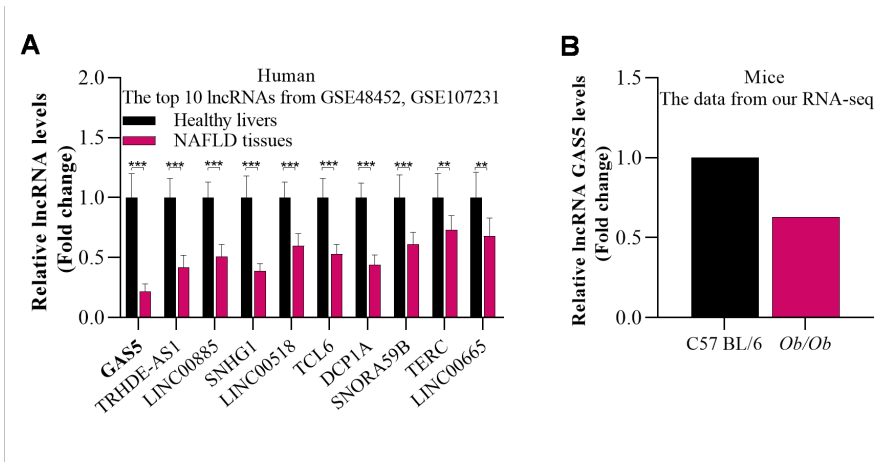


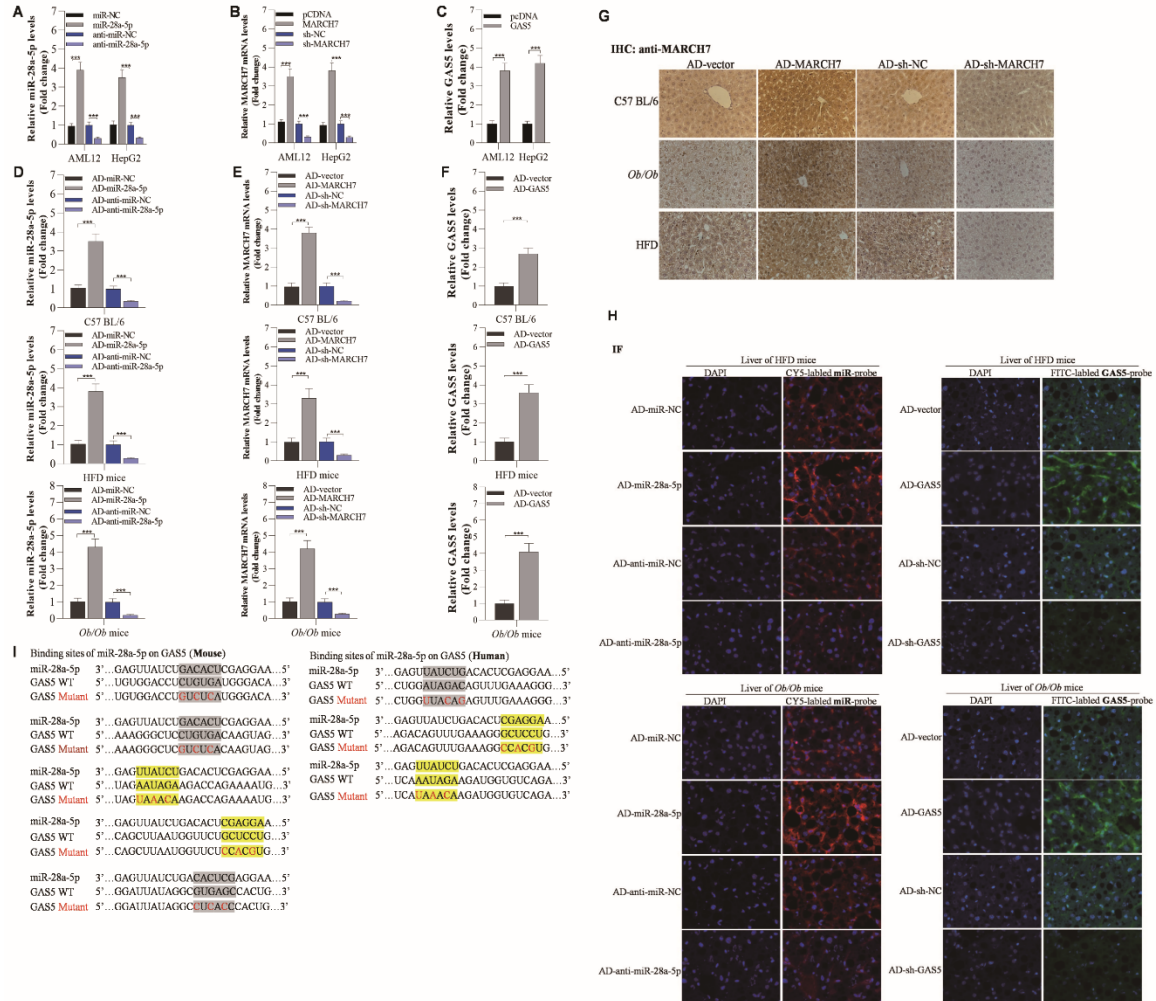
1 **Supplemental material**



2

3 **Fig. S1. GAS5 was the lncRNA with significantly changed expression identified by RNA-seq analysis**
4 **of liver samples. (A)** We first downloaded 2 human NAFLD-related microarray datasets, GSE48452
5 (containing 14 healthy control and 32 NAFLD tissue samples) and GSE107231 (containing 5 normal liver
6 and 5 NAFLD biopsy tissue samples), from Gene Expression Omnibus (GEO) and analysed the
7 downregulated lncRNAs in both datasets. Here, we reconfirmed the top 10 decreased lncRNAs by RT-qPCR
8 in our collected clinical specimens. **(B)** GAS5 expression was also decreased in the livers of NAFLD mice
9 as determined by lncRNA-seq analysis in our own database. The data are presented as the mean \pm S.D. of
10 three independent experiments. A p value of < 0.05 was considered statistically significant; and $*p < 0.05$,
11 $**p < 0.01$, $***p < 0.001$ assessed via a two-tailed t test for examining the significance of differences between
12 two groups; ns: not significant.

