

Supplementary Information

CRISPR prime editing for unconstrained correction of oncogenic *KRAS* variants.

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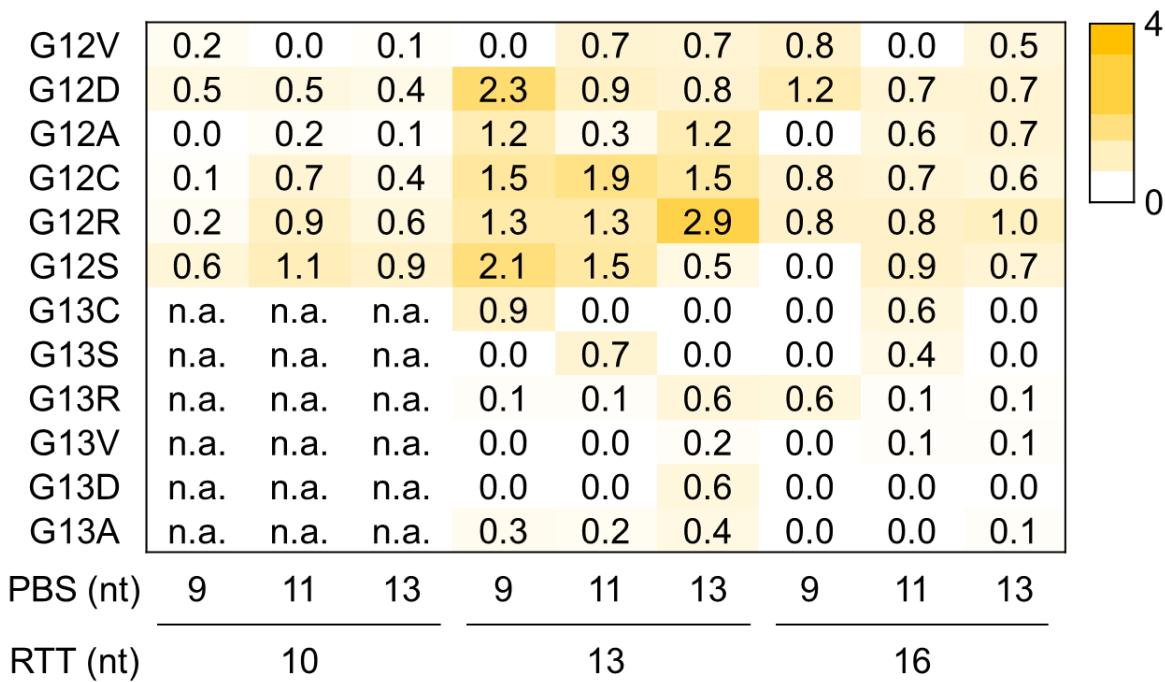
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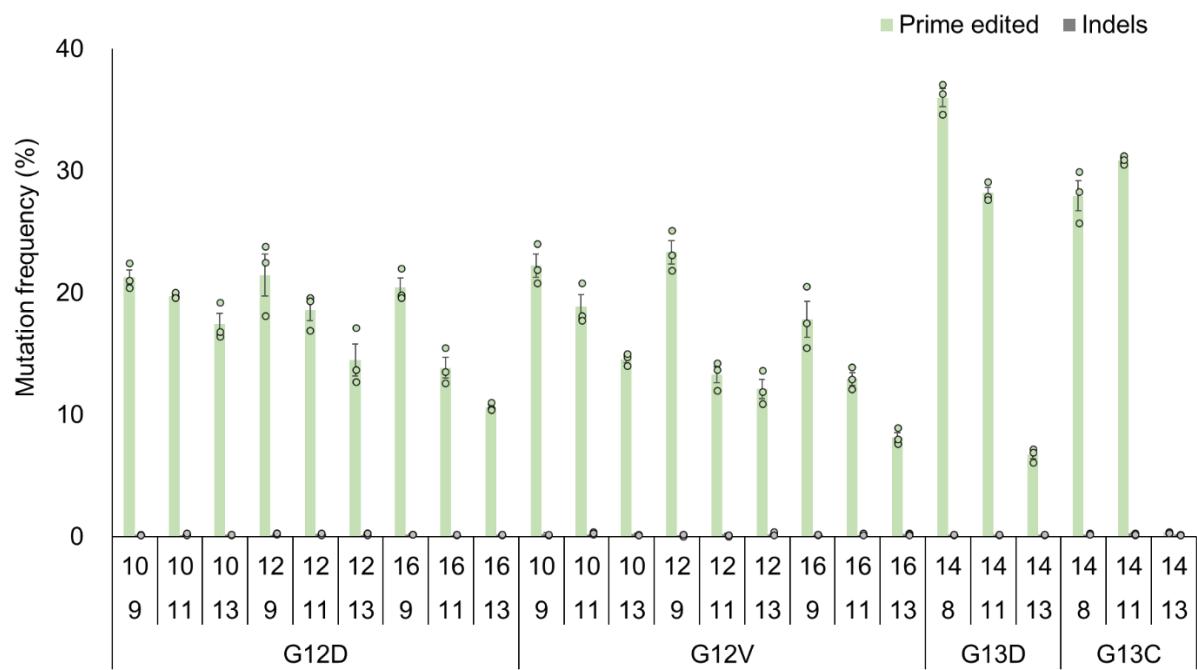
