

## Supplementary material for

# Genomic and transcriptomic analyses illuminate the molecular basis of the unique lifestyle of a tubeworm, *Lamellibrachia satsuma*

## Contents

Supplementary Figure S1. Phylogenetic relationships among four vestimentiferans and 13 metazoans.

Supplementary Figure S2. Molecular phylogenetic tree of *Hox* and *ParaHox* genes.

Supplementary Figure S3. Functional enrichment analysis of obturacular region-specific DEGs based on GO terms.

Supplementary Figure S4. Functional enrichment analysis of vestimental region-specific DEGs based on GO terms.

Supplementary Figure S5. Functional enrichment analysis of trunk region-specific DEGs based on GO terms.

Supplementary Figure S6. Gene expression levels of TLR and MyD88.

Supplementary Figure S7. Gene expression levels of lysozymes.

Supplementary Figure S8. Molecular phylogenetic tree of lysozyme genes.

Supplementary Figure S9. Molecular phylogenetic tree of globin subunit genes and expression levels of genes involved in heme biosynthesis.

Supplementary Figure S10. Gene expression levels of globin subunits.

Supplementary Figure S11. Gene expression levels of FIH-1 and FIH-1-like proteins.

Supplementary Figure S12. Molecular phylogenetic tree and expression levels of chitinase genes.

Supplementary Figure S13. Molecular phylogenetic tree and expression levels of TMC genes.

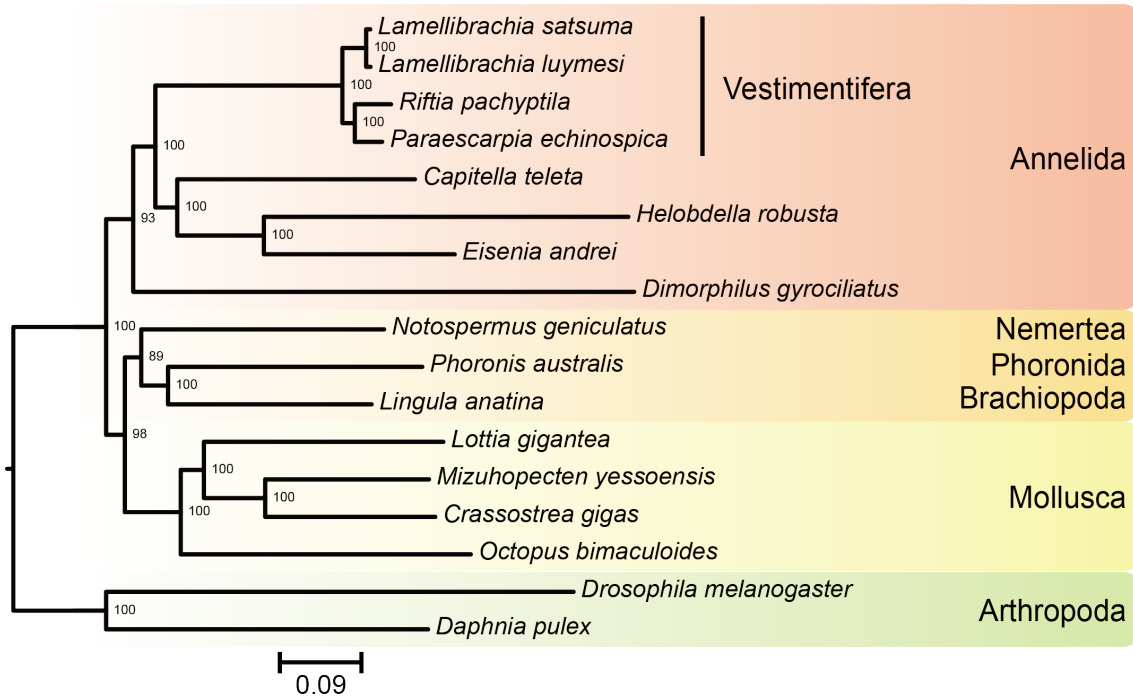
Supplementary Figure S14. Molecular phylogenetic tree and expression levels of C-type lectin genes.

Supplementary Figure S15. Molecular phylogenetic tree of chymotrypsin-like serine protease and titin-like protein genes.

Supplementary table footnotes

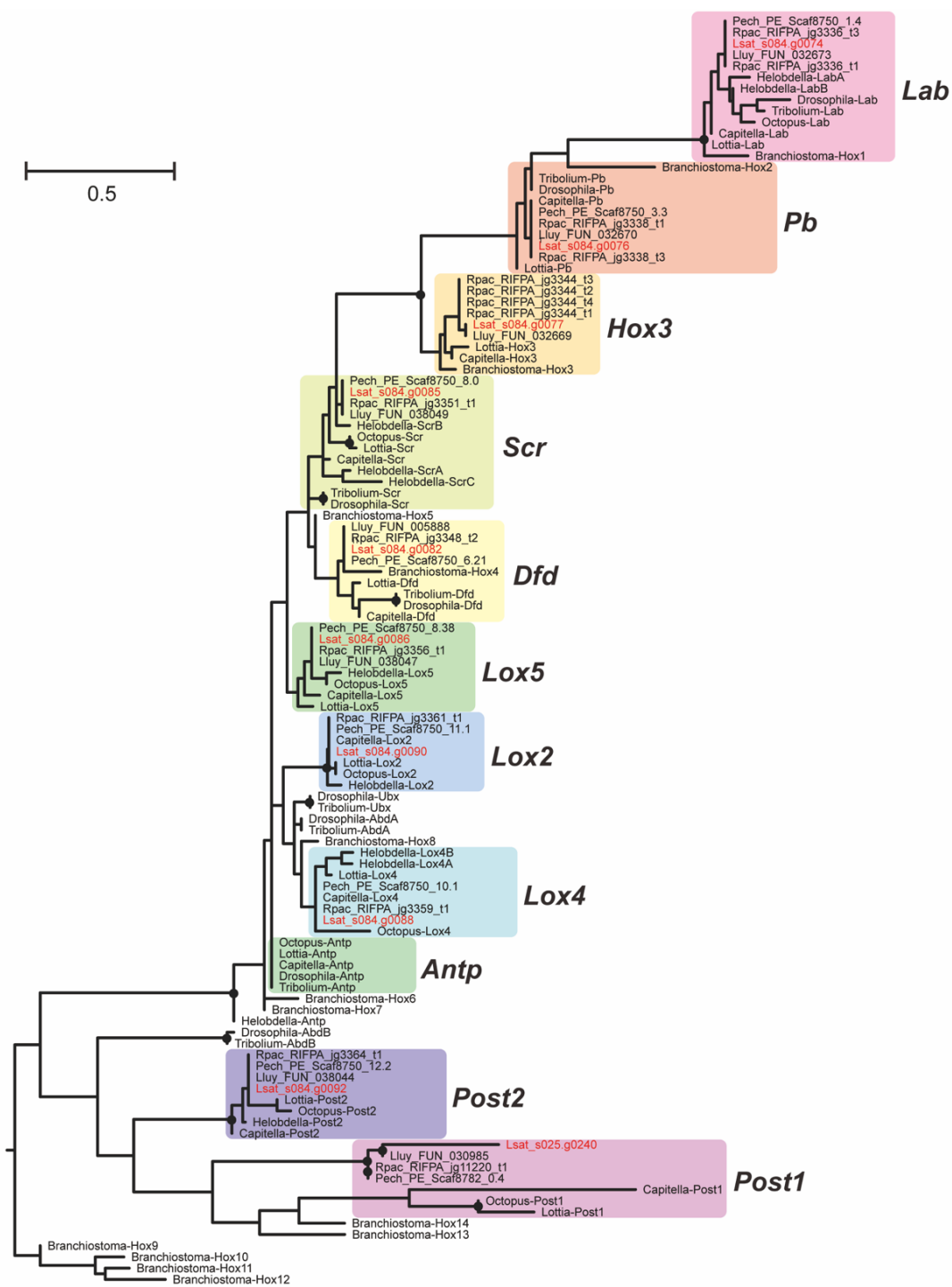
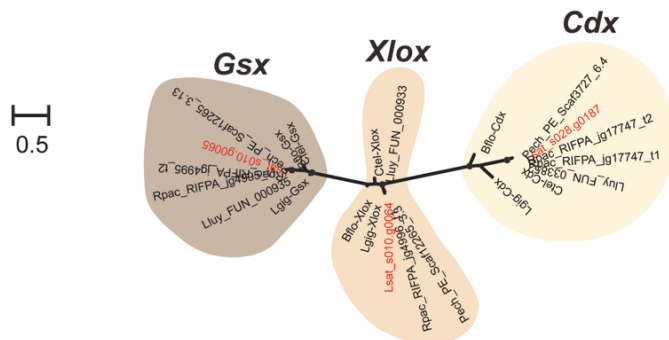
References

Supplementary Figures



Supplementary Figure S1.