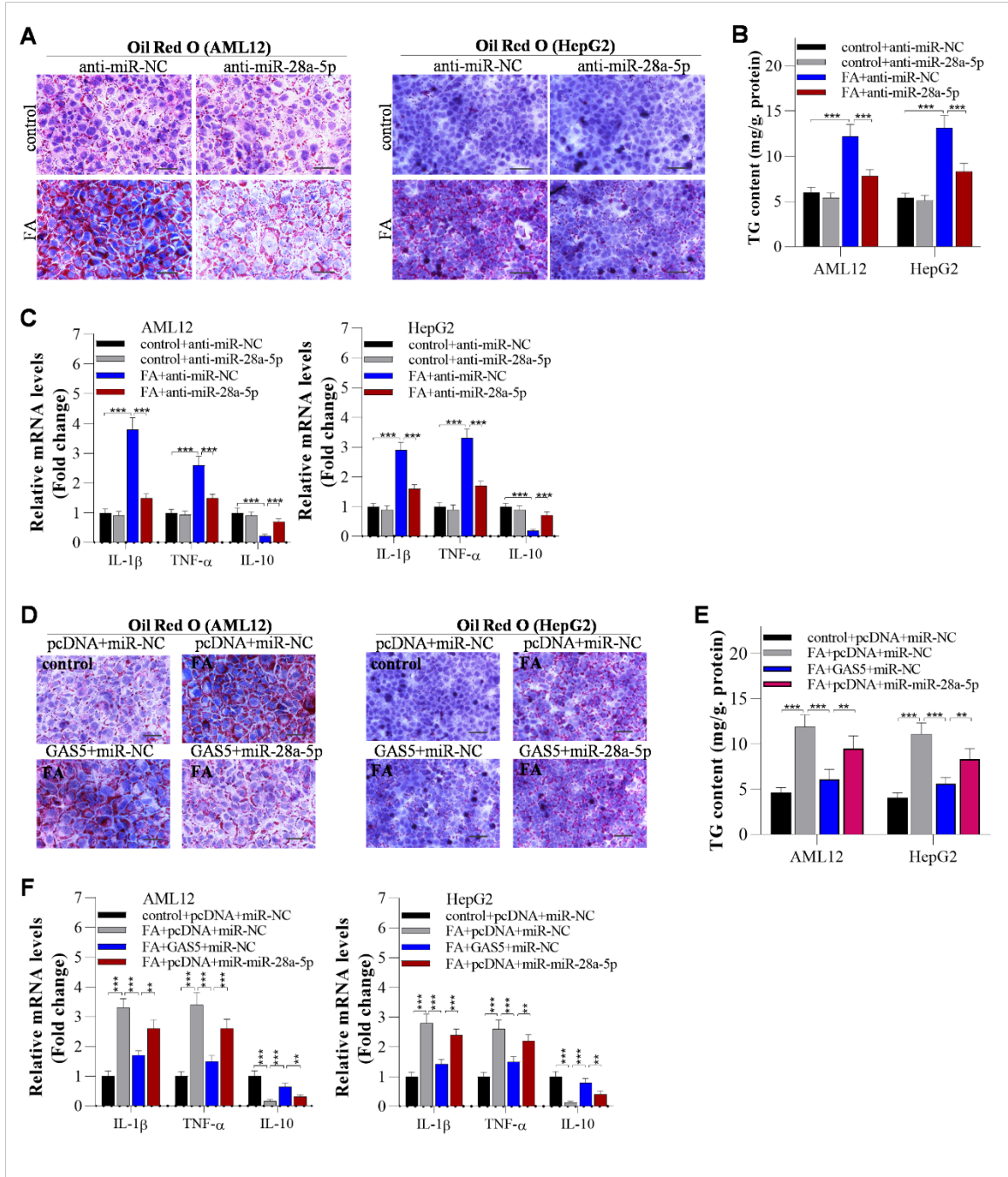
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14
 15 **Fig. S2. The expression of siRNAs, plasmids, and adenoviruses for knockdown or overexpression of**
 16 **GAS5, miR-28a-5p, and MARCH7 in AML12 cells, HepG2 cells and mouse livers.** In this study, AML12
 17 and HepG2 cells were transfected with miRNAs, plasmids, or lncRNAs for 12 h. In addition, mice were
 18 injected every 10 days with adenoviruses to decrease or increase the hepatic expression of GAS5, miR-28a-
 19 5p, and MARCH7. After 3 weeks of injection, the mice were prepared for further analysis. Herein, the
 20 expression of GAS5, miR-28a-5p, and MARCH7 *in vitro* and *in vivo* was measured using RT-qPCR analysis.
 21 (A) The expression of miR-28a-5p in AML12 and HepG2 cells. (B) The expression of MARCH7 in AML12
 22 and HepG2 cells. (C) GAS5 expression in AML12 and HepG2 cells. (D) The expression of miR-28a-5p in
 23 the livers of C57BL/6 mice, HFD-fed mice, and *Ob/Ob* mice after 3 weeks of the first injection. (E) The
 24 expression of MARCH7 in the livers of C57BL/6 mice, HFD-fed mice, and *Ob/Ob* mice. (F) The expression
 25 of GAS5 in the livers of C57BL/6 mice, HFD-fed mice, and *Ob/Ob* mice. (G) IHC staining of MARCH7
 26 levels in liver of mice after 3 weeks of injection. (H) Immunofluorescence detecting of GAS5 and miR-28a-
 27 5p in liver of mice after 3 weeks of injection. (I) Additionally, the binding sites between GAS5 and miR-28a-
 28 5p were predicted by sequence alignment, and there were two conserved binding sites between GAS5 and
 29 miR-28a-5p (yellow background). The data are presented as the mean \pm S.D. of three independent

30 experiments. Two-tailed Student's *t* test was used to determine the significance of differences between two
 31 groups (A-F). A *p* value of < 0.05 was considered statistically significant; and **p* < 0.05, ***p* < 0.01, ****p* <
 32 0.001; ^{ns}*p* > 0.05, ns: not significant.

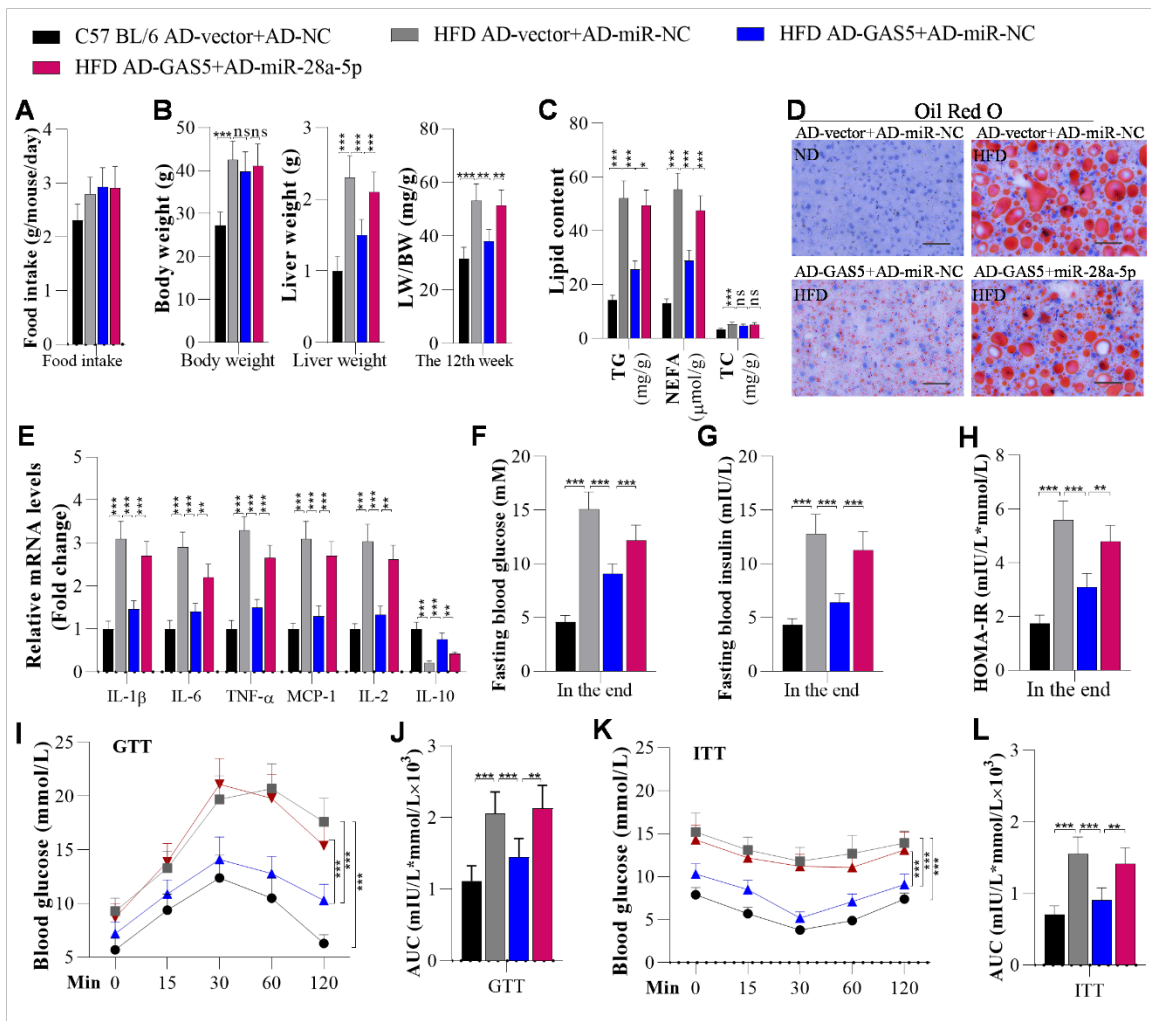
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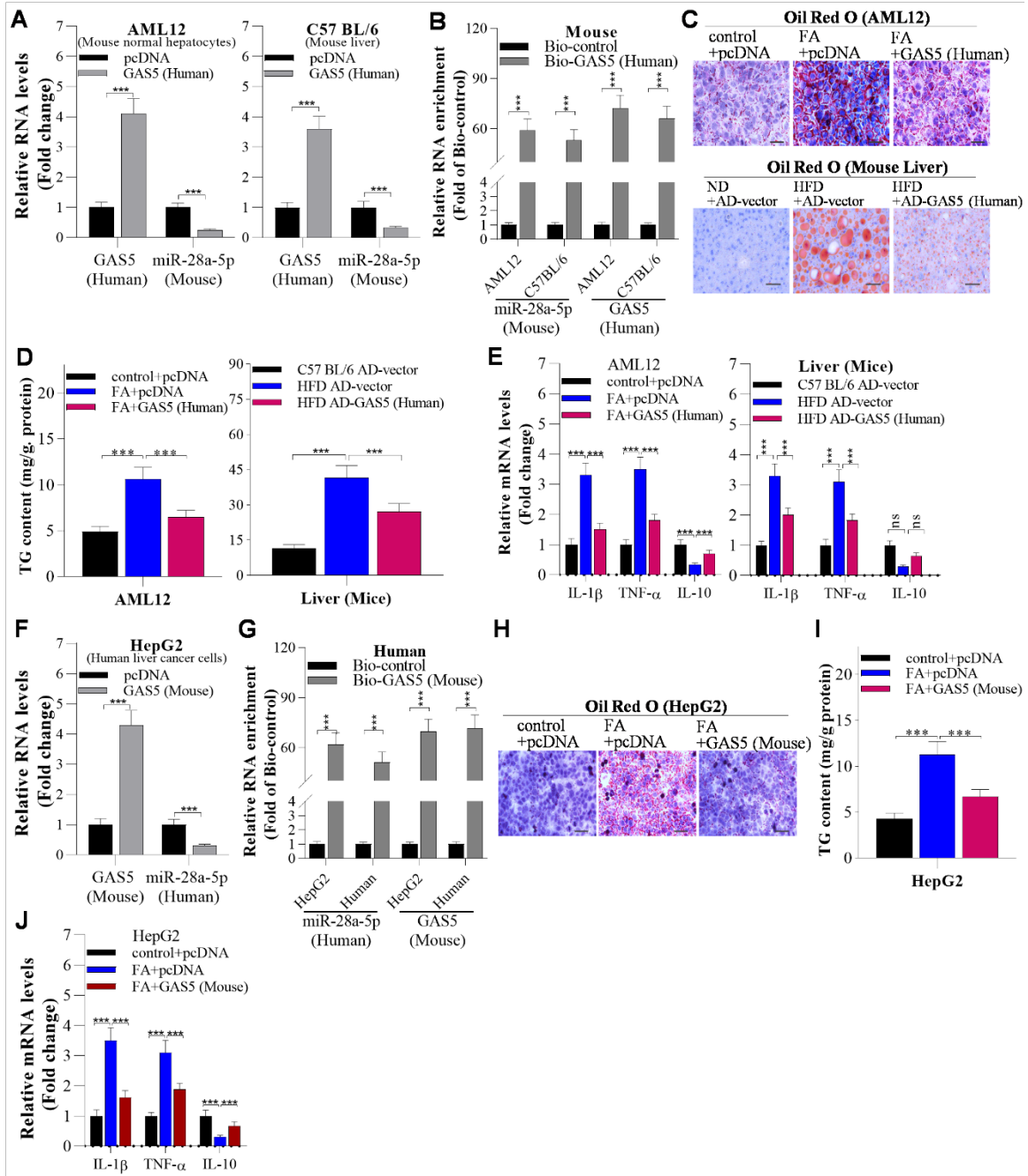
35 **Fig. S3. miR-28a-5p knockdown inhibits lipid deposition and inflammation in AML12 and HepG2 cells**
 36 **(A-C). Overexpression of miR-28a-5p significantly blocks GAS5 overexpression-induced inhibition of**
 37 **lipid accumulation in AML12 and HepG2 cells (D-F).**

