

Hedgehog signaling via its ligand DHH acts as cell fate determinant during skeletal muscle regeneration

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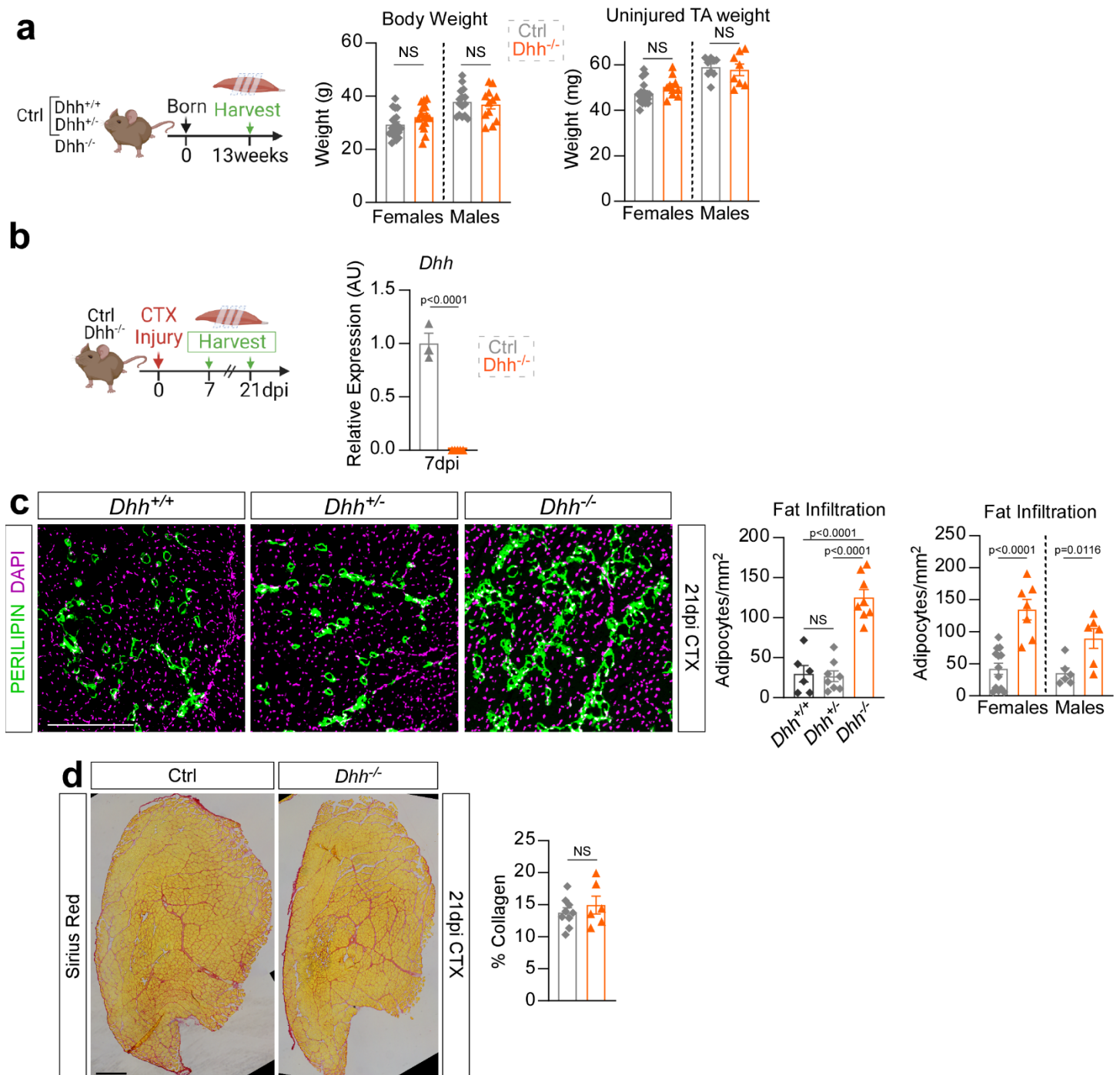
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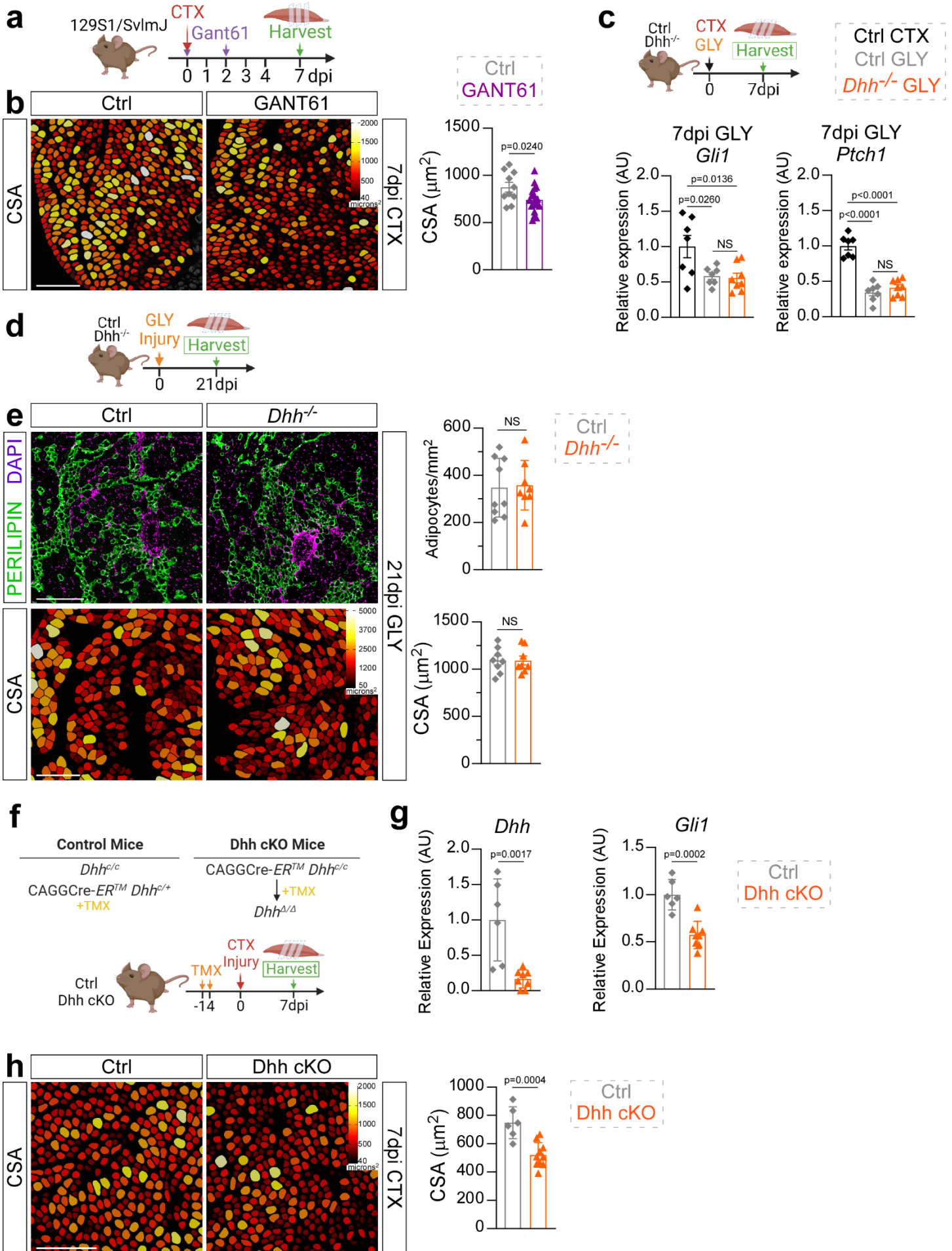
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Supplemental Figures and Table



