

Supplemental Figure 1. *Lp* knockdown significantly impairs oogenesis and affects Vg expression and localization. (A) Successful *Lp* knockdown as determined by RT-qPCR of *Lp* expression levels relative to *Rpl19* in the fat body of dsLacZ and dsLp females (REML variance component analysis: * = p < 0.05; **** = p < 0.0001; three biological replicates). (B) RT-qPCR of *Vg* expression levels relative to *Rpl19* in the fat body of dsLacZ and dsLp females (REML variance component analysis: ** = p < 0.01; **** = p < 0.0001; four biological replicates). (C) Western blot quantification from Figure 1C showing an accumulation of Vg in the fat body and a decrease of Vg in the ovaries upon *Lp* knockdown (REML variance component analysis: ** = p < 0.001; **** = p < 0.0001; three biological replicates). (D) Images of ovaries at 24 and 48h post blood meal showing that Lp depleted ovaries develop normally at first before degenerating by 48h; scale bar = 2mm.

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Supplemental Figure 2. *Vg* knockdown effects in fat body and ovaries. (A) Successful *Vg* knockdown as determined by RT-qPCR of *Vg* expression levels relative to *Rpl19* in the fat body (REML variance component analysis: * = p < 0.05; **** = p < 0.0001; three biological replicates). (B) Fold change in protein levels measured by Bradford assay in the ovaries of dsLacZ and dsVg females before blood meal and at 24h and 48h post blood meal (PBM); each dot is representative of three ovaries (REML variance component analysis by timepoint: ** = p < 0.01; **** = p < 0.001; three biological replicates). (C) Fold change in free amino acid levels in the ovaries of dsLacZ and dsVg females before blood meal and at 24h and 48h PBM; each dot is representative of five ovaries (REML variance component analysis by timepoint: * = p < 0.05; three biological replicates). (D) *Vg* knockdown by second target also results in an increase in *Lp* levels as determined by RT-qPCR (REML variance component analysis: * = p < 0.05; three biological replicates). (E) Western blot quantification from Figure 2G showing an accumulation of Lp in the fat body and ovaries upon *Vg* knockdown (REML variance component analysis: fat body – dsRNA: p < 0.01; * = p < 0.05; ovaries – dsRNA: p < 0.05; * = p < 0.05; three biological replicates).

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Supplemental Figure 3. Vg expression regulates Lp-mediated accumulation of lipids via TOR signaling. (A) Western blot quantification from Figure 3B showing an increase in phospho-S6K levels in the fat body upon Vg knockdown (REML variance component analysis: **** = p < 0.0001; three biological replicates). (B) Fold change in protein levels, measured by Bradford assay of the fat body are increased in dsVg females; each dot is representative of three fat bodies (REML variance component analysis by timepoint: **** = p < 0.0001; three biological replicates). (C) Western blot quantification from Figure 3D showing a decrease in phospho-S6K levels in the fat body upon rapamycin treatment (ANOVA; three biological replicates). (D) Western blot quantification from Figure 3E showing Lp protein levels upon Vg knockdown and rapamycin treatment (ANOVA; three biological replicates). (E) Triglyceride levels measured in dsLacZ and dsVg ovaries upon 0.5 μ l of 40 μ M rapamycin treatment at 72h post blood meal and normalized to