38 On the one hand (A-C), AML12 and HepG2 cells were incubated with FAs for 12 h after 12 h of transfection
 39 with the indicated anti-miR-NC and anti-miR-28a-5p constructs. (A) Oil Red O staining; Scale bar, 50 μ M.
 40 (B) ELISA of TG content. (C) RT-qPCR analysis of inflammation-related genes. On the other hand (D-F),
 41 we found that overexpression of miR-28a-5p blocks GAS5-controlled lipid deposition and inflammation in
 42 AML12 and HepG2 cells. (D) Oil Red O staining; Scale bar, 50 μ M. (E) ELISA of TG content. (F) RT-
 43 qPCR analysis of inflammation-related genes. All indicated data are shown as the means \pm S.D.s. A *p* value
 44 of < 0.05 was considered statistically significant; and **p* <0.05 , ***p* <0.01 , ****p* <0.001 assessed via a ANOVA
 45 with the Bonferroni post hoc test for comparisons among more than two groups (B, C, E, F); ns: not
 46 significant.



48
 49 **Fig. S4. Overexpression of miR-28a-5p significantly reduces GAS5 overexpression-mediated**
 50 **suppression of hepatic steatosis, inflammation, and insulin resistance in HFD-fed mice.**
 51 Mice were fed a ND or HFD for 12 weeks. Then, they were injected with adenoviruses every 10 days, and
 52 these mice were continuously fed a ND or HFD. After 3 weeks of the first injection, the mice were used for
 53 further analysis. (A) Food intake was measured every day during ND or HFD feeding (n=5/group). (B) The

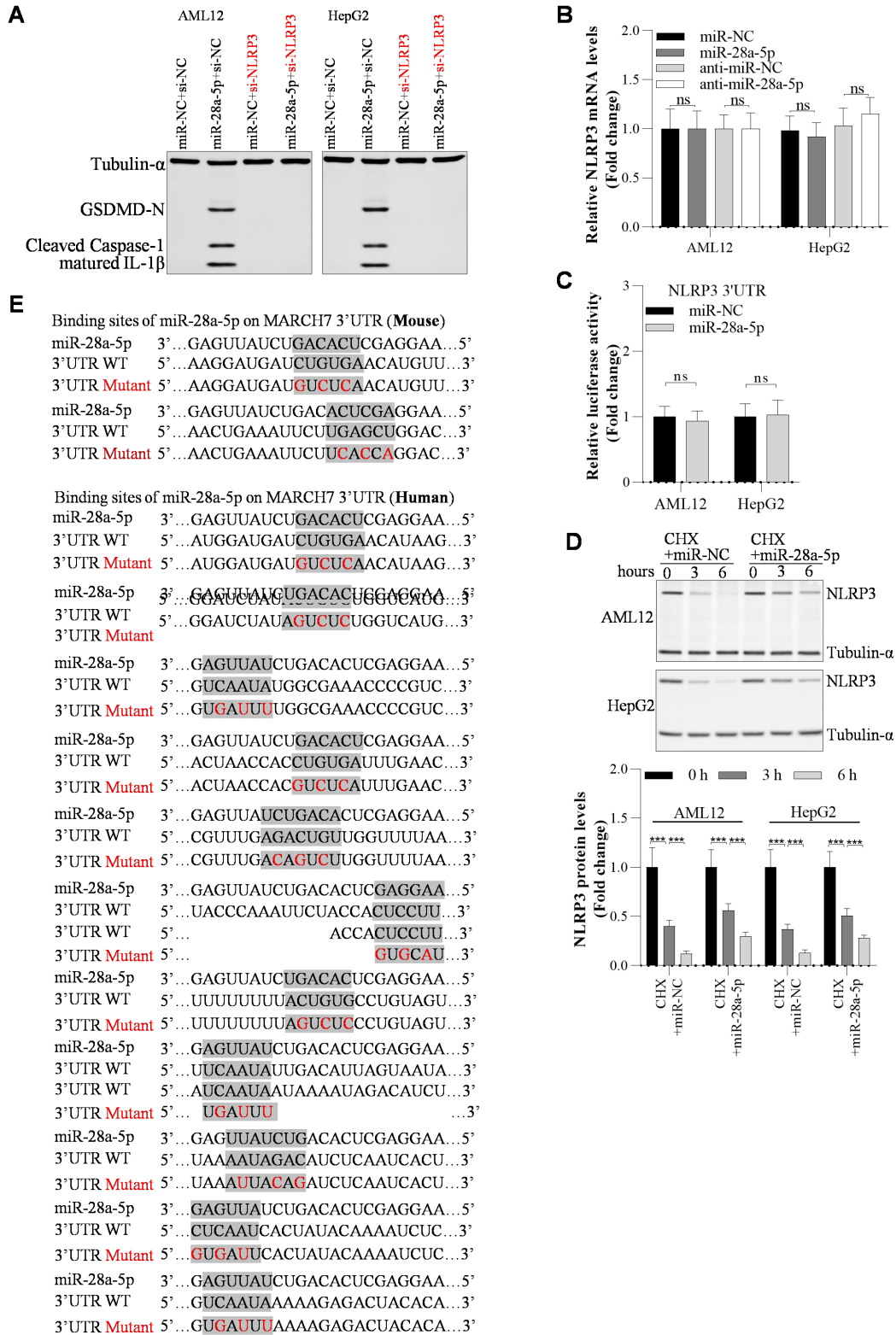
54 body weight, liver weight, and LW/BW ratios of C57BL/6 and HFD-fed mice after 3 weeks of injection of
55 adenoviruses (n=5/group). (C) TG, NEFA, and TC levels were measured using ELISA in the livers of
56 C57BL/6 and HFD-fed mice (n=5/group). (D) Oil Red O staining of liver sections from adenovirus-injected
57 C57BL/6 and HFD-fed mice (n=5/group); Scale bar, 50 μ M. (E) The mRNA levels of inflammation-related
58 genes analysed by RT-qPCR (n=5/group). (F) Fasting blood glucose levels of C57BL/6 and HFD-fed mice
59 (n=5/group). (G, H) Fasting insulin levels and HOMA-IR scores of adenovirus-injected C57BL/6 and HFD-
60 fed mice (n=5/group). (I-L) GTT and ITT of C57BL/6 and HFD-fed mice after 3 weeks of the first adenovirus
61 injection and the areas under the curve (AUCs) for the GTT and ITT (N=5/group). The data are presented as
62 the mean \pm S.D. of three independent experiments. A *p* value of < 0.05 was considered statistically significant;
63 and **p* <0.05 , ***p* <0.01 , ****p* <0.001 assessed via a ANOVA with the Bonferroni post hoc test for comparisons
64 among more than two groups (A-L); ns: not significant.
65