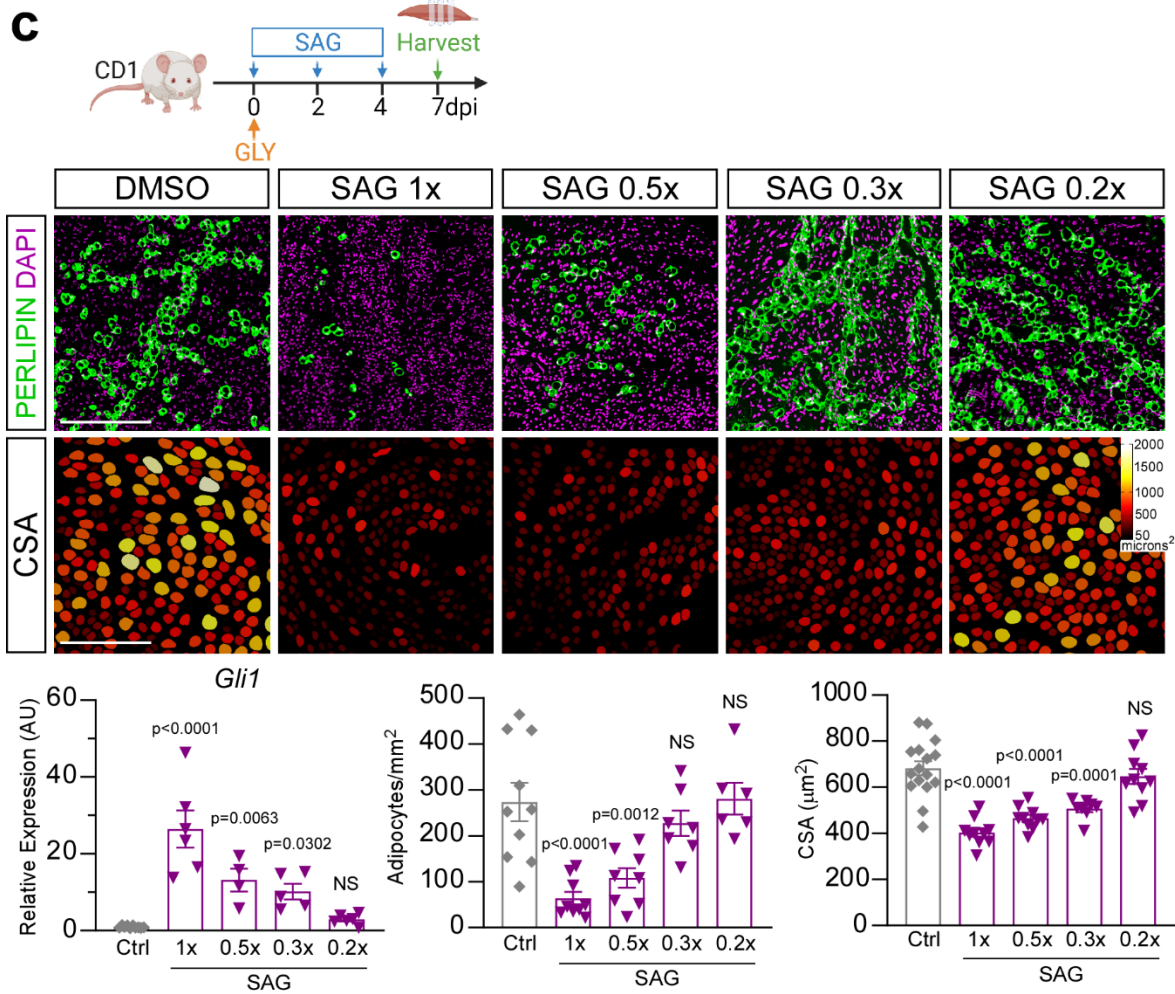
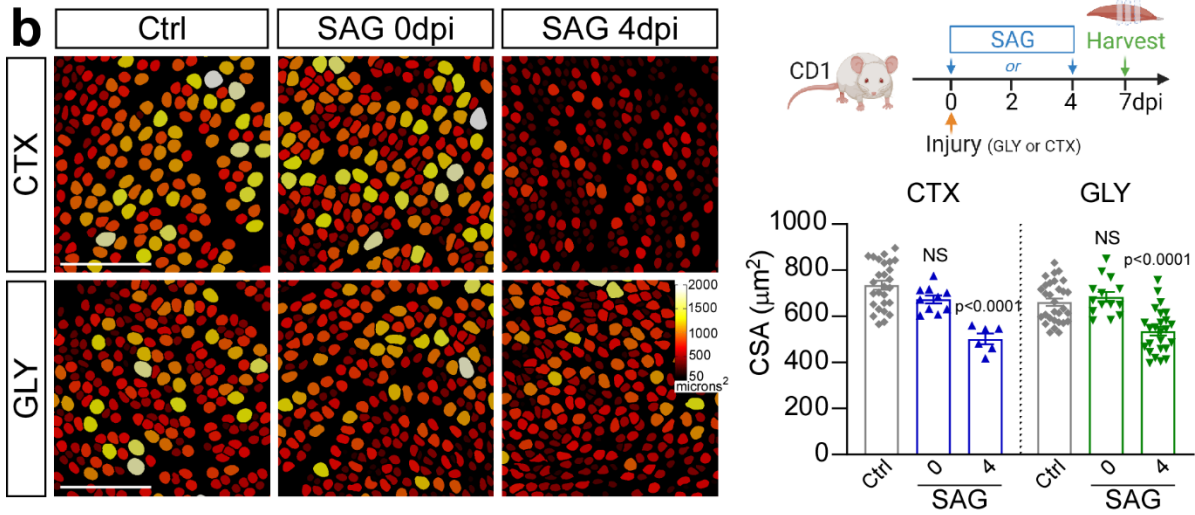
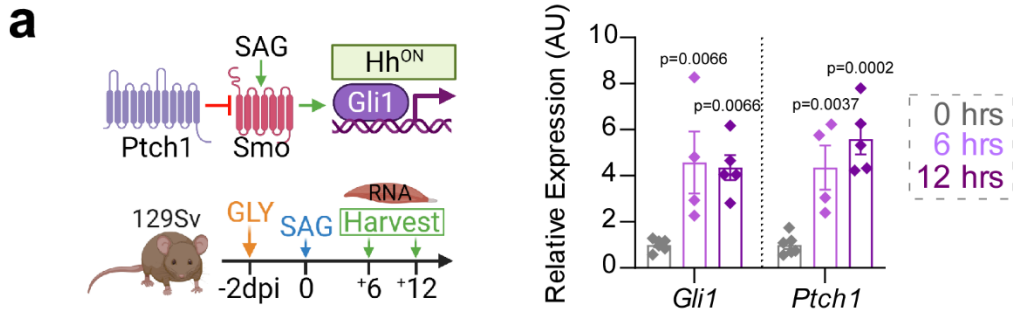
Supplemental Figure 1. Validation of the *Dhh*^{-/-} mouse model.