Amino acid	Percent	Decrease upon Vg	Significance	
	content in Vg	KD in mothers (fold change)		
Ser (S)	8.50%	2.2	0.09205	
Tyr (Y)	8.00%	52.6	0.00010	***
Phe (F)	7.70%	63.2	0.00000	****
Glu (E)	6.50%	1.2	0.33025	
Ala (A)	6.30%	2.0	0.02859	*
Asp (D)	6.30%	2.1	0.15012	
Lys (K)	6.20%	10.0	0.00000	****
Gln (Q)	6.10%	1.1	0.88614	
Val (V)	5.80%	3.1	0.00005	****
Asn (N)	5.60%	5.5	0.00006	****
Leu (L)	5.60%	3.7	0.00024	***
Gly (G)	4.40%	2.2	0.02060	*
Pro (P)	4.40%	1.4	0.01066	*
Thr (T)	4.40%	3.9	0.00015	***
Arg (R)	3.90%	2.9	0.00012	***
Ile (I)	3.20%	8.7	0.00001	****
His (H)	2.90%	1.0	0.69815	
Met (M)	2.20%	4.9	0.00010	***
Cys (C)	1.10%	Not detected		
Trp (W)	0.80%	13.2	0.00002	****

Supplemental Table 1: Vg amino acids and their decrease in embryos upon Vg depletion.



Supplemental Figure 4. *Vg* knockdown in females prevents embryo melanization and causes early embryonic arrest. (A) Light microscopy of embryos from dsLacZ- and dsVg-derived females at 3-5h post oviposition; scale bar = $200 \,\mu$ m. (B-C) Total lipids (B) and lipid classes (C) in dsLacZ- and dsVg-derived embryos 3-5h post oviposition as determined by mass spectrometry (unpaired t tests, followed by FDR correction: * = p < 0.05; ** = p < 0.01; four biological replicates).

Supplemental Table 4

Gene	Primer	Citation, if previously published	
Rpl19	FCCAACTCGCGACAAAACATTC	Werling, Shaw, Itoe et al.,	
-	R ACCGGCTTCTTGATGATCAGA	2019	
Lp	FCAGCCAGGATGGTGAGCTTAA	Werling, Shaw, Itoe et al.,	
	R CACCAGCACCTTGGCGTT	2019	
Vg	F CCGACTACGACCAGGACTTC	Werling, Shaw, Itoe et al.,	
	R CTTCCGGCGTAGTAGACGAA	2019	
ILP1/7	F GCAAAAAGTCCGAGAATCTACTGATGA		
	R CGAACGATCGTTCAATGTGTGGA		
ILP2	FCTACCTCTACGCCCAACAGC		
	R CGTGTACATAATCTGTGCGATAGTG		
ILP3/6	FGGTAAAGGTACTGTCCTTCCTG	Arsic and Guerin, 2008	
	R AGTATCTGCTGCGTGTTGTC		
ILP4	F TCTCCGAAAGAACACAGTTGA	Arsic and Guerin, 2008	
	R GGTTTCTGCCTGAACCACAT		
ILP5	F GTGGCACCAGGAGAGTCATT		
	R GCCCAGTACAGATGGCGTAT		

Supplemental Table 5. Details of statistical tests and outputs are summarized for each figure. For qRT-PCR, at least three independent biological replicates of a gene expression timecourse were analyzed, except for ILP1, where one replicate was excluded as an outlier. Effect test outputs are reported here. Multiple comparisons were calculated using pairwise Student's t tests at each timepoint followed by FDR correction (see Table S6). KD = knock down; rand = random effect; FDR = false discovery rate.

Fig	Comparison	Statistical test	Effect Test Outputs
1A	Egg number after Lp KD	Mann-Whitney	p<0.0001
1B	TAG levels after Lp KD (Ovaries)	Ln(x+1.1) transformation; Linear Mixed Model at each timepoint	(0h) dsRNA p=0.7964 replicate p=0.3755 (24h) dsRNA p<0.0001 replicate p=0.3400 (48h) dsRNA p<0.0001 replicate p=0.0109
1B	TAG levels after Lp KD (Midgut)	Ln(x+1.1) transformation; Linear Mixed Model at each timepoint	(0h) dsRNA p=0.2984 replicate p=0.3317 (24h) dsRNA p<0.0001 replicate p=0.3573 (48h) dsRNA p<0.0001 replicate p=0.3228
2A	Egg number after Vg KD	Mann-Whitney test	p<0.0001
2B	Fertility after Vg KD	Kruskal-Wallis test	dsLacZ vs dsVg: p<0.0001 dsLacZ vs dsVg#2: p<0.0001 dsVg vs dsVg#2: p>0.05
2 E	Triglycerides after Vg KD (ovaries 48h)	Unpaired t test on transformed data	p=0.0222
2F	Lp expression after Vg KD	4 th root transformation; Linear Mixed Model followed by 5 post-hoc t- tests (Table S6)	timepoint p<0.0001 dsRNA p<0.0001 dsRNA x timepoint p=0.0032 replicate[rand] p=0.3090
3A	Vg upon rapamycin treatment	Unpaired t test	p<0.0001

