

Supplementary Material

***O*-GlcNAcylation of RIPK1 Rescues Red Blood Cells from Necroptosis**

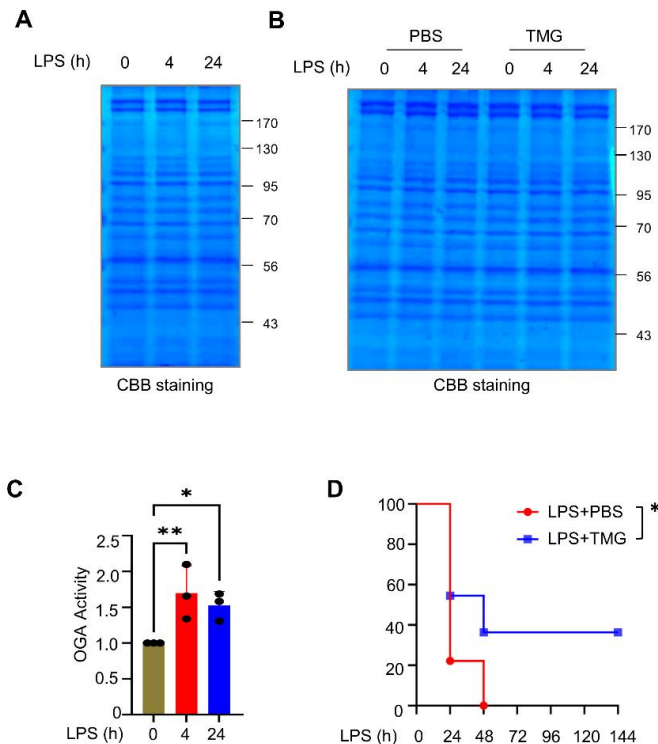
Junghwa Seo¹, Yeolhoe Kim^{1,2}, Suena Ji¹, Han Byeol Kim³, Hyeryeon Jung^{1,3}, Eugene C. Yi^{1,3}, Yong-ho Lee^{1,4}, Injae Shin^{1,5}, Won Ho Yang^{1,2*}, Jin Won Cho^{1,2*}

Correspondence: Jin Won Cho: chojw311@yonsei.ac.kr

Won Ho Yang: bionicwono@yonsei.ac.kr

1.1 Supplementary Figures

Supplementary Fig. 1



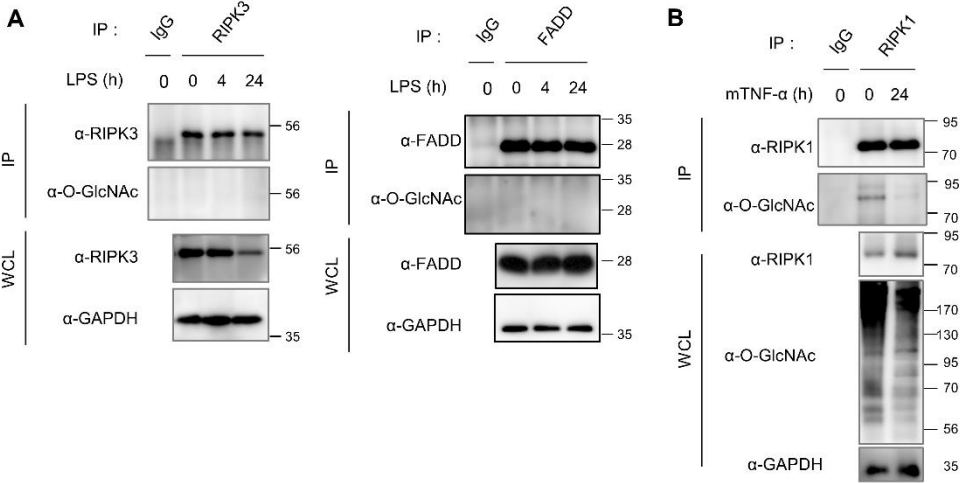
Supplementary Figure 1. OGA activity and lethality in LPS-injected mouse.