a) Experimental outline. Body weight (g) of 13-week-old *Dhh*^{-/-} (females, n=15 mice; males, n=12 mice) and ctrl mice (females, n=19 mice; males n=15 mice). Wet-weight of uninjured TAs (mg) of *Dhh*^{-/-} (females, n=10 TAs; males, n=8 TAs) and ctrl mice (females, n=17 TAs; males n=9 TAs). **b**) Experimental outline. RT-qPCR of *Dhh* expression 7 days after CTX injury of *Dhh*^{-/-} (n=5 TAs) and ctrl (n=3 TAs) mice. **c**) *Left*: Immunofluorescence of adipocytes (PERILIPIN⁺, green) 21 days post CTX injury of *Dhh*^{+/+}, *Dhh*^{+/-} and *Dhh*^{-/-} mice. Nuclei were visualized with DAPI (purple). Scale bars: 250 μ m. *Right*: Quantification of adipocytes per injured area (mm²) 21 days post CTX injury in *Dhh*^{+/+} (n=6 TAs), *Dhh*^{+/-} (n=8 TAs) and *Dhh*^{-/-} (n=8 TAs) mice. Adipocyte quantification of *Dhh*^{-/-} and ctrl mice separated by females (*Dhh*^{-/-} n=7 TAs; ctrl n=13 TAs) and males (*Dhh*^{-/-} n=6 TAs; ctrl n=6 TAs). **d**) *Left*: Histological Sirius red staining 21 days post CTX injury in *Dhh*^{-/-} and ctrl mice. Scale bars: 500 μ m. *Right*: Quantification of percent of collagen (red) over total TA area 21 dpi in *Dhh*^{-/-} (n=6 TAs) and ctrl mice (n=9 TAs). All data are represented as mean \pm SEM. An unpaired two-tailed t test or a one-way ANOVA followed by a Dunnet's multiple comparison was used.