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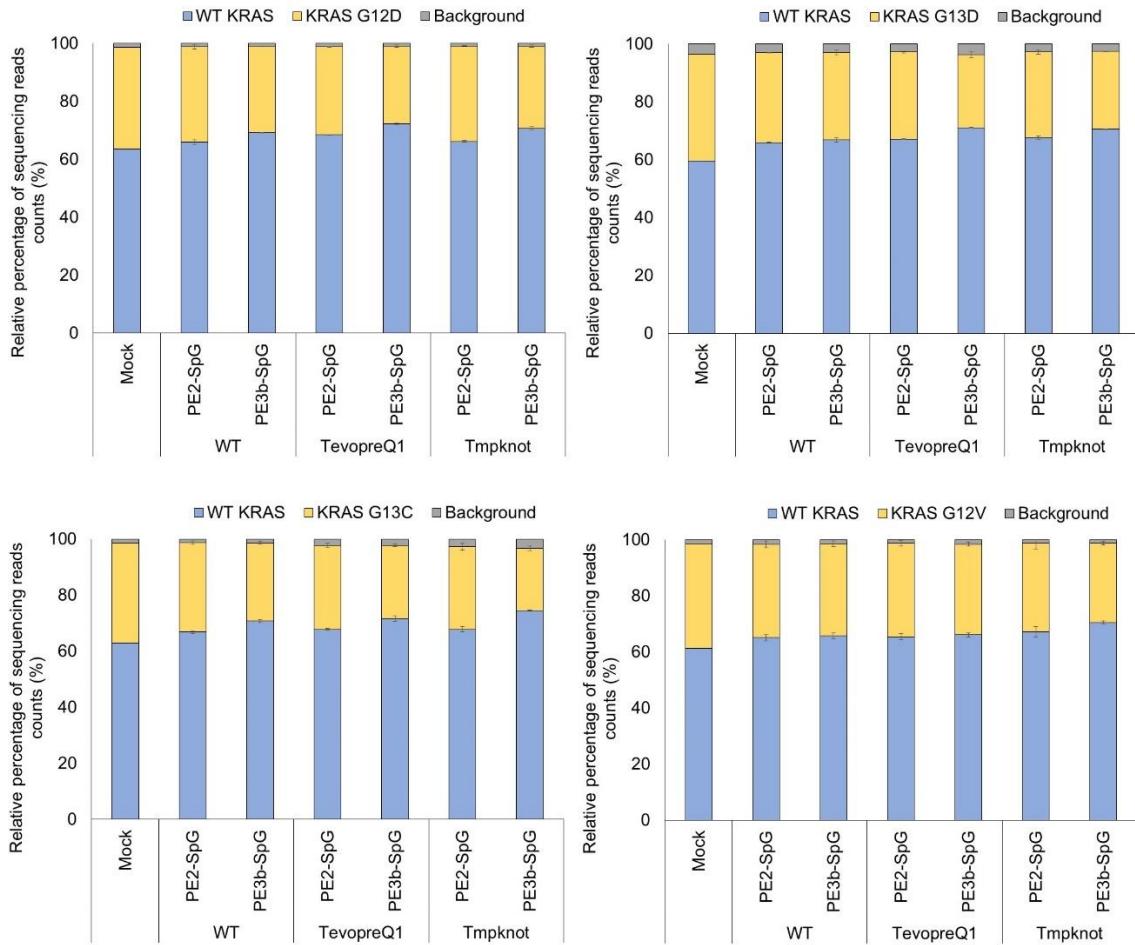
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Supplementary Figure 1. KRAS correction frequency of KRAS-#1 universal pegRNA in HEK293T/17-KRAS library cells.



