

Supporting Information for**De novo designed ice-binding proteins from twist-constrained helices**

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Section 1. Computational design details.

To generate helical bundle backbones we adapted a python script from Huang et al. (1) and uncouple major (ω_0) and minor (ω_1) twisting of the helices. Such that only straight helices of $\omega_0=0$ would be generated, with a fixed minor twist of $\omega_1 = 100, 99.2, 98.2$ or 97.2° per residue. After a hyperparameter scan to find optimal parameters for helix offset, phase, bundle radius we typically combinatorically generated 15,000-25,000 three-helix bundles backbones (see **supplementary Script 1** (params.dat) for bundle parameters). Each bundle was designed using two passes Rosetta fixed backbone layer design sequence (see **supplementary scripts** for details) and filtered using the packstat filter (2) with a cutoff of 0.3 to remove designs with poor hydrophobic packing in the protein interior. The designs that passed the filter are scored using the Rosetta REF2015 energy function (3). The top 250-500 scoring designs were selected for loop modelling. For each model Rosetta Remodel (4) using a custom blueprint was executed 500 times to design 2 or 3 residues to connect the 3 helices using the KIC protocol into a single chain bundle (see **supplementary scripts** for details). Approximately 30% loops resulted in closure. After loop modeling, designs were scored using REF2015 energy functional and the top 75-150 low scoring designs were selected. Rosetta FastRelax (5) was applied to backbones, and the Ca-RMSD to the unrelaxed model were calculated using PyRosetta (see **supplementary scripts** for details). Designs were again scored after FastRelax with REF2015 energy function, and selected by RMSD ($<1.5\text{\AA}$) and Rosetta score. Rosetta *ab initio* structure prediction was performed starting from primary sequence (see **supplementary scripts** for details). Designs that showed a clear folding funnel towards CA-RMSD $< 2.5\text{\AA}$ with a deep minima were selected for experimental characterization. All design and energy landscape calculations were performed on the WUR Anunna High Performance Computing cluster.

Section 2. Uniqueness of 11-mer for ice-binding.

Besides 11-mers, other (twist constrained) helices that produce straight alignment of threonines a limited number of alternative n-mers are possible (**Table S1**). For example, 7-mers with a helical twist of $\omega_1 = 102.9^\circ$ per residue will produce straight residue alignment, but requires large over-

twisting of +2.9° per residue. This likely leads to unstable bundles. Alternatively, 18-mer helices with $\omega_1 = 100^\circ$ per residue also produce aligned residues, but this will lead to extremely long helices. Moreover, 7-mer and 18-mer helices will have threonine spacings of 10.7 Å and 27.5 Å respectively, and these do not match the pyramidal ice lattice plane spacing, making 11-mers with $\omega_1 = 98.2^\circ$ per residue uniquely suitable for ice-binding.

Table. S1. Helix residue alignment table.

List of straight helix n-mers with associated twists that lead to alignment of residues (e.g. $n=7$ for 7-mer, $n=11$ for 11-mer). $\Delta\omega_1$: deviation the ideal helical twist (100° per residue). ω_1 : helical twist (° per residue) necessary to align two residues every n residues. m = cumulative rotation between the two aligned residues. t : number of rotations or turns. * indicates potential helical twists that can be constrained with ($\Delta\omega_1 < 3^\circ$ per residue) that can lead to aligned threonine residues ($n=7$, $n=11$ or $n=18$).

<i>n</i>	$\Delta\omega_1$ (° per residue)	ω_1 (° per residue)	<i>m</i> (°)	<i>t</i>
2	80	180.0	360	1
3	20	120.0	360	1
4	-10	90	360	1
5	44	144.0	720	2
6	20	120.0	720	2
*7	2.9	102.9	720	2
8	-10	90.0	720	2
9	20	120.0	1080	3
10	8	108.0	1080	3
*11	-1.8	98.2	1080	3
12	-10	90.0	1080	3
13	-16.9	83.1	1080	3
14	2.9	102.9	1440	4

15	-4.0	96.0	1440	4
16	-10	90.0	1440	4
17	5.9	105.9	1800	5
*18	0.0	100.0	1800	5
19	-5.3	94.7	1800	5
20	-10	90.0	1800	5

Section 3. Helix Twist in Type-I AFPs

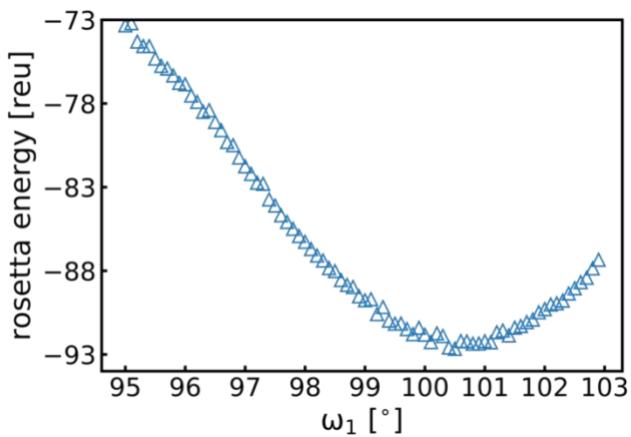


Fig. S1. Helix stability over minor helical twist. Single straight ($\omega_0=0$) alpha-helices of 44 residues were generated using a parametric design script modified from Huang et al. and designed using Rosetta fixed backbone application with amino acid sequence (TAAAAAAA)₄. Sequences were scored with the Rosetta energy function REF2015 with Rosetta energy units (reu) on the y-axis as a proxy for protein stability: lower reu score means higher stability. A clear minimum is observed at ~100-101° per residue for minor helical twisting of a straight helix.

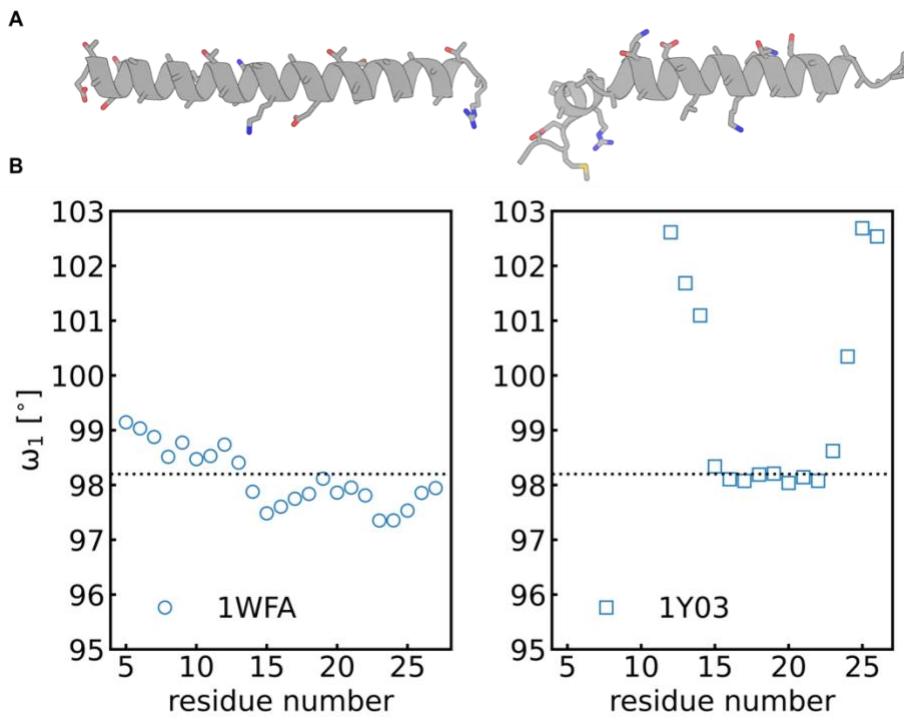


Fig. S2. Helical twisting for different various type-I ice-binding proteins. **A)** experimental structures of type-I ice-binding proteins. *Left*: x-ray crystal structure of wfAFP (pdb accession code: 1WFA) and *right* NMR solution structure of ssAFP (pdb accession code: 1Y03, state 1 shown). **B)** mean helical minor twist (ω_1) over 11 residues calculated with by HELANAL (6) in MDAnalysis (version: 0.20.1) vs. residue number. Dashed line shows expected ice-binding required twisting of $\omega_1 = 98.2^\circ$ per residue which leads to alignment of threonines. For ssAFP the N and C termini deviate from 98.2° per residue helical twist, likely due to flexibility.

Section 4. Energy landscape of TIP designs

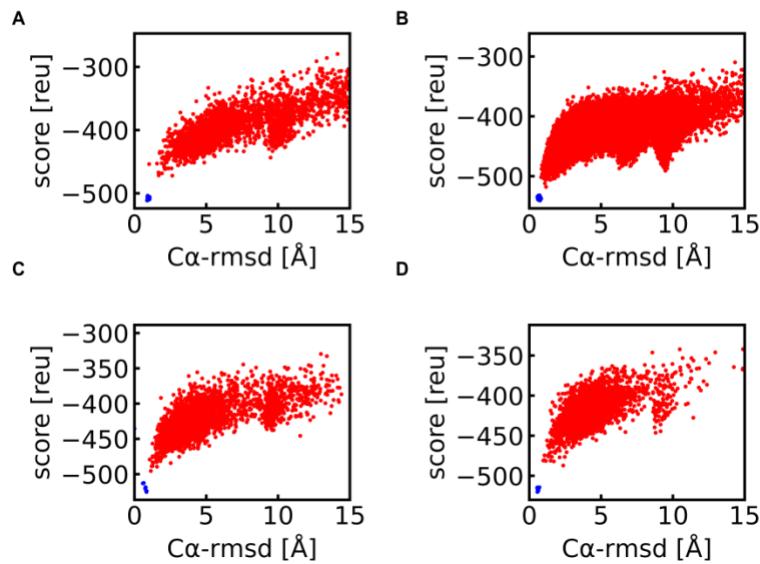


Fig. S3. Energy landscape of TIP designs. A) TIP-97, B) TIP-98, C) TIP-99_a, D) TIP-99_b. Score on y-axis shows the rosetta energy of the model in Rosetta energy units [reu] using the energy function REF2015(3) after folding from the extended chain using Rosetta AbinitioRelax simulations, and x-axis shows the C α -RMSD in [\AA] relative to the design model. Each red point represents an individual folding simulation starting with Rosetta AbinitioRelax. Blue points represent a Rosetta Relaxed model starting from the design model. Funnel-like plots which converge to a single deep minimum near C α -RMSD < 2.5 \AA likely demonstrate accurate folding of the design model.

