Cell Reports Medicine, Volume 4

Supplemental information

Tissue-resident memory CAR T cells with

stem-like characteristics display enhanced

efficacy against solid and liquid tumors

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Supplementary Figure 1. Optimization of CAR-T_{RM} culture conditions. Related to Figure **1.** (**A**) Schematic of optimizing TGF-β concentration. (**B**) Frequencies of CD103⁺ and CD103⁺CD39⁺ CD8⁺ CAR T-cells with 0, 0.2, 1, 2, 10 ng/mL TGF-β added during CAR T-cell manufacturing. (**C**) Fold CAR T-cell expansion during manufacturing with TGF-β concentration as indicated in panel **A**. (**D**) Schematic of optimizing duration of TGF-β exposure. (**E**) Frequencies of CD103⁺ and CD103⁺CD39⁺ CD8⁺ CAR T-cells with TGF-β added at different timepoints during CAR T-cell manufacturing. (**F**) Fold CAR T-cell expansion during manufacturing with TGF-β added at different timepoints with TGF-β added on different timepoints as indicated in panel **D**. One-way ANOVA, n = 4 biological replicates. *P < 0.05, *P < 0.01, ***P < 0.001, ns.: not significant.







Supplementary Figure 2. Characterization of CAR-T_{RM} cells. Related to Figure 1.

(A) M5 CAR T-cells were isolated after three consecutive tumor challenges and co-cultured with Capan-2 target cells at an effector to target ratio of 1:2. Cytotoxic capacity at 7-hours post- co-culture is shown (Mann-Whitney test, n = 5 biological replicates). (B) Cytokine production after a second challenge with AsPC1 tumor cells. (C) Fold CAR T-cell expansion during a restimulation assay (Mann-Whitney test, n = 4 biological replicates). Experiments in panels B-C were conducted using CAR T-cells manufactured from different healthy donors. Figures display representative results from one donor. (D, E) M5 CAR T-cells were incubated with Capan-2 target cells at an effector to target ratio of 1:3 in presence or absence of 2 ng/mL TGF β . The experiments were performed using CAR T-cells generated from individual healthy donors/biological replicates. (D) Cytolytic activity of CAR T-cells 4-days post-coculture. (E) Effector cytokine production 24-hours after co-incubation (one-way ANOVA, n = 4 biological replicates). (F) Frequencies of FOXP3⁺ and FOXP3⁺CD25⁺ CAR T-cells after manufacturing (paired *t*-test, n = 3 biological replicates). *P < 0.05, *P < 0.01, ***P < 0.001, ns.: not significant.

MSigDB_Hallmark_2020



Supplementary Figure 3. Top pathways enriched in CAR- T_{RM} cells. Related to Figure 2.

Pathway analysis using Enrichr was carried out with genes upregulated in $CD8^+CAR-T_{RM}$ cells compared to $CD8^+CAR-T_{CONV}$ cells (MSigDB_Hallmark_2020).



Supplementary Figure 4. Single-cell gene expression analysis of CAR-T_{CONV} and CAR-T_{RM} cells. Related to Figure 3. (A) Violin plots displaying expression of cluster-defining markers in CD8⁺ CAR T-cells. (B) UMAP plots illustrating expression levels of the same markers in the CD8⁺ CAR T-cell population. (C) Module scores for 'stem-like' (TCF1⁺ memory CD8⁺ T-cells) and 'non-stem-like' (TCF1⁻ memory CD8⁺ T-cells) T-cell signatures were calculated for CD8 clusters and CAR T-cell samples (GSE83978).



Supplementary Figure 5. Sustained resident memory phenotype of CAR-T_{RM} cells during chronic antigen stimulation. Related to Figure 4. Anti-mesothelin CAR T-cells were serially challenged with AsPC1 tumor cells, and frequencies of resident memory (CD103⁺CD49a⁺CD8⁺) T-cells were assessed over time.





D

Proliferative Capacity



Supplementary Figure 6. Exhaustion features in CAR-T_{RM} cells. Related to Figure 5.

(**A**, **B**) Exhaustion scores are compared between CAR-T_{CONV} and CAR-T_{RM} in each CD8⁺ T-cell cluster. (**A**) PMID26123020 (**B**) PMID31802004. (**C**) ATAC-seq tracks of *PRDM1*, *ID2*, and *TOX2*. Differentially accessible regions are highlighted in blue. (**D**) M5 CAR T-cells were serially challenged with AsPC1 cells at an E:T ratio of 2:1. Fold CAR T-cell expansion after the third round of antigen stimulation is shown (two-way ANOVA, *n* = 3 biological replicates). *P < 0.05, *P < 0.01, ***P < 0.001, ns.: not significant.



Supplementary Figure 7. CAR- T_{RM} cell immunophenotype and antitumor efficacy in pancreatic and prostate cancer models. Related to Figure 6. (A-C) Capan-2 pancreatic cancer xenograft model: (A) NSG mice engrafted with 4×10^6 Capan-2 cells received M5 CAR-T_{CONV} and CAR-T_{RM} cells labeled with near-infrared (NIR) fluorescent dye intravenously on day 33 post-engraftment. NIR intensity was monitored over time (Unpaired t-test, n = 3 biological replicates). (B) Frequencies of hCD45⁺CD8⁺ cells in tumors expressing CCR7 and CD62L are shown (Mann-Whitney test, n = 7 biological replicates). (C) Tumor-infiltrating CAR T-cells were reactivated with phorbol 12-myristate 13-acetate and ionomycin, followed by measurement of effector molecule elaboration (Mann-Whitney test, n = 12 biological replicates). (D-E) Intraosseous PC3-PSMA prostate tumor model: (D) Male NSG mice intrafemorally engrafted with 2×10^5 PC3-PSMA cells received 1×10^5 anti-PSMA CAR-T_{CONV} or CAR-T_{RM} cells, or control CD19-targeting CAR T-cells (19BBz) intravenously on day 21 post-implantation (n = 7biological replicates per group). Tumor burden was assessed by bioluminescent imaging after intraperitoneal injection of luciferin and quantification with the Xenogen IVIS Imaging System (one-way ANOVA). (E) Kaplan-Meier survival curves show prolonged survival in the CAR-T_{RM} group compared to CAR-T_{CONV} (Log-rank test). *P < 0.05, **P < 0.01, ***P < 0.001, ns: not significant.