardiomyopathy, and myasthenic features. *Neurology (Minneapolis)* 1978; **28**:1135-40.
- ⁷ Schotland D. Ultrastructural abnormalities in myotonic dystrophy including an unusual T system alteration. *J Neuropathol Exp Neurol* 1968; **27**:109-10.
- ⁸ Bergman RA, Afifi AK, Dunkle LM, Johns RJ. Muscle pathology in hypokalemic periodic paralysis with hyperthyroidism. *Hopkins Med J* 1970; **126**:88-94.
- ⁹ Bergman RA, Afifi AK, Dunkle LM, Johns RJ. Muscle pathology in hypokalemic periodic paralysis with hyperthyroidism. *Hopkins Med J* 1970; **126**:101-18.
- ¹⁰ Odor DL, Patel AN, Pearce LA. Familial hypokalemic periodic paralysis with permanent myopathy. *J Neuropathol Exp Neurol* 1967; **26**:98-114.
- ¹¹ Carpenter S, Karpati G, Robitaille Y, Melmed C. Cylindrical spirals in human skeletal muscle. *Muscle Nerve* 1979; **2**:282-7.
- ¹² Gallas M. Myopathy with hyperaldosteronism. *J Neurol Sci* 1977; **32**:337-45.
- ¹³ Meyers KR, Gilden DH, Rinaldi CF, Hansen JL. Periodic muscle weakness, normokalemia and tubular aggregates. *Neurology (Minneapolis)* 1972; **22**:269-79.
- ¹⁴ Johns TR, Campa JF, Adelman LS. Familial myasthenia with "tubular aggregates" treated with Prednisone. *Neurology (Minneapolis)* 1973; **23**:426.
- ¹⁵ Schiffer D, Giordana MT, Monga G, Mollo F. Histochemistry and electron microscopy of muscle fibres in a case of congenital paramyotonia. *J Neurol* 1976; **211**:125-33.
- ¹⁶ Chui L, Neustein H, Munsat T. Tubular aggregates in subclinical alcoholic myopathy. *Neurology (Minneapolis)* 1975; **25**:405-12.
- ¹⁷ Negro ADV, Angulo JM, Pomar JMR, Errasti CA. Tubular aggregates in skeletal muscle of chronic alcoholic patients. *Acta Neuropathol (Berl)* 1982; **56**:250-4.
- ¹⁸ Dunkle LM, Diggs CH, Bergman RA, Johns RJ. A light and electron microscopic study of a second case of hypokalemic periodic paralysis with hyperthyroidism. *Hopkins Med J* 1970; **126**:225-36.
- ¹⁹ Faugere MC, Pellissier JF, Toga M. Subsequent morphological changes in periodic paralysis: A study of seven cases. *Act Neuropathol (Berl)* 1981; **VII**:301-4.
- ²⁰ Dyken M, Zeman W, Rusche T. Hypokalemic periodic paralysis, children with permanent myopathic weakness. *Neurology (Minneapolis)* 1969; **19**:691-9.
- ²¹ Rosenberg NL, Neville HE, Ringel SP. Tubular aggregates: Their association with clinical symptomatology. *Neurology (NY)* 1983; **33**(Suppl 2):236.
- ²² Morgan-Hughes JA, Mair WGP, Lascelles PT. A disorder of skeletal muscle associated with tubular aggregates. *Brain* 1970; **93**:873-80.
- ²³ Brumback RA, Staton RD, Susag ME. Exercise-induced pain, stiffness, and tubular aggregates in skeletal muscle. *J Neurology, Neurosurg, Psychiatry* 1981; **44**:250-4.
- ²⁴ Smith R, Hughes R, Borensztajn J, *et al.* Focal muscle cramp in a young man, possible myopathy with tubular aggregates. *Chest* 1983; **84**:795-800.
- ²⁵ Lewis PD, Pallis C, Pearse AGE. "Myopathy" with tubular aggregates. *J Neurol Sci* 1971; **13**:381-8.
- ²⁶ Lazaro RP, Fenichel GM, Kilroy AW, *et al.* Cramps, muscle pain, and tubular aggregates. *Arch Neurol* 1980; **37**:715-7.
- ²⁷ Gibbels E, Henke U, Shadlich HJ, *et al.* Cylindrical spirals in skeletal muscle: A further observation with clinical, morphological and biochemical analysis. *Muscle Nerve* 1983; **6**:646-55.
- ²⁸ Bove KE, Iannaccone ST, Hilton PK, Samaha F. Cylindrical spirals in a familial neuromuscular disorder. *Ann Neurol* 1980; **7**:550-6.
- ²⁹ McDougall J, Wiles CM, Edwards RHT. Spiral membrane cylinders in the skeletal muscle of a patient with melorheosis. *Neuropathol Appl Neurobiol* 1980; **6**:69-74.
- ³⁰ Schiaffino S, Severin E, Cantini M, Sartore S. Tubular aggregates induced by anoxia in isolated rat skeletal muscle. *Lab Investigation* 1977; **37**:223-8.
- ³¹ Duchon LW. Changes in the electron microscopic structure of slow and fast skeletal muscle fibres of the mouse after the local injection of botulinum toxin. *J Neurol Sci* 1971; **14**:61-74.
- ³² Duchon LW. The local effects of tetanus toxin on the electron microscopic structure of skeletal muscle fibres of the mouse. *J Neurol Sci* 1973; **19**:169-77.
- ³³ Fardeau M, Tome FMS, Simon P. Muscle and nerve changes induced by perhexiline maleate in man and mice. *Muscle Nerve* 1979; **2**:24-36.
- ³⁴ Craig ID, Allen IV. Tubular aggregates in murine dystrophy heterozygotes. *Muscle Nerve* 1980; **3**:134-40.
- ³⁵ Cullen MJ, Weightman D. The ultrastructure of normal human muscle in relation to fibre type. *J Neurol Sci* 1975; **25**:43-56.
- ³⁶ Streter FA. Comparative studies on white and red muscle fractions. *Fed Proc* 1964; **23**:930-2.
- ³⁷ Price HM, Howes EL, Blumberg JA. Ultrastructural alteration of skeletal muscle fibres injury by cold: The acute degenerative changes. *J Lab Invest* 1964; **13**:1264-78.
- ³⁸ MacDonald RD, Rencastle NB, Humphrey JG. The myopathy of hyperkalemic period paralysis. *Arch Neurol* 1968; **19**:274-84.
- ³⁹ Mills KR, Edwards RHT. Investigation strategies for muscle pain. *J Neurol Sci* 1983; **58**:73-88.