Section 5. Comparison of Helical Twisting of TIP Models

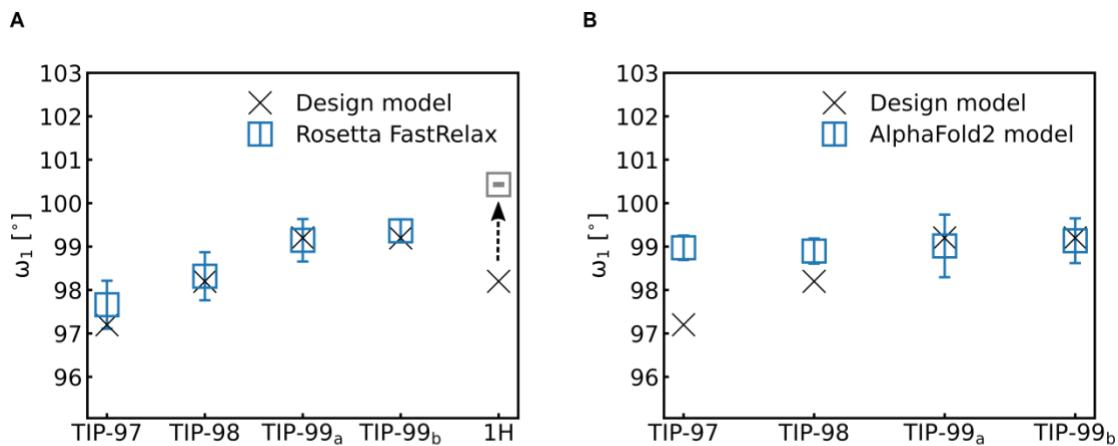


Fig. S4. Comparison helical twisting of ice-binding helix after flexible backbone Rosetta Relax. Y-axis shows the average helical minor twist (ω_1) of the ice-binding helix calculated by running average over 11 residues using HELANAL (6) in MDAnalysis (version: 0.20.1)

A) shows ω_1 for all characterized designs TIP-99_a, TIP-99_b, TIP-98 and TIP-97 after Rosetta Relax with flexible backbone. For all designs the twist of the ice-binding helix is maintained after relaxing with flexible backbone except for 1H which is a single alpha-helix $\omega_1=98.2^\circ$ per residue with sequence (TAAAAAAA)₄. As expected the 1H helix under-twisting rewinds to the optimal helix twisting of $\omega_1=100^\circ$ per residue because it is not constrained by supporting helices. **B**) shows ω_1 of the highest confidence (highest pLDTT ranking) AlphaFold2 model prediction. TIP-97 and TIP-98 show some deviation from the target twist the AF2 predictions. Error bars show standard deviation and X shows the designed ω_1 .

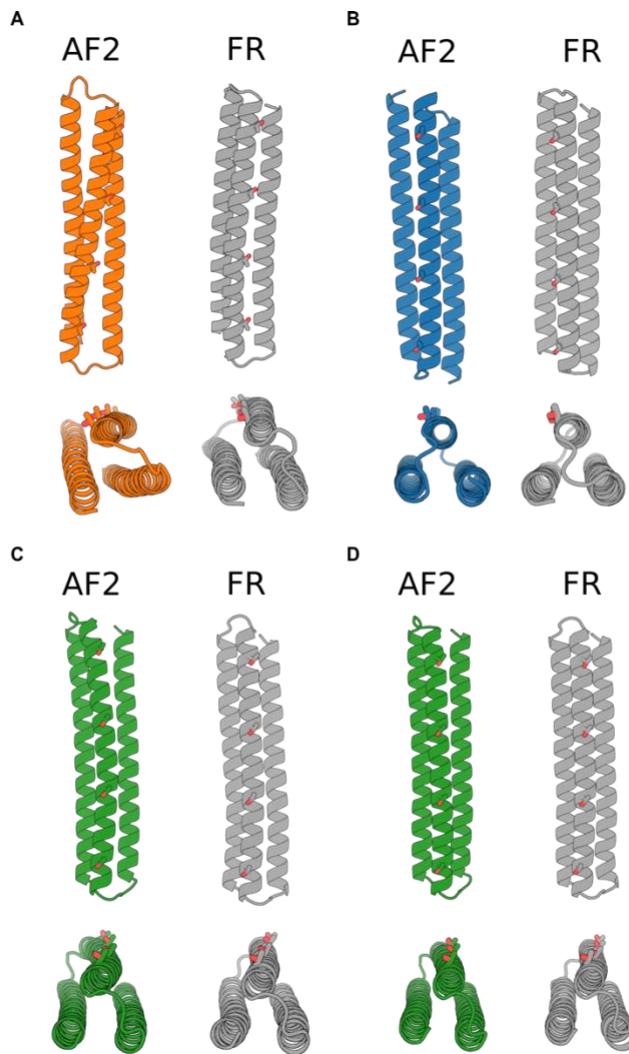


Fig. S5. AlphaFold2 predictions compared to Rosetta FastRelax models. Cartoons of **A)** TIP-97, **B)** TIP-98, **C)** TIP-99_a, **D)** TIP-99_b. AlphaFold2 (AF2) highest ranking pLDTT models (colored) predicted by AlphaFold 2 (7–11) are shown and compared to Rosetta FastRelax (FR) models (grey). Threonine residues on the ice-binding helix are shown as sticks. Noteably: TIP-97 is predicted by AF2 as a bundle with nonzero ω_0 major helix twist. And TIP-98 is predicted by AF2 to maintain the $\omega_1=98.2^\circ$ of the ice-binding helix, but has a different topology where the two stabilizing helices are reversed in order. TIP-99_a and TIP-99_b match the Rosetta relaxed structures well both in topology and twisting of the ice-binding helix.

Table S2. AlphaFold 2 per residue confidence scores (pLDDT) for the 5 parameter models.

A higher pLDDT score is better. For each TIP design, the highest ranking pLDTT model is underlined.

Design	Model 1	Model 2	Model 3	Model 4	Model 5
TIP-97	83.32	<u>84.60</u>	81.87	80.54	79.10
TIP-98	86.07	91.01	<u>92.79</u>	89.87	89.49
TIP-99 _a	90.46	<u>92.76</u>	91.29	90.47	89.63
TIP-99 _b	90.88	92.14	<u>92.20</u>	91.13	90.30

Section 6. Consensus Motif Type-I AFPs



Fig. S6. Logo diagram for 11-mer consensus motif computed from sequences of the type-I helical ice-binding proteins wfAFP and ssAFP. The dominant motif is TAAAAAA...AAA.

Section 7. TIP mutant studies

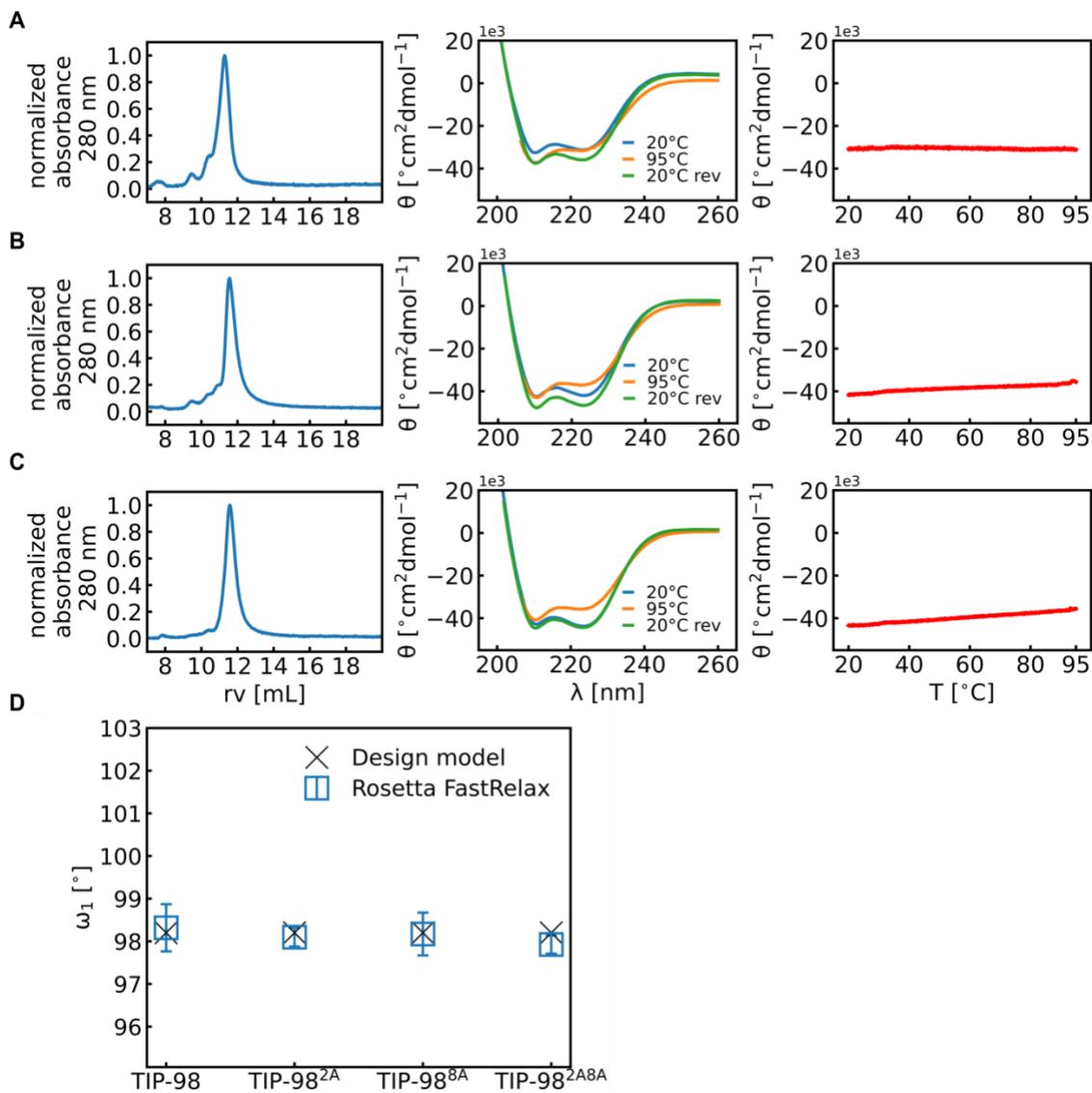


Fig. S7. Experimental characterization TIP-98 variants with expanded ice-binding motif. A) TIP-98^{2A} B) TIP-98^{8A} C) TIP-98^{2A8A} Left: size-exclusion chromatography on a Superdex 75 10/300 column in PBS+ (10 mM phosphate pH 7.4 + 300 mM NaCl), demonstrating proteins elute at the expected retention volume (rv) of ~12 mL, corresponding to a 17 kDa globular protein. Middle: circular dichroism spectra. Molar ellipticity (θ) versus wavelength (λ) at 20°C, 95°C and 20°C reversed (20°C rev) demonstrating alpha-helical secondary structure and high thermal melt with $T_m > 95^\circ\text{C}$, except for TIP-97. Right: molar ellipticity (θ) at 222 nm during temperature ramp from 20°C

to 95°C (1°C/min). D) Average helical minor twist (ω_1) of the ice-binding helix calculated by running average over 11 residues using HELANAL (6) after Rosetta Relax with flexible backbone.