Supplementary Figure 2. Optimization of pegRNAs to generate *KRAS* heterozygous HEK293T/17 cells. Error bar means s.e.m. of $n=3$ biological triplicate samples.



Supplementary Figure 3. KRAS correction frequencies in KRAS heterozygous HEK293T/17 cells. Target sites were amplified and analyzed by targeted-deep sequencing. Relative counts of sequencing reads were calculated from total number of sequencing reads of wild-type KRAS sequences and mutant KRAS sequences. Error bar means s.e.m. of $n=3$ biological triplicate samples.

KRAS G12V mutant alleles of HEK293T/17 heterogenous cells

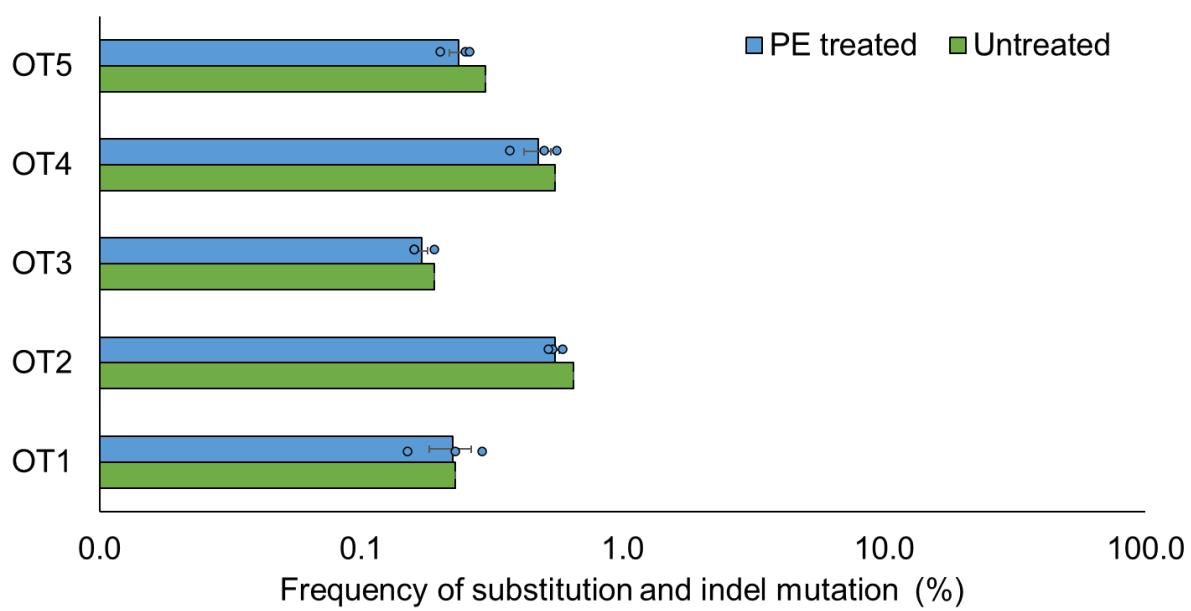
E Y K L V V V G A V G V G K S A L T
 5'-GAATATAAACCTGTGGTAGTTGGAGCTGTTGGCGTAGGCAAGAGTCCTTGACGA-3',
 3'-CTTATATTGAACACCATCACCTGACAACCGC~~A~~TCCGTTCTCACGGAACTGCT-5'

