VectorNet: collaborative mapping of arthropod disease vectors in Europe and surrounding areas since 2010

SUPPLEMENTARY INFORMATION

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# SUPPLEMENTARY INFORMANTION SECTION I: Supplementary Table S1: Contributor List

Contributors involved in all constituent projects. Key for Vector Group Column: C= Culicoides, M = Mosquitoes, S= Sand flies, T=Ticks, VEN = VectorNet Entomological Network Member, VGL = Vector Group Leader.

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Harrat, Z.AlgeriaLaboratoire éco-épidémiologie Parasitaire et Génétique des Populations, Institut Pasteur d'AlgérieVENHendrickx G.BelgiumAviaGIS, BelgiumSHenney J.GuernseyEnvironment GuernseyTHlavackova K.CzechiaCharles University Prague, CzechiaSHodzik A.AustriaUniversity of Veterinary Medicine ViennaTHøye TT.DenmarkDepartment of Bioscience and Arctic Research CentreCHubalek Z.CzechThe Czech Academy of Sciences RepublicTHuber K.FranceINRAeC	Hansford K.	UK	UK Health Security Agency	Т
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Høye TT. Denmark Department of Bioscience and Arctic Research Centre C   Hristescu D. Romania Institute for Diagnosis and Animal Health C   Hubalek Z. Czech Republic The Czech Academy of Sciences Republic T	Hodzik A.	Austria	University of Veterinary Medicine Vienna	Т
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Hubalek Z. Czech The Czech Academy of Sciences T   Republic INRAe C	Hristescu D.	Romania	Institute for Diagnosis and Animal Health	С
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	Huber K.	France	INRAe	С

Contributor	Country	Institution	Vector
			Group
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		Safety, Institute for Medical	
		Microbiology and Hygiene	
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Jääskeläinen A.	Finland	Independent	Т
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Janev-Holker, N.	Croatia	Croatian Institute of Public Health	VEN
Johnston C.	UK	UK Health Security Agency	Т
Jöst A.	Germany	Kommunale Aktionsgemeinschaft zur	М
		Bekämpfung der Schnakenplage, Speyer	
Jourdain, F.	France	Santé publique France	VEN
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Kalan K.	Slovenia	University of Primorska	М
Каро N.	Bosnia and	University of Sarajevo	Т
	Herzegovina		
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Karakus M.	Turkey	Hacettepe University, Ankara	S
Kasap O.	Turkey	Hacettepe University, Ankara	S
Kavur H.	Turkey	Cukurova University, Dept of Medical	S, M
Kenveres 7	Hungary	Acrida Conservational Research L P	м
Kenyeres Z.	rungary	Tapolca	141
Khalin A:	Russia	Zoological Institute, Russian Academy of	М
		Sciences, St. Petersburg	
Khallaayoune	Morocco	Institut Agronomique et Vétérinaire	С
К.		Hassan II	
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		Pharmacy	

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			Group
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		Medicine, Hamburg	
Kuhn C.	Germany	Umweltbundesamt, Berlin	М
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Larska M.	Poland	National Veterinary Research Institute	С
Lindstrom A.	Sweden	National Veterinary Research Institute	VEN
Lucientes J.	Spain	Facultad de Veterinaria. University of	С
		Zaragoza	
Madeira S.	Portugal	CIISA - Centro de Investigação	С
		Interdisciplinar em Sanidade Animal	
Mammadova	Azerbaijan	Azerbaijan State Agricultural University	T, M, VEN
К.			
Mancini G.	Italy	Istituto Zooprofilattico Sperimentale	С
		dell'Abruzzo e del Molise 'G. Caporale'	
Martínez-	Spain	Rede Galega de Vixilancia de Vectores	М
Barciela Y.		(ReGaViVec), Xunta de Galicia,	
		Department of Ecology and Animal	
		Biology, Faculty of Biology, University of	
		Vigo	
Mathieu B.	France	Institut de Parasitologie et de Pathologie	С
		Tropicale, Strasbourg	
McGinley L.	UK	UK Health Security Agency	Т
Meadows S.	Jersey	States of Jersey Department of	Т
		Environment	
Medlock J.	UK	UK Health Security Agency	М, Т,
			VGL, VEN
Michaelakis A.	Greece	Benaki Phytopathological Institute,	М
		Arhens	
Melashvili G.	Georgia	Agricultural University of Georgia	Т
Mihalca A. D.	Romania	University of Agricultural Sciences and	S, T, VGL,
		Veterinary Medicine of Cluj-Napoca	VEN
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		Parasitic Diseases, Sofia	VEN
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		(CIBERESP), Madrid	
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		l'Ambiente, Turin	
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Ν.		Sciences, University of Prishtina	

Contributor	Country	Institution	Vector
			Group
Müller P.	Switzerland	Swiss Tropical and Public Health Institute, Basel	М
Muñoz C.	Spain	University of Murcia	S
Murchie A.	United-	Agri-Food and Biosciences Institute	С
	Kingdom		
Muscat I.	Jersey	Jersey General Hospital's Pathology	т
		Laboratory	_
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Nițescu C.	Romania	Institute for Diagnosis and Animal Health	С
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Olafsson E.	Iceland	Icelandic Institute of Natural History	Т
Omeragic J.	Bosnia and Herzegovina	University of Sarajevo - Veterinary Faculty	T, VEN
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Ozber 1.	титкеу	Department of Parasitology, Izmir	
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Tchakarova S.	Bulgaria	National Diagnostic and Research	С		
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	Republic				
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Wieckowski F.	Alderney	Alderney Wildlife Trust	Т		
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## SUPPLEMENTARY INFORMATION SECTION II: Supplementary Table S2: Literature Search Protocols

Databases: Medline/Pubmed; Scopus; Embase; Google Scholar; Web of Science (if available to Institution)

More specialist institutional databases may also be used such as CORDIS, as well as, dependent on institutional access. Other published sources include: Online accessible dedicated web-sites such as those maintained by national public and veterinary health institutes and services (examples: <u>https://solidarites-sante.gouv.fr/sante-et-</u><u>environnement/risques-microbiologiques-physiques-et-chimiques/especes-nuisibles-et-parasites/article/cartes-de-</u><u>presence-du-moustique-tigre-aedes-albopictus-en-france-metropolitaine;</u> https://www.nvwa.nl/onderwerpen/muggen-knutten-en-teken) are checked continually for primary data sources.

#### **Time limits**

Default time limits: 2016-present

Vector group specific

Mosquitoes: 2013–present for RVF Mandate: *caspius, detritus, pipiens, albopictus, japonicus, vexans, theileri*, 1980– present for species not extracted before: *claviger, richiardii, annulata, coluzzii*, 2016–present for all others

Culicoides: 1980–present for countries not already covered during VectorNet 2014/18 (Austria, Croatia, Czech Republic, Denmark, Germany, Ireland, Norway, Slovakia and Sweden); 2016–present for all others

Ticks: All years for Hyalomma lusitanicum; 2016-present for all others

Sand flies: All years for langeroni, alexandri, kandelaki, major, 2016-present for all others

#### Minimum data requirements for eligibility

Data Source: a personal communication without an electronic document with the relevant data is not sufficient, but an unpublished report is fine as long as it is available for access.

Location: data with a precision to less than NUTS2 level or equivalent should not be considered, data with NUTS2 precision should be marked as such.

Vector species: data where samples are not identified to the level of species or species groups, except where currently allowed in the migrated VectorNet list, should not be considered: -Exclusion if identification is not based on reliable method within species complexes (e.g. *Anopheles maculipennis* complex: morphology accepted only for *An. sacharovi* adults, molecular methods only for all other cases).

Reported status or number of specimens caught: if the number of specimens caught is not reported, the reported status is a mandatory minimum data requirement.