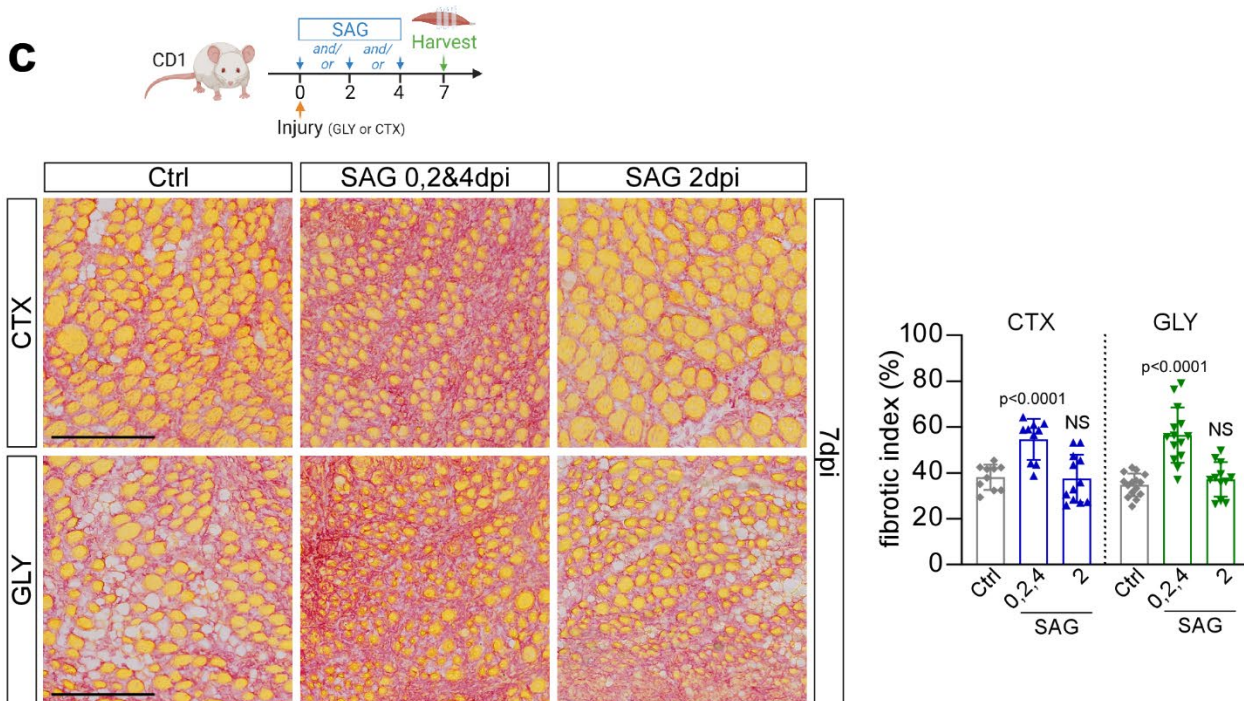
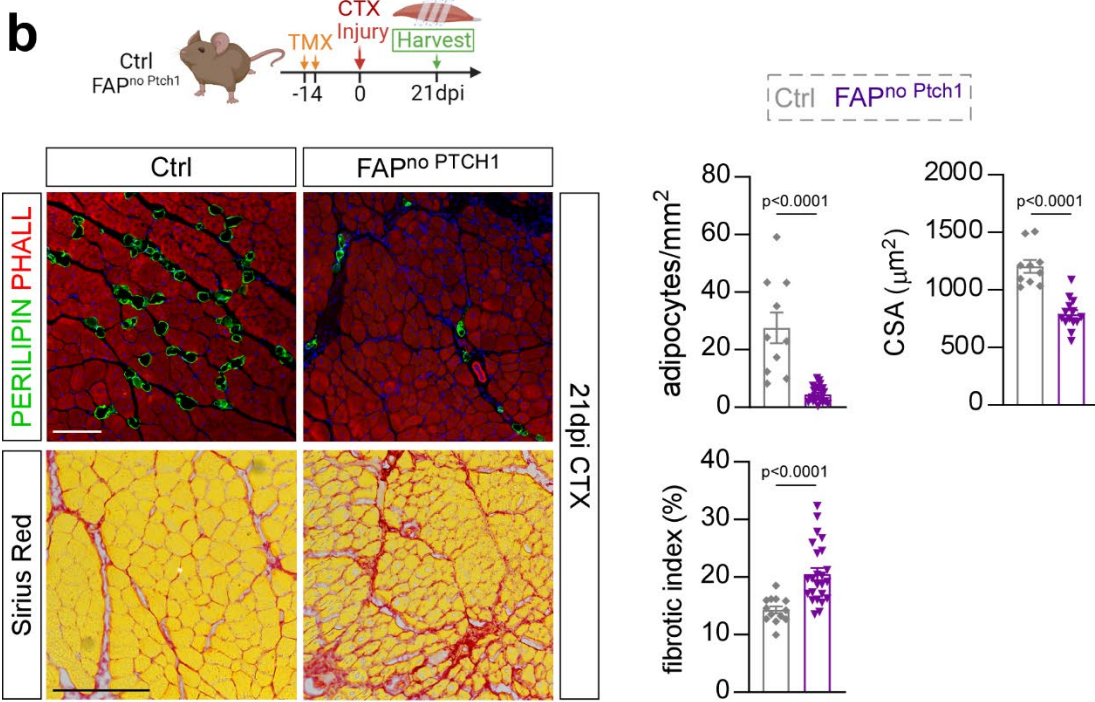
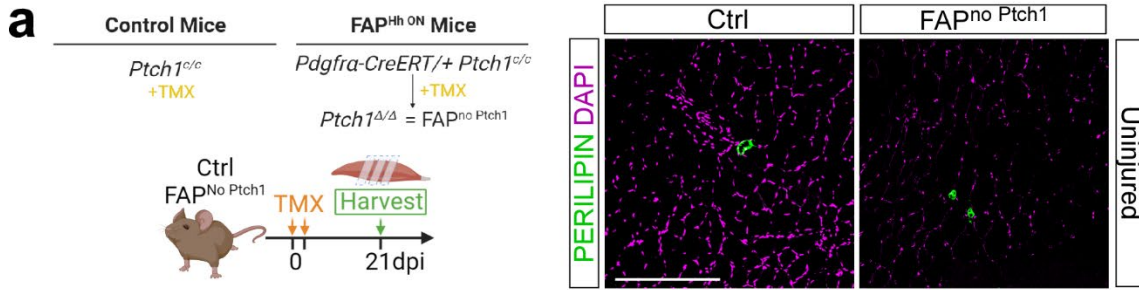
Supplemental Figure 2. Validation of DHH loss of function data and Dhh cKO mouse model.