↓
 Prime editing
 using the universal pegRNA
 containing a silent mutation

E Y K L V V V G A G G V G K S A L T
 5'-GAATATAAACCTGTGGTAGTTGGAGCAGGTGGCGTAGGCAAGAGTCCTTGACGA-3',
 3'-CTTATATTGAACACCATCACCTGTCACCGC~~A~~TCCGTTCTCACGGAACTGCT-5'

Untreated	TAGTTGGAGCTGGTGGCGTAGGCAA	(2434/4029)	60.4%
	TAGTTGGAGCTGTTGGCGTAGGCAA	(1519/4029)	37.7%
	TAGTTGGAGCAGGTGGCGTAGGCAA	(0/4029)	0.0%
	TAGTTGGAGCAGTTGGCGTAGGCAA	(0/4029)	0.0%
pegRNA treated	TAGTTGGAGCTGGTGGCGTAGGCAA	(2889/4738)	61.0%
	TAGTTGGAGCTGTTGGCGTAGGCAA	(1561/4738)	32.9%
	TAGTTGGAGCAGGTGGCGTAGGCAA	(206/4738)	4.3%
	TAGTTGGAGCAGTTGGCGTAGGCAA	(0/4738)	0.0%

Supplementary Figure 4. *KRAS* correction with pegRNA capable of inducing silent mutation in G12V heterozygous HEK293T/17 cells. The pegRNA could induce T:A to A:T silent mutation at alanine 11 residue and correct *KRAS* G12V mutation. The silent mutation and *KRAS* G12V mutation were highlighted in blue and red, respectively, and the spacer sequences of pegRNA were boxed. NGS sequencing results showed that *KRAS* correction was accompanied by the silent mutation, indicating that *KRAS* correction was induced by prime editing.



Supplementary Figure 5. Analysis of potential off-target effects of the universal epegRNA. Five potential-offtarget sites were selected *in silico* and the universal epegRNA was transfected into HEK293T/17-KRAS G12V cells with PEmax-SpG-hMLH1dn. Targeted-deep sequencing results showed that there were no significant off-target mutations in the five potential off-target sites. The potential off-target sites were listed in Supplementary Table 3. Error bar means s.e.m. of biological triplicate.

Supplementary Table 1. Substitution and indel frequencies at endogenous KRAS sites in HEK293T/17-KRAS library cells.

pegRNA names	PBS length (nt)	RTT length (nt)	Replicate 1		Replicate 2	
			Substitutions (%)	Indels (%)	Substitutions (%)	Indels (%)
Universal pegRNA #2-1	9	10	1.3%	0.1%	1.3%	0.1%
Universal pegRNA #2-2	11	10	1.3%	0.1%	1.3%	0.1%
Universal pegRNA #2-3	13	10	1.5%	0.1%	1.2%	0.0%
Universal pegRNA #2-4	9	13	1.3%	0.0%	1.2%	0.0%
Universal pegRNA #2-5	11	13	1.5%	0.1%	1.4%	0.0%
Universal pegRNA #2-6	13	13	1.4%	0.0%	1.5%	0.0%
Universal pegRNA #2-7	9	16	1.5%	0.0%	1.2%	0.0%
Universal pegRNA #2-8	11	16	1.4%	0.1%	1.6%	0.0%
Universal pegRNA #2-9	13	16	1.4%	0.1%	1.4%	0.0%
Mock			1.6%	0.1%		