Collection effort end date: If the source does not have a valid vector collection end date with accuracy less than a year, the data should not be considered.

**Search Terms:** The syntax of the search strings with the terms listed below needs to be built and adjusted dependent on the database. The terms and search strategy may be adjusted depending on the quality of the search results. The terms are listed in four lists: a 'datatype OR list', a 'Country/Region OR list', a 'Genus OR list' and a 'Species OR list'. The latter two are vector group-specific lists. In the searches, these are combined into search-strings according to the following principle:

#### Applicable to all Vector groups

*Terms in Title OR in Abstract*: 'Genus OR list' AND in all fields: 'Species OR list', AND in all fields: 'Datatype OR list' AND in all fields: 'Country/Region OR list'

*Datatype OR List*: distribution OR presence OR occurrence OR report\* OR spread OR dispers\* OR detect\* OR abundan\* OR densit\* OR number OR absen\* OR surv\* OR monitor\* OR introduc\* OR intercept\*

*Country/Region OR list:* Europe\* OR EU OR "Mediterranean Basin" OR "Mediterranean area" OR Balkan\* OR Scandinavia\* OR "Iberian peninsula" OR Aland OR Albania\* OR Andorra\* OR Atlantic OR Austria\* OR Belgi\* OR "Black sea" OR Bosnia\* OR Herzegovina OR Bulgaria\* OR Croatia\* OR Cypr\* OR "Czech Republic" OR Denmark OR Greenland OR German\* OR Spain OR Estonia\* OR Finland OR "Faroe islands" OR France OR Corsica\* OR Greece OR Gibraltar OR Hungary OR Iceland\* OR Ireland OR Italy OR Sicil\* OR Sardinia\* OR Kosov\* OR Latvia\* OR Liechtenstein OR Lithuania\* OR Luxembourg OR Macedonia\* OR FYROM OR Malta OR Monac\* OR Monegasqu\* OR Montenegr\* OR Netherlands OR Norway OR Poland OR Portug\* OR Slovenia\* OR Romania\* OR "San Marino" OR Serbia\* OR Slovakia\* OR Switzerland OR Sweden OR "United Kingdom" OR "British Isles" OR "Great Britain" OR Wales OR England OR Scotland OR Turk\* OR "Vatican city" OR Svalbard OR Israel\* OR Palestin\* OR Jordan\* OR Leban\* OR Syria\* OR Morocc\* OR Algeria\* OR Tunisia\* OR Libya\* OR Egypt\* OR "Western Sahara\*" OR Armenia\* OR Azerba\* OR Belarus OR Bielorussia\* OR Georgia\* OR Moldov\* OR Russia\* OR Yugoslavia\* OR Ukrain\* OR Ukrayin\* OR OR USSR OR SSSR OR "Soviet Union" OR British OR Irish OR Scottish OR Welsh OR "Channel Islands" OR Jersey OR Guernsey OR Sark OR French OR OR Greek OR Italian OR Spanish OR Swiss OR Swedish OR Transcaucasia\* OR CaucasusOR Danish OR Finnish OR Norwegian OR Baltic OR Czech\* OR Hungarian OR Polish OR Mediterranean OR Sahara OR OR Majorca\* OR Mallorca\* OR Minorca\* OR Ibiza OR Azores OR Canar\* OR Balearic\* OR "Member St\*" OR "North Africa\*" OR Kazakh\*OR Uzbek\* OR Karakalpakstan\* OR Crimea\* OR Nakhchivan\* OR Alsac\* OR Abkhazia\* OR Adjara\* OR Ossetia OR Athos OR Aosta\* Friuli\* Giulia\* Trentin\* Adige Südtirol\* OR "South Tyrol\*" Găgăuzia\* Transnistria\* Madeira\* Catalan Catalonia\* Adygea\* Bashkortostan\* Chechnya\* Chuvashia\* Dagestan\* Ingushetia\* Karbardin\* Kalmykia\* Karachay Komi\* Karelia "Mari El"\* Mordovia\* Tatarstan\* Udmurtia\* Nenets Vojvodin\* Metohija\* Srpska\* Andalusia\* Aragon\* Asturias\* "Balearic Islands" Basque Cantabria\* Castile\* Mancha\* León Extremadura\* Galicia\* Rioja\* Madrid Murcia\* Navarr\* Valencia\* Ceuta\* Melilla\* "Isle of Man"

### Terms applicable to individual Vector groups

## Genus OR list.

Mosquitoes: Aedes OR Stegomyia OR Ochlerotatus OR Anopheles OR Culex OR mosquito\* OR Culicidae OR Culiseta OR Coquillettidia

### Culicoides: Culicoides OR "biting midge\*"

Ticks: *Ixodes* OR *Dermacentor* OR *Rhipicephalus* OR *Hyalomma* or OR *Ornithodoros* OR "ticks" OR "soft ticks" OR "hard ticks"

Sand flies: Phlebotomus OR Leishmania OR sand flies OR "sand flies" OR "sand fly"

#### Species OR list:

Mosquitoes: *albopict*\* OR *aegypti* OR *atropalpus* OR *koreicus* OR *japonic*\* OR *caspius* OR *coluzzii* OR *detritus* OR *vexans* OR *plumbeus* OR *maculipennis* OR *messeae* OR *atroparvus* OR *sacharovi* OR *labranchiae* OR *superpictus* OR *modestus* OR *perexiguus* OR *univittatus* OR *pipiens* OR *molestus* OR *quinquefasciatus* OR *annulata* OR *claviger* OR *antennatus* OR *richiardii* OR *theileri* OR *torrentium* OR *tritaeniorhynchus* 

### Culicoides: N/A

Ticks: ricinus OR persulcatus OR reticulatus OR sanguineus OR marginatum OR erraticus OR lusitanicum

Sand flies: alexandri OR ariasi OR kandelakii OR major OR mascitii OR neglectus OR papatasi OR perniciosus OR perfiliewi OR tobbi OR sergenti OR similis OR langeroni

# SUPPLEMENTRAY INFORMATION SECTION III: Supplementary Table S3: Data Entry Worksheet and Column Descriptions

Key and Notes:

**Bold =Worksheet Title**; *Italic = Calculated Field*.

If Field Value selected with dropdown lists, column Validation Lookup sheet indicates "Lists". List definitions in Worksheet Lists

Worksheet	Column Name	Column Description	Validation Lookup sheet	Validation lookup sheet column	Permitted range
Data Entry		Main Data entry sheet			
DataEntry	SourceID	Unique ID, assigned on addition to dataset	None		
DataEntry	VectorCategory	Vector Group: Ticks, Culicoides midges, Sand flies, Invasive Mosquitoes, Native Mosquitoes.	Lists	В	In Lookup
DataEntry	VectorSpeciesName	Vector Name	Lists	B and D,F,H,J	In Lookup
DataEntry	Country	Country Code, validated against list in worksheet LocationCodes/Country	LocationCodesCountry		In Lookup
DataEntry	LocationCode	Administrative Unit Code, validated against list in worksheet LocationCodes/Counry	LocationCodesCountry		In Lookup
DataEntry	LocationName	Administrative Unit Name, calculated from previous column using worksheet CodesVLOOKUP	Codes VLOOKUP		In Lookup
DataEntry	Longitude E/W	Longitude East or West (E.W)	Lists	AT	In Lookup
DataEntry	Longitude (degrees)	Longitude degrees	None		
DataEntry	Longitude (minutes)	Longitude minutes	None		
DataEntry	Longitude (seconds)	Longitude seconds	None		
DataEntry	Longitude (decimal degrees) (X)	Calculated Latitude in decimal degrees calculated from previous columns	None		
DataEntry	Latitude N/S	Latitude East or West (E.W)	Lists	AV	In Lookup
DataEntry	Latitude (degrees)	Latitude degrees	None		
DataEntry	Latitude (minutes)	Latitude minutes	None		
DataEntry	Latitude (seconds)	Latitude seconds	None		
DataEntry	Latitude (decimal degrees) (Y)	Calculated Latitude in decimal degrees calculated from previous columns	None		
DataEntry	User Entry Longitude (X)(decimal degrees)	User entry Longitude in decimal degrees	None		