66
 67 **Fig. S5. Heterologous overexpression of GAS5 significantly suppresses lipid deposition and**
 68 **inflammation in AML12 and HepG2 cells.** Because there are two conserved binding sites between
 69 GAS5/miR-28a-5p between humans and mice, we hypothesized that the functions of the GAS5/miR-28a-5p
 70 axis were also conserved in the liver between these two species. (A) After overexpression of human GAS5
 71 in AML12 normal mouse hepatocytes for 12 h and in the livers of mice for one week, we examined the miR-
 72 28a-5p levels. (B) the Biotinylated GAS5 pull-down assay showed that human GAS5 interacted with mouse
 73 miR-28a-5p in AML12 cells and mouse liver tissues. (C, D, E) After overexpression of human GAS5 in
 74 AML12 cells treated with FAs for 24 h and the livers of C57BL/6 mice, we examined the lipid content and

75 inflammation; Scale bar, 50 μ M. **(F)** After overexpression of mouse GAS5 in HepG2 human liver cancer
76 cells for 12 hours, we determined the miR-28a-5p levels. **(G)** After overexpression of mouse GAS5 in human
77 liver cancer HepG2 cells, we confirmed that mouse GAS5 bound to human miR-28a-5p in HepG2 cells. **(H,**
78 **I, J)** After overexpression of mouse GAS5 inhibited lipid accumulation and inflammation in HepG2 cells,
79 we evaluated the lipid content and inflammation; Scale bar, 50 μ M. The data are presented as the mean \pm
80 S.D. of three independent experiments. A p value of < 0.05 was considered statistically significant; and
81 $*p<0.05$, $**p<0.01$, $***p<0.001$ assessed via a two-tailed t test for examining the significance of differences
82 between two groups (A, B, F, G) or ANOVA with the Bonferroni post hoc test for comparisons among more
83 than two groups (D, E, I, J); ns: not significant.

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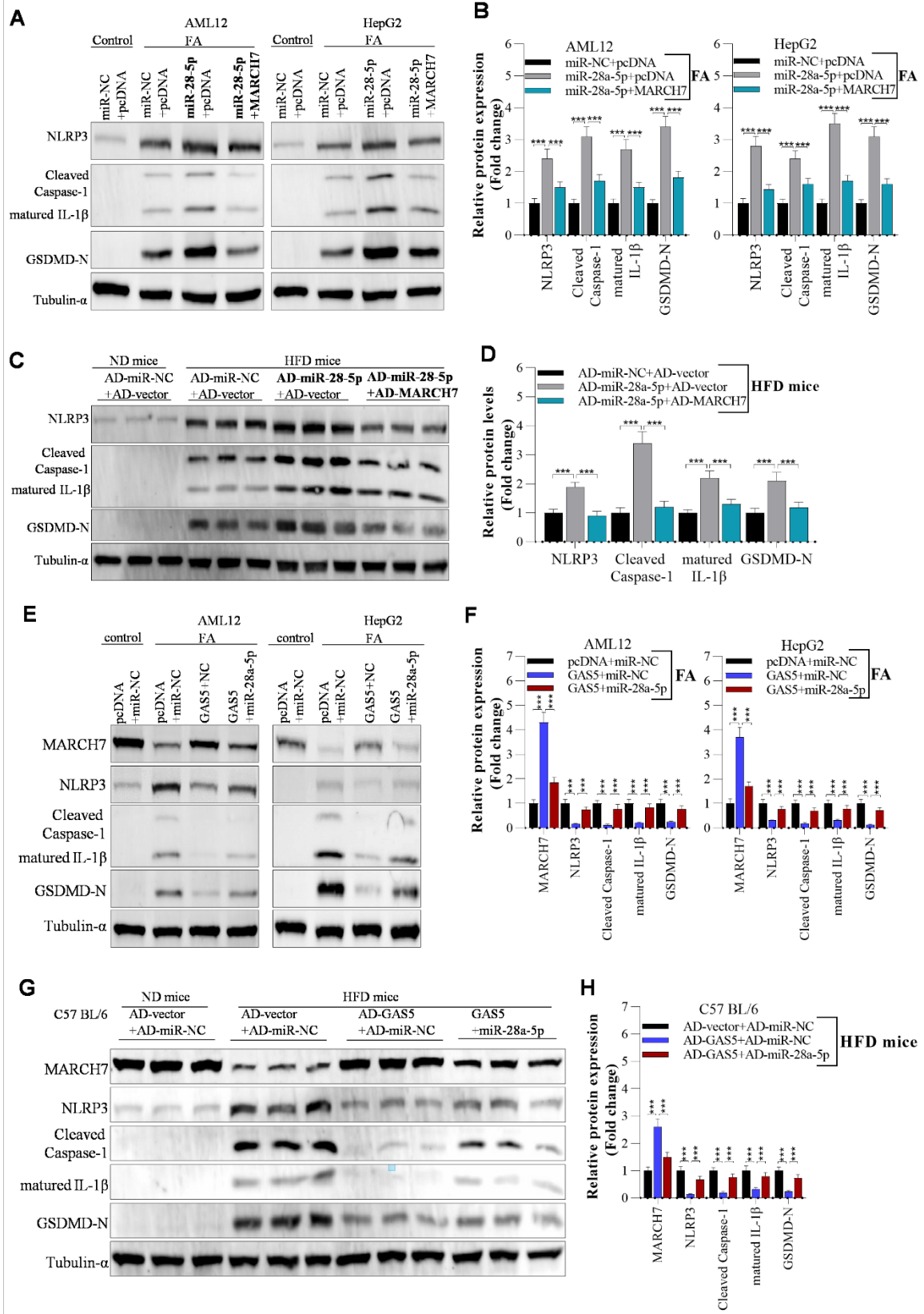
86 **Fig. S6. Interfering with NLRP3 expression blocks miR-28a-5p-mediated activation of pyroptosis. (A)**

87 Interfering with NLRP3 expression blocks miR-28a-5p-mediated activation of pyroptosis, as evidenced by

88 Western blot analysis of pyroptosis-associated proteins after 24 h of FA treatment in anti-miR-NC-, si-NC,

89 anti-miR-28a-5p, or si-NLRP3-transfected AML12 and HepG2 cells. **(B)** The hepatocytes were transfected
90 with miR-NC, miR-28a-5p, anti-miR-NC, anti-miR-28a-5p for 24 h, and they were collected to determine
91 NLRP3 mRNA expressions evidenced by RT-qPCR. **(C)** The hepatocytes were transfected with miR-NC,
92 miR-28a-5p, and co-transfected with NLRP3 3'UTR for 24 h. Then, they were prepared for luciferase
93 reporter assay. **(D)** miR-28a-5p significantly inhibited CHX-induced degradation of NLRP3 protein. They
94 were incubated with 30 μ M CHX along with miR-NC or miR-28a-5p transfection for 0 h, 3 h, 6 h. **(E)** The
95 potential binding sites of miR-28a-5p in the MARCH7 3'UTR (mouse or human) were examined by sequence
96 alignment. The data are presented as the mean \pm S.D. of three independent experiments. A *p* value of < 0.05
97 was considered statistically significant; and **p*<0.05, ***p*<0.01, ****p*<0.001 assessed via a two-tailed *t* test for
98 examining the significance of differences between two groups (B, C) or ANOVA with the Bonferroni post
99 hoc test for comparisons among more than two groups (D); ns: not significant.