a) Experimental outline. **b) Left:** Myofibers color-coded according to size (μm^2) of Gant61- and vehicle treated mice 7 days post CTX injury. Scale bar: 250 μm . **Right:** Average CSA (μm^2) of vehicle (n=10 TAs) and Gant61 (n=19 TAs) treated mice 7 days post CTX. **c) Top:** Experimental outline. **Bottom:** RT-qPCR of expression of *Gli1* and *Ptch1* 7 days post CTX in ctrl (n=7 TAs); and 7 days post GLY injury in ctrl (n=7 TAs) and *Dhh*^{-/-} (n=8 TAs) mice. **d)** Experimental outline. **e) Left:** Immunofluorescence of adipocytes (PERILIPIN⁺, green) and nuclei visualized with DAPI (purple); color-coded myofibers according to CSA of *Dhh*^{-/-} and ctrl mice 21 days post GLY. Scale bars: 250 μm . **Right:** Quantification of adipocytes per injured area (mm^2) 21 days post GLY in *Dhh*^{-/-} (n=8 TAs) and ctrl (n=9 TAs) mice. Average CSA (μm^2) of *Dhh*^{-/-} (n=8 TAs) and ctrl (n=8 TAs) mice 21 days after GLY injury. **f)** Experimental outline. **g)** RT-qPCR of expression of *Dhh* and *Gli1* 7 days post CTX in *Dhh* cKO (n=8 TAs) and ctrl (n=6 TAs) mice. **h) Left:** Color-coded myofibers according to CSA (μm^2) of *Dhh* cKO and ctrl 7 days post CTX injury. Scale bar: 250 μm . **Right:** Average CSA (μm^2) of *Dhh* cKO (n=10 TAs) and ctrl (n=6 TAs) mice 7 days post CTX injury. All data are represented as mean \pm SEM. An unpaired two-tailed t test or a one-way ANOVA followed by a Dunnett's multiple comparison was used.