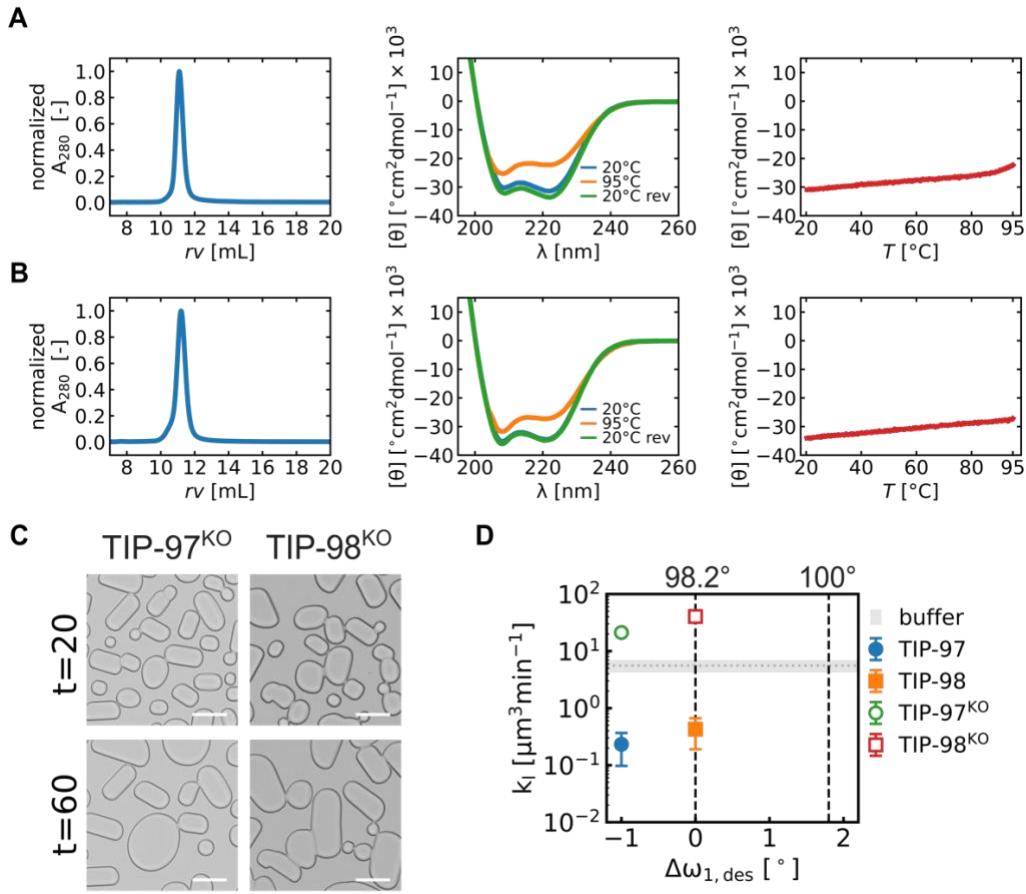


Fig. S8. Experimental characterization and activity of TIP knock out (KO) proteins. Biochemical characterization **A)** TIP-97^{KO}, **B)** TIP-98^{KO}, where ^{KO} designates that all 4 ice-binding threonines are mutated to glutamic acid. Left: size-exclusion chromatography on a Superdex 75 10/300 column in PBS+ (10 mM phosphate pH 7.4 + 300 mM NaCl), demonstrating proteins elute at the expected retention volume (rv) of ~12 mL, corresponding to a ~17 kDa globular protein. Middle: circular dichroism spectra. Molar ellipticity (θ) versus wavelength (λ) at 20°C, 95°C and 20°C reversed (20°C rev) demonstrating alpha-helical secondary structure and high thermal stability. Right: molar ellipticity (θ) at 222 nm during temperature ramp from 20°C to 95°C (1°C/min). **C)** Microscopy image of ice crystals at 20 min. (top) and after 60 min. (bottom) exposure to -7°C in presence of TIPs at 40 μM , demonstrating ice-recrystallization inhibition (IRI) for TIP-97^{KO} and TIP-

98^{KO} . Scale bar: 20 μ m. **D)** Quantitative IRI analysis. Speed of growth of ice-crystals [$\mu\text{m}^3\text{min}^{-1}$] during Ostwald ripening at a fixed protein concentration of 40 μM for TIP- 97^{KO} and TIP- 98^{KO} and 60 μM for TIP-97 and TIP-98 as a function of the deviation from proposed ideal twist angle.

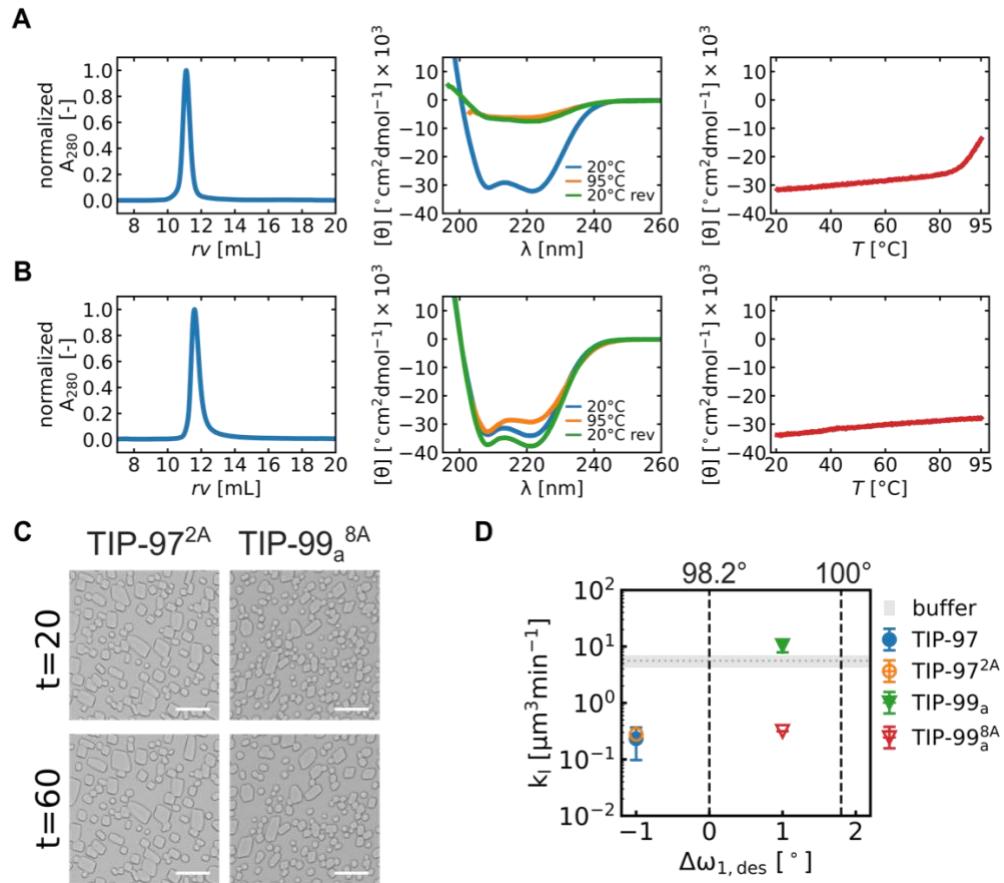


Fig. S9. Experimental characterization and activity of TIP alanine mutants. Biochemical characterization **A)** TIP- 97^{2A} , **B)** TIP- 99_a^{8A} . Left: size-exclusion chromatography on a Superdex 75 10/300 column in PBS+ (10 mM phosphate pH 7.4 + 300 mM NaCl), demonstrating proteins elute at the expected retention volume (rv) of ~12 mL, corresponding to a ~17 kDa globular protein. Middle: circular dichroism spectra. Molar ellipticity (θ) versus wavelength (λ) at 20°C, 95°C and 20°C reversed (20°C rev) demonstrating alpha-helical secondary structure and high thermal stability. Right: molar ellipticity (θ) at 222 nm during temperature ramp from 20°C to 95°C (1°C/min). **C)** Microscopy image of ice crystals at 20 min. (top) and after 60 min. (bottom) exposure to -7°C in presence of TIPs at 60 μM , demonstrating ice-recrystallization inhibition (IRI) for TIP- 97^{2A} and TIP-

99_a^{8A} . Scale bar: 20 μ m. **D)** Quantitative IRI analysis. Speed of growth of ice-crystals [$\mu\text{m}^3\text{min}^{-1}$] during Ostwald ripening at a fixed protein concentration of 60 μ M for TIP-97^{2A} and TIP- 99_a^{8a} , TIP-97 and TIP- 99_a and 60 μ M as a function of the deviation from proposed ideal twist angle.