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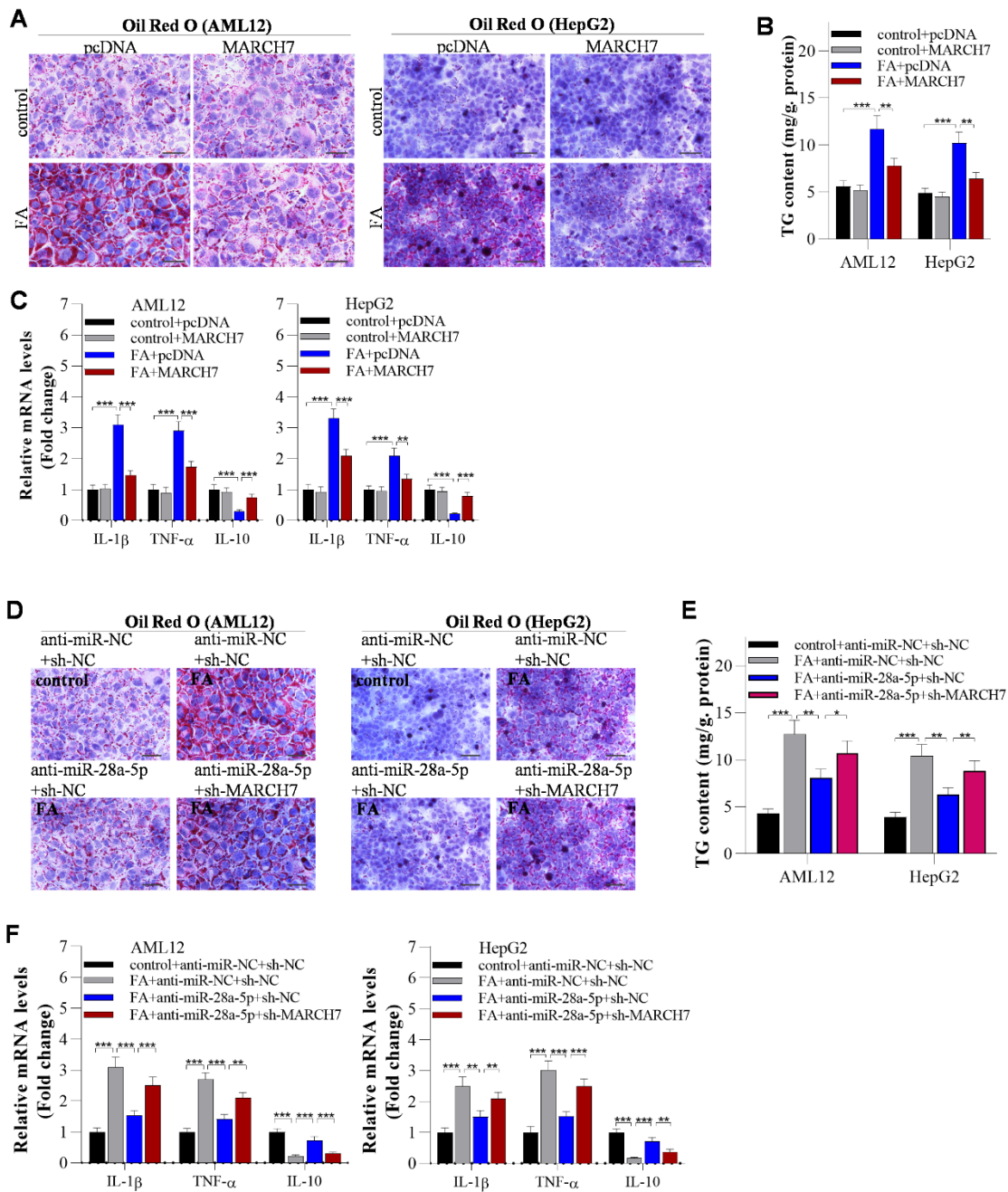


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Fig. S7. MARCH7 overexpression reversed the miR-28a-5p overexpression-induced enhancement of