Worksheet	Column Name	Column Description	Validation Lookup sheet	Validation lookup sheet column	Permitted range
DataEntry	User Entry Latitude (Y) (decimal degrees)	User entry Latitude in decimal degrees	None		
DataEntry	PrecisionLocation	Point type or polygon	Lists	L	In Lookup
DataEntry	VectorLifeStage	Vector life stages: adult, nymph, pupa, eggs, Unknown	Lists	AL	In Lookup
DataEntry	VectorSex	Vector sex	Lists	АН	In Lookup
DataEntry	NumberOfVectorsCaug ht	Number of vectors caught:	None		
DataEntry	CollectionPlaceID	Collection place	None		
DataEntry	CollectionEffortStartDa te	Start date for collection effort	None		
DataEntry	CollectionEffortEndDat e	End date for collection effort	None		
DataEntry	VectorCollectionMeth od	Collection method used (permitted methods customised for vector type)	Lists	B: AL: N, P,R, T, V, X	In Lookup
DataEntry	CollectionEffortP1Valu e	Collection effort as number of units in following column	None		
DataEntry	UnitsEffortP1	Calculated from preceding columns using worksheet VLookupTableUnitsVectorCollect	VLookupTableUnitsVectorC ollect		In Lookup
DataEntry	CollectionEffortP2Valu e	Collection effort as number of units in following column	None		
DataEntry	UnitsEffortP2	Calculated from preceding columns using worksheet VLookupTableUnitsVectorCollect	VLookupTableUnitsVectorC ollect		In Lookup
DataEntry	VectorHostSpecies	Vector Host Species	Lists	AP	In Lookup
DataEntry	Host Bodypart	Host Bodypart not validated	None		
DataEntry	VectorIdentificationM ethod	Method used to identify vector species	Lists	АВ	In Lookup
DataEntry	ShelteredEnvironment	Whether sample site was sheltered or not	Lists	AR	In Lookup
DataEntry	ReportedDistributionSt atus	Reported distribution status: Present, Absent, Anticipated Absent, Introduced, Unknown, No Data	Lists	В, Z	In Lookup
DataEntry	PathogenName	Name of Pathogen for which test carried out	Lists	AD	In Lookup
DataEntry	PathogenDetectionMe thod	Method used to detect pathogen	Lists	AF	In Lookup
DataEntry	NumberOfVectorTeste d	Number of vectors tested	None		

Worksheet	Column Name	Column Description	Validation Lookup sheet	Validation lookup sheet column	Permitted range
DataEntry	PositivePathogenDetec tion	Whether pathogen detected or not	None		
DataEntry	PathogenComment	Additional information about pathogen	None		
DataEntry	SourceType	Type of information source	None		
DataEntry	PublicationTitle	Publication Title	None		
DataEntry	Author	Publication Author	None		
DataEntry	YearOfPublication	Publication Year	None		
DataEntry	DOI	Publication DOI	None		
DataEntry	VectorNetFieldStudyID	Vectornet field study ID if applicable	None		
DataEntry	URL	Reference URL	None		
DataEntry	NotesFromDataExpert	Option Notes from Validator	None		
DataEntry	Data expert name	Data expert: name and family name	None		
DataEntry	CollectionEffortStartDa te_ddmmyyyy	Calculated standardised date for start of Collection effort from CollectionEffortStartDate (x)	None		
DataEntry	CollectionEffortEndDat e_ddmmyyyy	Calculated standardised date for start of Collection effort from CollectionEffortEndDate (x)	None		
DataEntry	Submitter email (first row only)	email of data submitter	None		
Name2Geocode		Worksheet with Tool to identify Location code from Name			
Name2Geocode	LOCATION NAME	User entered Location Name			
Name2Geocode	LOCATION CODE	Location code returned by Tool			
Name2Geocode	LOCATION TYPE	Location Administrative Unit type returned by Tool			
Lists		Look Up tables as used in Column: Validation lookup sheet column			
Lists	VectorCategoryCode	Look Up for Vector category Code			
Lists	Mosquito	Look Up for mosquito species names			
Lists	Sandfly	Look Up for sandfly species names			
Lists	Culicoides	Look Up for Culicoides species names			

Worksheet	Column Name	Column Description	Validation Lookup sheet	Validation lookup sheet column	Permitted range
Lists	Tick	Look Up for Tick species names			
Lists	PrecisionCoordinate	Look Up for precision of coordinates entered			
Lists	TrapTypeName	Look Up for trap type			
Lists	VectorCollectionMeth odName	Look Up for collection method			
Lists	CollectionMosquitoAd ults	Look Up for collection method of Adult Mosquitoes			
Lists	CollectionMosquitoLar vae	Look Up for collection method of Larval Mosquitoes			
Lists	CollectionMosquitoNy mph_Pupa	Look Up for collection method of Nymphal or Pupal Mosquitoes			
Lists	CollectionMosquitoEgg s	Look Up for collection method of Mosquito Eggs			
Lists	Vector Distribution Stat us Invasive	Look up for Vector Status Invasive or Native			
Lists	VectorIdentificationM ethod	Look up for Vector Identification Method			
Lists	PathogenNameMosqui to	Look up for Pathogens transmitted by Mosquitoes			
Lists	PathogenDetectionMe thod	Look up for Pathogen Detection Methods			
Lists	VectorSex	Look Up Vector Sex			
Lists	VectorLifeStage	Look Up for Vector Life Stage			
Lists	VectorInformationSour ceType	Look Up Type of Vector Information Source			
Lists	VectorHostSpeciesNa me	Look Up Vector Host Species Name			
Lists	ShelteredEnvironment				
Lists	Longitude E/W	Look Up for Longitude Abbreviations			
Lists	Latitude N/S	Look Up for Latitude Abbreviations			
LocationCodesby Country			Look Up for Location Codes pairs, first as Admin Unit Co Names by Country	and Names: Columns in de, second as Admin Unit	
CodesVLOOKUP		Look Up for Location codes and Names, formatted for VLOOKUP function			

Worksheet	Column Name	Column Description	Validation Lookup sheet	Validation lookup sheet column	Permitted range
CodesVLOOKUP	LocationCode	Admin Unit Code			
CodesVLOOKUP	LocationName	Admin Unit Name			
Geocodes					
VLookupTableUn itsVectorCollect		Lookup for Sample Effort Units Specification			
VLookupTableUn itsVectorCollect	LOOKUPTABLEUNITS				
VLookupTableUn itsVectorCollect	CollectionTickAdultsNy mphsLarvae				
VLookupTableUn itsVectorCollect	CONCATENATED				
VLookup Table Un its Vector Collect	Unit1				
VLookup Table Un its Vector Collect	Unit2				
VLookupTableUn itsVectorCollect	Unit3				

## SUPPLEMENTRAY INFORMATION SECTION IV: Supplementary Table S4: Quality Indicators

Processes that ensure the quality of the database work on different levels: a) A standardised search string is created to ensure the accuracy and consistency in the literature review; b) A data entry template is created for complete, accurate and consistent data entry, and c) Entry restrictions are put in place within the data entry template to ensure valid and accurate data entry

Random data quality checks on 1% of the submitted data are performed after data uploads, and the quality criteria completeness, unique-ness, validity, accuracy and consistence are checked based on quality indicators for these qualities. The Table lists the fields to be included in the QI assessment: which are evaluated together and which not. The data entry tool comes with a manual to ensure ease of use and correct use. The quality indicators are calculated after the new data entry system is implemented, tested and validated, and the backlog of data has been entered into the new system. The quality indicators are calculated database and apart from uniqueness, all indicators are only calculated for the new data.