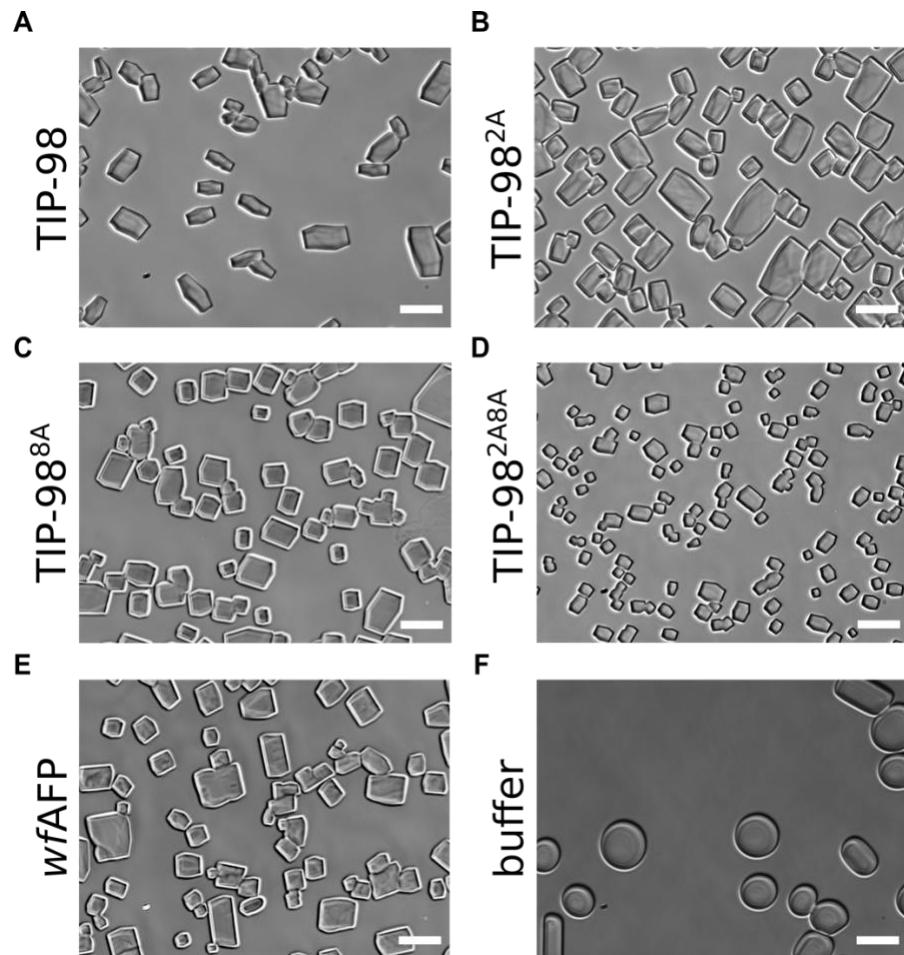


Fig. S10. Full views of ice shaping experiment. Images of blunt-end ice crystals after 1 minute of cooling with 0.2°C per minute (from -4.3 °C to -4.5 °C) in buffer PBS+ (10 mM phosphate pH 7.4 + 300 mM NaCl) at 50 μ M protein concentration. Scale bar: 50 μ m.

Section 8. Protein sequences

Table S3. proteins sequences of experimentally characterized proteins.

Capitalized sequence is the designed sequence. N-terminally a methionine is added and C-terminally a short serine-glycine linker followed by a tryptophan for extinction at 280nm and a histidine tag for affinity purification. For the variants of TIP-98^{2A}, TIP-98^{8A}, TIP-98^{2A8A}, the alanine mutations are underlined.

Design name	Protein sequence
TIP-99 _a	mEEEAKKKIDDLLTKARREVKKAIKTAREVAKRASKKIEELERRNEDKEAA ATKMEAILRAVKTTMKALIEALRTQMQKAACAMKTIVKAEPSEELKKV DAIKDMRRLVEEAIREMEKLARELEKQAREAQKRTsggwhhhhhh
TIP-99 _b	mEEEVREKLKRMEKKFDDSLKAERKIREIIKEAEKKLKTLLKKRNGPYEAV VTTLRAILKAVETKIRAIKALKTELDALIKAMETILKAHDKNDELKKEVE DIIKKMRDKLTKLIRKAKELLDRLLKKAKKVQDETsggwhhhhhh
TIP-98	mEEEALKKLKDTVKEALKRLKELVDRAKKLKETVKRAEELKKLGDEAT ITKAKAKLRAIVTCAEADLRALVTKAEEAKLKAIVTEASANGVSEEALERLE RILREALKRLKKILKEALERLKKILKTAERLDRNsggwhhhhhh
TIP-97	mKEEFEEKKARTLYKRFKTEMRRITTAERQLTKTARKILTDLRRKKDGGEA DLTKAKATLKAQITTIARFRARATKLIAQINAELTKDVAKMPVDEEMQKR LEKILKLTLEETARRLLKTMTRMQQTLEKIITTMDREsggwhhhhhh
TIP-98 ^{2A}	mEEEALKKLKDTVKEALKRLKELVDRAKKLKETVKRAEELKKLGDEAT <u>ITA</u> AKAKLRAIV <u>TAA</u> EADLRALV <u>TAA</u> EAKLKAIV <u>TA</u> ASANGVSEEALERLE RILREALKRLKKILKEALERLKKILKTAERLDRNsggwhhhhhh
TIP-98 ^{8A}	mEEEALKKLKDTVKEALKRLKELVDRAKKLKETVKRAEELKKLGDA <u>AT</u> ITKAKAK <u>LAA</u> IVTCAEAD <u>LAA</u> LVTKAEAK <u>LAA</u> IVTEASANGVSEEALERLE RILREALKRLKKILKEALERLKKILKTAERLDRNsggwhhhhhh
TIP-98 ^{2A8A}	mEEEALKKLKDTVKEALKRLKELVDRAKKLKETVKRAEELKKLGDA <u>AT</u> <u>ITA</u> AKAK <u>LAA</u> IV <u>TAA</u> EAD <u>LAA</u> LV <u>TAA</u> EAK <u>LAA</u> IV <u>TA</u> ASANGVSEEALERLE RILREALKRLKKILKEALERLKKILKTAERLDRNsggwhhhhhh

TIP-97 ^{KO}	mKEEFEKKARTLYKRFKTEMDRRITTAERQLTKTARKILTDLRRKKDGGEA DLEKAKATLKAQIETIIARFRARAEKLIAQINAELDVAKMPVDEEMQKR LEKILKLEETARRLLKMTTRMQQTLEKIITTMDREsggwhhhhhh
TIP-98 ^{KO}	mEEEALKLKDTVKEALKRLKELVDRALKKLKVRAEKLKLGKDEAT IEKAKAKLRAIVEKAADLRALVEKAEAKLKAIVEEASANGVSEEALERLE RILREALKRLKKILKEALERLKKILKTAERLDRNsggwhhhhhh
TIP-97 ^{2A}	mKEEFEKKARTLYKRFKTEMDRRITTAERQLTKTARKILTDLRRKKDGGEA DLTA <u>A</u> KATLKAQIT <u>A</u> IIARFRAR <u>A</u> LIAQINAEL <u>A</u> DVAKMPVDEEMQKR LEKILKLEETARRLLKMTTRMQQTLEKIITTMDREsggwhhhhhh
TIP-99 _a ^{8A}	mEEEVREKLKRMEKKFDDSLEKAERKIREIIKEAEKKLKT <u>L</u> KKRNGPY <u>A</u> V VTTLRAILAAVETKIRAI <u>I</u> AALKTELDALIA <u>A</u> METILKAHDKN <u>E</u> LKKEVE DIKKMRDKLT <u>K</u> IRKAKELL <u>D</u> RLKKAKKVQDETsggwhhhhhh

Section 9. Crystallographic data

Table S4. Crystallographic data collection and refinement statistics

Design	TIP-99 _a
Resolution range	49.94 - 2.35 (2.434 - 2.35)
Space group	P 21 21 21
Unit cell	54.582 67.427 74.326 90 90 90
Total reflections	157388 (16229)
Unique reflections	11928 (1172)
Multiplicity	13.2 (13.8)
Completeness (%)	99.62 (100.00)
Mean I/sigma(I)	13.80 (1.55)
Wilson B-factor	60,27
R-merge	0.1145 (1.379)
R-meas	0.1193 (1.431)
R-pim	0.03301 (0.3815)
CC1/2	0.999 (0.753)
CC*	1 (0.927)
Reflections used in refinement	11887 (1173)
Reflections used for R-free	553 (49)
R-work	0.2610 (0.3181)
R-free	0.3189 (0.3946)
CC (work)	0.964 (0.575)
CC (free)	0.930 (0.316)
macromolecules	2165
RMS(bonds)	0,002
RMS(angles)	0,39

Ramachandran favored (%)	97,74
Ramachandran allowed (%)	2,26
Ramachandran outliers (%)	0
Rotamer outliers (%)	0,43
Average B-factor	83,35
macromolecules	83,35