PE systems	PBS length (nt)	RTT length (nt)	RNA scaffolds	Replicate 1		Replicate 2	
				Substitutions (%)	Indels (%)	Substitutions (%)	Indels (%)
PE2-SpG	13	16	Wild-type	0.0%	0.0%	0.6%	0.0%
PE3b-SpG	13	16	Wild-type	0.8%	0.0%	0.6%	0.0%
PE2-SpG	13	16	TevopreQ1	0.8%	0.0%	0.5%	0.0%
PE3b-SpG	13	16	TevopreQ1	0.5%	0.0%	0.6%	0.1%
PE2-SpG	13	16	Tmpknot	0.6%	0.2%	0.6%	0.1%
PE3b-SpG	13	16	Tmpknot	0.5%	0.0%	0.6%	0.1%
Mock				0.8%	0.1%		

Supplementary Table 2. Prime editing and indel frequencies in HEK293T/17-KRAS heterogenous cells.

PE systems	PBS length (nt)	RT length (nt)	RNA scaffolds	G12D		G12V		G13D		G13C	
				% of wild-type KRAS sequences	Indels (%)	% of wild-type KRAS sequences	Indels (%)	% of wild-type KRAS sequences	Indels (%)	% of wild-type KRAS sequences	Indels (%)
PE2-SpG	13	16	Wild-type	66.6	0.0	65.1	0.0	66.1	0.0	66.1	0.0
PE3b-SpG	13	16	Wild-type	68.8	0.0	65.8	0.0	67.4	0.0	69.2	0.0
PE2-SpG	13	16	TevopreQ1	68.6	0.0	65.5	0.0	68.3	0.0	67.1	0.0
PE3b-SpG	13	16	TevopreQ1	71.0	0.0	66.2	0.0	71.2	0.0	70.1	0.0
PE2-SpG	13	16	Tmpknot	67.2	0.0	67.2	0.0	68.8	0.0	66.5	0.0
PE3b-SpG	13	16	Tmpknot	69.9	0.0	70.6	0.0	71.2	0.2	72.1	0.0
Mock				62		61.2		59.5		62.9	

Supplementary Table 3. List of potential off-target sites.

Target mutations	Target sequences (5' to 3')		PAM	Chr.	Mismatches (bp)
c.35T>G (G12V)	ON	CGTCAAGGCACTTTGCCCTA	CG	chr12	0
	OT1	CGTCActGCACTTTGCCCTA	GG	chr19	2
	OT2	aGaCAAGGCACCTtTTGCCCTA	TG	chr4	3
	OT3	CGTCAtGGgACTCTTtCCCTA	CG	chr10	3
	OT4	aGTCAAGGCACTCcTaCCCTA	TG	chr6	3
	OT5	CGTCActGCACTCTaGCCCTA	GG	chr5	3

Supplementary Table 4. KRAS-mutant target sequences used in HEK293T/17-KRAS library cells.

KRAS mutation	KRAS mutant target sequence(5' to 3')	Barcode sequence
G12V	ATGACTGAATATAAACTTGTGGTAGTTGGAGCTGtTGGCGTAGGCAAGAGTCCTTGACG	CGAGTAAT
G12D	ATGACTGAATATAAACTTGTGGTAGTTGGAGCTGaTGGCGTAGGCAAGAGTCCTTGACG	TCTCCGGA
G12A	ATGACTGAATATAAACTTGTGGTAGTTGGAGCTGcTGGCGTAGGCAAGAGTCCTTGACG	AATGAGCG
G12C	ATGACTGAATATAAACTTGTGGTAGTTGGAGCTtGTGGCGTAGGCAAGAGTCCTTGACG	GGAATCTC
G12R	ATGACTGAATATAAACTTGTGGTAGTTGGAGCTcGTGGCGTAGGCAAGAGTCCTTGACG	TTCTGAAT
G12S	ATGACTGAATATAAACTTGTGGTAGTTGGAGCTaGTGGCGTAGGCAAGAGTCCTTGACG	ACGAATTc
G13C	ATGACTGAATATAAACTTGTGGTAGTTGGAGCTGGTtGCGTAGGCAAGAGTCCTTGACG	AGCTTCAG
G13S	ATGACTGAATATAAACTTGTGGTAGTTGGAGCTGGTaGCGTAGGCAAGAGTCCTTGACG	GCGCATTA
G13R	ATGACTGAATATAAACTTGTGGTAGTTGGAGCTGGTcGCGTAGGCAAGAGTCCTTGACG	CATAGCCG
G13V	ATGACTGAATATAAACTTGTGGTAGTTGGAGCTGGTtCGTAGGCAAGAGTCCTTGACG	TTCGCGGA
G13D	ATGACTGAATATAAACTTGTGGTAGTTGGAGCTGGTaCGTAGGCAAGAGTCCTTGACG	GCGCGAGA
G13A	ATGACTGAATATAAACTTGTGGTAGTTGGAGCTGGGCGTAGGCAAGAGTCCTTGACG	CTATCGCT