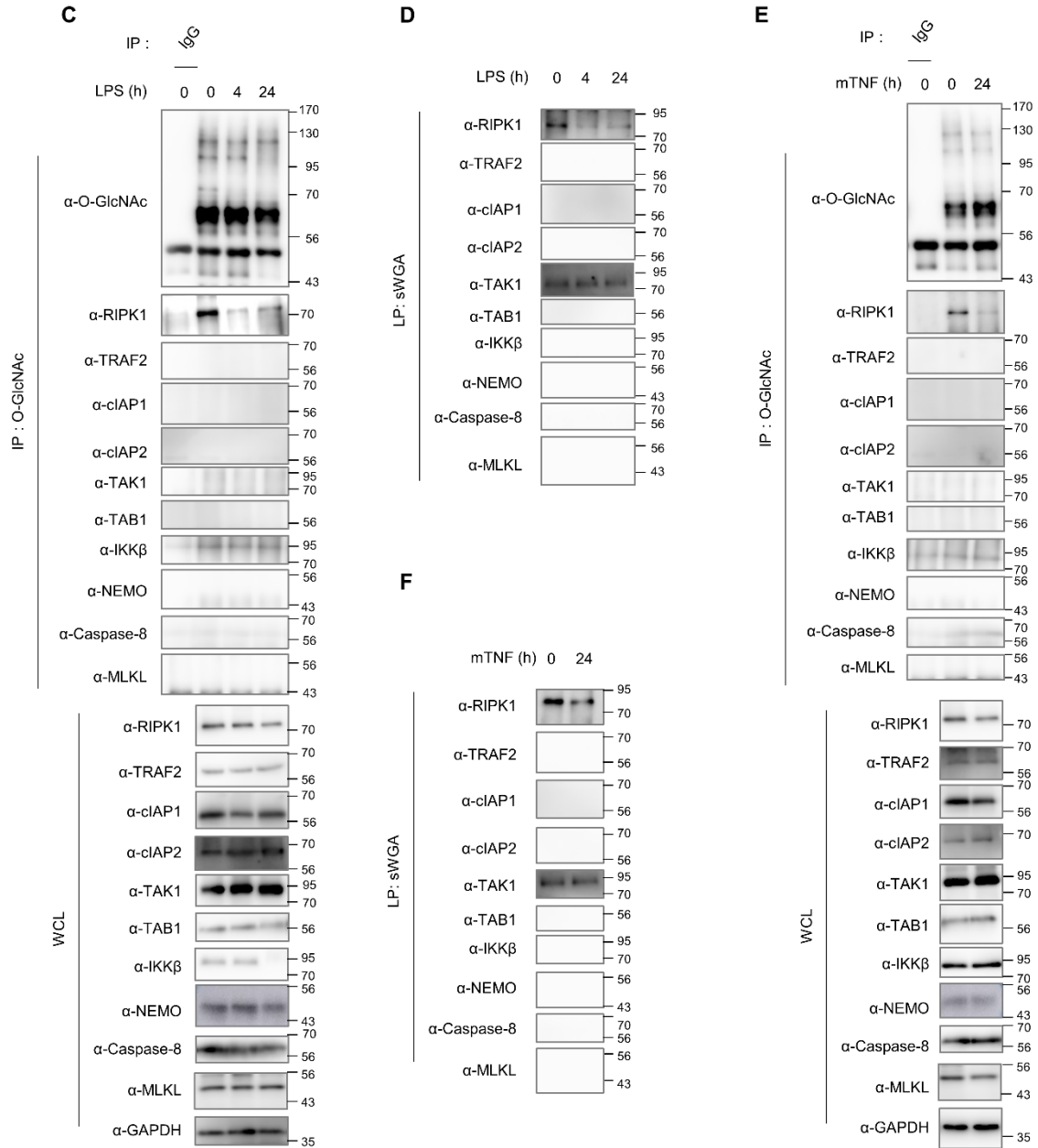
(A, B) Coomassie blue staining of total protein levels in lysates from Fig. 1A or Fig. 1B.