## Completeness

Completeness is expressed by the number of fields that are filled in. An analysis per column are made over the database to evaluate the extracted data and the data entry template. Which fields are checked can be found in the Table below. The QI for uniqueness is the proportion of complete records, as compared to the data filled from the random quality check.

## Uniqueness

Uniqueness is assessed for the following: species, coordinates, VectorLifeStage, VectorSex, CollectionPlaceID, CollectionEffortStartDate, VectorCollectionMethod, VectorHostSpecies, Host Bodypart and source as a measure for the uniqueness. A duplicate constitutes a record where all these fields are the same. If the source is different (a different document) but the rest is the same, this is not counted as a duplicate. Duplicates are marked in the database for easy filtering but are not automatically removed. The QI for uniqueness is the proportion of duplicate records.

## Validity and Correctness

There are specific restrictions built into the data entry template (e.g. location data entered with a dot and not a comma etc.). The QI for validity is the proportion of valid records.

## Correctness

The correctness of the data is different than validity. An entered cell can be valid according to the restrictions in the data entry template but can be not correct, e.g. the species name is correctly spelled but is the wrong species. This is checked during the data quality check, and the QI for correctness is the proportion of correct records.

## Accuracy and Consistency

Accuracy and consistency of new uploads is ensured by using defined upload templates. Within the template excel file there are several limitations and lists baked in to ensure the consistency of the data upload. The QI for accuracy and consistency is the proportion of records that comply with the template lists (out of those that have pre-defined lists).

Field Name	Comment
SourceID	Not considered during the QI evaluation.
VectorCategory	Evaluated under the QI assessment.
VectorSpeciesName	Evaluated under the QI assessment.
Country	Evaluated under the QI assessment.
LocationCode, Location Name	These are linked together and assessed as one.
Longitude E/W, Longitude (degrees),	
Longitude (minutes), Longitude (seconds),	
Longitude (decimal degrees) (X), Latitude	
N/S, Latitude (degrees)2, Latitude	Fuel used to eath on an thou and dueliantee
(minutes), Latitude (seconds), Latitude	Evaluated together as they are duplicates.
(decimal degrees) (Y), User Entry Longitude	
(X) (decimal degrees), User Entry Latitude	
(Y) (decimal degrees)	
PrecisionLocation	Evaluated under the QI assessment.
VectorLifeStage	Evaluated under the QI assessment.
VectorSex	Evaluated under the QI assessment.
NumberOfVectorsCaught	Evaluated under the QI assessment.
CollectionPlaceID	Evaluated under the QI assessment.
CollectionEffortStartDate	These are linked together and are assessed as one.
CollectionEffortEndDate	
VectorCollectionMethod	Evaluated under the QI assessment.
CollectionEffortP1Value, UnitsEffortP1	These are linked together and are assessed as one.
	These are linked together and are assessed as one. These are optional
CollectionEffortP2Value, UnitsEffortP2	and do not count towards completeness.
VectorHostSpecies, Host Bodypart	These are linked together and are assessed as one.
VectorIdentificationMethod	Evaluated under the QI assessment.
ShelteredEnvironment	Evaluated under the QI assessment.
Reported Distribution Status	Evaluated under the QI assessment.
PathogenName	Evaluated under the QI assessment.
PathogenDetectionMethod	Evaluated under the QI assessment.
NumberOfVectorTested	Evaluated under the QI assessment.
PositivePathogenDetection	Evaluated under the QI assessment.
PathogenComment	Not included in the QI assessment
PublicationTitle. Author, Year of Publication,	
Author, Source Type	These are linked together and are assessed as one.
DOI	Evaluated under the QI assessment.
VectorNetFieldStudyID	Not included in the QI assessment
URL	Not included in the QI assessment
NotesFromDataExpert	Not included in the QI assessment
Data expert name and surname for each	Not included in the OL concernment
row	Not included in the QL assessment
Submitter email (first row only)	Not included in the QI assessment

Table: Overview of the Quality Indicators different fields in the data entry template.

## SUPPLEMENTRAY INFORMATION SECTION V: Supplementary Figures S1 – S45: Maps

Surveillance	Distribution: Invasive Mosquitoes
Supplementary Figure S1: Invasive Mosquitoes	Supplementary Figure S7: Aedes aegypti
Supplementary Figure S2: Native Mosquitoes	Supplementary Figure S8: Ae. albopictus
Supplementary Figure S3: Biting Midges	Supplementary Figure S9: Ae. atropalpus
Supplementary Figure S4: Sand flies	Supplementary Figure S10: Ae. japonicus
Supplementary Figure S5: Ticks	Supplementary Figure S11: Ae. koreicus
Supplementary Figure S6 a, b: Comparison of surveillance	
and distributions status	
Distribution: Native Mosquitoes	Distribution: Biting Midges
Supplementary Figure S12: Aedes caspius	Supplementary Figure S21: Culicoides chiopterus
Supplementary Figure S13: Ae. detritus/coluzzii	Supplementary Figure S22: C. dewulfi
Supplementary Figure S14: Ae. vexans	Supplementary Figure S23: C. imicola
Supplementary Figure S15: Anopheles maculipennis s.l.	Supplementary Figure S24: C. kingi
Supplementary Figure S16: An. plumbeus	Supplementary Figure S25: C. newsteadi
Supplementary Figure S17: An. superpictus	Supplementary Figure S26: C. pulicaris /lupicaris
Supplementary Figure S18: Coquillettidia richiardii	Supplementary Figure S27: C. punctatus.
Supplementary Figure S19: Culex modestus	Supplementary Figure S28: C. obsoletus/scoticus
Supplementary Figure S20: Cx. pipiens/torrentium	
Distribution: Sand flies	Distribution: Ticks
Supplementary Figure S29: Phlebotomus alexandri	Supplementary Figure S39: Dermacentor reticulatus
Supplementary Figure S30: P. ariasi	Supplementary Figure S40: Hyalomma lusitanicum
Supplementary Figure S31: P. mascittii	Supplementary Figure S41: H. marginatum
Supplementary Figure S32: P. neglectus	Supplementary Figure S42: Ixodes persulcatus
Supplementary Figure S33: P. papatasi	Supplementary Figure S43: I. ricinus
Supplementary Figure S34: P. perfiliewi	Supplementary Figure S44: Ornithodoros erraticus
Supplementary Figure S35: P. perniciosus	Supplementary Figure S45: Rhipicephalus
Supplementary Figure S36: P. sergenti	sanguineus
Supplementary Figure S37: P. similis	
Supplementary Figure S38: P. tobbi	

## **Vector Group Surveillance Activity Maps**

**Figures S 1 to 5** present the summaries of reported surveillance activities from 2015 to 2019 for each vector group along with commentary descriptions. Two maps are presented for each group: a) Seasonal active surveillance = the number of months per year for which active surveillance (so not including passive surveillance through Citizen Science reporting) is reported and b) surveillance effort. For each admin unit, surveillance was coded by type with the highest code reflecting the highest effort as shown on the map legend. The highest surveillance type that occurred in an administrative area during 2015–2019 is depicted, with lower type categories contributing to the effort score with a lower weight as presented in the table at the end of the commentary text. The highest type-code that occurred during the period 2015–2019 decides the colour (hue) of the admin unit. The frequency with which it occurred decides the saturation of the colour: a deeper colour indicates more frequent surveillance. If lower surveillance types also occurred in the same unit, these also contribute to the saturation, but with a lower weight than the highest code which decides the hue.