Supplementary Table 5. Sequences of gRNA for PE3b, pegRNAs and epegRNAs used in this study.

gRNA or pegRNA name	Spacer sequence (5' to 3')	PAM	3' extension sequence (5' to 3')	PBS length (nt)	RTT length (nt)
G12D mutant pegRNA #1	AAACTTGTGGTAGTTGGAGC	TGG	GCCAtCAGCTCCAACCTACC	9	10
G12D mutant pegRNA #2	AAACTTGTGGTAGTTGGAGC	TGG	GCCAtCAGCTCCAACCTACCA	11	10
G12D mutant pegRNA #3	AAACTTGTGGTAGTTGGAGC	TGG	GCCAtCAGCTCCAACCTACCAA	13	10
G12D mutant pegRNA #4	AAACTTGTGGTAGTTGGAGC	TGG	ACGCCAtCAGCTCCAACCTACC	9	12
G12D mutant pegRNA #5	AAACTTGTGGTAGTTGGAGC	TGG	ACGCCAtCAGCTCCAACCTACCA	11	12
G12D mutant pegRNA #6	AAACTTGTGGTAGTTGGAGC	TGG	ACGCCAtCAGCTCCAACCTACCAA	13	12
G12D mutant pegRNA #7	AAACTTGTGGTAGTTGGAGC	TGG	GCCTACGCCAtCAGCTCCAACCTACC	9	16
G12D mutant pegRNA #8	AAACTTGTGGTAGTTGGAGC	TGG	GCCTACGCCAtCAGCTCCAACCTACCA	11	16
G12D mutant pegRNA #9	AAACTTGTGGTAGTTGGAGC	TGG	GCCTACGCCAtCAGCTCCAACCTACCAA	13	16
G12V mutant pegRNA #1	AAACTTGTGGTAGTTGGAGC	TGG	GCCAAcCAGCTCCAACCTACC	9	10
G12V mutant pegRNA #2	AAACTTGTGGTAGTTGGAGC	TGG	GCCAAcCAGCTCCAACCTACCA	11	10
G12V mutant pegRNA #3	AAACTTGTGGTAGTTGGAGC	TGG	GCCAAcCAGCTCCAACCTACCAA	13	10
G12V mutant pegRNA #4	AAACTTGTGGTAGTTGGAGC	TGG	ACGCCAaCAGCTCCAACCTACC	9	12
G12V mutant pegRNA #5	AAACTTGTGGTAGTTGGAGC	TGG	ACGCCAaCAGCTCCAACCTACCA	11	12
G12V mutant pegRNA #6	AAACTTGTGGTAGTTGGAGC	TGG	ACGCCAaCAGCTCCAACCTACCAA	13	12
G12V mutant pegRNA #7	AAACTTGTGGTAGTTGGAGC	TGG	GCCTACGCCaCAGCTCCAACCTACC	9	16
G12V mutant pegRNA #8	AAACTTGTGGTAGTTGGAGC	TGG	GCCTACGCCaCAGCTCCAACCTACCA	11	16
G12V mutant pegRNA #9	AAACTTGTGGTAGTTGGAGC	TGG	GCCTACGCCaCAGCTCCAACCTACCAA	13	16
G13C mutant pegRNA #1	CTTGTGGTAGTTGGAGCTGG	TGG	TGCCTACGCaACCAGCTCCAAC	8	14
G13C mutant pegRNA #2	CTTGTGGTAGTTGGAGCTGG	TGG	TGCCTACGCaACCAGCTCCAAC	11	14
G13C mutant pegRNA #3	CTTGTGGTAGTTGGAGCTGG	TGG	TGCCTACGCaACCAGCTCCAAC	13	14