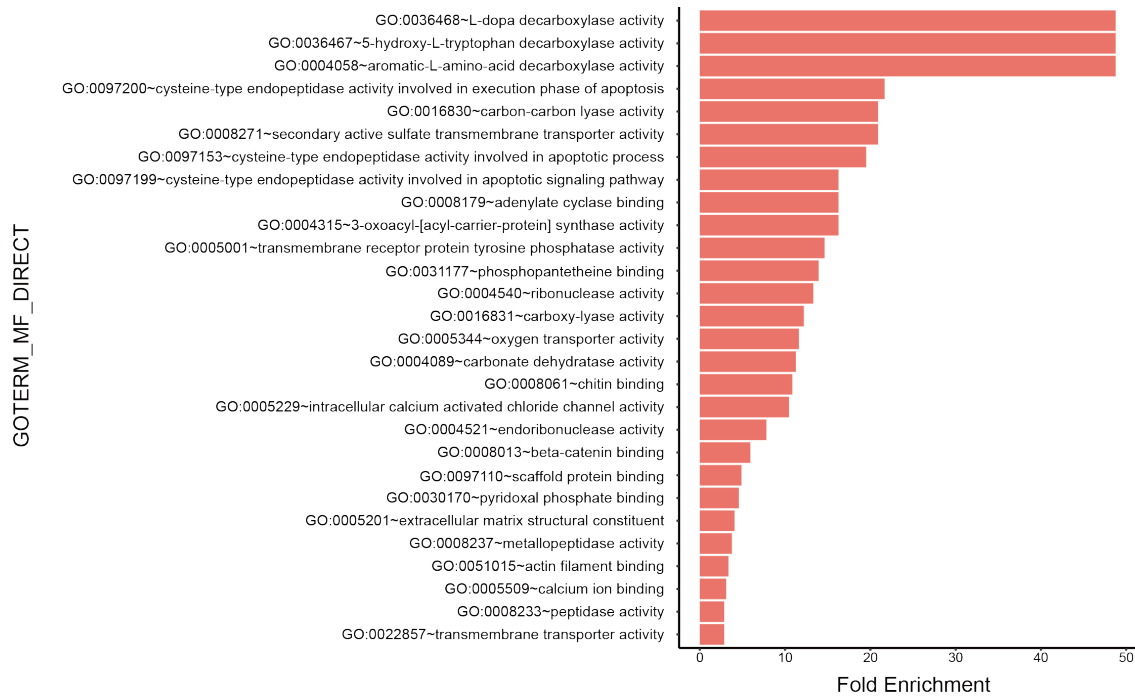
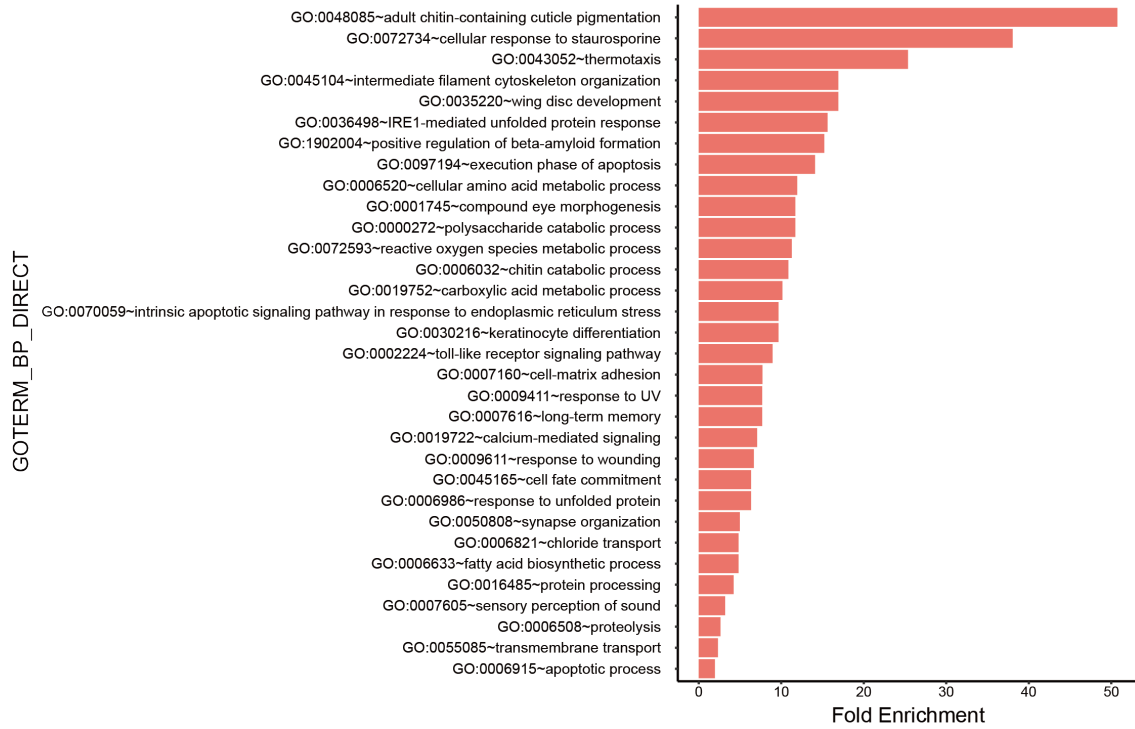
Phylogenetic relationships among four vestimentiferans and 13 metazoans. Concatenated amino acid sequences of 163 single-copy OGs were aligned, gap-trimmed, and used for construction of a maximum likelihood phylogenetic tree. Numbers on branches indicate bootstrap values.

**A****B**

Supplementary Figure S2.

Molecular phylogenetic tree of *Hox* and *ParaHox* genes. (A) Molecular phylogenetic tree of *Hox* genes of four vestimentiferans and seven non-vestimentiferan metazoans. In order to extract *Hox* candidate genes, gene models of four vestimentiferans were searched using BLAST against *Hox* genes of *Capitella teleta*, *Helobdella robusta*, *Lottia gigantea*, *Octopus bimaculoides*, *Drosophila melanogaster*, *Tribolium castaneum*, and *Branchiostoma floridae*<sup>1,2</sup>. Amino acid sequences of candidate genes were aligned, gap-trimmed, and used for construction of a maximum likelihood phylogenetic tree. Although previous studies identified *Hox3* in the *Paraescarpia* genome and *Lox4* and *Lox2* in the *L. hymesii* gene models<sup>3</sup>, our study and another study were not able to identify clear orthologs of these genes<sup>4</sup>. (B) Molecular phylogenetic tree of *ParaHox* genes of four vestimentiferans and three non-vestimentiferan metazoans. In order to extract *ParaHox* candidate genes, gene models of four vestimentiferans were searched using BLAST against *ParaHox* genes of *C. teleta*, *L. gigantea*, and *B. floridae*<sup>5,6</sup>. Amino acid sequences of candidate genes were aligned, gap-trimmed, and used for construction of a maximum likelihood phylogenetic tree. For genes of non-vestimentiferan metazoans, taxonomy and definitions are shown. Circles on branches indicate bootstrap values higher than 80%. *L. satsuma* genes are highlighted in red.

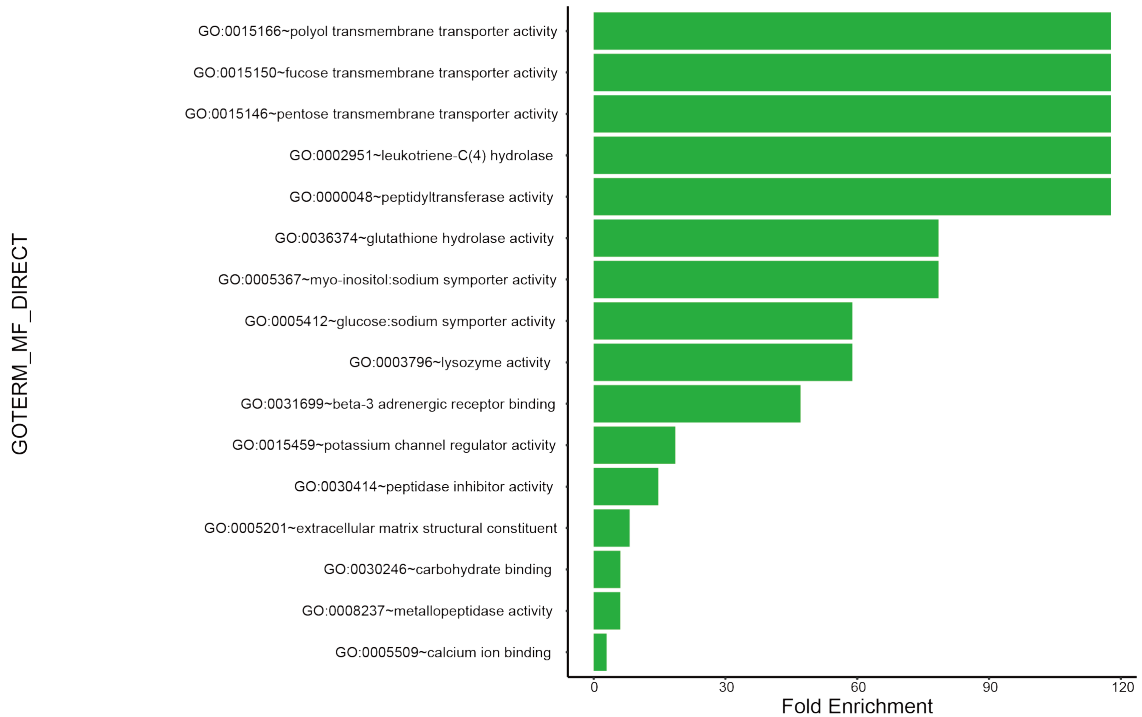
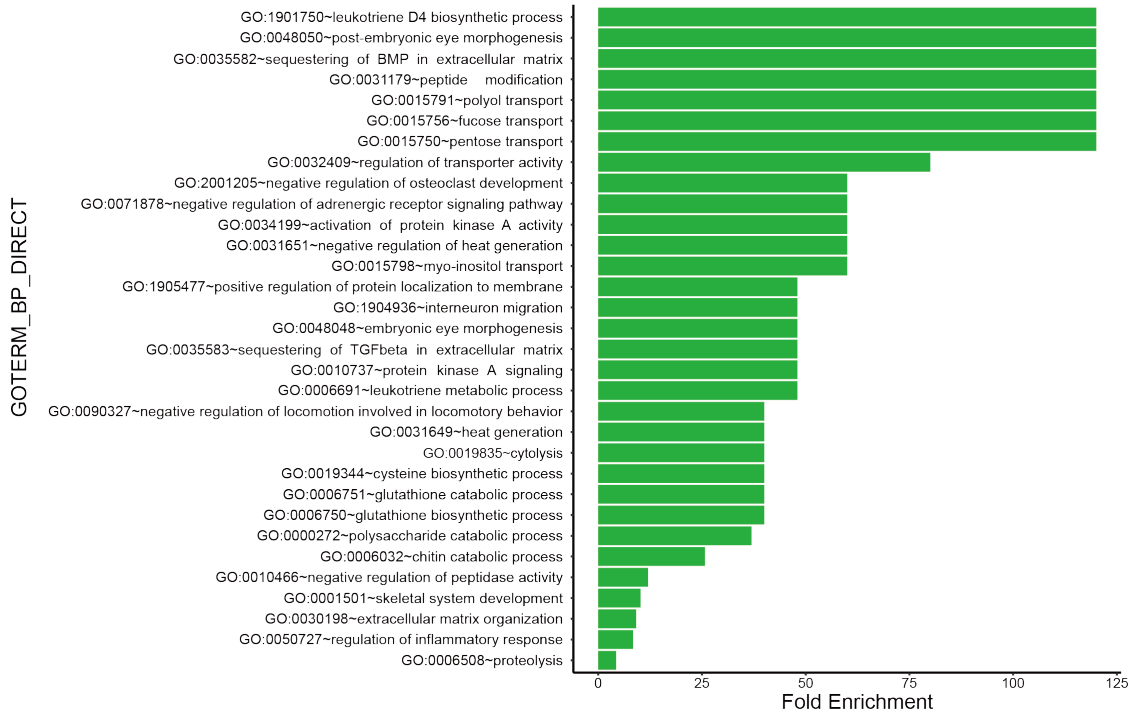
## Obturacular region