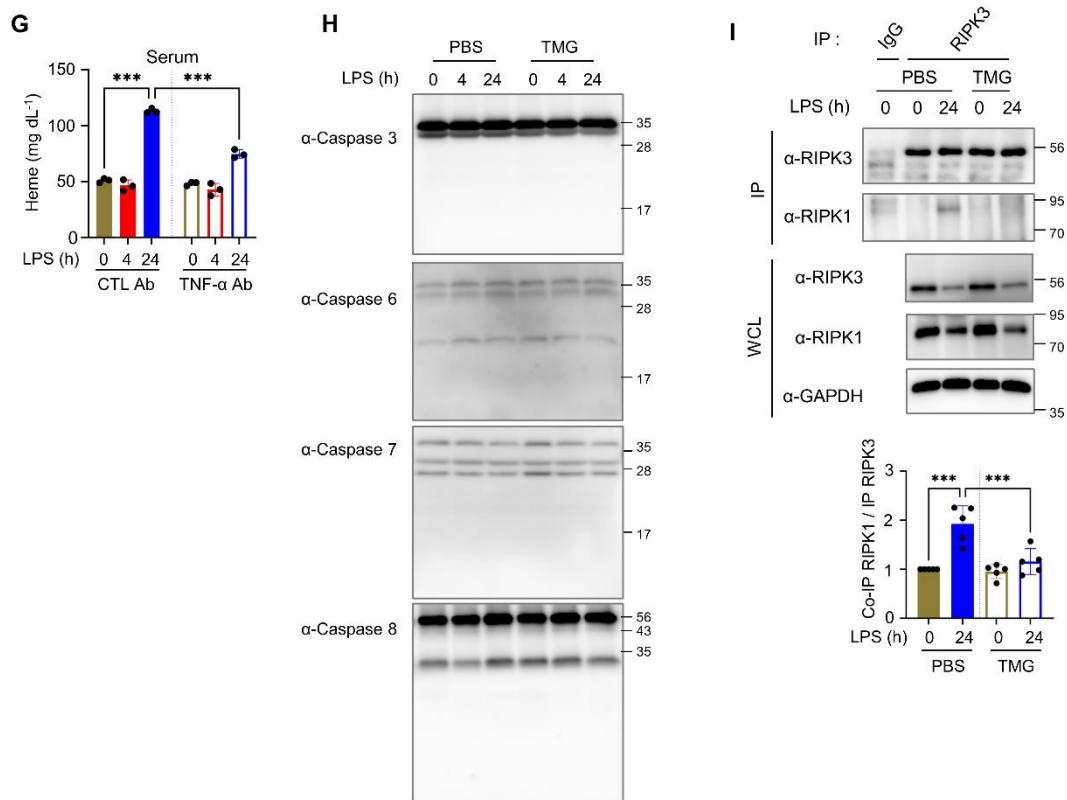
(C) Measurement of OGA activity of erythrocytes following LPS injection (A; one-way ANOVA with Sidak's multiple comparisons test; n=3)

(D) Survival measurement showing the effect of TMG administration on LPS-injected mice. Survival analysis is presented as a Kaplan–Meyer survival curve and was evaluated using a log-rank (Mantel–Cox) test; 20 mg kg⁻¹ LPS, 40 mg kg⁻¹ TMG, or the same volume of PBS (as a control) (LPS + PBS group [n]=9), (LPS + TMG group [n]=11). statistical significance was annotated as *P < 0.05, **P < 0.01.

Supplementary Fig. 2







Supplementary Fig. 2. Investigation of *O*-GlcNAcylation and RIPK3-RIPK1 interaction of RIPK3 in erythrocytes.

(A) Immunoprecipitation (IP) assay to determine the *O*-GlcNAcylation of endogenous RIPK3 (left) and FADD (right) in erythrocytes isolated from LPS (10 mg kg⁻¹ day⁻¹)-injected mice during the indicated times. (A: the same amount of RIPK3 or FADD protein was immunoprecipitated from erythrocyte whole cell lysate.)

(B) Immunoprecipitation (IP) assay to determine the *O*-GlcNAcylation of endogenous RIPK1 in erythrocytes isolated from TNF-α (250 μg kg⁻¹) -injected mice.