G13D mutant pegRNA #1	CTTGTGGTAGTTGGAGCTGG	TGG	TGCCTACGtCACCAAGCTCCAAC	8	14
G13D mutant pegRNA #2	CTTGTGGTAGTTGGAGCTGG	TGG	TGCCTACGtCACCAAGCTCCAAC	11	14
G13D mutant pegRNA #3	CTTGTGGTAGTTGGAGCTGG	TGG	TGCCTACGtCACCAAGCTCCAAC	13	14
Universal pegRNA #1-1	TATAAACTTGTGGTAGTTGG	AG	AccAGCTCCAACCTACCA	9	10
Universal pegRNA #1-2	TATAAACTTGTGGTAGTTGG	AG	AccAGCTCCAACCTACCAAG	11	10
Universal pegRNA #1-3	TATAAACTTGTGGTAGTTGG	AG	AccAGCTCCAACCTACCAAGTT	13	10
Universal pegRNA #1-4	TATAAACTTGTGGTAGTTGG	AG	GccAccAGCTCCAACCTACCA	9	13
Universal pegRNA #1-5	TATAAACTTGTGGTAGTTGG	AG	GccAccAGCTCCAACCTACCAAG	11	13
Universal pegRNA #1-6	TATAAACTTGTGGTAGTTGG	AG	GccAccAGCTCCAACCTACCAAGTT	13	13
Universal pegRNA #1-7	TATAAACTTGTGGTAGTTGG	AG	TACGccAccAGCTCCAACCTACCA	9	16
Universal pegRNA #1-8	TATAAACTTGTGGTAGTTGG	AG	TACGccAccAGCTCCAACCTACCAAG	11	16
Universal pegRNA #1-9	TATAAACTTGTGGTAGTTGG	AG	TACGccAccAGCTCCAACCTACCAAGTT	13	16
Universal pegRNA #2-1	CGTCAAGGCACTCTGCCTA	CG	ggTggCGTAGGCAAGAGTG	9	10
Universal pegRNA #2-2	CGTCAAGGCACTCTGCCTA	CG	ggTggCGTAGGCAAGAGTGCC	11	10
Universal pegRNA #2-3	CGTCAAGGCACTCTGCCTA	CG	ggTggCGTAGGCAAGAGTGCTT	13	10
Universal pegRNA #2-4	CGTCAAGGCACTCTGCCTA	CG	GCTggTggCGTAGGCAAGAGTG	9	13
Universal pegRNA #2-5	CGTCAAGGCACTCTGCCTA	CG	GCTggTggCGTAGGCAAGAGTGCC	11	13
Universal pegRNA #2-6	CGTCAAGGCACTCTGCCTA	CG	GCTggTggCGTAGGCAAGAGTGCTT	13	13
Universal pegRNA #2-7	CGTCAAGGCACTCTGCCTA	CG	GGAGCTggTggCGTAGGCAAGAGTG	9	16
Universal pegRNA #2-8	CGTCAAGGCACTCTGCCTA	CG	GGAGCTggTggCGTAGGCAAGAGTGCC	11	16
Universal pegRNA #2-9	CGTCAAGGCACTCTGCCTA	CG	GGAGCTggTggCGTAGGCAAGAGTGCTT	13	16
PE3b Correction gRNA #1	GCACTTGCCTACGCCACC	AG	N/A	N/A	N/A
PE3b Correction gRNA #2	GTGGTAGTGGAGCTGGTGG	CG	N/A	N/A	N/A
pegRNA-silent mutation	CGTCAAGGCACTCTGCCTA	CG	GGAGCaggTggCGTAGGCAAGAGTGCTT	13	16