Supplementary Figure S3.

Functional enrichment analysis of obturacular region-specific DEGs based on Gene Ontology (GO) terms. GO terms with  $p$ -values  $< 0.05$  are shown.

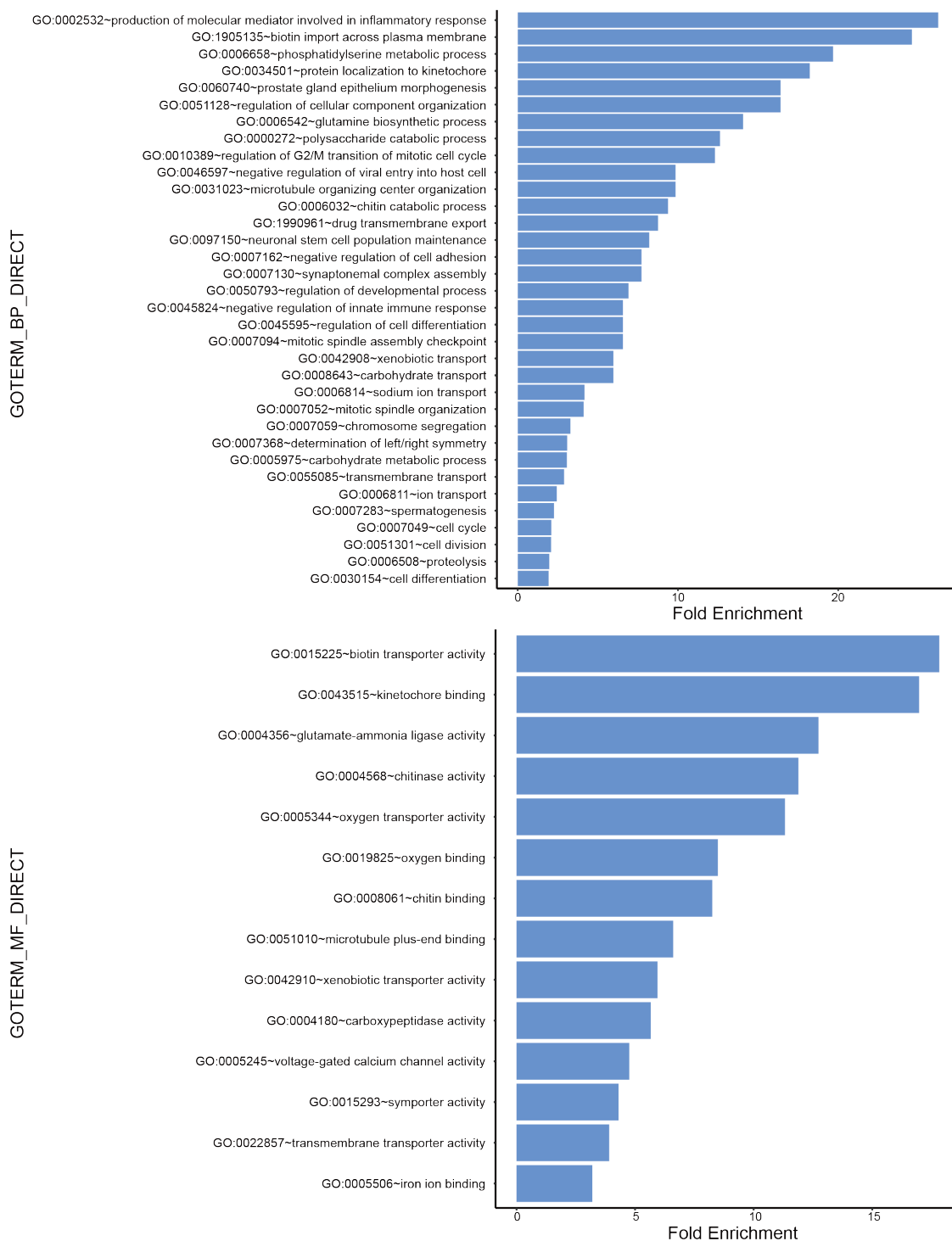
## Vestimental region



Supplementary Figure S4.

Functional enrichment analysis of vestimental region-specific DEGs based on GO terms. GO terms with  $p$ -values  $< 0.05$  are shown.

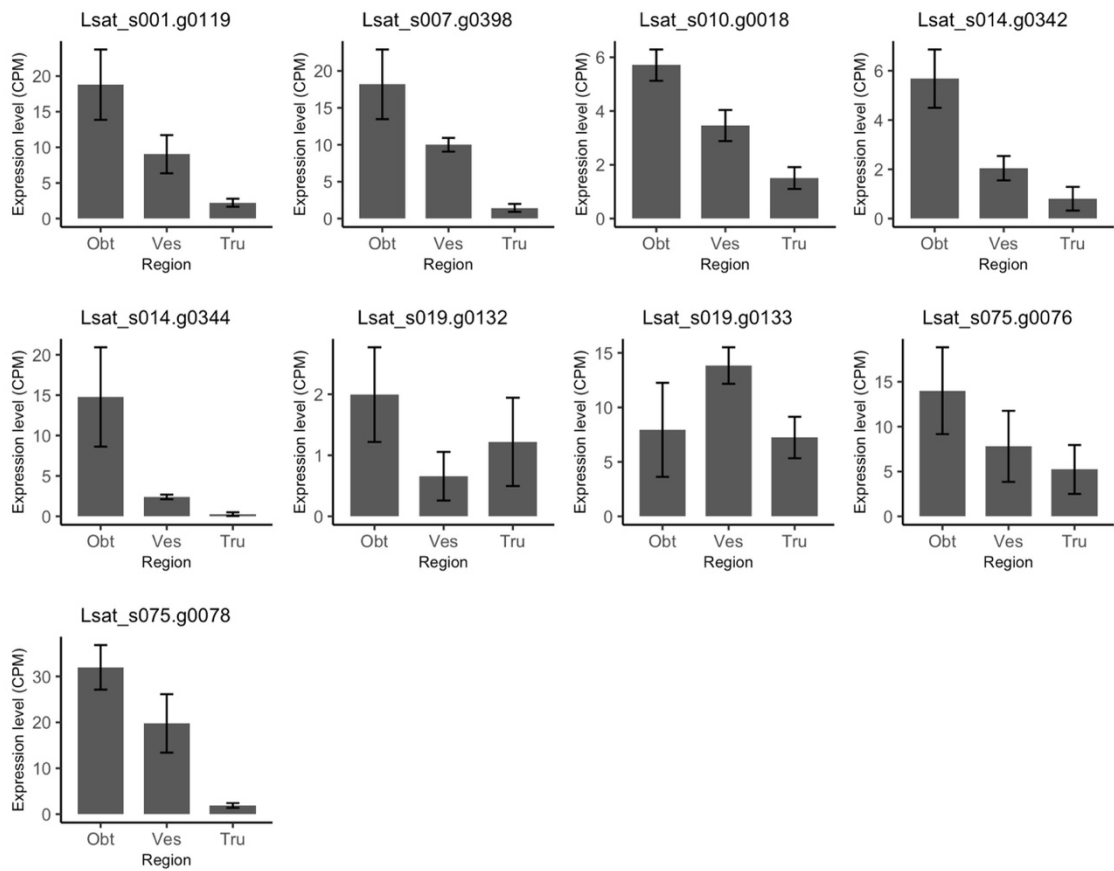
## Trunk region



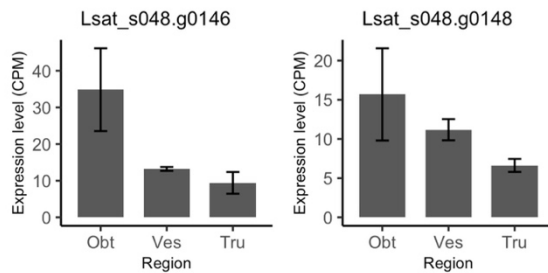
Supplementary Figure S5.