Section 10. Design and Analysis Scripts

Script S1. Parametric bundle backbone scripts

Example of parameters (params.dat) used to generate ~17,500 TIP-99 helices. params.dat adapted from Huang et al. (1) Python (generate_helix.py) script adapted from Huang et al.(1) to generate helices edited such that in the generation of helices major superhelical (ω_0) and minor helix twist (ω_1) are de-coupled, and $\omega_1=98.2^\circ$ per residue for all helices.

```
params.dat

TIP-99          # base name of output structures
44             # number of residues per helix
6.0 7.0 4      # supercoil radius range (low, high, number of samples)
3              # number of helices
3              # number of helices in asymmetric unit
1 -1 1         # orientation of each helix in asymmetric unit
200 250 9      # phase of first helix (low, high, number of samples)
170 200 3      # phase of second helix (low, high, number of samples)
200 250 9      # phase of third helix (low, high, number of samples)
-3.0 3.0 6     # z-offset of first helix (low, high, number of samples)
0.0 0.0 1       # z-offset of second helix (low, high, number of
samples)
-3.0 3.0 3     # z-offset of third helix (low, high, number of samples)
A B C          # chain names
0 1 2          # chain order

generate_helix.py (for generation of helices with  $\omega_1=99.2^\circ$  per residue)

#####Start of script#####
#!/usr/bin/env python
from os import system,popen
import string
from sys import argv
import math
from math import sin,cos,sqrt
def isint(x): return type(x) is int
def isfloat(x): return type(x) is float
def isnum(x): return isint(x) or isfloat(x)
def isiter(x): return hasattr(x,"__iter__")
def islist(x): return type(x) is list
def istuple(x): return type(x) is tuple
def isvec(x): return type(x) is Vec
def ismat(x): return type(x) is Mat
def isxform(x): return type(x) is Xform
class Vec(object):
    """a Vector like xyzVector<Real> in rosetta
    >>> v = Vec(1,2,3)
    >>> print v, 10*v
    (1.000000,2.000000,3.000000) (10.000000,20.000000,30.000000)
    multiplication is a dot prod at the moment
```

```

>>> v*v
14.0
>>> assert Vec(1,0,-0) == Vec(1,-0,0)
"""

def __init__(self,x=0.0,y=None,z=None):
    if y is None:
        if isnum(x):
            self.x,self.y,self.z = (float(x),)*3
        elif isvec(x):
            self.x,self.y,self.z = x.x,x.y,x.z
        elif isiter(x):
            i = iter(x)
            self.x,self.y,self.z =
i.next(),i.next(),i .next()
            else: raise NotImplementedError
        elif z is not None:
            assert isnum(x) and isnum(y) and isnum(z)
            self.x,self.y, self .z = float(x),float (y),
float (z)
            else: raise NotImplementedError
        assert isfloat(self .x)
        assert isfloat(self .y)
        assert isfloat(self .z)
    def dot(u,v):      assert isvec(v); return
u.x*v.x+u.y*v.y+u.z*v.z
    def normdot(u,v): assert isvec(v); return min(1.0,max(-
1.0,u.dot(v)/u.length()/v.length()))
    def angle(u,v):
        assert isvec(v)
        d = u.normdot(v)
        if d > 1.0-EPS: return 0.0;
        if d < EPS-1.0: return pi
        return acos(d)
    def angle_degrees(u,v): return degrees(u.angle(v))
    def lineangle(u,v):
        assert isvec(v);
        if u.length() < SQRTEPS or v.length < SQRTEPS: return
0.0
        ang = absacos( u.normdot(v) )
        return ang if ang < pi/2.0 else pi-ang
    def lineangle_degrees(u,v):
        if isvec(v): return degrees(u.lineangle(v))
        raise NotImplementedError
    def length(u):      return sqrt(u.dot(u))
    def length_squared(u): return      u.dot(u)
    def distance(u,v):      assert isvec(v); return (u-
v).length()
    def distance_squared(u,v): assert isvec(v); return (u-
v).length_squared()
    def cross(u,v): assert isvec(v); return Vec(u.y*v.z-
u.z*v.y,u.z*v.x-u.x*v.z,u.x*v.y-u.y*v.x)
    def __mul__(u,a):
        if isnum(a): return Vec(u.x*a,u.y*a,u.z*a)
        elif isvec(a): return u.dot(a)
        else: return a.__rmul__(u)
    def __rmul__(u,a): return u*a
    def __add__(u,v):

```

```

        if isvec(v): return Vec(u.x+v.x,u.y+v.y,u.z+v.z)
        return v.__radd__(u)
    def __sub__(u,v):
        if isvec(v): return Vec(u.x-v.x,u.y-v.y,u.z-v.z)
        return v.__rsub__(u)
    def __neg__(u): return Vec(-u.x,-u.y,-u.z)
    def __div__(u,a): return u*(1.0/a)
    def __str__(self): return "%f,%f,%f"%(self.x,self.y,self.z)
    def __repr__(self): return "Vec( %f, %f, %f
)%(self.x,self.y,self.z)
    def normalize(u):
        l = u.length()
        u.x /= l
        u.y /= l
        u.z /= l
    def normalized(u):
        v = Vec(u)
        v.normalize()
        return v
    def outer(u,v):
        assert isvec(v)
        return Mat( u.x*v.x, u.x*v.y, u.x*v.z,
                    u.y*v.x, u.y*v.y,
                    u.y*v.z,
                    u.z*v.x, u.z*v.y,
                    u.z*v.z      )
    def __eq__(self,other):
        assert isvec(other)
        return ( abs(self.x-other.x) < EPS and
                abs(self.y-other.y) < EPS and
                abs(self.z-other.z) < EPS )
    def rounded(self,sd):
        return Vec(round(self.x,sd), round(self.y,sd),
round(self.z,sd) )
    def unit(v):
        if abs(v.x) > SQRTEPS: return v/v.x
        elif abs(v.y) > SQRTEPS: return v/v.y
        elif abs(v.z) > SQRTEPS: return v/v.z
    def __len__(v):
        return 3
    def abs(v):
        return Vec(abs(v.x),abs(v.y),abs(v.z))
    def __getitem__(v,i):
        if i is 0: return v.x
        if i is 1: return v.y
        if i is 2: return v.z
        raise IndexError
    def tuple(v):
        return (v.x,v.y,v.z)
    def key(v):
        return v.abs().unit().rounded(6).tuple()
Ux = Vec(1,0,0)
Uy = Vec(0,1,0)
Uz = Vec(0,0,1)
V0 = Vec(0,0,0)
def randvec(n=None):
    if n is None: return Vec(gauss(0,1),gauss(0,1),gauss(0,1))

```

```

        return [Vec(gauss(0,1),gauss(0,1),gauss(0,1)) for i in
range(n)]
def randveccube(r=1.0):
    return Vec(uniform(-1,1),uniform(-1,1),uniform(-1,1))*r
def randvecball(r=1.0):
    v =randveccube(r)
    while v.length_squared() > r*r: v = randveccube(r)
    return v
def randnorm(n=None):
    """
    >>> assert abs(randnorm().length()-1.0) < 0.0000001
    """
    if n is None: return randvec().normalized()
    return (randvec().normalized() for i in range(n))
def coplanar(x1,x2,x3,x4):
    """
    >>> u,v,w = randvec(3)
    >>> a,b,c = (gauss(0,10) for i in range(3))
    >>> assert      coplanar(u, v, w, u + a*(u-v) + b*(v-w) + c*(w-
u) )
    >>> assert not coplanar(u, v, w, u + a*(u-v) + b*(v-w) + c*(w-
u) + randvec().cross(u-v) )
    """
    return abs((x3-x1).dot((x2-x1).cross(x4-x3))) < SQRTEPS
def rmsd(l,m):
    """
    >>> l,m = randvec(6),randvec(6)
    >>> rmsd(l,l)
    0.0
    """
    rmsd = 0.0
    for u,v in izip(l,m):
        rmsd += u.distance_squared(v)
    return sqrt(rmsd)
class Mat(object):
    """docstring for Mat
    >>> m = Mat(2,0,0,0,1,0,0,0,1)
    >>> print m
    Mat[ (2.000000,0.000000,0.000000),
(0.000000,1.000000,0.000000), (0.000000,0.000000,1.000000) ]
    >>> print m*m
    Mat[ (4.000000,0.000000,0.000000),
(0.000000,1.000000,0.000000), (0.000000,0.000000,1.000000) ]
    >>> print Mat(*range(1,10)) * Mat(*range(10,19))
    Mat[ (84.000000,90.000000,96.000000),
(201.000000,216.000000,231.000000), (318.000000,342.000000,366.000000)
]
    >>> assert Mat(0.0,1.0,2.0,3,4,5,6,7,8) == Mat(-
0,1,2,3,4,5.0,6.0,7.0,8.0)
    >>> print Mat(100,2,3,4,5,6,7,8,9).det()
-297.0
    >>> m = Mat(100,2,3,4,5,6,7,8,9)
    >>> assert m * ~m == Iimat
    """
    def __init__(self, xx=None, xy=None, xz=None, yx=None, yy=None,
yz=None, zx=None, zy=None, zz=None):
        super(Mat, self).__init__()

```

```

        if xx is None: # identity default
            self.xx, self.xy, self.xz = 1.0,0.0,0.0
            self.yx, self.yy, self.yz = 0.0,1.0,0.0
            self.zx, self.zy, self.zz = 0.0,0.0,1.0
        elif xy is None and ismat(xx):
            self.xx, self.xy, self.xz = xx.xx, xx.xy, xx.xz
            self.yx, self.yy, self.yz = xx.yx, xx.yy, xx.yz
            self.zx, self.zy, self.zz = xx.zx, xx.zy,
xx.zz
        elif yx is None and isvec(xx) and isvec(xy) and
isvec(xz):
            self.xx, self.xy, self.xz = xx.x, xy.x, xz.x
            self.yx, self.yy, self.yz = xx.y, xy.y, xz.y
            self.zx, self.zy, self.zz = xx.z, xy.z, xz.z
        elif isnum(xx):
            self.xx, self.xy, self.xz = float(xx),
float(xy), float(xz)
            self.yx, self.yy, self.yz = float(yx),
float(yy), float(yz)
            self.zx, self.zy, self.zz = float(zx),
float(zy), float(zz)
        else:
            raise NotImplementedError
        assert isfloat(self.xx) and isfloat(self.xy) and
isfloat(self.xz)
        assert isfloat(self.yx) and isfloat(self.yy) and
isfloat(self.yz)
        assert isfloat(self.zx) and isfloat(self.zy) and
isfloat(self.zz)
    def row(m,i):
        assert isint(i)
        if i is 0: return Vec(m.xx,m.xy,m.xz)
        elif i is 1: return Vec(m.yx,m.yy,m.yz)
        elif i is 2: return Vec(m.zx,m.zy,m.zz)
        else: assert 0 <= i and i <= 2
    def col(m,i):
        assert isint(i)
        if i is 0: return Vec(m.xx,m.yx,m.zx)
        elif i is 1: return Vec(m.xy,m.yy,m.zy)
        elif i is 2: return Vec(m.xz,m.yz,m.zz)
        else: assert 0 <= i and i <= 2
    def rowx(m): return m.row(0)
    def rowy(m): return m.row(1)
    def rowz(m): return m.row(2)
    def colx(m): return m.col(0)
    def coly(m): return m.col(1)
    def colz(m): return m.col(2)
    def __invert__(m):
        """
        >>> from random import random
        >>> for i in range(10):
        ...     m =
random()*Mat(random(),random(),random(),random(),random(),random(),
random(),random())
        ...
        assert ~m*m == Iimat
        ...
        assert m*~m == Iimat
        """