Each map also shows the weighting table in the right-hand column, interpreted as follows:

		Highest level				
Level	1	2	3	4	5	
1	1	0	0	0	0	
2		1	1	0.5	0.25	
3			1	0.5	0.25	
4				1	0.5	
5					1	

All Level one (citizen science) scores are unweighted or discounted if reported to include active surveillance. All other level two and three surveillance scores are unweighted. If the most intensive surveillance in a polygon is level 4 or 5, scores for the less intensively sampled periods are progressively reduced as sampling level falls.

**Figures S 6, a and b** show a comparison between surveillance activities and distribution status to assess where the two match (i.e. there is surveillance and data, or where there is no surveillance and no data). A mismatch implies that either there are no data available or that the data have not been located.

**Tables S 5 to 43**: Each table presents a brief description of the species, and the VectorNet map for March 2021, with commentary on the distributions shown. Each table also provides the means to compare the VectorNet (VN) map with the GBIF or VectorBase (VBASE) records using a simplified map showing the presence and absence, each as a single category, with the point records from GBIF or VectorBase overlain. A summary table of the number of presence and absence polygons with and without GBIF or VectorBase records is also provided. An example interpretation is shown as follows:

GBIF or VectorBase Present	VectorNet Present	VectorNet Absent	VectorNet No Data	Total
No	22	807	666	1495
Yes	2	9	0	11
Total	24	816	666	1506

There are 11 polygons with GBIF or VectorBase records. There are no GBIF or VectorBase records in 22 polygons where VectorNet records the species as present, 807 polygons where VectorNet records the species as absent and 666 polygons where VectorNet has no data. There are GBIF or VectorBase records in 2 polygons where VectorNet records the species and present, 9 polygons where VectorNet records the species as absent, and 0 records where VectorNet has no data.

**Figures S 7 to 45** show the VectorNet March 2021 distribution mapd of the species / species group and a comparison with records from an alternative database (GBIF or Vectorbase, depending on which was the most populated for the species / species group).



Surveillance for invasive mosquitoes is concentrated in Western Europe, the Balkans, the Caucasus and the western part of North Africa, and the reported surveillance effort is generally high.

There is a need for strengthening the low intensity of surveillance efforts in countries where *Ae. albopictus* and other invasive species might be present but undetected in many administrative units. Assessing abundance and seasonality should be prioritised to help prevent/control possible disease outbreaks in areas were *Ae. albopictus* is well established and that are receptive to pathogen introduction, such as southern France and parts of Greece.

Invasive mosquito surveillance by citizen science is predominant in northern and central Europe whilst various forms of active surveillance tend to be most common in southwestern Europe and the Balkans. The main surveillance activities are for *Ae. albopictus* for its presence (introduction and spread), and less frequently for its abundance as part of disease control. Given the current known distributions, there are obvious gaps in the reported surveillance data in Italy, Turkey, and Crimea.

#### Effort Weighting

	Highest level				
Level	1	2	3	4	5
1	1	0	0	0	0
2		1	1	0.5	0.25
3			1	0.5	0.25
4				1	0.5
5					1

2 = active surveillance for species presence in sheltered environment only (points of entry like greenhouses or airport building)

3 = active surveillance for species presence through sampling/trapping (with or without passive surveillance)

4 = active surveillance for species presence, abundance and seasonality through sampling/trapping (with or without passive surveillance)

5 = active surveillance for species presence and pathogen infection through sampling/trapping (with or without passive surveillance)



The surveillance coverage appears to be quite patchy in some countries, possibly because mosquito control activities and their associated surveillance are often implemented over relatively small areas. The current datasets suggest that there are surveillance data gaps in some countries and that there is a need for strengthening the intensity of surveillance efforts in areas were mosquito-borne pathogens are known to circulate or be at risk of circulating.

Native mosquito surveillance techniques are very diverse and include: (1) presence data only, mainly based on citizen science projects; and (2) abundance and seasonality data acquired in areas where pest species occur and are controlled (e.g. parts of France) or more widely for scientific purposes. These abundance and seasonality data are associated with pathogen screening, both in areas where disease outbreaks are more frequent (West Nile infections in particular) and in currently disease-free countries.

#### Effort Weighting

	Highest level					
Level	1 2 3 4					
1	1	0	0	0		
2		1	0.5	0.25		
3			1	0.5		
4				1		

#### Surveillance Effort Categories

1 = passive surveillance for species presence through citizen science

2 = active surveillance for species presence through sampling/trapping (with or without passive surveillance)

3 = active surveillance for species presence, abundance and seasonality through sampling/trapping (with or without passive surveillance)

4 = active surveillance for species presence and pathogen infection through sampling/trapping (with or without passive surveillance)

The highest surveillance type that occurred in an administrative area during 2015–2019 is depicted, with lower type categories contributing to the effort score with a lower weight. Weights were as follows:



The most sustained annual surveillance is reported in Spain, Italy and the Balkans.

Midge surveillance, more than for other vector groups, has varied considerably during the past 15 years: most *Culicoides* surveillance activities in the VectorNet geographical area were initiated in response to bluetongue epizootics, and in parts of Spain, due to the African horse sickness outbreaks in 1987–1990.

Midge surveillance methodology varies considerably according to country. The most widespread method is full species identification (level 5), though this is not the case in Western Europe. Data for midge surveillance activities have been acquired for fewer countries than is the case for the other vector categories, but where data are available, surveillance occurs over a wide range, and is comparatively intensive. Data are still to be acquired for some countries, where information

#### Effort Weighting

		Highest level				
Level	1	2	3	4	5	
1	1	0.5	0.33	0.25	0.2	
2		1	0.66	0.5	0.4	
3			1	0.75	0.6	
4				1	0.8	
5					1	

#### Surveillance Effort Categories

1 = surveillance, with midges counted only to assess the activity threshold (*e.g.* more or less than 5 parous females), without species identification.

2 = surveillance implemented, with midges counted and identified into two taxa: C. imicola and 'other species'

3 = surveillance, with midges counted and identified into four taxa: C. imicola, other species of the Avaritia subgenus (= 'the Obsoletus group'), species of the Culicoides subgenus (= 'the Pulicaris group') and 'other species'

4 = surveillance, with midges counted and identified to species for the Culicoides subgenus: C. imicola, other species of the Avaritia subgenus (= 'the Obsoletus group'), at least C. newsteadi, C. pulicaris/C. lupicaris and C. punctatus and 'other species'

5 = surveillance, with midges counted and identified to species for at least the Avaritia and the Culicoides subgenera: results including at least C. imicola, C. obsoletus/C. scoticus, C. dewulfi, and C. chiopteru,s, as well as C. newsteadi, C. pulicaris/C. *lupicaris* and *C. punctatus*.



3 = surveillance for adult sand flies with sticky traps and light traps, identified at species level (with or without counting the numbers)

The highest surveillance type that occurred in an administrative area during 2015–2019 is depicted, with lower type categories contributing to the effort score with a lower weight. Weights were as follows:



Tick surveillance methodology is variable, ranging from citizen science methods in the north and west of Europe, to the more intensive vegetation and animal monitoring in the eastern countries and in Portugal. Though data from some major countries is missing, it is clear that tick surveillance is very widespread but that the surveillance effort employed tends towards the lower end of the recorded spectrum.

The most activity for surveillance across Europe relates to the main risks posed by Ixodes ricinus and Lyme/tick borne encephalitis or Hyalomma marginatum and CCHF.

There are data gaps for some of the larger countries of central, northern and western Europe, where the available distribution data suggests that surveillance does take place. Surveillance activity in the Balkans is good for some countries, whilst in others there are no data or only local/regional surveillance.