103 **pyroptosis.** First, AML12 and HepG2 cells were transfected with overexpression plasmids for 12 h. Then,
104 the cells were incubated with FAs for 12 h. The cells were collected for further analysis. **(A, B)** Western blot
105 analysis of the expression of pyroptosis-related molecules in AML12 and HepG2 cells after 24 h of
106 transfection with miR-NC, pcDNA, miR-28a-5p, or pcDNA-MARCH7. **(C, D)** Western blot analysis of the
107 expression of pyroptosis-related molecules in the livers of C57BL/6 mice after adenovirus injection for 7
108 days. All the data are shown as the means \pm S.D.s. The data are presented as the mean \pm S.D. of three
109 independent experiments. **(E, F)** Western blot analysis of pyroptosis-related gene expression in AML12 and
110 HepG2 cells after 24 h transfection with pcDNA, miR-NC, GAS5, and another 12 h of FA treatment. **(G, H)**
111 Western blot analysis of pyroptosis-related gene expression in the livers of HFD-fed mice after 7 days of
112 injection with adenoviruses. The data are presented as the mean \pm S.D. of three independent experiments. A
113 *p* value of < 0.05 was considered statistically significant; and **p* <0.05 , ***p* <0.01 , ****p* <0.001 assessed via a
114 ANOVA with the Bonferroni post hoc test for comparisons among more than two groups (B, D, F, H); ns:
115 not significant.
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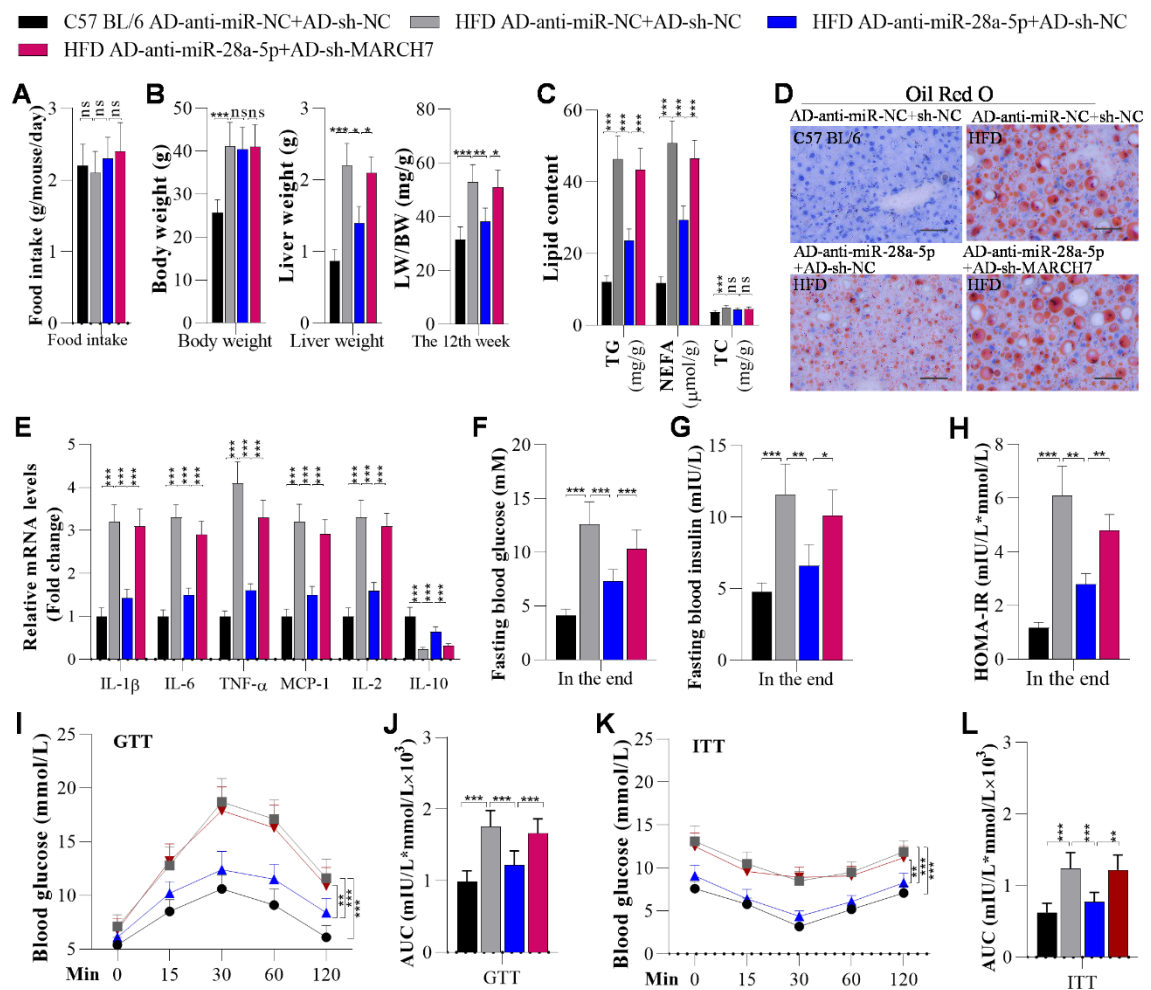
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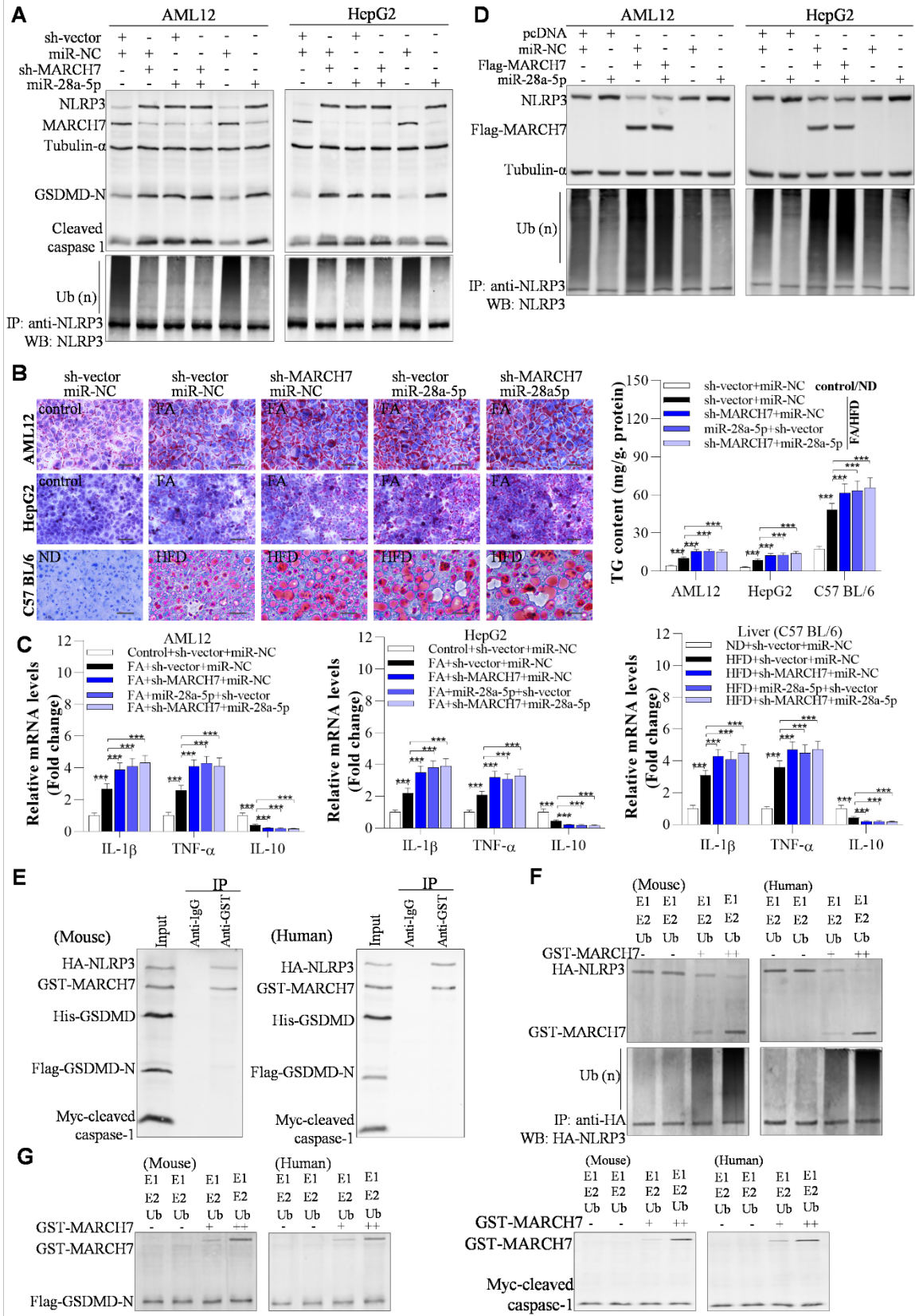
Fig. S8. Overexpression of MARCH7 attenuates lipid deposition in hepatocytes. AML12 and HepG2 cells were transfected with the indicated plasmids and miRNAs for 12 h and incubated with FAs for another 12 h. First, (A, D) Oil Red O staining of lipid droplets in hepatocytes; Scale bar, 50 μ m. (B, E) ELISA of TG content in AML12 and HepG2 cells. (C, F) RT-qPCR assay of inflammation-related gene expression. Afterwards, we found that MARCH7 overexpression significantly inhibited FA-induced hepatic lipid accumulation and that interfering with MARCH7 expression reversed the miR-28a-5p-mediated suppression of pyroptosis in AML12 and HepG2 cells. The data are presented as the mean \pm S.D. of three independent experiments. A p value of < 0.05 was considered statistically significant; and * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$

126 assessed via a ANOVA with the Bonferroni post hoc test for comparisons among more than two groups (B,
 127 C, E, F); ns: not significant.
 128



129
 130 **Fig. S9. Knockdown of MARCH7 reverses anti-miR-28a-5p-mediated suppression of hepatic steatosis,**
 131 **inflammation, and insulin resistance in HFD-fed mice.** Mice were fed a ND or HFD for 12 weeks. Then,
 132 they were injected *once a week* with the adenoviruses AD-anti-miR-28a-5p, AD-sh-NC, AD-anti-miR-28a-
 133 5p, and AD-anti-sh-MARCH7, *and* these mice were continuously fed a ND or HFD. After 3 weeks of the
 134 first injection, the mice were used for further experiments. (A) The food intake of ND- and HFD-mice
 135 (n=5/group). (B) The body weights, liver weights, and LW/BW ratios of C57BL/6 and HFD-fed mice
 136 (n=5/group). (C) TG, NEFA, and TC levels were measured using ELISA in the livers of C57BL/6 and HFD-
 137 fed mice (n=5/group). (D) Oil Red O staining of mouse liver sections (n=5/group); Scale bar, 50 μ M. (E)
 138 The mRNA levels of inflammation-related genes were measured by RT-qPCR (n=5/group). (F) Fasting blood
 139 glucose levels of C57BL/6 and HFD-fed mice (n=5/group). (G, H) Fasting insulin levels and HOMA-IR
 140 scores of adenovirus-injected C57BL/6 and HFD-fed mice (n=5/group). (I-L) GTT and ITT of C57BL/6 and
 141 HFD-fed mice after 3 weeks of the first adenovirus injection and the areas under the curve (AUCs) for the