Functional enrichment analysis of trunk region-specific DEGs based on GO terms. GO terms with  $p$ -values  $< 0.05$  are shown.

## TLR



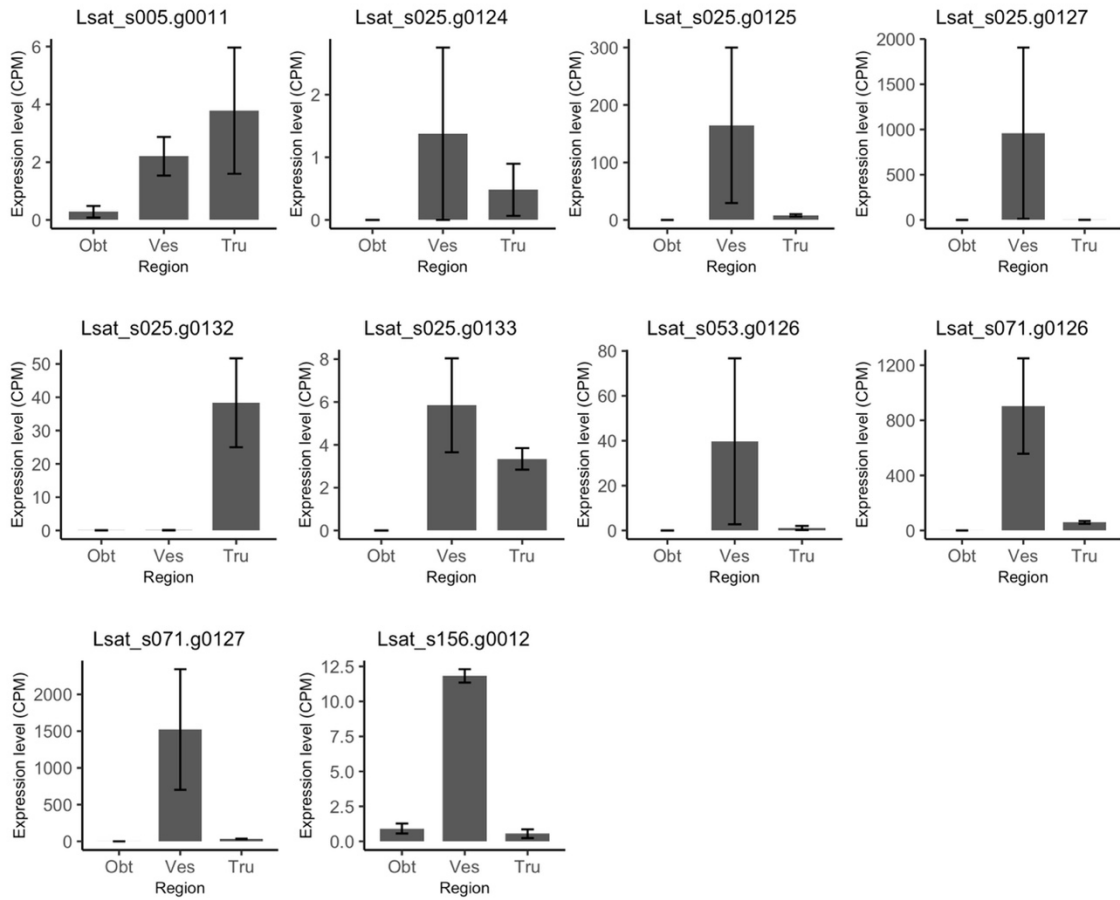
## MyD88



Supplementary Figure S6.

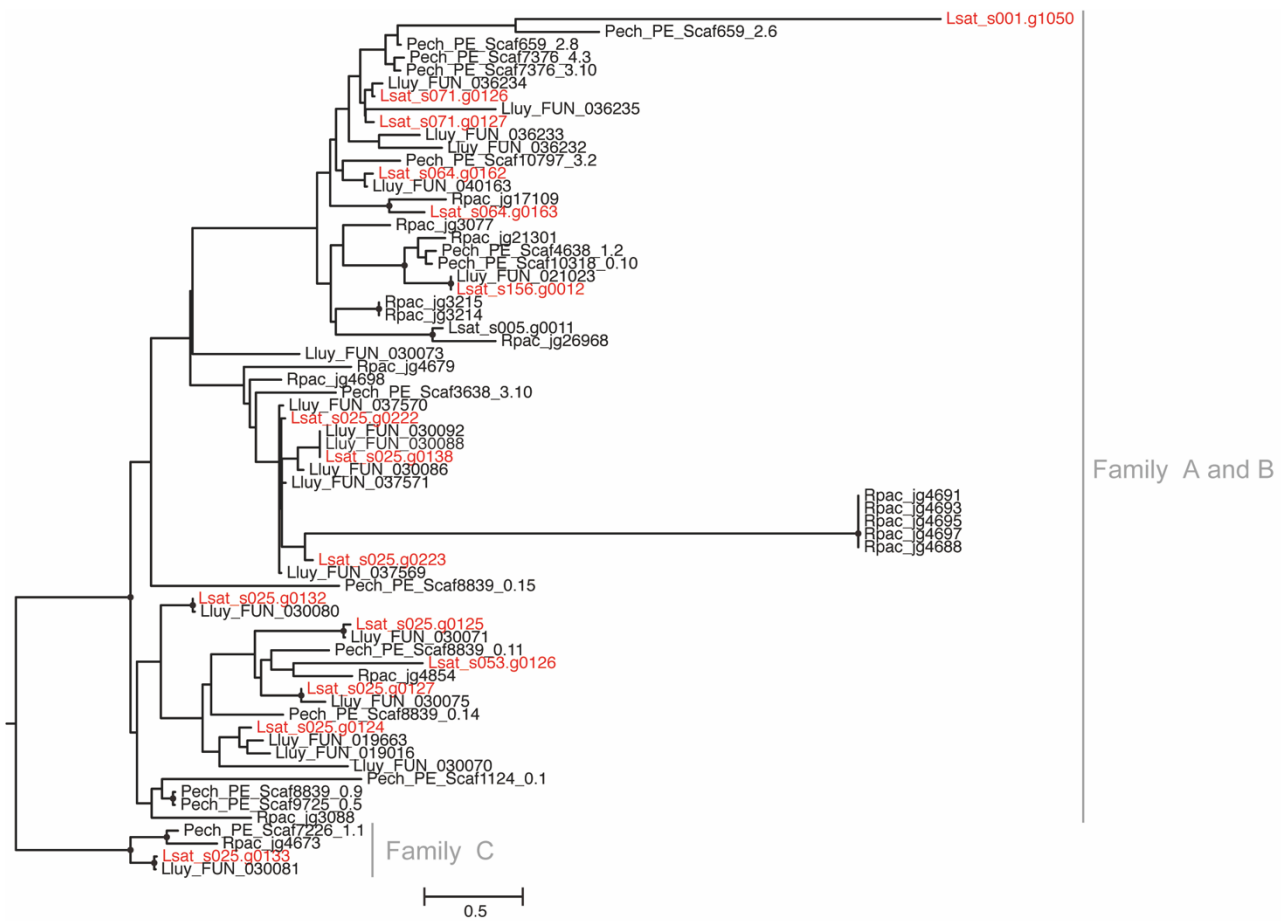
Gene expression levels (TMM-normalized CPMs) of TLR and MyD88. Data are shown as means  $\pm$  SEs ( $n = 3$ ). Obt, obturator region; Ves, vestimental region; Tru, trunk region.





Supplementary Figure S7.