Effort Weighting							
	Highest level						
Level	1	2	3	4			
1	1	0	0	0			
2		1	1	0.5			
3			1	05			

1

1 = passive surveillance through citizen science

- 2 = active surveillance through animal surveys (with/without passive surveillance)
- 3= active surveillance through vegetation surveys (with/without passive surveillance)
- 4= active surveillance through vegetation AND animal surveys (with/without passive surveillance)



Supplementary Figure S6a, Comparison of surveillance and distributions status for vector groups

Supplementary Figure S6b, Comparison of surveillance and distributions status for vector group representative species



Comparison status

Μ	at	chi	ng

data 'complete', no reported surveillance

data incomplete, surveillance reported

### Supplementary Figure S7 Aedes aegypti

The yellow fever mosquito *Ae. aegypti* originates from Africa but was historically present over decades in all countries surrounding the Mediterranean Basin, where it established following introductions via ships, but from where it disappeared in the mid-twenty Century (cf Schaffner & Mathis, 2014 doi: 10.1016/S1473-3099(14)70834-5.). *Aedes aegypti* is the major worldwide vector species for dengue, chikungunya, Zika and yellow fever viruses (cf Schaffner et al., 2013 <u>https://doi</u>.org/10.1111/1469-0691.12189). The species was associated with the large outbreak of dengue fever that occurred in 2012 in the Portuguese Autonomous Region of Madeira (cf Sousa et al 2012 Euro Surveill.





This invasive species has a pan global distribution. Within the VectorNet region, it has become established along the Black sea coast and on Madeira. Its establishment is not yet confirmed in Egypt, but is highly probable. Introductions to other places occur from time to time but control actions and/or climatic conditions do not lead to establishment of a viable mosquito population; such reports disappear from the maps if older than 5 years. Exceptional introduction events (e.g. single specimens or indoor records) are not reported by VectorNet (i.e. France, Germany, UK). The high number of negative areas stem from the widespread surveillance of Ae. albopictus.

Occurrences reported by GBIF in northern France, Germany, Israel, Turkey, UK) are not substantiated. Large VectorNet yellow or red areas (Egypt and Russia) are due to the large size of the mapped admin units, while they refer to much more localised populations.

#### Supplementary Figure S8: Aedes albopictus

The tiger mosquito *Ae. albopictus* is native to south-east Asia and is invading the European continent since the 1990s. It was first introduced into Albania (late 1970s) and Italy (1990s), from where it has progressively spread to neighbouring countries (cf Medlock et al 2015 doi:10.1017/S0007485315000103). This major invasive species is a nuisance species and a significant vector of dengue, chikungunya and Zika viruses, and of *Dirofilaria* parasites (cf Schaffner et al., 2013,



#### Supplementary Figure S9: Aedes atropalpus

The American rock pool mosquito *Ae. atropalpus* is native to Eastern North America. Although originally limited to rock pool areas, its utilisation of used tyres as an alternative larval development site resulted in its spread in North America and introductions into the European continent (cf Medlock et al 2015 doi:10.1017/S0007485315000103). This species is not considered to be an important vector of pathogens in the field, despite isolation of West Nile virus in field populations. Laboratory experiments demonstrate some competence to transmit several encephalitis viruses (cf Scholte et al., 2012, European Mosquito Bulletin 30 (2012), 1-14).



#### Supplementary Figure S10: Aedes japonicus

The Asian rock pool mosquito or Asian bush mosquito *Ae. japonicus* originates from East Asia where four subspecies occur, but only one of these, *Ae. japonicus japonicus*, is invasive in Europe. It is nowadays widely present in Northern America and has been successfully spreading throughout the European continent since the early 2000s (cf Medlock et al 2015 doi:10.1017/S0007485315000103). The species is not considered to be an important disease vector in its native Asian range. It may also be involved in West Nile virus transmission, and shows some level of vector competence for other viruses such as dengue and chikungunya (cf Schaffner et al., 2013, https://doi.org/10.1111/1469-0691.12189).



## Supplementary Figure S11: Aedes koreicus

The Korean mosquito *Ae. koreicus* is native from South-East Asia and was encountered for the first time in Europe in 2008, and later in Italy. The species has been suggested to be a possible vector for Japanese encephalitis virus in parts of Russia; it may contribute to the transmission of *Dirofilaria* parasites (cf Medlock et al 2015 doi:10.1017/S0007485315000103)



#### Supplementary Figure S12: Aedes caspius

The native species *Aedes caspius* is the primary European salt marsh mosquito that develops in coastal or inland brackish marshes, and, in southern Europe, in rice fields and flood plains (cf Becker et al., 2010). DOI: 10.1007/978-3-540-92874-4. The species is responsible for intense nuisance in the areas where it proliferates, and therefore is frequently targeted by control programmes (e.g. France, Greece, Italy, Spain). *Aedes caspius* may contribute to the transmission of Rift Valley fever, West Nile and Tahyna viruses; it may also play a role in the spread of tularaemia and myxomatosis (cf Becker et al., 2010, DOI: 10.1007/978-3-540-92874-4)



#### Supplementary Figure S13: Aedes detritus/coluzzii

*Aedes detritus/coluzzii* is a complex of two sibling species which have been described based on molecular studies; no morphological diagnostic features are described. To date, only few studies have investigated the populations with molecular tools. Therefore, they are mapped together. *Aedes detritus/coluzzii* is the second European salt marsh mosquito that develops in coastal or inland brackish marshes (cf Becker et al., 2010,. DOI: 10.1007/978-3-540-92874-4)). The species is responsible of intense nuisance in the areas where it proliferates, and is therefore frequently targeted by control programmes (e.g. France, Italy, Spain) The species It is a potential vector for RVF (cf Moutailler S, et al, http://doi.org/10.1089/vbz.2008.0009.



#### Supplementary Figure S14: Aedes vexans s.l,

Aedes vexans is mapped as sensu lato reflecting the presence of cryptic species in Scandinavia, and lack of documented molecular differentiation of *Ae. vexans vexans* and *Ae. vexans arabiensis* in North Africa and the Middle East. It is the primary European floodwater mosquito, which can generate huge nuisance along flood plains. It develops in temporary freshwater bodies such as flooded meadows, poplar cultures, willow and reed areas (Becker et al., 2010). This pest species is targeted by control programmes in many areas (e.g. Danube, Rhine, Rhone floodplains) (Schaffner et al., 2001). *Aedes vexans* has many attributes of an ideal vector species; it was found naturally infected by several encephalitis viruses and is involved in the transmission of Rift Valley fever and Tahyna viruses (cf Becker et al., 2010,. DOI: 10.1007/978-3-540-92874-



### Supplementary Figure S15: Anopheles maculipennis s.l.

Anopheles maculipennis s.l. is a species complex of up to eight species which are difficult to distinguish by morphology. The all develop mainly in semi-permanent or permanent water bodies, as long as the aquatic vegetation allows the larvae to hide from predators. This complex includes several species confirmed as historical (when malaria was endemic to the European subcontinent) significant vectors of malaria (i.e. *An. atroparvus, An. labranchiae, An. messeae, An. sacharovi*), their role being different according to the geographical region considered. Some members of the complex can also be involved in the transmission of Batai, Tahyna and West Nile viruses, canine filariae, tularaemia bacteria and Myxoma virus (cf Becker et al., 2010,. DOI: 10.1007/978-3-540-92874-4)).



## Supplementary Figure S16: Anopheles plumbeus

Anopheles plumbeus is a container-breeding species, developing mainly in rainwater-filled tree holes. It occasionally proliferates in unused cesspits (filed by rain water and animal dung residue) generating severe but localised nuisance. The species does not (yet) play an important vector role in Europe as it is highly competent for *Plasmodium falciparum*, and this malaria pathogen is rare in Europe (cf Becker et al., 2010,. DOI: 10.1007/978-3-540-92874-4).