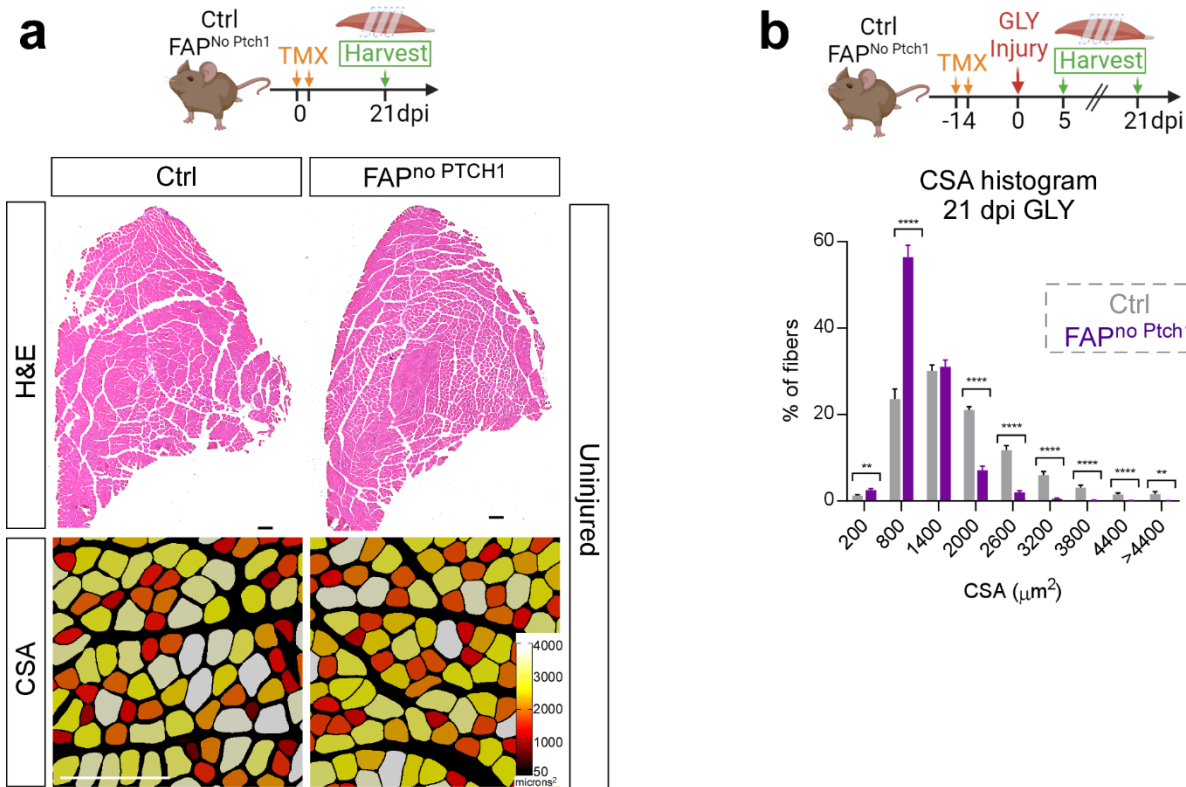
Supplemental Figure 3. SAG influences adipogenesis and myogenesis in a dose- and time-dependent manner.

a) Left: Experimental outline. **Right:** After 2 days post GLY injury, RT-qPCR of *Gli1* and *Ptch1* at 0 hrs (n=6 TAs), 6 hrs (n=4 TAs) and 12 hrs (n=5 TAs) following SAG administration. **b) Left:** Myofibers color-coded according to size (μm^2) 7 days post CTX or GLY in ctrl and SAG- treated (at 0 dpi or 4 dpi) mice. Scale bar: 250 μm . **Right:** Average CSA (μm^2) 7 days post injury with vehicle control (CTX n=27 TAs & GLY n=30 TAs), SAG at 0 dpi (CTX: n=10 TAs & GLY: n=14 TAs) and SAG at 4 dpi (CTX: n=6 TAs & GLY: n=25 TAs). To note, these time points are part of the experiment described in the main Figure 3 and, thus, the same controls were used. **c) SAG** was administered at 0-, 2- and 4dpi at varying concentrations and TAs harvested at 7 days post GLY injury. Immunofluorescence of adipocytes (PERILIPIN⁺, green) and nuclei visualized with DAPI (purple). Color-coded myofibers based on cross sectional area (CSA). Scale bars: 250 μm . **Bottom:** RT-qPCR of *Gli1* expression 7 days post GLY after treatment with DMSO control (n=10 TAs), SAG 1x (n=6 TAs), SAG 0.5x (n=4 TAs), SAG 0.3x (n=5 TAs) and SAG 0.2x (n=6 TAs). Quantification of adipocytes per injured area (mm^2) 7 days post GLY injury of DMSO control (n=10 TAs), SAG 1x (n=9 TAs), SAG 0.5x (n=8 TAs), SAG 0.3x (n=7 TAs), and SAG 0.2x (n=6 TAs). Average CSA (μm^2) 7 days after GLY injury of DMSO control (n=16 TAs), SAG 1x (n=10 TAs), SAG 0.5x (n=11 TAs), SAG 0.3x (n=8 TAs), and SAG 0.2x (n=10 TAs). All data are represented as mean \pm SEM. One- way ANOVA followed by a Dunnet's multiple comparison was used.