3C	Lp mRNA levels upon rapamycin treatment	ANOVA	LacZ Control vs. LacZ Rapamycin p=0.9956 LacZ Control vs. Vg Control p=0.0002 LacZ Control vs. Vg Rapamycin p=0.2500 LacZ Rapamycin vs. Vg Control p=0.0003 LacZ Rapamycin vs. Vg Rapamycin p=0.3469 Vg Control vs. Vg Rapamycin
4D	Embryo	Unpaired t test	p=0.0226 p=0.0496
S1A	Lp expression after Lp KD	4 th root transformation; Linear Mixed Model followed by 5 post- hoc t-tests (Table S6)	timepoint p<0.0001 dsRNA p<0.0001 dsRNA x timepoint p=0.0004 replicate[rand] p=0.478
S1B	Vg expression after Lp KD	4 th root transformation; Linear Mixed Model followed by 5 post- hoc t-tests (Table S6)	timepoint p<0.0001 dsRNA p=0.044 dsRNA x timepoint p=0.0001 replicate[rand] p=0.536 replicate x timepoint[rand] p=0.033
S1C	Lp expression after Vg KD (Ovaries)	Ln(x+1) transformation; Linear Mixed Model followed by 3 post- hoc t-tests (Table S6)	timepoint p<0.0001 dsRNA p=0.0103 dsRNA x timepoint p=0.0192 replicate p=0.3836
S1C	Lp expression after Vg KD (Fatbody)	No transformation; Generalized Linear Model followed by 3 post- hoc t-tests (Table S6)	timepoint p<0.0001 dsRNA p=0.0148 dsRNA x timepoint p<0.0001 replicate p=0.0012
S2A	Vg expression after Vg KD	8 th root transformation; Linear Mixed Model followed by 5 post- hoc t-tests (Table S6)	timepoint p<0.0001 dsRNA p<0.0001 dsRNA x timepoint p=0.0068 replicate[rand] p=0.6864
S2B	Protein levels after Vg KD (Ovaries)	No suitable transformation; Generalized Linear Mixed Model at each timepoint	(0h) dsRNA p=0.8072 replicate p=0.4399 (24h) dsRNA p=0.0044 replicate p=0.0002 (48h) dsRNA p<0.0001 replicate p=0.0092