142 GTT and ITT (N=5/group). The data are presented as the mean \pm S.D. of three independent experiments. A
143 p value of < 0.05 was considered statistically significant; and * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$ assessed via a
144 ANOVA with the Bonferroni post hoc test for comparisons among more than two groups (A-L); ns: not
145 significant.
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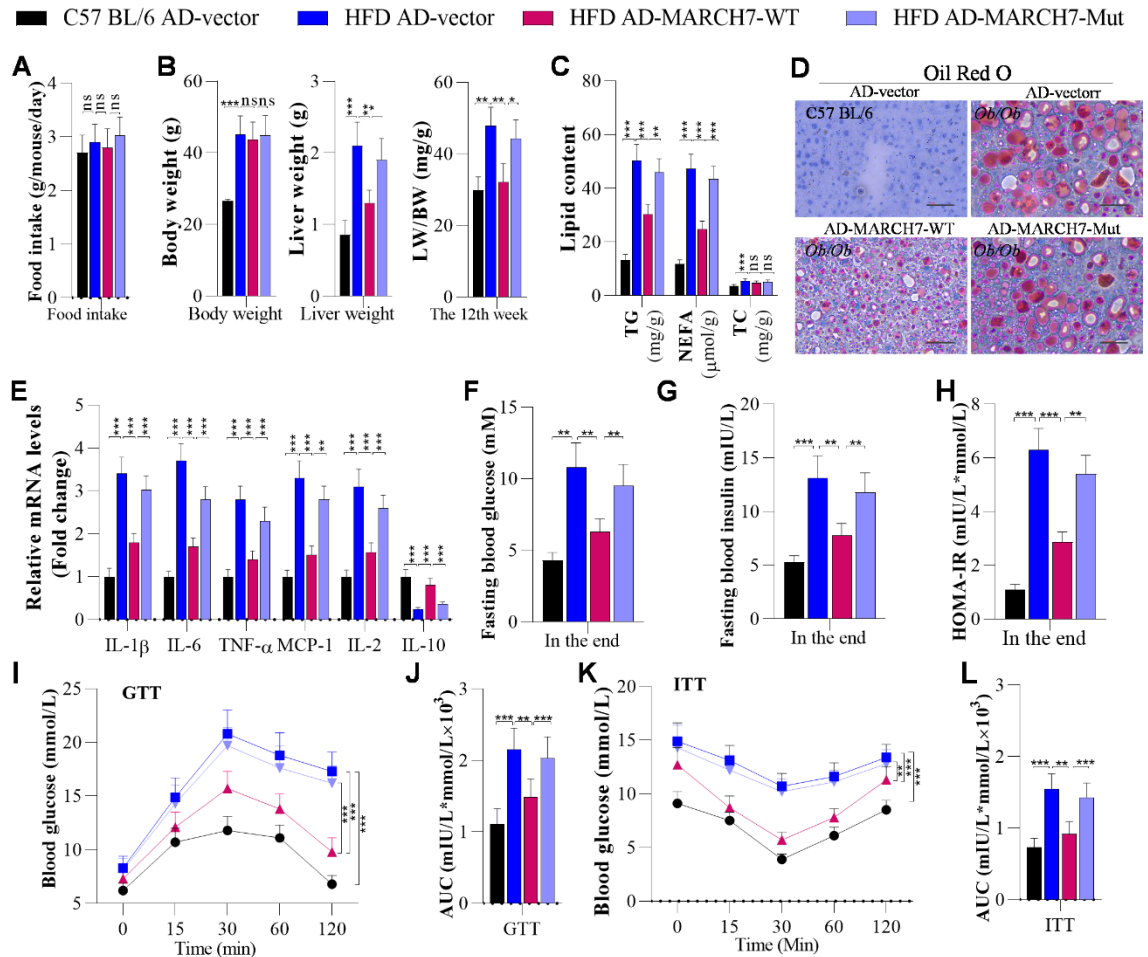


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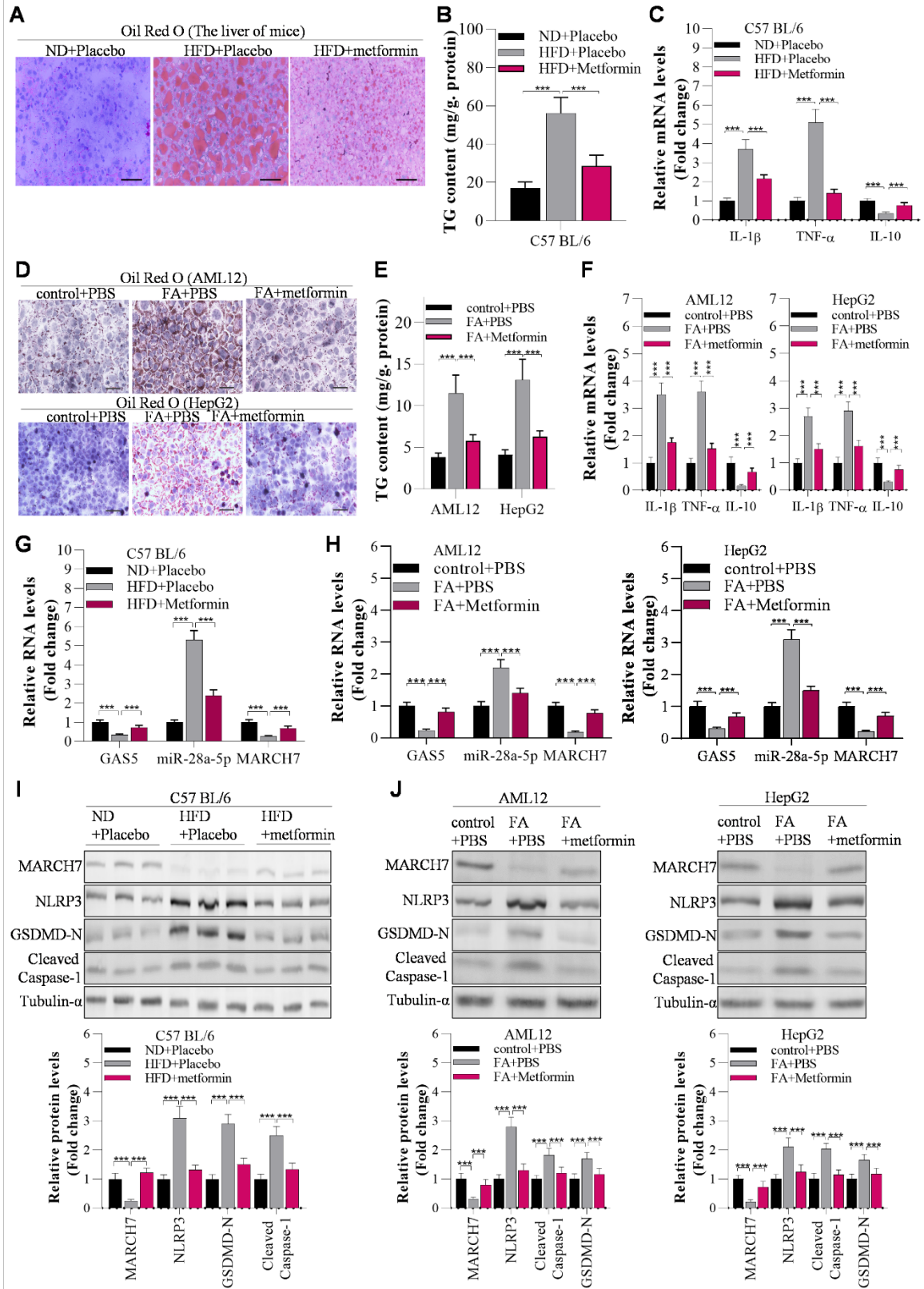
Fig. S10. MARCH7 was required for miR-28a-5p-induced inhibition of NLRP3 ubiquitination-

149 **mediated degradation to promote pyroptosis.** (A) After MARCH7 knockdown for 48 h, we overexpressed
150 miR-28a-5p in AML12 and HepG2 cells for another 48 h. Then, we harvested them for Western blot analysis
151 of pyroptosis and for analysis of NLRP3 ubiquitination after 12 h of incubation with 20 μ M MG132. (B, C)
152 After MARCH7 knockdown for 48 h, we overexpressed miR-28a-5p in AML12 and HepG2 cells for another
153 48 h, and they were incubated with FAs for 8 h; Scale bar, 50 μ M. In addition, mice were fed a ND or HFD
154 for 8 weeks. Then, they were injected with adenoviruses *every 10 days*, and these mice were continuously
155 fed a ND or HFD. After the first injection for 3 weeks, the mice were used for Oil Red O staining, TG
156 measurement, and RT-qPCR. (D) After MARCH7 overexpression for 48 h, we overexpressed miR-28a-5p
157 in AML12 and HepG2 cells for another 48 h, and we collected them for analysis of NLRP3 ubiquitination
158 after 12 h of incubation with 20 μ M MG132. (E) The purified proteins were prepared by Sangon (Shanghai)
159 and mixed together in cell lysis buffer, and the MARCH7-bound proteins were identified using a GST pull-
160 down assay. (F, G) The purified proteins were mixed together in cell lysis buffer following the instructions
161 of an *in vitro* ubiquitination assay kit (VIVA Bioscience, Amyjet Scientific Inc, Wuhan, China) and an *in vitro*
162 ubiquitination assay protocol (DOI: 10.21769/BioProtoc.928); the effects of GST-MARCH7 on the levels of
163 HA-NLRP3, Flag-GSDMD-N, and Myc-caspase-1 were examined by western blotting; and the
164 ubiquitination of HA-NLRP3 was analysed. The data are presented as the mean \pm S.D. of three independent
165 experiments. A *p* value of < 0.05 was considered statistically significant; and **p* <0.05 , ***p* <0.01 , ****p* <0.001
166 assessed via a ANOVA with the Bonferroni post hoc test for comparisons among more than two groups (B,
167 C); ns: not significant.
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Fig. S11. The MARCH7 mutant fails to suppress hepatic steatosis, inflammation, and insulin resistance in HFD-fed mice. Mice were fed a ND or HFD for 12 weeks. Then, they were injected every 10 days with the adenoviruses AD-vector, AD-MARCH7-WT, and AD-MARCH7-Mut, and these mice were continuously fed a ND or HFD. After 3 weeks of the first injection, the mice were used for analysis. (A) The food intake of ND- and HFD-mice (n=5/group). (B) The body weights, liver weights, and LW/BW ratios of C57BL/6 and HFD-fed mice (n=5/group). (C) Hepatic TG, NEFA, and TC levels were measured using ELISA (n=5/group). (D) Oil Red O staining of mouse liver sections (n=5/group); Scale bar, 50 μ M. (E) The mRNA levels of inflammation-related genes analysed by RT-qPCR (n=5/group). (F) Fasting blood glucose levels of C57BL/6 and HFD-fed mice (n=5/group). (G, H) Fasting insulin levels and HOMA-IR scores of C57BL/6 and HFD-fed mice (n=5/group). (I-L) GTT and ITT of C57BL/6 and HFD-fed mice and the areas under the curve (AUCs) for the GTT and ITT (N=5/group). The data are presented as the mean \pm S.D. of three independent experiments. A *p* value of < 0.05 was considered statistically significant; and **p*<0.05, ***p*<0.01, ****p*<0.001 assessed via a ANOVA with the Bonferroni post hoc test for comparisons among more than two groups (A-L); ns: not significant.