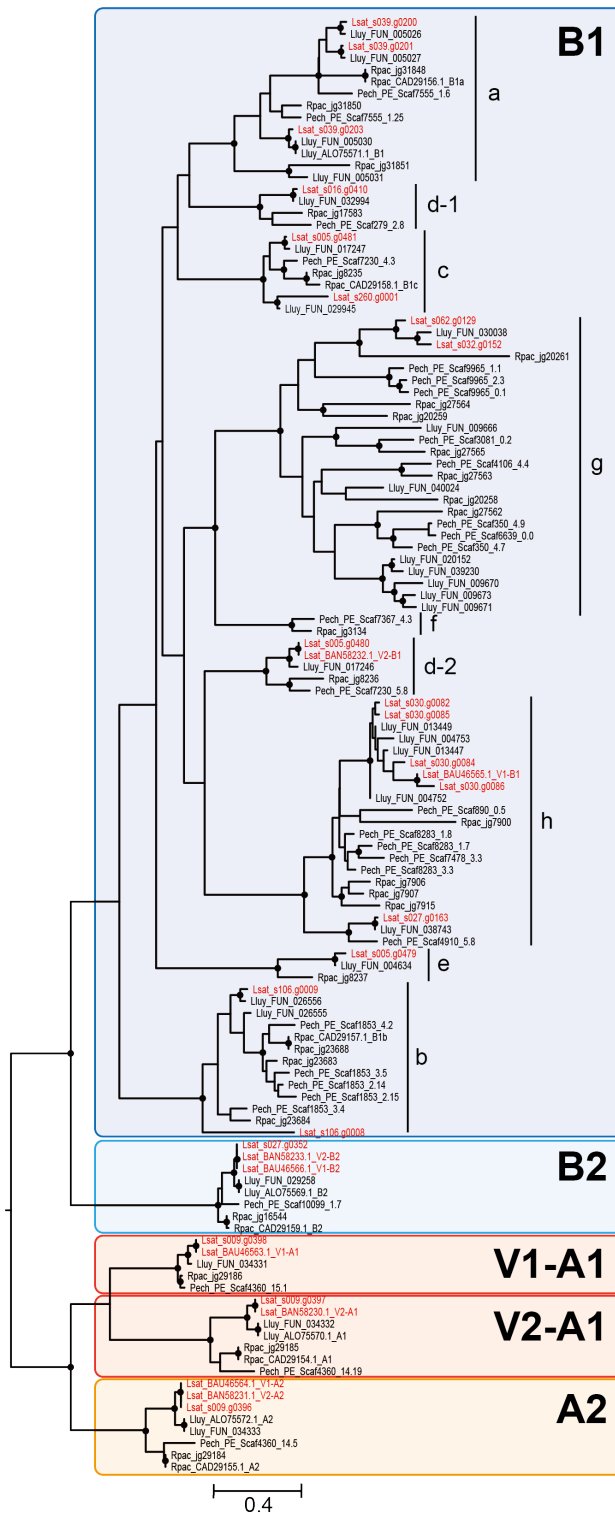
Gene expression levels (TMM-normalized CPMs) of lysozymes. Data are shown as means  $\pm$  SEs (n = 3). Obt, obturacular region; Ves, vestimental region; Tru, trunk region.



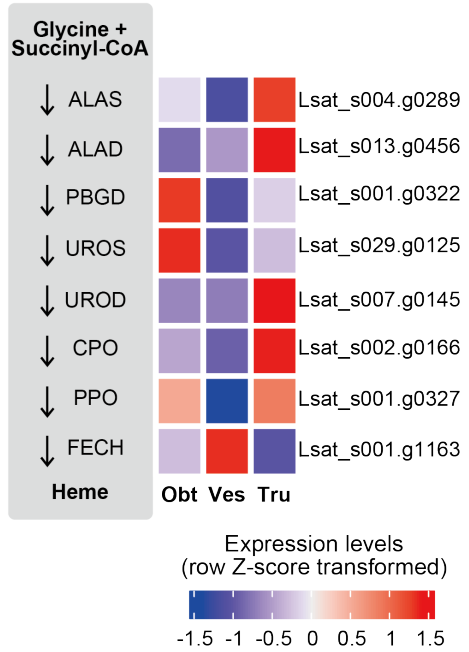
Supplementary Figure S8.

Molecular phylogenetic analysis of lysozyme genes in genomes of vestimentiferans. The maximum likelihood phylogenetic tree was constructed using aligned and gap-trimmed amino acid sequences. *L. satsuma* genes are highlighted in red. Circles on branches indicate bootstrap values higher than 80%.

**A**



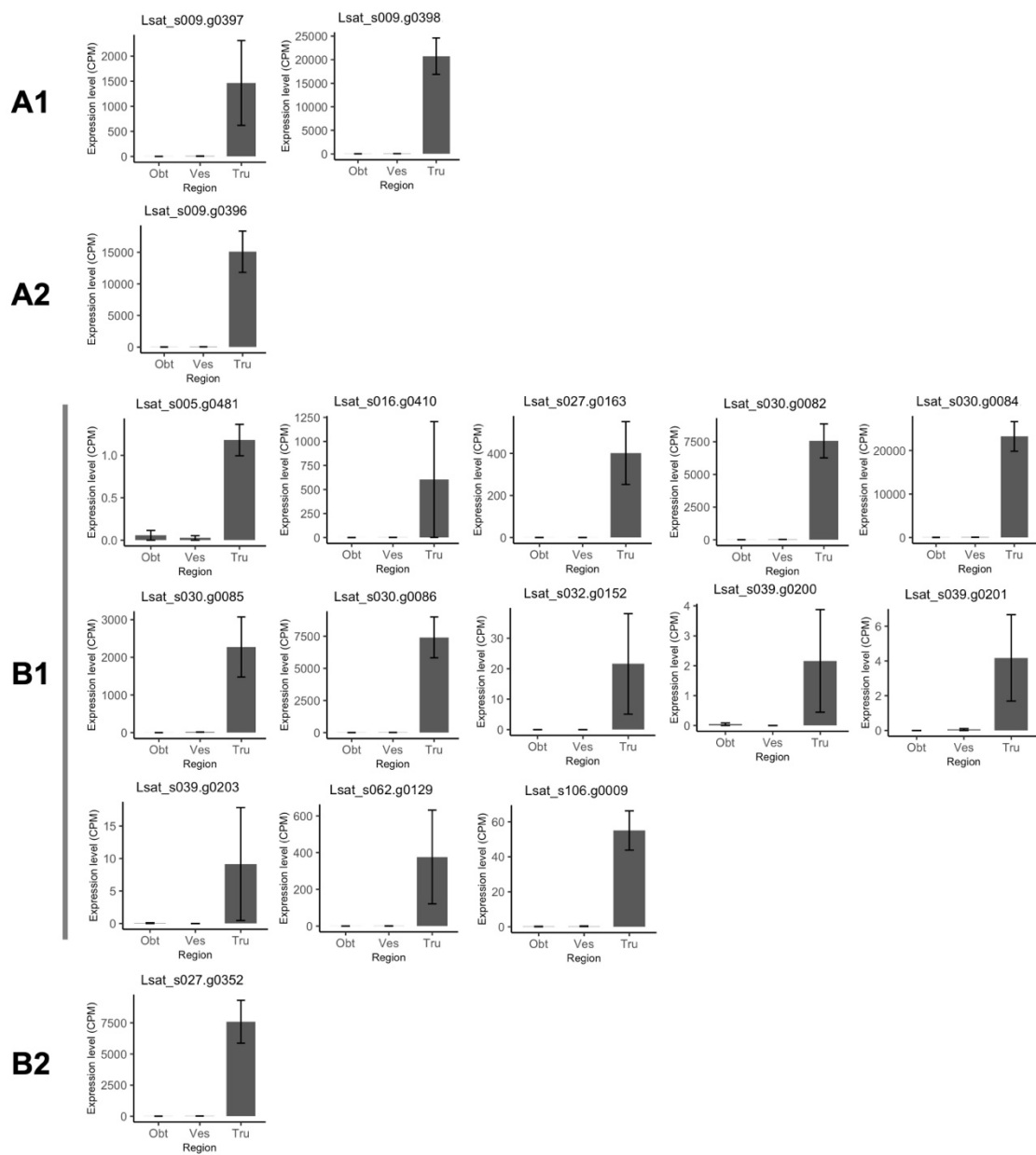
**B**



Supplementary Figure S9.

Molecular phylogenetic tree of globin subunit genes and expression levels of genes involved in heme biosynthesis. (A) Molecular phylogenetic tree of globin subunit genes of four vestimentiferans. The maximum likelihood phylogenetic tree was constructed using aligned and gap-trimmed amino acid sequences. For genes retrieved from the NCBI GenBank and UniProt/Swiss-Prot databases, taxonomy, accession number, and definitions are shown, separated by underscores. *L. satsuma* genes are highlighted in red. Circles on branches indicate bootstrap values higher than 80%. “a” – “h” correspond to eight paralogous groups proposed in a previous study<sup>4</sup>. Note that paralogous group d<sup>4</sup> was divided into two independent clades (d-1 and d-2); therefore, the total number of groups is nine. (B) Expression levels of putative genes involved in heme biosynthesis. The heme biosynthetic pathway and involved enzymes are shown on the left. A heat map of expression levels of the genes corresponding to the enzymes (row Z-score transformed) is shown on the right.