## Supplementary Figure S17: Anopheles superpictus

Anopheles superpictus breeds in slow running or standing water in river beds or in irrigated rice fields. The species is reported as being an important vector of malaria in Middle East and a secondary vector in other regions where it occurs; it may also play a role in the transmission of *Dirofilaria immitis* (cf Becker et al., 2010,. DOI: 10.1007/978-3-540-92874-4).



#### Supplementary Figure S18: Coquillettidia richiardii

*Coquillettidia richiardii* breeds in permanent water bodies with erected aquatic vegetation such as reeds and bulrush (*Typha* sp.). Larvae and pupae live submerged and obtain oxygen from the aerenchym of aquatic plants and move very little. Adults can be very numerous, generating a severe nuisance to humans and domestic animals within short distance from their breeding site. They may be involved in the transmission of several pathogens, including Batai, Tahyna and West Nile viruses (cf Becker et al., 2010,. DOI: 10.1007/978-3-540-92874-4).



## Supplementary Figure S19: Culex modestus

*Culex modestus* is a wetland mosquito that breeds mainly in rice fields, irrigation canals and semi-permanent marshes (Becker et al., 2010). this species sometimes generates significant nuisance for human and other mammals in the immediate neighbourhood of breeding places. *Culex modestus* is considered to be an important vector of West Nile virus in some places (e.g. Camargue, southern France) and is involved in the transmission of Tahyna and Myxoma viruses. Moreover, wild females have been found infected by tularaemia bacteria, Lednice and Sindbis viruses, and *Dirofilaria immitis*.



#### Supplementary Figure S20: Culex pipiens: Group

The *Culex pipiens* group comprises the Pipiens species complex and the sibling species *Cx. torrentium* and has a global distribution. In Europe, the Near East and North Africa, the Pipiens complex is represented by *Cx. pipiens*, form *pipiens*, which develops in the wild in clear water, bites mainly birds and diapauses in winter as an adult; and the form *molestus*, which develops in more polluted underground water throughout the year and bites mainly mammals. Females can be a major nuisance indoors and at night, in particular where underground breeding places exist. *Culex torrentium* is a northern species, common in high altitudes and rare or absent in the Mediterranean Basin. Larvae of both species develop in any man-made container as well as in a wide range of natural standing water bodies (e.g. ponds, ditches, marshes, rock pools, tree holes). *Culex pipiens* is considered an important vector of West Nile virus and avian plasmodia, and both *Cx. pipiens* and *Cx. torrentium* contribute to the transmission of Sindbis virus (cf Becker et al., 2010,. DOI: 10.1007/978-3-540-92874-4)



## Supplementary Figure S21: Culicoides chiopterus

*Culicoides chiopterus* is a species of the subgenus *Avaritia*, with a Holarctic distribution. This species has been identified in the Palearctic zone from Europe to Japan. Immatures of *C. chiopterus* develop exclusively in cattle and horse dung. Adults have been recorded feeding on a variety of mammals including ruminants, equids, suids, rabbits and humans. *Culicoides chiopterus* has been implicated in the transmission of veterinary pathogens including bluetongue and Schmallenberg viruses



## Supplementary Figure S22: Culicoides dewulfi

*Culicoides dewulfi* is a species belonging to the subgenus *Avaritia*, with a Palearctic distribution, including northern Russia. This species has a biology and an ecology very similar to *C. chiopterus*. Indeed immatures develop exclusively in cattle and horse dung. Moreover, this species has been recorded feed on a variety of mammals, including ruminants, equids or also humans. *Culicoides dewulfi* has been implicated in the transmission of bluetongue and Schmallenberg viruses in Europe.



#### Supplementary Figure S23: Culicoides imicola

*Culicoides imicola* is a species of the *Avaritia* subgenus. This Afrotropical species is widespread in Africa, in the Mediterranean basin and in the Middle East, and is occasionally recorded in the Far East (*e.g.* India). Immatures develop in a wide range of semi-aquatic habitats that are usually associated with livestock productions, in areas next to pond shorelines and irrigation channels. This adult feeds opportunistically on a wide variety of mammals including ruminants, equids, humans (rarely) and exotic host species in zoos. *Culicoides imicola* is a well-known vector of economically important livestock viruses such as bluetongue virus affecting domestic and wild ruminants or African horse sickness (AHS) virus affecting equids. The ability of culicoides to transmit these orbiviruses was first demonstrated in this species in the 1940s.



### Supplementary Figure S24: Culicoides kingi

*Culicoides kingi* belongs to the Schultzei group, which is affiliated to the Remmia or the Wirthomyia subgenera depending on author. This species has an Afrotropical distribution, with records in Africa, in the Sahel and the Maghreb. Immatures have been observed to emerge from the edge of lakes or ponds in Senegal, whereas adults attack livestock such as horses or ruminants. The virus of the epizootic hemorrhagic disease has been isolated from *C. kingi* in Sudan and Nigeria (cf Mellor, p. *et al.* doi: 10.1017/s0022172400065190.) This species is also associated with the transmission of *Onchocerca gutturosa* in cattle.



## Supplementary Figure S25: Culicoides newsteadi

*Culicoides newsteadi s.l.* belongs to the *Culicoides* subgenus. Cryptic diversity has been suggested within this taxon by molecular investigations (cf This <u>https://doi.org/10.1111/j.1365-2915.2012.01050.x</u>) species has a Palearctic distribution. Immatures are associated with brackish marshes, and adults have been recorded feed on the different domestic ruminants (cattle, goats, sheep), but also on humans, and sporadically on domestic birds. *Culicoides newsteadi s.l.* is considered a possible bluetongue and Schmallenberg viruses vector, due to its ecological habits and to virus isolation/viral genome detections from field-collected individuals in Mediterranean regions.



## Supplementary Figure S26: Culicoides pulicaris s.l./Culicoides lupicaris

The taxon *Culicoides pulicaris s.l./Culicoides lupicaris* includes species of the subgenus *Culicoides*, namely *C. pulicaris s.l.* that may include cryptic diversity (cf <u>https://doi.org/10.1111/j.1365-2915.2012.01050.x</u>) and *C. lupicaris*. Some authors consider *C. lupicaris* as a synonym of *Culicoides delta* although evidences suggest two distinct valid species (cf

https://www.inhs.illinois.edu/files/5014/6532/8290/CulicoidesSubgenera.pdf). These species are grouped because diagnostic wing patterns cannot be distinguished. These species have a Palearctic distribution. Immatures have been found at the edges of ponds, whereas adults have been recorded feeding on ruminants (cattle and deer) and on horses. *Culicoides pulicaris s.l.* is considered a bluetongue and Schmallenberg viruses vector.



## Supplementary Figure S27: Culicoides punctatus

*Culicoides punctatus s.l.* belongs to the subgenus *Culicoides*. New species have been proposed after morphological detailed and genetic investigations of individuals firstly identified as *C. punctatus* (cf

https://onlinelibrary.wiley.com/doi/full/10.1111/mve.12228) . This species has a Palearctic distribution. Immatures have been found at the edges of ponds, whereas adults have been recorded feed on a variety of mammals, including cattle, goats, deer and zoo large mammals, equids, suids or humans. *Culicoides punctatus s.l.* is considered a possible bluetongue and Schmallenberg viruses vector, due to its ecological habits and to virus isolation/viral genome detections from field-collected individuals.