```



```

                abs(self.yx-other.yx) < EPS and
                abs(self.yy-other.yy) < EPS and
                abs(self.yz-other.yz) < EPS and
                abs(self.zx-other.zx) < EPS and
                abs(self.zy-other.zy) < EPS and
                abs(self.zz-other.zz) < EPS )

def rotation_axis(R):
    """
    >>> axis ,ang = randnorm(),uniform(-pi,pi)
    >>> axis2,ang2 =
rotation_matrix(axis,ang).rotation_axis()
    >>> assert abs( abs(ang) - abs(ang2) ) < EPS
    >>> assert axis == axis2 * copysign(1,ang*ang2)
    """
    cos_theta = sin_cos_range((R.trace()-1.0)/2.0);
    if cos_theta > -1.0+EPS and cos_theta < 1.0-EPS:
        x = ( 1.0 if R.zy > R.yz else -1.0 ) * sqrt(
max(0.0, ( R.xx - cos_theta ) / ( 1.0 - cos_theta ) ) )
        y = ( 1.0 if R.xz > R.zx else -1.0 ) * sqrt(
max(0.0, ( R.yy - cos_theta ) / ( 1.0 - cos_theta ) ) )
        z = ( 1.0 if R.yx > R.xy else -1.0 ) * sqrt(
max(0.0, ( R.zz - cos_theta ) / ( 1.0 - cos_theta ) ) )
        theta = acos( cos_theta );
        assert abs( x*x + y*y + z*z - 1 ) <= 0.01
        return Vec(x,y,z),theta
    elif cos_theta >= 1.0-EPS: return Vec(1.0,0.0,0.0),0.0
    else:
        nnT = (R+Imat)/2.0
        x,y,z = 0.0,0.0,0.0;
        if nnT.xx > EPS:
            x = sqrt( nnT.xx )
            y = nnT.yx / x
            z = nnT.zx / x
        elif nnT.yy > EPS:
            x = 0
            y = sqrt(nnT.yy)
            z = nnT.zy / y
        else:
            assert( nnT.zz > EPS );
            x = 0
            y = 0
            z = sqrt( nnT.zz )
            assert abs( x*x + y*y + z*z - 1.0 ) <= 0.01
            return Vec( x, y, z ),pi
def euler_angles(self):
    FLOAT_PRECISION = 1e-5
    if self.zz >= 1 - FLOAT_PRECISION:
        e1 = math.acos( sin_cos_range( self.xx ) )
        e2 = 0.0
        e3 = 0.0
        return Vec(e1,e2,e3)
    if self.zz <= -1 + FLOAT_PRECISION:
        e1 = math.acos( sin_cos_range( self.xx ) )
        e2 = 0.0
        e3 = math.pi
        return Vec(e1,e2,e3)

```

```

        pos_sin_theta = math.sqrt( 1 - self.zz*self.zz ) #
sin2theta = 1 - cos2theta.
        # two values are possible here : my convention is
to use positive theta only .
        # corresponding theta between [0, pi /2] -> [0,90]
since st > 0
        # and asin returns value between [-pi /2, pi /2]
e3 = math.asin( pos_sin_theta )
        # decide whether the actual positive theta is
between [ pi /2, pi [ using the value of cos ( theta )
        # which happens to be the matrix element self . zz
(and is thus signed ) .
if self.zz < 0:
        e3 = math.pi - e3
# e1 = math.atan2( -UU(1,3), UU(2,3) )      # between -
Pi and Pi -> [-180,180]
        # e2 = math.atan2( UU(3,1), UU(3,2) )      # between -
Pi and Pi -> [-180, 180]
        # this is atan( sin_phi * c , cos_phi * c ) as
opposed to Alex ' s atan ( -sin_phi * c, -cos_phi * c ) .
e1 = math.atan2( self.zx, -self.zy )
e2 = math.atan2( self.xz, self.yz )
return Vec(e1,e2,e3)
def from_euler_angles(self,euler):
    phi   = euler.x()
    psi   = euler.y()
    theta = euler.z()
    cos_phi   = math.cos( phi )
    sin_phi   = math.sin( phi )
    cos_psi   = math.cos( psi )
    sin_psi   = math.sin( psi )
    cos_theta = math.cos( theta )
    sin_theta = math.sin( theta )
    self.xx =  cos_psi * cos_phi - cos_theta * sin_phi *
sin_psi
    self.xy =  cos_psi * sin_phi + cos_theta * cos_phi *
sin_psi
    self.xz =  sin_psi * sin_theta
    self.yx = -sin_psi * cos_phi - cos_theta * sin_phi *
cos_psi
    self.yy = -sin_psi * sin_phi + cos_theta * cos_phi *
cos_psi
    self.yz =  cos_psi * sin_theta
    self.zx =  sin_theta * sin_phi
    self.zy = -sin_theta * cos_phi
    self.zz =          cos_theta
    return self
Imat = Mat(1,0,0,0,1,0,0,0,1)
class Xform(object):
    """Coordinate frame like rosetta Xform, behaves also as a
rosetta Stub
    >>> x = Xform(R=Imat,t=Uz)
    >>> print x
    Xform( Mat[ (1.000000,0.000000,0.000000),
(0.000000,1.000000,0.000000), (0.000000,0.000000,1.000000) ],
(0.000000,0.000000,1.000000) )
    >>> assert (x*x) == Xform(R=Imat,t=2*Uz)

```

```

>>> x =
Xform(R=rotation_matrix_degrees(Vec(1,0,0), 90.0), t=Vec(0,0,0))
    >>> print x
    Xform( Mat[ (1.000000,0.000000,0.000000), (0.000000,0.000000,-
1.000000), (0.000000,1.000000,0.000000) ], (0.000000,0.000000,0.000000)
)
    >>> assert x*x*x*x == Ixform
    >>> x.t = Ux
    >>> assert x*x*x*x == Xform(R=Iimat, t=4*Ux)
    >>> x.t = Uz
    >>> print x
    Xform( Mat[ (1.000000,0.000000,0.000000), (0.000000,0.000000,-
1.000000), (0.000000,1.000000,0.000000) ], (0.000000,0.000000,1.000000)
)
    >>> assert x == Xform(R=rotation_matrix_degrees(Ux, 90.0), t=Vec(0, 0, 1))
    >>> assert x*x ==
Xform(R=rotation_matrix_degrees(Ux, 180.0), t=Vec(0, -1, 1))
    >>> assert x*x*x ==
Xform(R=rotation_matrix_degrees(Ux, 270.0), t=Vec(0, -1, 0))
    >>> assert x*x*x*x ==
Xform(R=rotation_matrix_degrees(Ux, 0.0), t=Vec(0, 0, 0))
    >>> assert x*x*x*x*x ==
Xform(R=rotation_matrix_degrees(Ux, 90.0), t=Vec(0, 0, 1))
    >>> assert x*x*x*x*x*x ==
Xform(R=rotation_matrix_degrees(Ux, 180.0), t=Vec(0, -1, 1))
    >>> assert x*x*x*x*x*x*x ==
Xform(R=rotation_matrix_degrees(Ux, 270.0), t=Vec(0, -1, 0))
    >>> assert x*x*x*x*x*x*x*x ==
Xform(R=rotation_matrix_degrees(Ux, 0.0), t=Vec(0, 0, 0))
    >>> x =
Xform(rotation_matrix_degrees(Vec(1,2,3), 123), Vec(5,7,9))
    >>> assert ~x * x == Ixform
    >>> assert x * ~x == Ixform
    Frames / RTs are interchangable:
    >>> fr = Xform(rotation_matrix_degrees(Vec(1,2,3),
65.64), t=Vec(3,2,1))
    >>> to =
Xform(rotation_matrix_degrees(Vec(7,5,3), 105.44), t=Vec(10,9,8))
    >>> x = to/fr
    >>> assert to/Ixform == to
    >>> assert Ixform/fr == ~fr
    >>> assert (to * ~fr) * fr == to
    >>> assert x * fr == to
    >>> a1 = randnorm()
    >>> b1 = randnorm()
    >>> ang = uniform(0,1)*360.0-180.0
    >>> a2 = rotation_matrix_degrees(a1.cross(randnorm()), ang) * a1
    >>> b2 = rotation_matrix_degrees(b1.cross(randnorm()), ang) * b1
    >>> assert abs(angle(a1,a2) - angle(b1,b2)) < EPS
    >>> xa = Xform().from_two_vecs(a1,a2)
    >>> xb = Xform().from_two_vecs(b1,b2)
    >>> assert xa.tolocal(a1) == xb.tolocal(b1)
    >>> assert xa.tolocal(a2) == xb.tolocal(b2)
    >>> assert ~xa*a1 == ~xb*b1
    >>> assert ~xa*a2 == ~xb*b2
    >>> assert xb/xa*a1 == b1

```

```

>>> assert xb/xa*a2 == b2
add/sub with Vecs:
>>> X = randxform()
>>> u,v = randvec(2)
>>> assert isxform(u+X) and isxform(X+u) and isxform(u-X) and
isxform(X-u)
>>> assert X*v+u == (u+X)*v
>>> assert X*(v+u) == (X+u)*v
>>> assert Xform(u)*X*v == (u+X)*v
>>> assert X*Xform(u)*v == (X+u)*v
>>> assert X*v-u == (u-X)*v
>>> assert X*(v-u) == (X-u)*v
mul,div with Mats:
>>> R = randrot()
>>> assert isxform(R*X) and isxform(X*R)
>>> assert R*X*u == (R*X)*u == R*(X*u)
>>> assert X*R*u == (X*R)*u == X*(R*u)
>>> assert Xform(R)*X*u == Xform(R)*(X*u)
>>> assert X*Xform(R)*u == X*(Xform(R,V0)*u)
>>> assert X/X*v == v
mul/div Xforms:
>>> Y = randxform()
>>> assert isxform(X/Y) and isxform(X*Y)
>>> assert X/Y*v == X~Y*v
these don't work yet:
>>> axis,ang,cen = randnorm(),uniform(-pi,pi),randvec()
#doctest: +SKIP
    >>> X = rotation_around(axis,ang,cen)
#doctest: +SKIP
    >>> axis2,ang2,cen2 = X.rotation_center()
#doctest: +SKIP
    >>> assert abs( abs(ang) - abs(ang2) ) < EPS
#doctest: +SKIP
    >>> assert axis == axis2 * copysign(1,ang*ang2)
#doctest: +SKIP
    >>> print cen
#doctest: +SKIP
    >>> print cen2
#doctest: +SKIP
    >>> x = Xform( Mat( Vec( 0.816587, -0.306018,
0.489427 ), Vec( 0.245040, 0.951487, 0.186086 ), Vec( -0.522629, -
0.032026, 0.851959 ) ), Vec( 1.689794, 1.535762, -0.964428 ) )
    >>> assert repr(x) == "Xform( Mat( Vec( 0.816587, -0.306018,
0.489427 ), Vec( 0.245040, 0.951487, 0.186086 ), Vec( -0.522629, -
0.032026, 0.851959 ) ), Vec( 1.689794, 1.535762, -0.964428 ) )"
    """
def __init__(self, R=None, t=None):
    super(Xform, self).__init__()
    if isvec(R) and t is None: R,t = Iimat,R
    self.R = R if R else Iimat
    self.t = t if t else V0
    assert ismat(self.R) and isvec(self.t)
def from_four_points(s,cen,a,b,c):
    s.t = cen
    e1 = (a-b).normalized()
    e3 = e1.cross(c-b).normalized()
    e2 = e1.cross(e3).normalized()