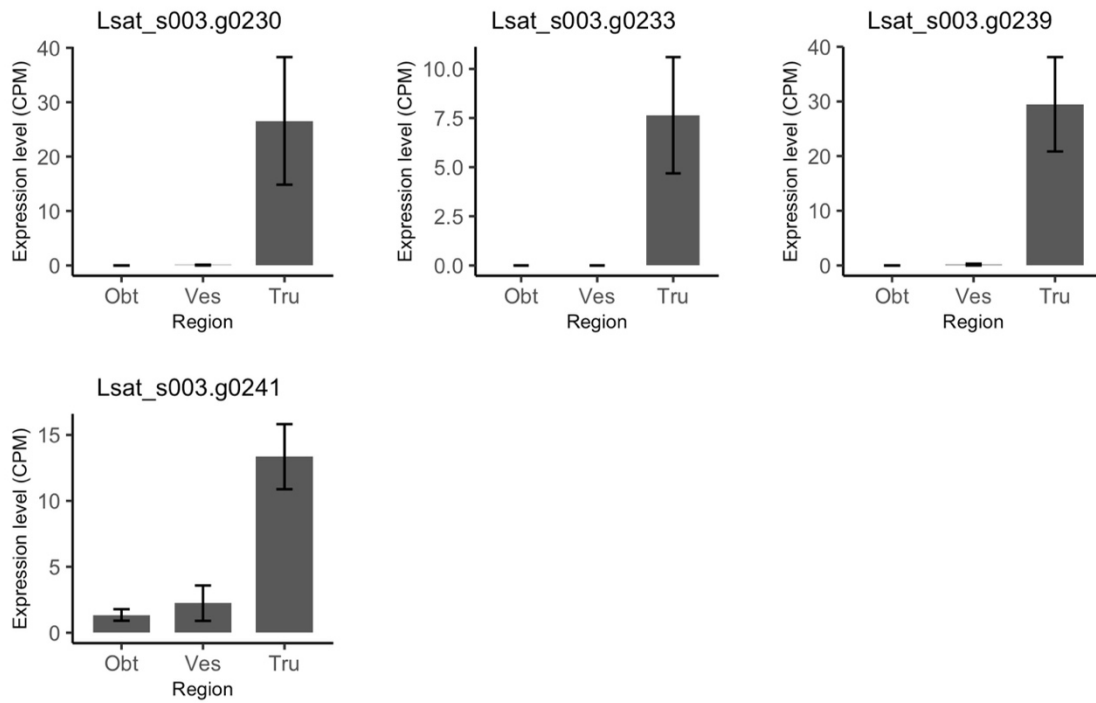
Lsat, *L. satsuma*; Lluy, *L. luyesi*; Rpac, *R. pachyptila*; Pech, *P. echinospica*; A1-B2, globin A1-B2 chain (subunit); V1, V2, V1 and V2 hemoglobin; Obt, obturacular region; Ves, vestimental region; Tru, trunk region.



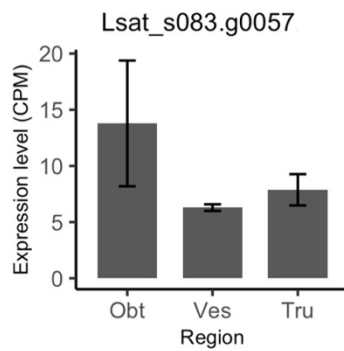
Supplementary Figure S10.

Gene expression levels (TMM-normalized CPMs) of globin subunits. Data are shown as means  $\pm$  SEs ( $n = 3$ ). A1-B2, globin A1-B2 chain (subunit); Obt, obturacular region; Ves, vestimental region; Tru, trunk region.

## FIH-1-like



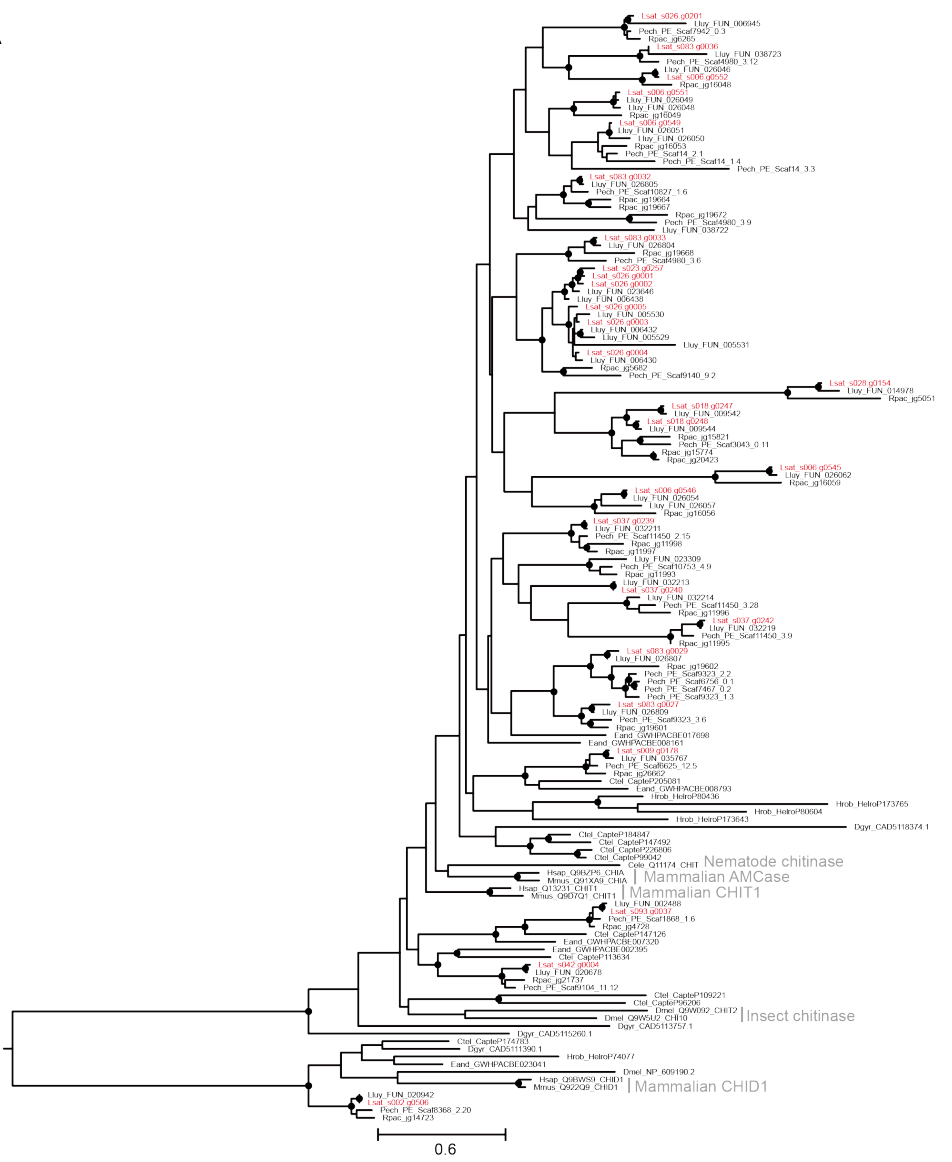
## FIH-1



Supplementary Figure S11.

Gene expression levels (TMM-normalized CPMs) of FIH-1 and FIH-1-like proteins. Data are shown as means  $\pm$  SEs (n = 3). Obt, obturacular region; Ves, vestimental region; Tru, trunk region.

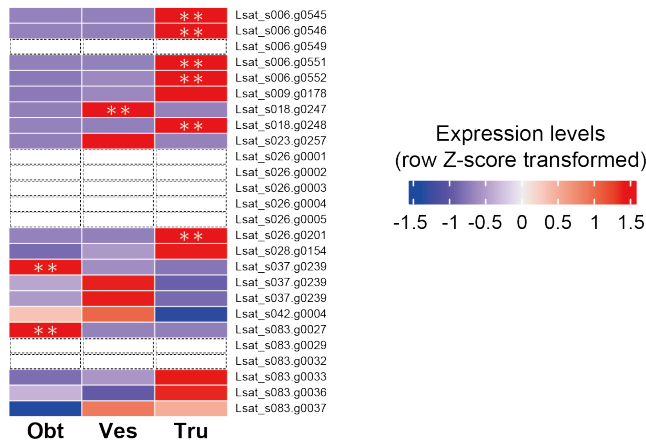
**A**



**Chitinase**

**CHID**

**B**

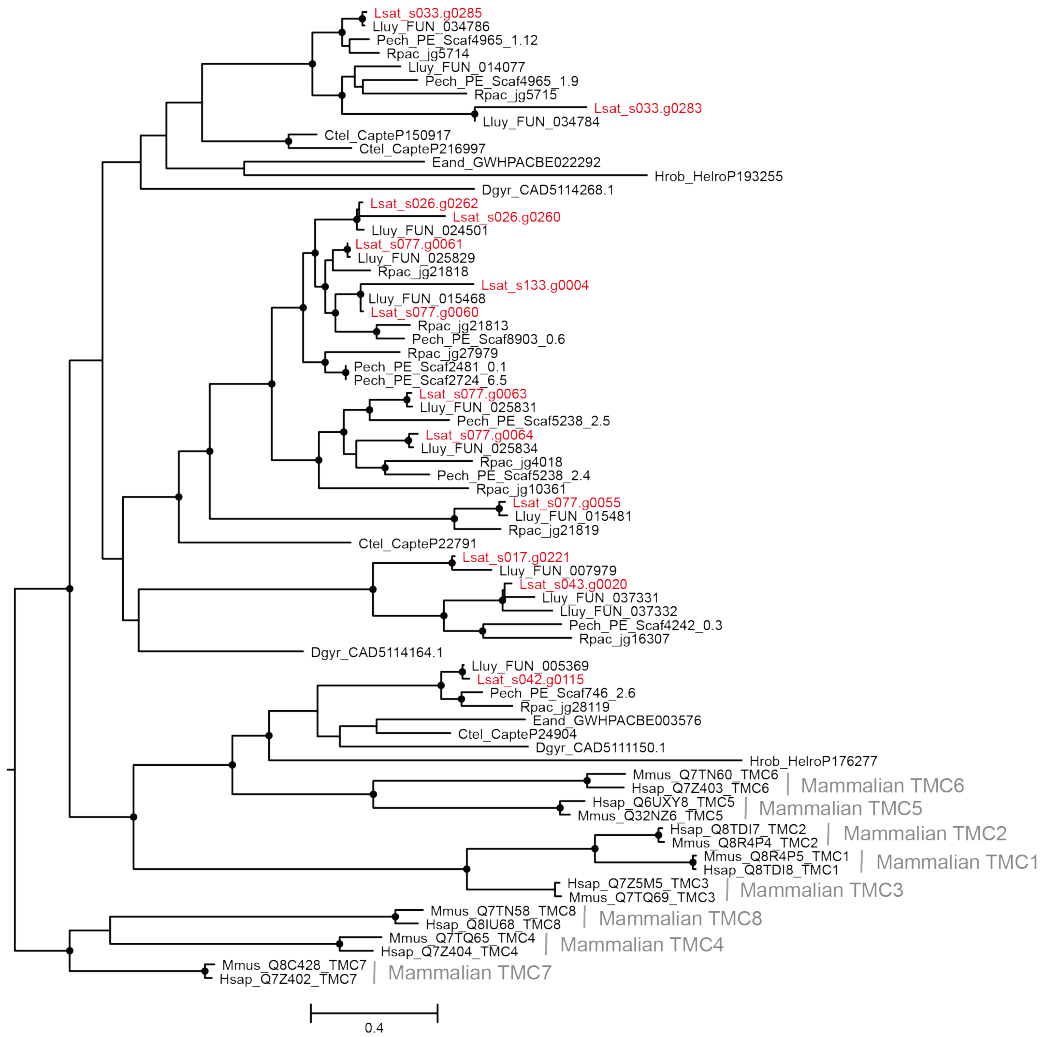


Supplementary Figure S12.