			(0h) dsRNA p=0.5067
		Square root	replicate p=0.4232
630	Amino acid	transformation;	(24h) dsRNA p=0.8833
S2C	levels after Vg KD	Linear Mixed Model	replicate p=0.4843
	(Ovaries)	at each timepoint	(48h) dsRNA p=0.0163
			replicate p=0.3562
		4 th root transformation:	timepoint p=0.0004
	Lp expression	Linear Mixed Model	dsRNA p=0.1804
S2D	after Vg KD#2	followed by 3 post-	dsRNA x timepoint p=0.0409
		hoc t-tests (Table S6)	replicate[rand] p=0.9751
		Square root $(x+0.03)$	
	Vg expression	transformation:	timepoint p<0.0001
S2E	after Lp KD	Linear Mixed Model	dsRNA p=0.0100
5-12	(Ovaries)	followed by 3 post-	dsRNA x timepoint p=0.1130
	(O varies)	hoc t-tests (Table S6)	replicate p=0.8073
		No transformation:	timepoint p=0.0004
	Vg expression	Linear Mixed Model	dsRNA p=0.0025
S2E	after Lp KD	followed by 3 post-	dsRNA x timepoint $p=0.5277$
	(Fatbody)	hoc t-tests (Table S6)	replicate $p=0.6241$
			(0h) dsRNA p=0.4585
		Cube root	replicate p=0.3221
	Protein levels	transformation.	(24h) dsRNA p<0.0001
S3A	after Vg KD	Linear Mixed Model	replicate p=0 3232
	(Fat body)	at each timepoint	(48h) ds RNA p < 0.0001
		at each timepoint	replicate $p=0.3260$
		No transformation:	
	pS6K	Generalized	timepoint p<0.0001
S3B	expression	Linear Model	dsRNA p=0.1251
502	after Vg KD	followed by 4 post-	dsRNA x timepoint p<0.0001
	(Fatbody)	hoc t-tests (Table S6)	replicate p=0.0084
			LacZ Control vs. LacZ
			Rapamycin
			p=0.9976
			LacZ Control vs. Vg Control
			p<0.0001
	Phospho S6K		LacZ Control vs. Vg Rapamycin
GAG	protein levels upon		p=0.0072
S3C	rapamycin	ANOVA	LacZ Rapamycin vs. Vg Control
	treatment		p<0.0001
			LacZ Rapamycin vs. Vg
			Rapamycin
			p=0.0100
			Vg Control vs. Vg Rapamycin
			p=0.0102
COD	Lp protein		LacZ Control vs. LacZ
53D	levels upon	ANUVA	Rapamycin

n>0 0000
ol vs. Vg Control
p=0.0185
p=0.0105
n=0.2184
p=0.2104
m = 0.0172
p=0.0172
Rapamycin vs. vg
Rapamycin
p=0.2044
vs. Vg Rapamycin
p=0.4834
Control vs. LacZ
Rapamycin
P=0.9860
ol vs. Vg Control
p=0.0030
s. Vg Rapamycin
p=0.2565
vin vs. Vg Control
p=0.0018
Rapamycin vs. Vg
Rapamycin
p=0.1618
vs. Vg Rapamycin
p=0.2566
dsRNA p=0.8672
eplicate p=0.3832
dsRNA p=0.1087
eplicate p=0.3257
dsRNA p=0.0064
eplicate p=0.3201
dsRNA p=0.0027
eplicate p=0.4067
dsRNA p=0.7177
eplicate p=0.7615
dsRNA p=0.6672
eplicate p=0.4125
dsRNA p=0.2457
eplicate p=0.3536
dsRNA p=0.8222
eplicate p=0.7415
- ·
nepoint p<0.0001
dsRNA p=0.426
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	ILP2 expression after Vg KD	arcsine transformation; Linear Mixed Model No post-hoc t-testing	timepoint p=0.0089 dsRNA p=0.920
	ILP3 expression after Vg KD	No transformation; Linear Mixed Model No post-hoc t-testing	timepoint p=0.248 dsRNA p=0.367
	ILP4 expression after Vg KD	No transformation; Linear Mixed Model No post-hoc t-testing	timepoint p=0.002 dsRNA p=0.408
	ILP5 expression after Vg KD	5 th root transformation; Linear Mixed Model No post-hoc t-testing	timepoint p=0.002 dsRNA p=0.241
S4B	Embryo total lipids	Unpaired t test	p=0.1606

Supplemental Table 6. Post-hoc testing for significant differences using an FDR of 0.05. See

Supplemental Table 5.