epegRNA name	Spacer sequence (5' to 3')	PAM	3' extension sequence (5' to 3')	PBS length (nt)	RTT length (nt)	Linker
TevopreQ1 epegRNA #2-9	CGTCAAGGCACTCTGCCTA	CG	GGAGCTggTggCGTAGGCAAGAGTGCTT	13	16	CTTAAGTA
Tmpknot epegRNA #2-9	CGTCAAGGCACTCTGCCTA	CG	GGAGCTggTggCGTAGGCAAGAGTGCTT	13	16	TAATTATA

Supplementary Table 6. Primer sequences used in this study.

Target sites	Primer name	Forward primers	Reverse primers
Endogenous KRAS	1st	CTTAAGCGTCGATGGAGGAG	CCCTGACATACTCCAAGGA
	2nd	ACACTCTTCCCTACAGGACGCTTTCCGATCTAGGCCGCTGAAATGACTG GTGTACGGTGGGGGGCTAT	GTGACTGGAGTTCAGACGTGTGCTCTCGATCTTGAAAATGGTCAGAGAACCC GTCCGCTCTCGAGGGTACTA
(HEK293T/17-KRAS library cells)	1st		GTGACTGGAGTTCAGACGTGTGCTCTCGATCTTGAAAATGGTCAGAGAACCC
	2nd	ACACTCTTCCCTACAGGACGCTTTCCGATCTTGCAAGCTAGTCCCAGCTA	GTGACTGGAGTTCAGACGTGTGCTCTCGATCTTGAAAATGGTCAGAGAACCC GTGACTGGAGTTCAGACGTGTGCTCTCGATCTTGAAAATGGTCAGAGAACCC
Universal pegRNA #2 OT1	2nd	ACACTCTTCCCTACAGGACGCTTTCCGATCTTGCAAGCTAGTCCCAGCTA	GTGACTGGAGTTCAGACGTGTGCTCTCGATCTTGAAAATGGTCAGAGAACCC GTGACTGGAGTTCAGACGTGTGCTCTCGATCTTGAAAATGGTCAGAGAACCC
Universal pegRNA #2 OT2	2nd	ACACTCTTCCCTACAGGACGCTTTCCGATCTTGCAAGCTAGTCCCAGCTA	GTGACTGGAGTTCAGACGTGTGCTCTCGATCTTGAAAATGGTCAGAGAACCC GTGACTGGAGTTCAGACGTGTGCTCTCGATCTTGAAAATGGTCAGAGAACCC
Universal pegRNA #2 OT3	2nd	ACACTCTTCCCTACAGGACGCTTTCCGATCTTGCAAGCTAGTCCCAGCTA	GTGACTGGAGTTCAGACGTGTGCTCTCGATCTTGAAAATGGTCAGAGAACCC GTGACTGGAGTTCAGACGTGTGCTCTCGATCTTGAAAATGGTCAGAGAACCC
Universal pegRNA #2 OT4	2nd	ACACTCTTCCCTACAGGACGCTTTCCGATCTTGCAAGCTAGTCCCAGCTA	GTGACTGGAGTTCAGACGTGTGCTCTCGATCTTGAAAATGGTCAGAGAACCC GTGACTGGAGTTCAGACGTGTGCTCTCGATCTTGAAAATGGTCAGAGAACCC
Universal pegRNA #2 OT5	2nd	ACACTCTTCCCTACAGGACGCTTTCCGATCTTGCAAGCTAGTCCCAGCTA	GTGACTGGAGTTCAGACGTGTGCTCTCGATCTTGAAAATGGTCAGAGAACCC GTGACTGGAGTTCAGACGTGTGCTCTCGATCTTGAAAATGGTCAGAGAACCC