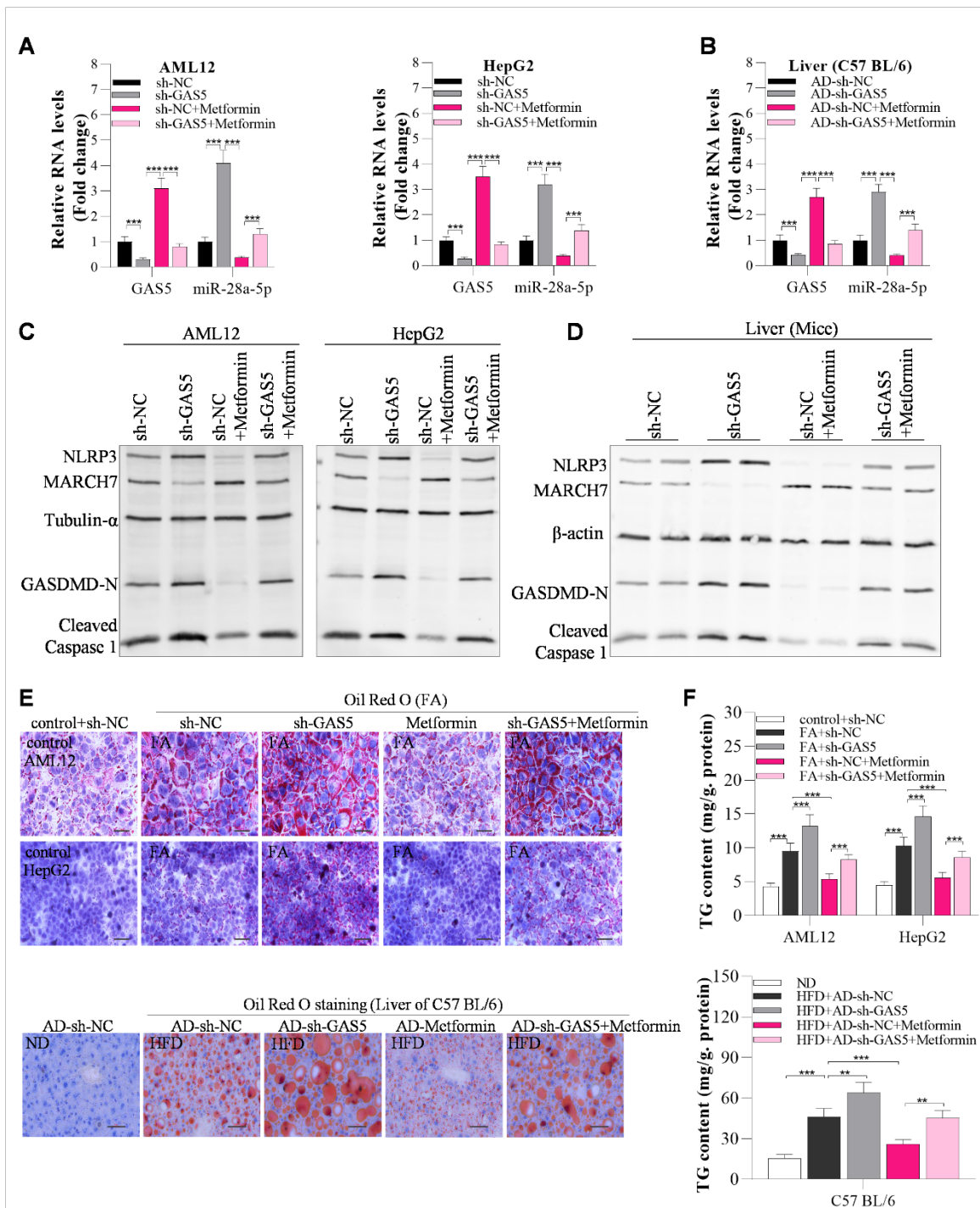


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Fig. S12. Metformin exerts anti-NAFLD activity by affecting the levels of GAS5, miR-28a-5p,

187 **MARCH7, and NLRP3.** C57BL/6 mice (N=5) were administered daily with 200 mg metformin per
188 kilogram of body weight for a month accompanied by 12 weeks of ND or HFD feeding. AML12 and HepG2
189 cells were incubated with FAs for 6 h, and they were incubated with both 10 nM metformin and FA for
190 another 6 h. (A) Oil Red O staining of mouse liver sections; Scale bar, 50 μ M. (B) The TG content was
191 measured using ELISA in the livers of 12-week-old ND- or HFD-fed C57BL/6 mice (n=8/group). (C) RT-
192 qPCR analysis of inflammatory factors in the livers of mice. (D) Oil Red O staining of hepatocytes (AML12
193 and HepG2); Scale bar, 50 μ M. (E) The TG content was measured using ELISA in hepatocytes. (F) RT-
194 qPCR analysis of inflammatory genes in hepatocytes. (G) RT-qPCR analysis of the expression of miR-28a-
195 5p, MARCH7, and GAS5 in the livers of mice. (H) RT-qPCR analysis of the expression of miR-28a-5p,
196 MARCH7, and GAS5 in hepatocytes. (I) Western blot analysis of MARCH7-mediated pyroptosis genes in
197 the livers of mice. (J) Western blot analysis of MARCH7-mediated pyroptosis genes in hepatocytes. The
198 data are presented as the mean \pm S.D. of three independent experiments. The one-way analysis of variance
199 (ANOVA) with the Bonferroni post hoc test was conducted for comparisons among more than two groups
200 (B, C, E-J). A *p* value of < 0.05 was considered statistically significant; and **p* < 0.05 , ***p* < 0.01 , ****p* < 0.001 ;
201 ^{ns}*p* > 0.05 , ns: not significant.
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Fig. S13. Knockdown of GAS5 significantly inhibited the effects of metformin on pyroptosis and hepatic lipid deposition. After 48 h of transfection, AML12 and HepG2 cells were incubated with 10 nM metformin for another 6 h. After one week of injection of AD-GAS5, C57BL/6 mice were administered 200 mg metformin per kilogram of body weight daily for one month. Then, AML12 cells, HepG2 cells, and mouse livers were collected for further experiments. (A, B) RT-qPCR was used to determine the effects of metformin on miR-28a-5p levels in GAS5-knockdown AML12 cells, HepG2 cells, and mouse livers. (C, D)

210 Western blot analysis of the effects of metformin on pyroptosis in GAS5-knockdown AML12 cells, HepG2
211 cells, and mouse livers. (E, F) Oil Red O staining was used to examine the effects of metformin on lipid
212 accumulation in GAS5-knockdown AML12 cells, HepG2 cells, and mouse livers; Scale bar, 50 μ M. The data
213 are presented as the mean \pm S.D. of three independent experiments. The one-way analysis of variance
214 (ANOVA) with the Bonferroni post hoc test was conducted for comparisons among more than two groups
215 (A, B, F). A p value of < 0.05 was considered statistically significant; and * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$;
216 ^{ns} $p > 0.05$, ns: not significant.
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