(C, E) Immunoprecipitation (IP) assay to examine the *O*-GlcNAcylation of indicated proteins in erythrocytes isolated from LPS (10 mg kg⁻¹ day⁻¹) or TNF-α (250 μg kg⁻¹) -injected mice using an *O*-GlcNAc antibody.

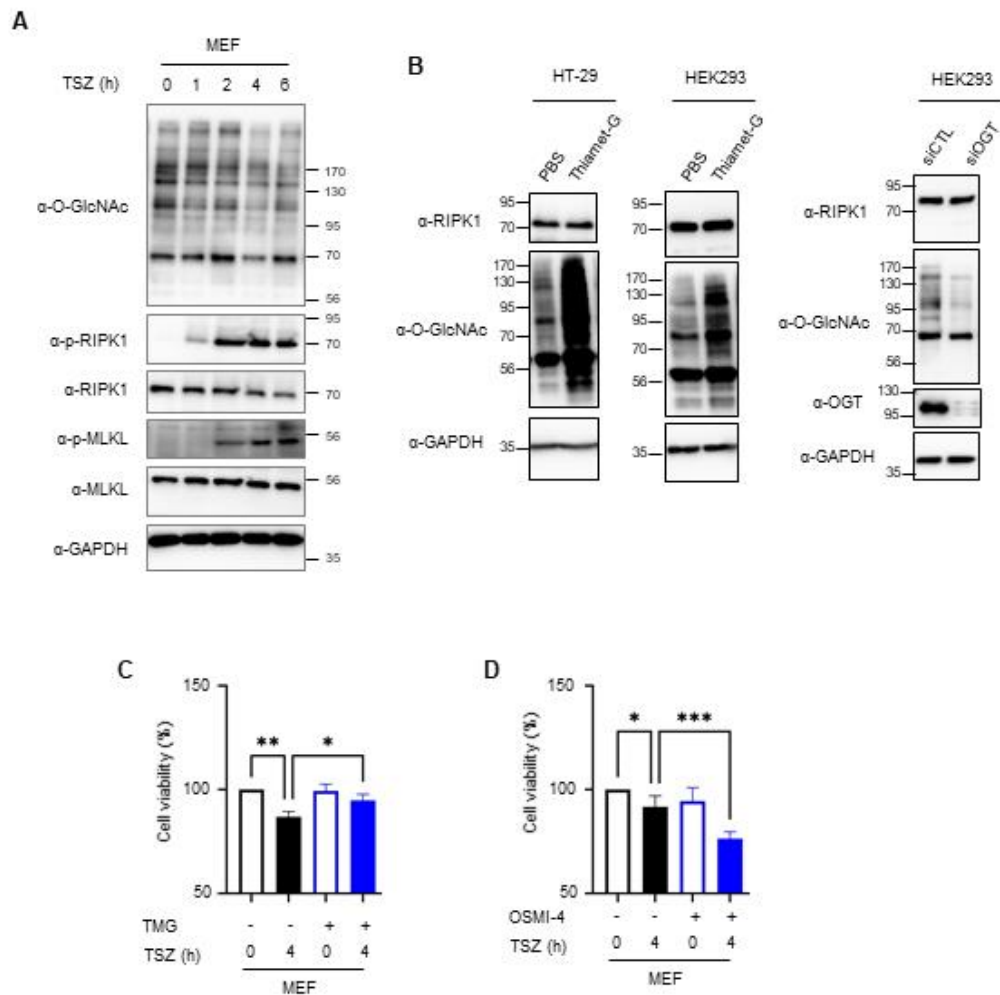
(D, F) Succinylated-wheat germ agglutinin (sWGA) assay to examine the *O*-GlcNAcylation of indicated proteins in erythrocytes isolated from LPS (10 mg kg⁻¹ day⁻¹) or TNF-α (250 μg kg⁻¹) -injected mice.

(G) Heme assay to measure the concentration of free heme released into the serum. Serum was harvested at the indicated times after i.p. injection with 10 mg kg⁻¹ day⁻¹LPS and 10 mg kg⁻¹ day⁻¹ TNF-α antibody or control antibody.