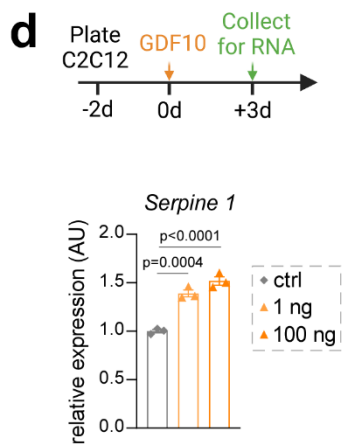
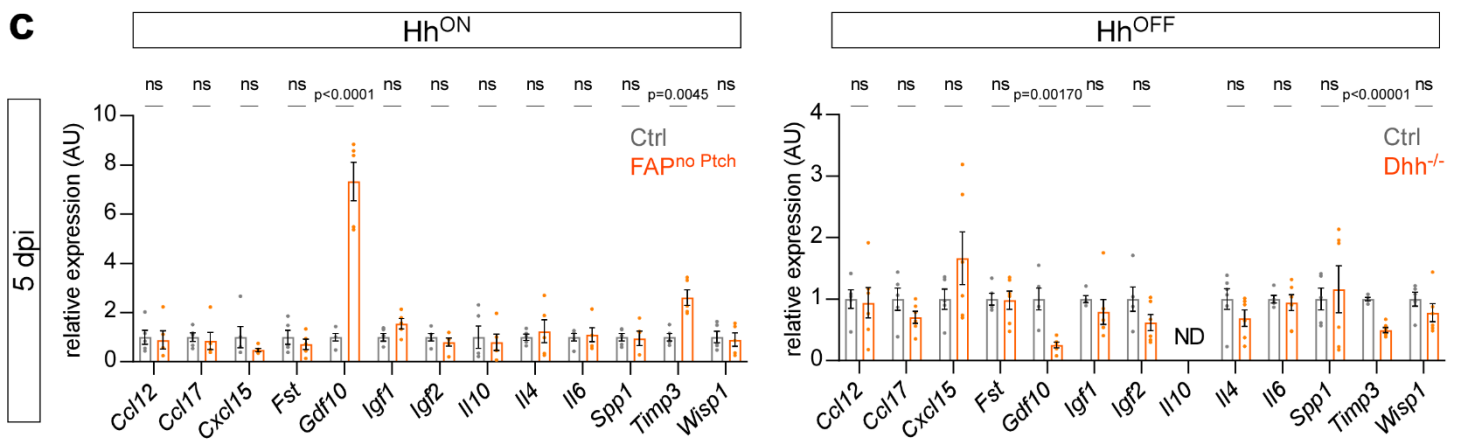
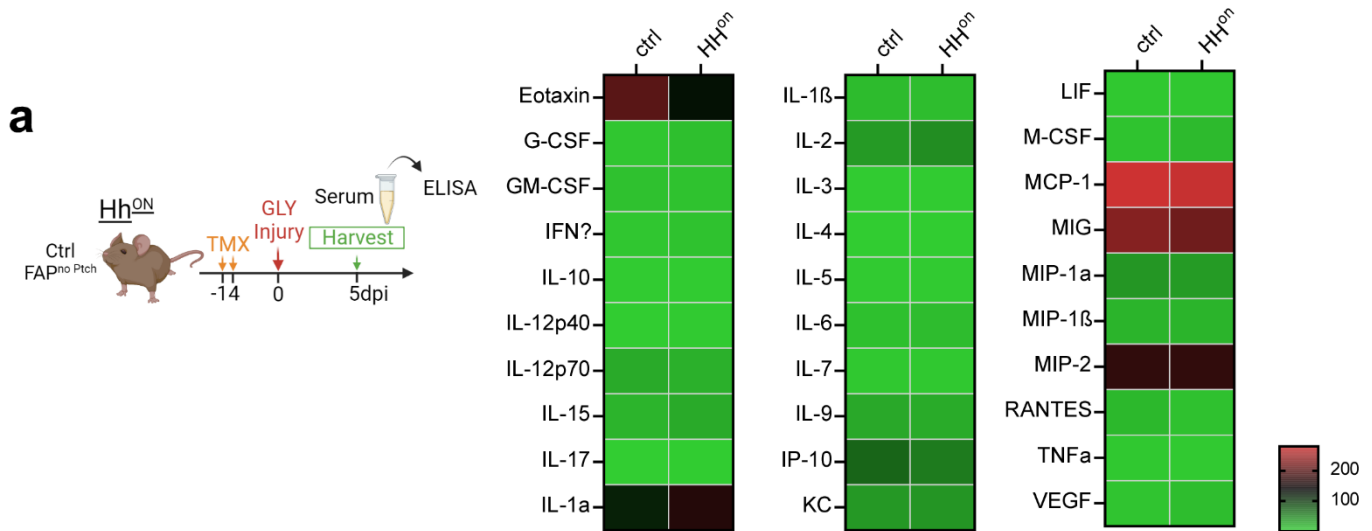
Supplemental Figure 4. Genetic and pharmacological Hh activation prevents IMAT formation but causes fibrosis and impairs myogenesis.

a) Left: Experimental outline. **Right:** Immunofluorescence of PERILIPIN⁺ adipocytes (green) of uninjured FAP^{no P^{ch1}} and control mice (n=5 for each). Scale bars: 250 μ m. **b)** Immunofluorescence of PERILIPIN⁺ adipocytes (green) and PHALLOIDIN⁺ myofibers (scale bar: 100 μ m) and histological Sirius red staining (scale bar: 250 μ m) 21 days post CTX injury in FAP^{no P^{ch1}} and control mice. **Right:** Average CSA (μ m²) 21 days post CTX of ctrl (n=10 TAs) and FAP^{no P^{ch1}} (n=14 TAs) mice. Quantification of adipocytes per injured area (mm²) 21 days post CTX injury of ctrl (n=10 TAs) and FAP^{no P^{ch1}} (n=22 TAs) mice. Quantification of percent of collagen (red) of total TA area 21 dpi CTX in FAP^{no P^{ch1}} (n=19 TAs) and ctrl (n=24 TAs). **c) Left:** Sirius red staining 7 days after CTX (*top*) or GLY (*bottom*) injury after vehicle control, SAG at 0-, 2- and 4 dpi, or SAG at 2 dpi. Scale bars: 250 μ m. **Right:** Quantification of percent of Sirius red-positive collagen (red) over total TA area 7 days after injury from mice treated with vehicle control (CTX: n=10 TAs & GLY: n=15 TAs), SAG at 0-, 2- and 4 dpi (CTX: n=10 TAs & GLY: n=14 TAs) or SAG at 2 dpi (CTX & GLY: n=12 TAs). **c)** All data are represented as mean \pm SEM. An unpaired two-tailed t test or a one-way ANOVA followed by a Dunnet's multiple comparison was used.