#### Supplementary Figure S28: Culicoides obsoletus s.l./Culicoides scoticus

The taxon *Culicoides obsoletus s.l./Culicoides scoticus* (cf doi: 10.1186/s13071-020-04114-1) includes species of the subgenus *Avaritia*, namely *C. scoticus* (that may include cryptic diversity), *C. montanus*, *C. obsoletus s.s.* and few other species that were identified after molecular investigations. The status of *C. montanus* as valid species is still uncertain. *Culicoides scoticus* is almost always grouped with *C. obsoletus s.l.* in routine identification process, because morphological separation of females is uncertain, though males can be separated accurately. Adult females of this taxon feed on a wide variety of mammalian hosts and appear to be opportunistic in preference. Feeding on avian hosts (*e.g.* ducks and chickens) has been recorded for *C. obsoletus s.l.* but is relatively rare. This taxon has been strongly implicated in the transmission of bluetongue and Schmallenberg viruses, and *C. obsoletus s.l.* may have played a role in the transmission of African horse sickness virus.



The taxon is present virtually everywhere in Europe, except in Iceland. Absence records are located in the Extramadura region in Spain, and occasionally in Maghreb. C. obsoletus s.l. is the most abundant species of biting midge in most of northern Europe, and catches of over ten thousand individuals in a single night of light trapping are frequently reported. It has been found at altitudes of up to 1,200 m. Its abundance and cryptic diversity decreases in Mediterranean regions, especially in Northern Africa. . C. obsoletus s.l. has a Holarctic distribution, although this status as a single species is under investigation, whereas C. scoticus is restricted to the western Palearctic zone. Immature stages of C. obsoletus s.l./C. scoticus require a semi-aquatic environment and organic enrichment. Habitats of this taxon identified include deciduous leaf litter, compost heaps, the dungsoil interface, marginal vegetation in open water and silage and also, for C. scoticus, tree fungi.

Two thirds of the GBIF records are located in the VectorNet presence polygons. Most of the GBIF records fall in the VectorNet polygons with no data are located in the Unitedkingdom where VectorNet data are scarce and from Estonia where there is no VectorNet records, suggesting the existence of unpublished datasets in these countries. Interesting GBIF records are observed in Turkey and in the Caucasus, where very little information is yet available in the VectorNet database.





















### Supplementary Figure S39: Dermacentor reticulatus

Dermacentor reticulatus is known as the ornate cow tick. It frequently infests cattle and horses, and can also be found on dogs. It is generally considered a 'winter tick', active in northern Europe during the late winter / early spring, often before the other ticks become active. It can be found in central Europe in river valleys, and in north-western Europe in sand dune habitats and coastal grasslands. The tick is easily collected by flagging vegetation as well as from animal surveys. *D. reticulatus*, and similar species *D. marginatus* are considered competent vectors of CCHFV (cf Bazanov, B.A. et al. doi: 10.1089/vbz.2016.2075. epub 2017), TBEV and are important vectors of rickettsiae that cause TIBOLA/DEBONEL (*R. sibirica, R. slovaca*).



## Supplementary Figure S40: Hyalomma lusitanicum

*Hyalomma lusitanicum* is a tick of the western Mediterranean. It has recently been implicated in the transmission of CCHFV in Spain. It can be found in woodland, steppe and scrub habitat in Iberia. It adopts a hunter strategy, like other *Hyalomma* ticks. It feeds on a variety of large and medium sized mammals such as wild and domestic ungulates. Immature stages feed on lagomorphs, and rarely birds, though it is transported on dogs.



#### Supplementary Figure S41: Hyalomma marginatumCCHF

*Hyalomma marginatum* is the primary European vector of CCHFV. Adult stages are frequently found on cattle where it has a predisposition to biting around the perineum and underbelly. They also often bite humans. The immature stages are found on birds and lagomorphs. The immature stages are nidifugous and are often carried by migratory birds to distant (northern) locations. Here, the ability of the engorged nymph to moult to the adult stage is climate dependent. This species does not quest so are normally sampled by examining the animal hosts, rather than more traditional flagging techniques used for other tick species such as *Ixodes* and *Dermacentor*. In addition, in contrast to other genera, including *Ixodes*, the *Hyalomma* ticks can survive in more arid environments, and are not restricted to humid environments, often afforded by forested habitats.



## Supplementary Figure S42: Ixodes persulcatus

*Ixodes persulcatus* is very similar morphologically to *Ixodes ricinus*, as it occurs in similar woodland habitats though it is found in more grassy areas. Its populations tend to peak earlier in the year. It is also a vector of *Borrelia burgdorferi s.l.* and TBEV, along with other tick-borne pathogens such as *Anaplasma*.



#### Supplementary Figure S43: Ixodes Ricinus

*Ixodes ricinus* is widespread across Europe. Principally a tick of habitats that afford a moist microclimate like forests, it is found in grazed grasslands, moorland/heathland/montane habitats, mosaic habitats with agriculture, grassland and pasture, as well as urban parks. It's ubiquity and wide host range makes it an ideal vector for several pathogens. Larvae, nymphs and adults can be found on a wide range of animal host. Humans are often bitten, as are companion animals such as dogs and cats, and so records of tick bites from these species are numerous. In more sylvatic habitats, all stages of tick can be found on large ungulates such as deer, cattle and sheep, with the immature tick stages infesting a range of small and medium sized mammals and birds. It is the primary vector of *Borrelia burgdorferi* (the causative agent of Lyme borreliosis), Tick-borne encephalitis virus and other pathogens such as *Babesia, Anaplasma, Borrelia miyamotoi, Ricketssia* and Louping ill virus.



The distribution is essentially panregional, but acquired records are biased to west central and southern areas, with relatively few data records from eastern regions, so extrapolations are likely to be less accurate to east. Records of introduction (yellow) are seen in the far north of its range. There are few absence records.

*Ixodes ricinus* is limited in its distribution where there is an absence of hosts or a microclimate that limits their survival. For example, southern parts of Europe are too dry for their survival. High altitude areas of the Alps of central Europe are too cold for their survival, although they are being found at high altitude. In northern regions, there is a climatic limit, but the absence of abundant hosts is a limiting factor in places like Iceland.

This species is one of the most prolifically recorded by GBIF. Ninety percent of the GBIF occurrences are in VectorNet areas defined as present with about ten percent in areas with No Data, most noticeably in Norway and to a lesser extent in Russia and the Baltic countries. There are no GBIF records in VectorNet polygons defined as absent.

*Ixodes ricinus* remains one of the best recorded species of tick in Europe. For some records in southern Europe, these are now being questioned as other similar species (*I. gibbosus. I. inopinatus*).

## Supplementary Figure S44: Ornithodorus erraticus

The Ornithodorus erraticus complex includes a number of similar species and is a subject of much debate. They are associated with warm blooded animals, such as ungulates, carnivores, rodents and insectivores. Species of the O. erraticus complex are often found on pigs, with some experimental evidence of a role in the transmission of African swine fever virus.



#### Supplementary Figure S45: Rhipicephalus sanguineus group

*R. sanguineus* s.l. (the brown dog tick) has two defined lineages, one that is temperate, the other tropical. They are primarily ticks of dogs, particularly in urban areas, and are one of the most common ticks on dogs globally, associated with their homes and shelters. They can be found in the cracks and crevices on walls of rooms where dogs spend the night and have an ability to survive indoors in cooler climes. *R. turanicus* is morphologically similar to *R. sanguineus*, although it is more of a sylvatic species in grassland and meadows, semi-desert and steppe habitats, feeding on a range of mammals and birds, with a possible role in rickettsial transmission. The brown dog tick can transmit both human and animal diseases such as Mediterranean spotted fever and canine babesiosis and ehrlichiosis.



# SUPPLEMENTRAY INFORMATION SECTION VI: Supplementary Figure S46: Citations



Supplementary Figure S46: Citation Numbers to November 2021