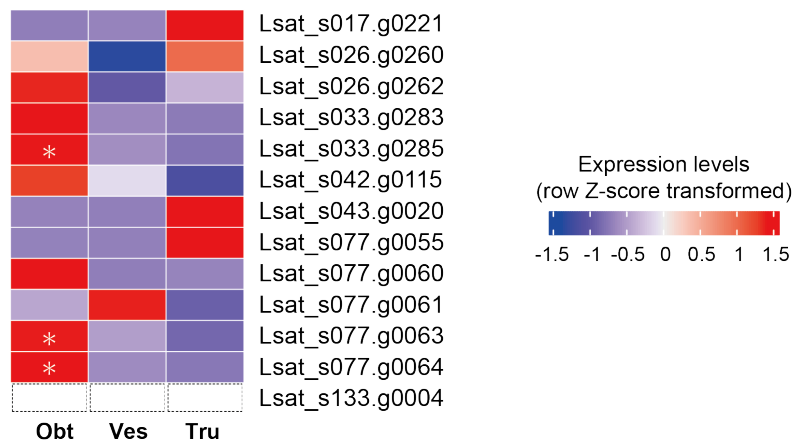
Molecular phylogenetic tree and expression levels of chitinase genes. (A) Molecular phylogenetic tree of the chitinase family in genomes of four vestimentiferans. The maximum likelihood phylogenetic tree was constructed using aligned and gap-trimmed amino acid sequences. For genes retrieved from the UniProt database, taxonomy, accession number, and definitions are shown. *L. satsuma* genes are highlighted in red. Circles on branches indicate bootstrap values higher than 80%. (B) Heat map of expression levels (row Z-score transformed) of *L. satsuma* genes belonging to the chitinase family. AMCase, acidic mammalian chitinase; CHIT, chitotriosidase; CHID, chitinase domain-containing protein; Lsat, *L. satsuma*; Lluy, *L. luyesi*; Rpac, *R. pachyptila*; Pech, *P. echinospica*; Ctel, *C. teleta*; Hrob, *Helobdella robusta*; Eand, *Eisenia andrei*, Dgyr, *Dimorphilus gyrocoliatus*; Cele, *Caenorhabditis elegans*; Hsap, *Homo sapiens*; Mmus, *Mus musculus*; Dmel, *Drosophila melanogaster*; Obt, obturacular region; Ves, vestimental region; Tru, trunk region; \*\*, region-specific DEGs.



**A**



**B**

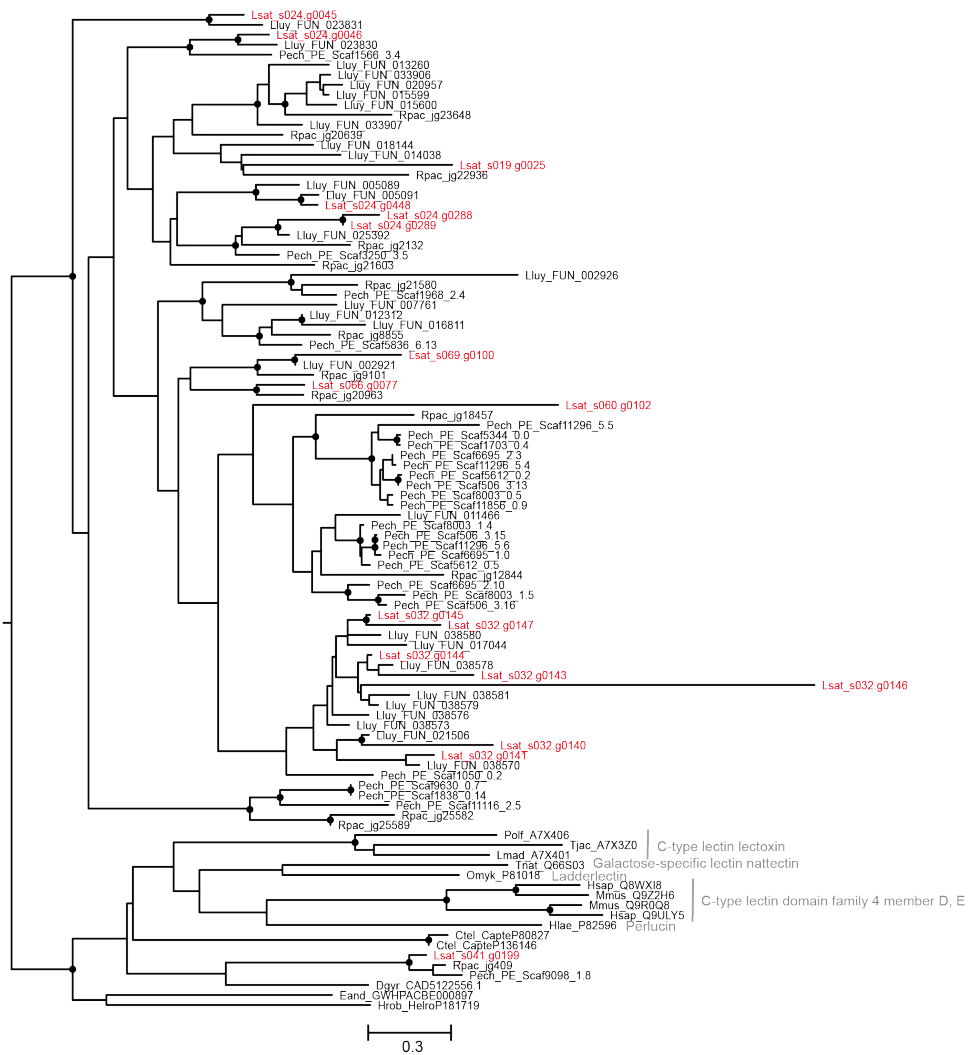


Supplementary Figure S13.

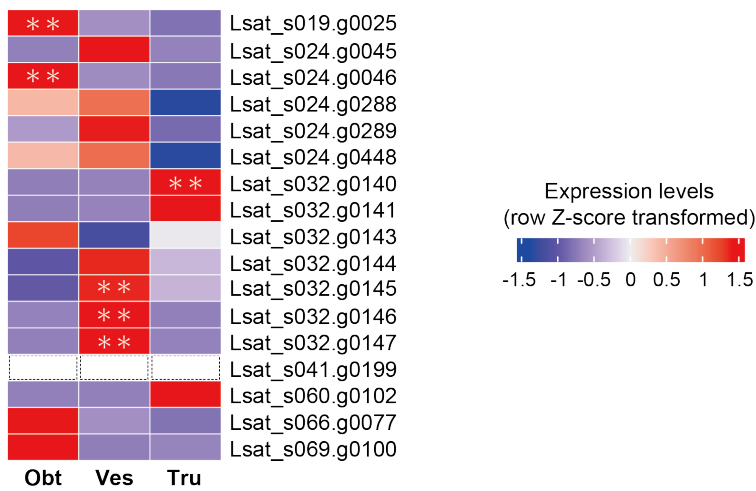
Molecular phylogenetic tree and expression levels of TMC genes. (A) Molecular phylogenetic tree of the TMC family in the genomes of four vestimentiferans. The maximum likelihood phylogenetic tree was constructed using aligned and gap-trimmed amino acid sequences. For genes retrieved from the UniProt database, taxonomy and accession number, and definitions are shown. *L. satsuma* genes are highlighted in red. Circles on branches indicate bootstrap values higher than 80%. (B) Heat map of expression levels (row Z-score transformed) of *L. satsuma* genes belonging to the TMC family.

Lsat, *L. satsuma*; Lluy, *L. luymesii*; Rpac, *R. pachyptila*; Pech, *P. echinospica*; Ctel, *C. teleta*; Hrob, *Helobdella robusta*; Eand, *Eisenia andrei*, Dgyr; *Dimorphilus gyrocolatus*; Hsap, *Homo sapiens*; Mmus, *Mus musculus*; Obt, obturacular region; Ves, vestimental region; Tru, trunk region; \*, genes with expression levels significantly higher in a specific region than other regions (FDR < 0.05).