Supplemental Figure 5. Ectopic Hh activation does not impact muscle homeostasis.

a) Experimental outline. Color-coded myofibers based on cross sectional area (CSA) of uninjured FAP^{no Ptch1} and control mice. Scale bar: 250 μm . H&E staining of uninjured FAP^{no Ptch1} and control mice. Scale bar: 500 μm . **b)** *Top:* Experimental outline. *Middle:* Fiber number distribution, displayed as percent of total fibers, based on their CSA (μm^2) in FAP^{no Ptch1} (n=18) and ctrl mice (n=14) 21 days post GLY injury. Source data are provided as a Source Data file. All data are represented as mean \pm SEM. A two-way ANOVA followed by Tukey's multiple comparison test was used. A p value less than 0.05 was considered statistically significant where: ** p \leq 0.01, *** p \leq 0.001 and **** p \leq 0.0001. Source data are provided as a Source Data file.



Supplemental Figure 6. Screen for FAP-specific and Hh-induced factors that control adipogenesis and myogenesis.

a) Enzyme-linked immunosorbent assay (ELISA) of whole muscle protein lysates from FAP^{no Ptch1} and ctrl mice 5 days post GLY injury. **b)** Experimental outline. **c)** RT-qPCR of whole muscle RNA from FAP^{no Ptch1} (n=5 TAs) and ctrls (n=5 TAs); and *Dhh*^{-/-} (n=6 TAs) and ctrls (n=4-6) 5 days post injury for the known FAP targets: *Ccl2*, *Ccl7*, *Cxcl5*, *Fst*, *Gdf10*, *Igf1*, *Igf2*, *Il10*, *Il4*, *Spp1*, *Timp3* and *Wisp1*. **d)** RT-qPCR of *Serpine 1* from RNA isolated from C2C12 cells (n= 3 replicates per experimental group) 3 days after induction in ctrl versus rGDF10-treated cells (1 ng & 100 ng). All data are represented as mean ± SEM. A one-way ANOVA followed by a Dunnet's multiple comparison was used.

Supplementary Table

Supplementary Table 1: List of primers used for RT-qPCR

Gene	Forward	Reverse
<i>Hprt</i>	CATAACCTGGTTCATCATCGC	TCCTCCTCAGACCGCTTTT
<i>Pde12</i>	ACCTTTTGGGTGCCAGTAGA	CCAGAGGTCATCTGTCCTTCA
<i>Dhh</i>	CGATGGCTAGAGCGTTCAC	GTACCCAAC TACAACCCCGA
<i>Timp3</i>	TAGACCAGAGTGCCAAAGGG	CCAGGATGCCTTCTGCAAC
<i>Gli1</i>	GGTGCTGCCTATAGCCAGTGTCCCTC	GTGCCAATCCGGTGGAGTCAGACCC
<i>Ptch1</i>	AATTCTCGACTCACTCGTCCA	CTCCTCATATTTGGGGCCTT
<i>Gdf10</i>	CCAAATCCTTTGACGCCTACT	GCTCTGACGATGCTCTGGAT
<i>Acta2</i>	GACAGAGGCACCACTGAACC	ACAGCACAGCCTGAATAGCC
<i>Ccl12</i>	TTAAAAACCTGGATCGGAACCAA	GCATTAGCTTCAGATTTACGGGT
<i>Ccl17</i>	GCTGCTTTCAGCATCCAAGTG	CCAGGGACACCGACTACTG
<i>Cxcl15</i>	GTTCCATCTCGCCATTCATGC	GCGGCTATGACTGAGGAAGG
<i>Fst</i>	GCCTCCTGCTGCTGCTACTC	TTATACAGGACCTGGCAGCG
<i>Igf1</i>	CTGAGCTGGTGGATGCTCTT	TCATCCACAATGCCTGTCTG
<i>Igf2</i>	GTGCTGCATCGCTGCTTAC	ACGTCCCTCTCGGACTTGG
<i>Il10</i>	AGCATTTGAATTCCTGGGT	TTTTCACAGGGGAGAAATCG
<i>Il4</i>	GGTCTCAACCCCCAGCTAGT	GCCGATGATCTCTCTCAAGTGAT
<i>Il6</i>	TAGTCCTTCCTACCCCAATTTCC	TTGGTCCTTAGCCACTCCTTC
<i>Spp1</i>	AGCAAGAACTCTTCCAAGCAA	GTGAGATTCGTCAGATTCATCCG
<i>Wisp1</i>	GTGTGATGATGACGCAAGGA	ATGCAGTTCTCATACCGTTGC
<i>Serpine 1</i>	GTGAATGCCCTCTACTTCAGTG	GCTGCCATCAGACTTGTGGAA