2F	p-value	FDR-adjusted	Significant?
Lp after Vg KD		p-value	
dsLacZ – dsVg at 0h	0.6414	0.6414	No
dsLacZ – dsVg at 12h	0.0731	0.0914	No
dsLacZ – dsVg at 24h	4.13 x 10 ⁻⁶	2.07 x 10 ⁻⁵	Yes
dsLacZ – dsVg at 36h	0.0020	0.0050	Yes
dsLacZ-dsVg at 48h	0.0450	0.0750	No
S1B	p-value	FDR-adjusted	Significant?
Vg after Lp KD		p-value	
dsLacZ – dsLp at 0h	0.9198	0.9198	No
dsLacZ – dsLp at 12h	0.7704	0.9198	No
dsLacZ – dsLp at 24h	0.0029	0.0048	Yes
dsLacZ – dsLp at 36h	0.0028	0.0048	Yes
dsLacZ – dsLp at 48h	7.55 x 10 ⁻⁵	3.78 x 10 ⁻⁴	Yes
S1A	p-value	FDR-adjusted	Significant?
Lp after Lp KD		p-value	
dsLacZ – dsLp at 0h	2.64 x 10 ⁻⁵	6.60 x 10 ⁻⁵	Yes
dsLacZ – dsLp at 12h	4.36 x 10 ⁻⁷	2.18 x 10 ⁻⁶	Yes
dsLacZ – dsLp at 24h	0.0138	0.0173	Yes
dsLacZ – dsLp at 36h	0.8884	0.8884	No
dsLacZ – dsLp at 48h	2.67 x 10 ⁻⁴	4.45 x 10 ⁻⁴	Yes
S1C (Ovaries)	p-value	FDR-adjusted p-	Significant?
		value	
dsLacZ – dsVg at 0h	0.7766	0.7766	No
dsLacZ – dsVg at 24h	0.6119	0.7766	No
dsLacZ – dsVg at 48h	0.0009	0.0027	Yes
S1C (Fat body)	p-value	FDR-adjusted p-	Significant?
		value	
dsLacZ – dsVg at 0h	0.7447	0.7447	No
dsLacZ – dsVg at 24h	0.1780	0.2670	No
dsLacZ – dsVg at 48h	2.86 x 10 ⁻¹⁰	8.56 x 10 ⁻¹⁰	Yes
S2A	p-value	FDR-adjusted	Significant?
Vg after Vg KD		p-value	
dsLacZ - dsVg at 0h	0.9000	0.9000	No
dsLacZ – dsVg at 12h	0.0057	0.0115	Yes
dsLacZ – dsVg at 24h	2.94 x 10 ⁻⁵	1.47 x 10 ⁻⁴	Yes
dsLacZ – dsVg at 36h	0.0069	0.0115	Yes
dsLacZ – dsVg at 48h	0.5501	0.6877	No

S2D	p-value	FDR-adjusted	Significant?
Lp after Vg KD#2		p-value	
dsLacZ – dsVg at 0h	0.3785	0.5678	No
dsLacZ – dsVg at 24h	0.0098	0.0294	Yes
dsLacZ – dsVg at 48h	0.8191	0.8191	No
S2E (Ovaries)	p-value	FDR-adjusted p-	Significant?
		value	
dsLacZ – dsVg at 0h	0.9392	0.9392	No
dsLacZ – dsVg at 24h	0.0210	0.0315	Yes
dsLacZ – dsVg at 48h	0.0177	0.0315	No
S2E (Fat body)	p-value	FDR-adjusted p-	Significant?
		value	
dsLacZ – dsVg at 0h	0.1927	0.1927	No
dsLacZ – dsVg at 24h	0.0133	0.0399	Yes
dsLacZ – dsVg at 48h	0.0298	0.0447	Yes
S3A (Fat body)	p-value	FDR-adjusted p-	Significant?
		value	
dsLacZ – dsVg at 0h	0.9058	0.9058	No
dsLacZ-dsVg at 12h	0.1829	0.3658	No
dsLacZ – dsVg at 24h	8.06 x 10 ⁻²⁵	3.22 x 10 ⁻²⁴	Yes
dsLacZ-dsVg at 48h	0.6257	0.8343	No