**A**



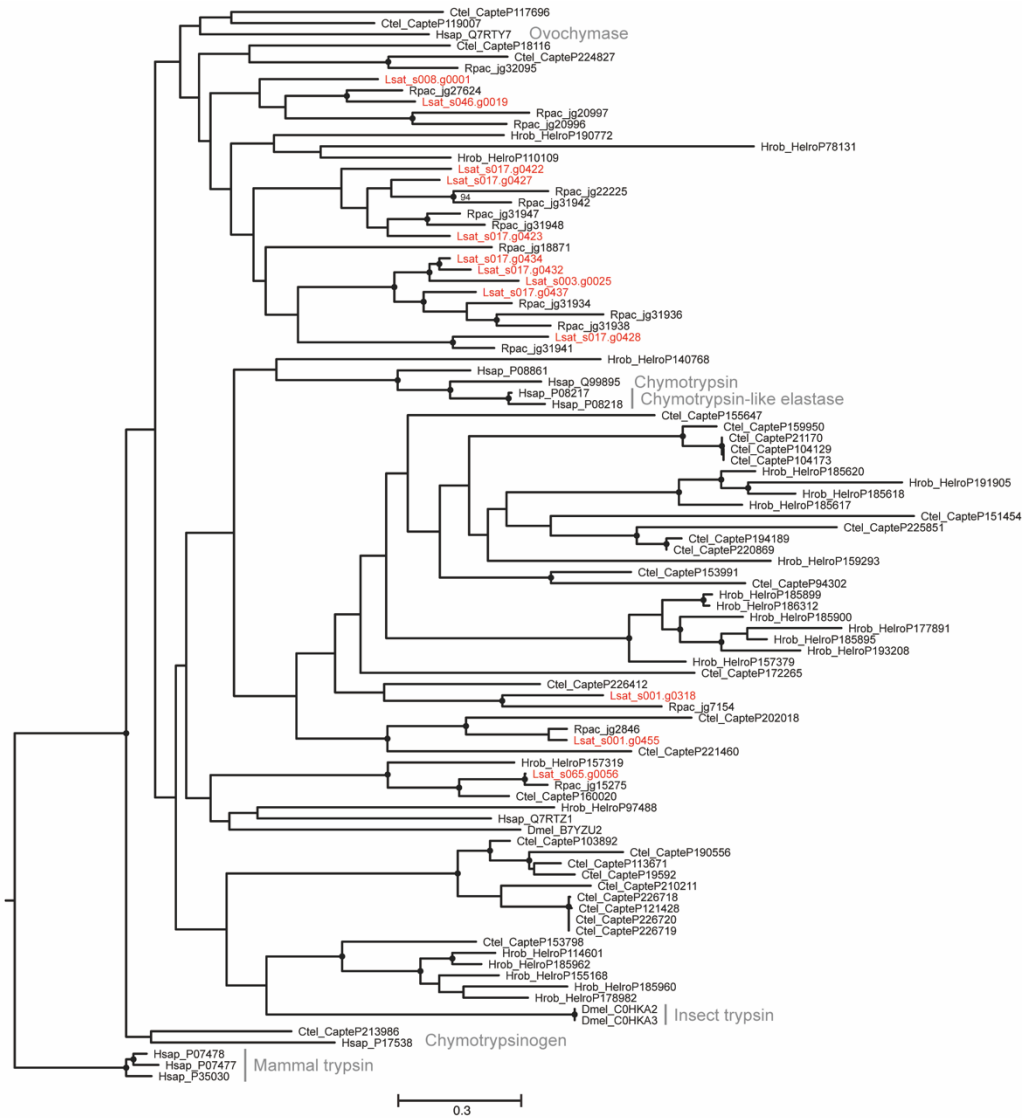
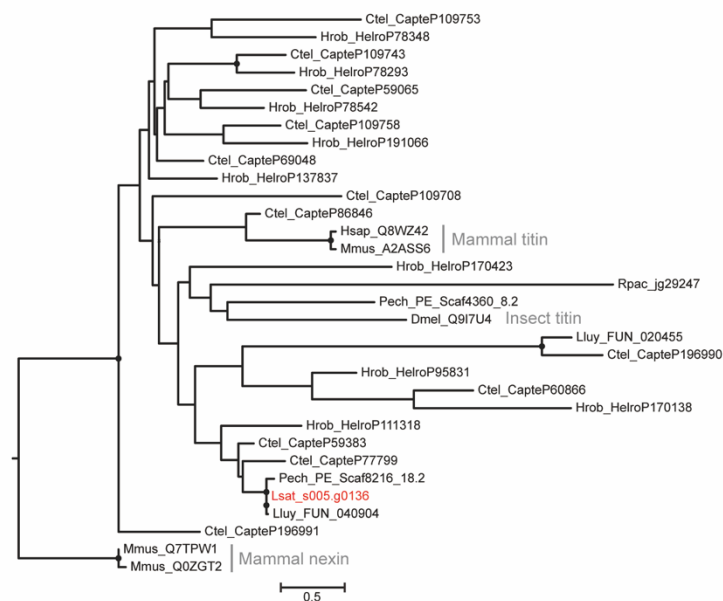
**B**



Supplementary Figure S14.

Molecular phylogenetic tree and expression levels of C-type lectin genes. (A) Molecular phylogenetic tree of the C-type lectin family in the genomes of four vestimentiferans. The maximum likelihood phylogenetic tree was constructed using aligned and gap-trimmed amino acid sequences. For genes retrieved from the UniProt database, taxonomy and accession numbers are shown. *L. satsuma* genes are highlighted in red. Circles on branches indicate bootstrap values higher than 80%. (B) Heat map of expression levels (row Z-score transformed) of *L. satsuma* genes belonging to the C-type lectin family.

Lsat, *L. satsuma*; Lluy, *L. luyesi*; Rpac, *R. pachyptila*; Pech, *P. echinospica*; Ctel, *C. teleta*; Hrob, *Helobdella robusta*; Eand, *Eisenia andrei*, Dgyr; *Dimorphilus gyrociliatus*; Polf, *Philodryas olfersii* (Green snake); Tjac, *Thrasops jacksonii* (Jackson's black tree snake); Lmad, *Leioheterodon madagascariensis* (Malagasy giant hognose snake); Tnat, *Thalassophryne nattereri* (Copper Joe toadfish); Omyk, *Oncorhynchus mykiss* (Rainbow trout); Hlae, *Haliotis laevis* (Smooth Australian abalone); Hsap, *Homo sapiens*; Mmus, *Mus musculus*; Obt, obturacular region; Ves, vestimental region; Tru, trunk region; \*\*, region-specific DEGs.

**A****B**

Supplementary Figure S15.

(A) Molecular phylogenetic tree of chymotrypsin-like serine protease genes.

(B) Molecular phylogenetic tree of titin-like protein genes.

The maximum likelihood phylogenetic tree was constructed using aligned and gap-trimmed amino acid sequences. For genes retrieved from the UniProt database, taxonomy and accession numbers are shown. *L. satsuma* genes are highlighted in red. Circles on branches indicate bootstrap values higher than 80%. Lsat, *L. satsuma*; Rpac, *R. pachyptila*; Ctel, *C. teleta*; Hrob, *Helobdella robusta*; Hsap, *Homo sapiens*; Mmus, *Mus musculus*; Dmel, *Drosophila melanogaster*.

## Supplementary table footnotes

### Supplementary Table S1.

Statistics of gene models used in this study. For gene models downloaded from the NCBI RefSeq or GenBank database, accession numbers are shown. Only the longest transcript variants of each gene were counted. The longest transcript variants of each gene were translated into amino acid sequences, and then used for the analysis with BUSCO v.5.3.2.

S, complete and single-copy BUSCOs; D, complete and duplicated BUSCOs.

### Supplementary Table S2.

Statistics of RNA-seq, quality trimming, and mapping data.

### Supplementary Table S3.

Read counts for RNA-seq quantified with Salmon v.1.8.0

### Supplementary Table S4.

TPMs of RNA-seq samples calculated with EdgeR v.3.36.0.

### Supplementary Table S5.

CPMs of RNA-seq samples calculated with EdgeR v.3.36.0.

### Supplementary Table S6-8

Pairwise comparison of CPMs between obturacular and vestimental region, obturacular and trunk region, and trunk and vestimental region, respectively.

### Supplementary Table S9.

Statistics of the region-specific DEGs.

## References

1. Simakov, O., Marletaz, F., Cho, S.-J., et al. 2013, Insights into bilaterian evolution from three spiralian genomes. *Nature*, **493**, 526–31.
2. Albertin, C. B., Simakov, O., Mitros, T., et al. 2015, The octopus genome and the evolution of cephalopod neural and morphological novelties. *Nature*, **524**, 220–4.
3. Sun, Y., Sun, J., Yang, Y., et al. 2021, Genomic Signatures Supporting the Symbiosis and Formation of Chitinous Tube in the Deep-Sea Tubeworm *Paraescarpia echinospica*. *Mol. Biol. Evol.*, **38**, 4116–34.
4. de Oliveira, A. L., Mitchell, J., Girguis, P., and Bright, M. 2022, Novel Insights on Obligate Symbiont Lifestyle and Adaptation to Chemosynthetic Environment as Revealed by the Giant Tubeworm Genome. *Mol. Biol. Evol.*, **39**, msab347.
5. Fröblius, A. C., and Seaver, E. C. 2006, *ParaHox* gene expression in the polychaete annelid *Capitella* sp. I. *Dev. Genes Evol.*, **216**, 81–8.
6. Brooke, N. M., Garcia-Fernández, J., and Holland, P. W. H. 1998, The *ParaHox* gene cluster is an evolutionary sister of the *Hox* gene cluster. *Nature*, **392**, 920–2.