(H) Western blot analysis showing the levels of Caspase-3,6,7 and 8 after LPS injection in erythrocytes.

(I) Left: Co-IP assay showing the interaction between RIPK3 and RIPK1 in erythrocytes after a 24h LPS ($10 \text{ mg kg}^{-1} \text{ day}^{-1}$) injection. Right: graph showing the amount of co-immunoprecipitated RIPK1 normalized with immunoprecipitated RIPK3. (**G, I**; one-way ANOVA with Sidak's multiple comparisons tests; (G: n=3) (I: n=5). Data are presented as the mean \pm standard deviation (SD); statistical significance was annotated as ***P < 0.001.

Supplementary Fig. 3



Supplementary Fig. 3. The decline of *O*-GlcNAcylation following TSZ stimulation and regulation of cell viability by TMG or OSMI-4 treatment in MEF cells.

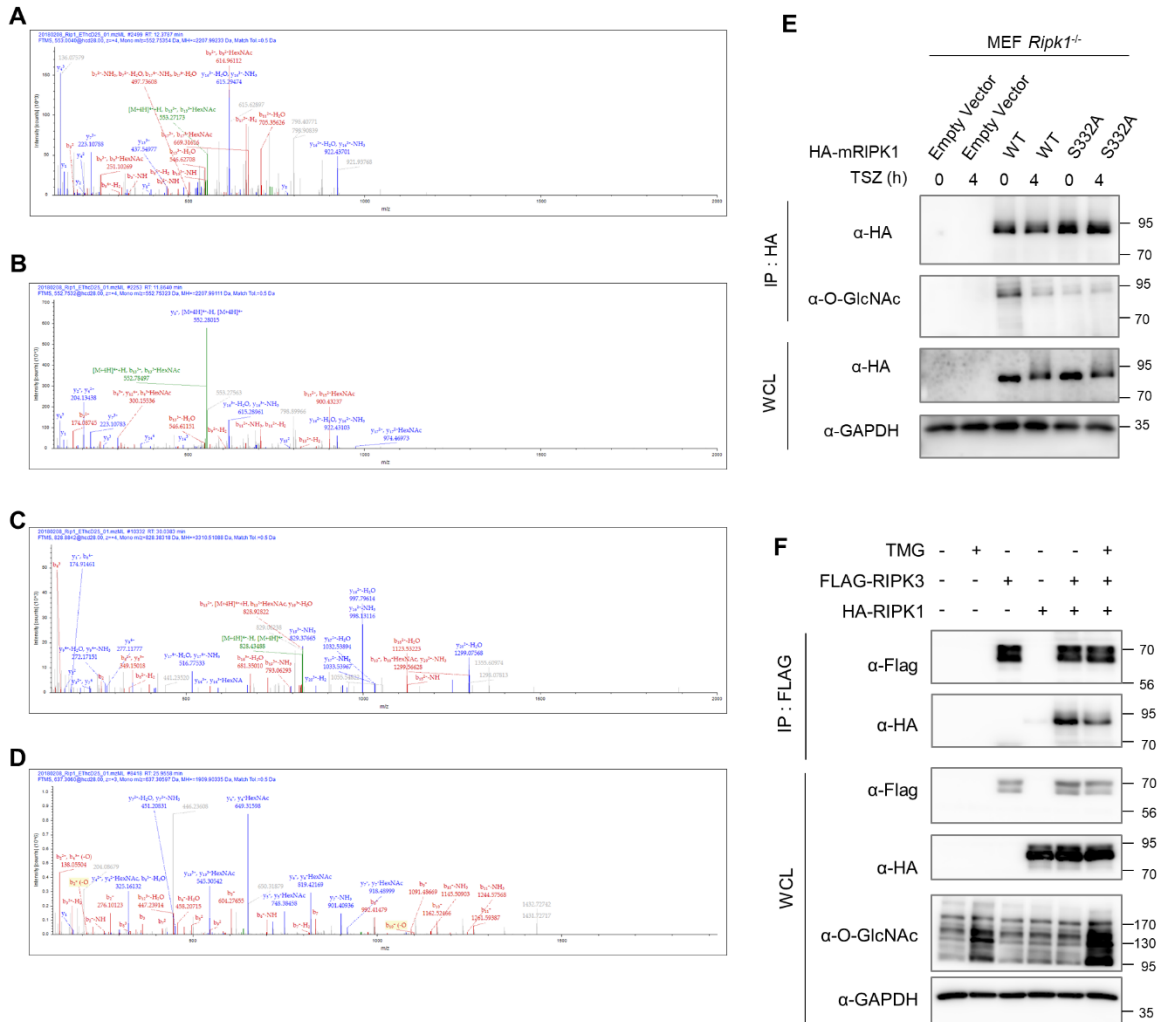
(A) Western blot analysis showing the levels of *O*-GlcNAcylation after TSZ stimulation in MEF cells.