```

```

        s.R = Mat(e1.x,e2.x,e3.x,e1.y,e2.y,e3.y,e1.z,e2.z,e3.z)
        return s
    def from_two_vecs(s,a,b):
        e1 = a.normalized()
        e2 = projperp(a,b).normalized()
        e3 = e1.cross(e2)
        return Xform(
            Mat(e1.x,e2.x,e3.x,e1.y,e2.y,e3.y,e1.z,e2.z,e3.z),V0)
    def tolocal(s,x): return s.R.transposed() * (x - s.t)
    def toglobal(s,x): return (s.R * x) + s.t
    def __invert__(self):
        R = ~self.R
        t = R * -self.t
        return Xform(R,t)
    def __mul__(X,o):
        if isvec(o): return X.R * o + X.t
        elif isxform(o): return Xform(X.R*o.R,X.R*(o.t) + X.t)
        elif ismat(o): return Xform(X.R*o,X.t)
        elif islist(o): return [X*x for x in o]
        elif istuple(o): return tuple([X*x for x in o])
        elif isiter(o): return (X*x for x in o)
        else: return o.__rmul__(X)
    def __rmul__(X,o):
        if ismat(o): return Xform(o*X.R,o*X.t)
        raise NotImplementedError
    def __div__(X,o):
        if isxform(o): return X*~o
        return o.__rdiv__(X)
    def __add__(X,v):
        if isvec(v): return Xform( X.R, X.t + X.R*v )
        return v.__radd__(X)
    def __radd__(X,v):
        if isvec(v): return Xform( X.R, X.t + v )
        raise NotImplementedError
    def __sub__(X,v):
        if isvec(v): return Xform( X.R, X.t - X.R*v )
        return v.__rsub__(X)
    def __rsub__(X,v):
        if isvec(v): return Xform( X.R, X.t - v )
        raise NotImplementedError
    def __eq__(self,other): return self.R==other.R and
self.t==other.t
        def __str__(self): return "Xform( %s, %s )" %
(str(self.R),str(self.t))
        def __repr__(self): return "Xform( %s, %s )" %
(repr(self.R),repr(self.t))
        def __eq__(X,Y):
            assert isxform(Y)
            return X.R == Y.R and X.t == Y.t
    def rotation_axis(X): return X.R.rotation_axis()
    def pretty(self):
        a,r = self.rotation_axis()
        if self.t.length() > EPS: return "Xform( axis=%s,
ang=%f, dir=%s, dis=%f
)"%(str(a),degrees(r),str(self.t.normalized()),self.t.length())
        else: return "Xform( axis=%s, ang=%f, dir=%s, dis=%f
)"%(str(a),degrees(r),str(V0),0)

```

```

Ixform = Xform(Imat,V0)
def stub(cen=None, a=None, b=None, c=None):
    if cen is None: cen = a
    if c is None: cen,a,b,c = cen,cen,a,b
    return Xform().from_four_points(cen,a,b,c)
def enumerate_combinations(list):
    num = len(list)
    ncomb=1
    counter=[]
    for i in range(num):
        ncomb=ncomb*list[i]
        counter.append(0)
    add = 1
    combs=[]
    for j in range(ncomb):
        combs.append(string.join(map(lambda x: str(counter[x]),
range(num))))
        for i in range(num):
            counter[i]=counter[i]+add
            if counter[i]==list[ i ]:
                counter[i]=0
            else:
                break
    return(combs)
def
Generate_Pdbs(Nres,num_chain,num_to_output,w,R,orientation,helix_phase,
delta_z,output_file_name,chain_name, chain_order):
    deg_to_rad= math.pi/180.
# chain parameters
ph = 360/num_chain
phase=[]
for i in range(num_chain):
    phase.append(i*ph)
chain_set=['A','B','C','D','E','F','G','H','I ','J ','K',
'L','M','N','O','P','Q','R','S ','T','U','V','W','X','Y','Z']
chain_num=chain_set[0:num_chain]
R1=2.26
#rise per residue d fixed . this constrains pitch ( alpha )
d=1.51
z1=0.
z2=0.
line='ATOM      7  CA   GLY A    2          5.520    2.352   1.361  1.00
20.00
last=line[54:-1]
atom_num=1
res_num=1
Res_id=[]
CA_list=[]
alpha=math.asin(R*w*deg_to_rad/d)
for iter in range(num_to_output):
    CA_list.append([])
    Res_id.append([])
    chain = chain_name[iter]
    orient = orientation[iter ]
    #w1=100-w
    w1=99.2 # decouple w1 and w and fix w1 to 99.2 (3 full turns per 11
residues). Change to 98.2 for TIP-98 or 97.2 for TIP-97 etc.

```

```

if orient == 1:
    res_num=0
else :
    res_num=Nres+1

supercoil_phase=phase[iter]+delta_z[iter]*math.tan(alpha) / (R*deg_to_rad)
)
    for t in range(Nres+2):      ## need two extra residues to guide
placement of 1 st and last residue
        a0=(w*(t-1)+supercoil_phase)*deg_to_rad
        if orient == 1:
            a1=(w1*(t-1)+helix_phase[iter])*deg_to_rad    # set ref point for
phase to be along supercoil radius
        else:
            a1=(w1*t-w1*Nres-helix_phase[iter])*deg_to_rad
            x=R*math.cos(a0) + R1*math.cos(a0) * cos(a1) -
R1*cos(alpha)*sin(a0)*sin(a1)
            y=R*math.sin(a0) + R1*sin(a0)*cos(a1) +
R1*cos(alpha)*cos(a0)*sin(a1)
            if w==0:
                z=d*t+delta_z[iter]
            else:
                z= R*w*t*deg_to_rad/math.tan(alpha)-
R1*sin(alpha)*sin(a1)+delta_z[iter]
            CA_list[iter].append( (res_num,Vec(x,y,z)) )
            Res_id[iter].append(res_num)
            atom_num=atom_num+1
            if orient == 1:
                res_num=res_num+1
            else:
                res_num=res_num-1
# convert CA trace to full backbone model by superimposing on
ideal template
# by matching 3 consecutive CA atoms
# set up ideal template
stub_file=map(string.split,open('ideal.pdb','r ') . readlines ())
atom=[]
for line in stub_file:
    atom.append( (Vec(float(line[6]),float( line [7]) , float ( line
[8])) ))
ideal_stub=stub(atom[6],atom[1],atom[11])
#now make full backbone pdb
full_pdb=open(output_file_name,'w')
atom_num=1
res_num=0
for counter in range(num_to_output):
    iter=int(chain_order[counter])
    chain=chain_name[iter]
    CA_chain_u=CA_list[iter]
    CA_chain = sorted(CA_chain_u, key = lambda res: res[0])
    for res in range(1,Nres+1):
        res_num=res_num+1
        actual_stub=stub(CA_chain[res][1],CA_chain[res-
1][1],CA_chain[res+1][1])
        transform=actual_stub * ~ideal_stub
    # N
        coords=transform*atom[5]

```

```

        full_pdb.write('ATOM %6d N GLY %s %3d
%8.3f%8.3f%8.3f%s\n'%(atom_num,chain,res_num,coords.x,coords.y,coords.z
,last))
        atom_num=atom_num+1
# CA (use actual CA from trace rather than superimposed one)
        coords=CA_chain[res][1]
        tcoords=transform*atom[6]
#     print coords , tcoords , ' CA'
        full_pdb.write('ATOM %6d CA GLY %s %3d
%8.3f%8.3f%8.3f%s\n'%(atom_num,chain,res_num,coords.x,coords.y,coords.z
,last))
        atom_num=atom_num+1
# NH
        coords=transform*atom[7]
        full_pdb.write('ATOM %6d H GLY %s %3d
%8.3f%8.3f%8.3f%s\n'%(atom_num,chain,res_num,coords.x,coords.y,coords.z
,last))
        atom_num=atom_num+1
# C
        coords=transform*atom[8]
        full_pdb.write('ATOM %6d C GLY %s %3d
%8.3f%8.3f%8.3f%s\n'%(atom_num,chain,res_num,coords.x,coords.y,coords.z
,last))
        atom_num=atom_num+1
# O
        coords=transform*atom[9]
        full_pdb.write('ATOM %6d O GLY %s %3d
%8.3f%8.3f%8.3f%s\n'%(atom_num,chain,res_num,coords.x,coords.y,coords.z
,last))
        atom_num=atom_num+1
        start_d=Vec.distance(atom[8],atom[6])
        end_d = Vec.distance(transform*atom[8],transform*atom[6])
    return()
def input_params(input):
    output_file_prefix=input[0][0]
    Nres = int(input[1][0])
    w0_begin = float(input[2][0])
    w0_end = float(input[2][1])
    w0_iter=int(input[2][2])
    R0_begin = float(input[3][0])
    R0_end = float(input[3][1])
    R0_iter = int(input[3][2])
    num_chain=int(input[4][0])
    num_to_output=int(input[5][0])
    orientation=[]
    for i in range(num_to_output):
        orientation.append( int(input[6][i] ) )
    phase=[]
    for i in range(num_to_output):    ## restrict phase and delta_z to
switch sign in sym_mates
        phase.append( (float(input[7+i][0]),float(input[7+i ][1])
,int(input[7+i ][2]) ))
    z_list=[]
    for i in range(num_to_output):
        l=7+num_to_output+i
        z_list.append( ( float(input[l ][0]) , float (input[ l ][1])
,int(input[ l ][2]) ) )