(B) Western blot analysis representing the RIPK1 protein levels upon altering *O*-GlcNAcylation status in HT-29 or HEK 293 cells.

(C, D) CellTiter-Glo assay to measure the cell viability of MEFs in necroptosis following TSZ stimulation. 1 μ M TMG (C) or 2 μ M OSMI-4 (D) were pretreated before a 4h TSZ stimulation. The y-axis starts at 50%. (C: n=3) (D: n=8)

(C, D; one-way ANOVA with Sidak's multiple comparisons test). Data are presented as the mean \pm standard deviation (SD); statistical significance was annotated as * $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$.

Supplementary Fig. 4



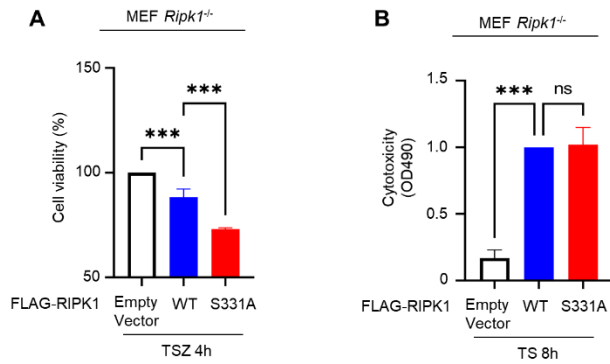
Supplementary Fig. 4. Mass spectrometry (MS) analysis to identify *O*-GlcNAcylation sites of RIPK1.

(A, B, C, D) Mass spectrometry (MS) analysis to identify *O*-GlcNAcylation sites in RIPK1. EThcD spectra of *O*-Glycopeptide (A, B, C) NVIKMKSSDFLESAG or (D) DCVAVPSSRSNSA from human RIPK1 is shown. The site of *O*-GlcNAc modification was identified as (A) serine 14, (B) serine 15, (C) serine 20, and (D) serine 330, although the moderate or low reliability. The b and y fragments detected are as indicated in the sequence.

(E) IP assay comparing *O*-GlcNAcylation levels between mouse WT RIPK1 and mouse S332A RIPK1.

(F) Co-IP analysis showing that RIPK1-RIPK3 complex formation was attenuated by TMG in HEK293 cells. 10 μg of each DNA or empty vector (-) was overexpressed for 24h.

Supplementary Fig. 5



Supplementary Fig. 5. Measurement of cell viability in RIPK1 WT or RIPK1 S331A overexpressing MEF *Ripk1*^{-/-} cells.

(A) CellTiter-Glo assay to measure the viability of MEF *Ripk1*^{-/-} cells in necroptosis following TSZ stimulation. Before a 4h TSZ stimulation, 10 μ g of each DNA construct was overexpressed for 20–24h. The y-axis starts at 50%. (one-way ANOVA with Sidak's multiple comparisons test, n=4).

(B) LDH release assay reflecting the cytotoxicity of MEF *Ripk1*^{-/-} cells in apoptosis following an 8h TS stimulation. Before stimulation, 10 μ g of each DNA construct was transfected. (one-way ANOVA with Sidak's multiple comparisons test, n=4).

Data are presented as the mean \pm standard deviation (SD); statistical significance was annotated as ***P < 0.001.