```

```

chain_name=[]
for i in range(num_to_output):
    chain_name.append(input[l+1][i])
chain_order=[]
for i in range(num_to_output):
    chain_order.append(input[l+2][i])
print ' ##### SUPERHELIX PARAMS ##### \n'
print ' output file prefix: %s \n' %(output_file_prefix)
print ' helix length: %s \n' %(Nres)
print ' starting twist: %s ending twist: %s number of samples: %s
\n' %(w0_begin,w0_end,w0_iter)
    print ' starting R0: %s ending R0: %s number of smaples: %s
\n' %(R0_begin,R0_end,R0_iter)
    print ' number of chains: %s \n' %(num_chain)
    print ' number of chains in assymetric unit: %s \n' %
(num_to_output)
    print ' ##### individual helix parameters ##### \n'
    print ' ORIENTATION CHAIN CHAIN_ORDER PHASE (start, end, number)
Z OFFSET (start, end, number) \n '
    for i in range(num_to_output):
        print '%s %s %s %s %s %s %s %s
\n' %(orientation[i],chain_name[i],chain_order[i],phase[i][0],phase[i][1],
phase[i][2],z_list[i][0],z_list[i][1],z_list[i][2])
        # sample p1 evenly , w and R around the input values
        number_of_combinations=w0_iter*R0_iter
        for i in range(num_to_output):

number_of_combinations=number_of_combinations*phase[i][2]*z_list[i][2]
        print ' NUMBER OF PDBS TO GENERATE: %s
\n' %number_of_combinations
        combinations=[]
        w0_inc = (w0_end - w0_begin)/max(w0_iter-1,1)
        R0_inc = (R0_end - R0_begin)/max(R0_iter-1,1)
        w0=[]
        R0=[]
        ph=[]
        z = []
        for i in range(w0_iter):
            w0.append( w0_begin+w0_inc*i)
        for i in range(R0_iter):
            R0.append( R0_begin+R0_inc*i)
        for j in range(num_to_output):
            p_inc=(phase[j][1] - phase[j][0])/max(phase[j][2]-1,1)
            ph.append([])
            for i in range(phase[j][2]) :
                ph[j].append(phase[j][0]+p_inc*i)
        for j in range(num_to_output):
            z_inc=(z_list[j][1] - z_list[j][0]) /max(z_list[j][2]-1,1)
            z.append([])
            for i in range(z_list[j][2]) :
                z[j].append(z_list[j][0]+z_inc*i)

return(Nres,num_chain,num_to_output,orientation,R0,w0,ph,z,output_file_
prefix,chain_name,chain_order)
#####
input_file=argv[1]
tag = argv[2]

```

```

input =map(string.split,open(input_file,'r')) . readlines ())
Nres,num_chain,num_to_output,orientation,R0,w0,ph,z,output_file_prefix,
chain_name,chain_order = input_params(input)
items=[]
items.append(len(w0))
items.append(len(R0))
for p in ph:
    items.append(len(p))
for zz in z:
    items.append(len(zz))
combos=enumerate_combinations(items)
for combo in combos:
    id=map(int,string.split(combo))
    helix_phase=[]
    delta_z []
    for i in range(num_to_output):
        helix_phase.append(ph[i][id[2+i]])
        delta_z.append(z[i][id[2+num_to_output+i]])
    ## to get close to C2 symmetry for axis in x-y plane, switch
sign of phase and delta_z of sym mates
    out_file_name='%.2f%.2f'%(tag,w0[id[0]],R0[id[1]])
    for i in range(num_to_output):
        out_file_name=out_file_name+'_'+%.2f%helix_phase[i]
    for i in range(num_to_output):
        out_file_name=out_file_name+'_'+%.2f%delta_z[i]
    out_file_name=out_file_name+'.pdb'
Generate_Pdbs(Nres,num_chain,num_to_output,w0[id[0]],R0[id[1]],orientation,
helix_phase,delta_z,out_file_name,chain_name,chain_order)
#####
End of script#####

```

Execution: python2.7 generate_helix.py <design_name> params.dat

Script S2. Rosetta Design scripts

Simplified design script and score weight (.wts) adapted from Huang et al. (1) to design helix bundle backbones.

```

design.xml

<ROSETTASCRIPTS>
<SCOREFXNS>
    <ScoreFunction name="soft" weights="/input/soft_rep_trp_ala"/>
    <ScoreFunction name="hard" weights="ref2015">
        <Reweight scoretype="coordinate_constraint" weight="0.5" />
    </ScoreFunction>
    <ScoreFunction name="hard_bb" weights="/input/bb_only">
        <Reweight scoretype="coordinate_constraint" weight="2." />
        <Reweight scoretype="cart_bonded" weight="0.5" />
    </ScoreFunction>
</SCOREFXNS>
<RESIDUE_SELECTORS>
    <Or name="IBP">
        <Index resnumss="47"/>
        <Index resnumss="50"/>

```

```

<Index resnumss="54"/>
<Index resnumss="58"/>
<Index resnumss="61"/>
<Index resnumss="65"/>
<Index resnumss="69"/>
<Index resnumss="72"/>
<Index resnumss="76"/>
<Index resnumss="80"/>
<Index resnumss="83"/>
<Index resnumss="87"/>
</Or>
<Layer name="core" select_core="true"
use_sidechain_neighbors="false" ball_radius="2.5" />
<Not name="not_core" selector="core" />
<Layer name="boundary" select_boundary="true"
use_sidechain_neighbors="false" ball_radius="2.5" />
<Not name="not_boundary" selector="boundary" />
<Layer name="surface" select_surface="true"
use_sidechain_neighbors="false" ball_radius="2.5" />
<Not name="not_surface" selector="surface" />
</RESIDUE_SELECTORS>

<TASKOPERATIONS>
<IncludeCurrent name="current"/>
<LimitAromaChi2 name="arochi" />
<ExtraRotamersGeneric name="ex1_ex2" ex1="1" ex2="1"/>
<OperateOnResidueSubset name="surface_res_allowed"
selector="surface"> <RestrictAbsentCanonicalAASRLT
aas="DESTNQKRH"/>
</OperateOnResidueSubset>
<OperateOnResidueSubset name="notsurface" selector="not_surface">
<PreventRepackingRLT/>
</OperateOnResidueSubset>
<OperateOnResidueSubset name="core_res_allowed" selector="core">
<RestrictAbsentCanonicalAASRLT aas="VAILMF"/>
</OperateOnResidueSubset>
<OperateOnResidueSubset name="notcore" selector="not_core">
<PreventRepackingRLT/>
</OperateOnResidueSubset>
<OperateOnResidueSubset name="boundary_res_allowed"
selector="boundary">
<RestrictAbsentCanonicalAASRLT aas="VAILYDESTNQKRH"/>
</OperateOnResidueSubset>
<OperateOnResidueSubset name="notboundary" selector="not_boundary">
<PreventRepackingRLT/>
</OperateOnResidueSubset>
<OperateOnResidueSubset name="restrict_IBP" selector="IBP">
<RestrictToRepackingRLT/>
</OperateOnResidueSubset>
<RestrictAbsentCanonicalAAS name="ala_only" resnum="0" k
eep_aass="A" />
</TASKOPERATIONS>

<FILTERS>
<PackStat name="packstat" threshold="0.3" confidence="1"/>
</FILTERS>

```

```

<MOVERS>
    <AddConstraintsToCurrentConformationMover name="add_cst"
        use_distance_cst="0" max_distance="12" coord_dev="2.5"
        min_seq_sep="8" />
    <ClearConstraintsMover name="clearconstraints"/>
    <PackRotamersMover name="transform_sc" scorefxn="hard"
        task_operations="ala_only"/>
    <PackRotamersMover name="softpack_core" scorefxn="soft"
        task_operations="core_res_allowed,notcore,current,arochi,restrict_IBP"/>
    <PackRotamersMover name="softpack_surface" scorefxn="soft"
        task_operations="surface_res_allowed,notsurface,current,arochi,restrict_IBP"/>
    <PackRotamersMover name="hardpack_surface" scorefxn="hard"
        task_operations="surface_res_allowed,notsurface,current,arochi,ex1,restrict_IBP"/>
    <PackRotamersMover name="hardpack_core" scorefxn="hard"
        task_operations="core_res_allowed,notcore,current,arochi,ex1_ex2,restrict_IBP"/>
    <PackRotamersMover name="softpack_boundary" scorefxn="soft"
        task_operations="boundary_res_allowed,notboundary,current,arochi,restrict_IBP"/>
    <PackRotamersMover name="hardpack_boundary" scorefxn="hard"
        task_operations="boundary_res_allowed,notboundary,current,arochi,ex1_ex2,restrict_IBP"/>
    <MinMover name="hardmin_bb" scorefxn="hard_bb"
        type="lbfgs_armijo_nonmonotone" tolerance="0.0001" chi="1" bb="1"
        bondangle="1" bondlength="1" jump="all" cartesian="1"/>
    <MinMover name="hardmin_sonly" scorefxn="hard" chi="1" bb="0"
        bondangle="0" bondlength="0"/>
    <MutateResidue name="50T" target="50" new_res="THR"/>
    <MutateResidue name="60T" target="61" new_res="THR"/>
    <MutateResidue name="70T" target="72" new_res="THR"/>
    <MutateResidue name="80T" target="83" new_res="THR"/>
</MOVERS>
<PROTOCOLS>
    <!--mutate all residues in the pose to alanines-->
    <Add mover="transform_sc"/>
    <!--minimize the backbone with constraints -->
    <Add mover="add_cst"/>
    <Add mover="hardmin_bb"/>
    <Add mover="clearconstraints"/>
    <!--apply threonine mutations-->
    <Add mover="50T"/>
    <Add mover="60T"/>
    <Add mover="70T"/>
    <Add mover="80T"/>
    <!--layer design round 1-->
    <Add mover="softpack_core"/>
    <Add mover="softpack_boundary"/>
    <Add mover="softpack_surface"/>
    <Add mover="hardmin_sonly"/>
    <!--layer design round 2-->
    <Add mover="hardpack_core"/>
    <Add mover="hardpack_boundary"/>
    <Add mover="hardpack_surface"/>
    <!--remove designs with packstat < 0.3-->

```

```

        <Add filter="packstat"/>
    </PROTOCOLS>
    </ROSETTASCRIPTS>

soft_rep_trp_ala.wts

ETABLE FA_STANDARD_SOFT
METHOD_WEIGHTS ref 1.0 -0 -0.699 -0.654 1.652 -0.449 0.89 0.526 -
0.749 0.226 0.328 -0.877 0.214 -0.834 -0.797 -0.492 -0.743 0.177 2.0
1.077
pro_close 1.0
fa_atr 0.8
fa_rep 0.657
fa_sol 0.648
fa_intra_rep 0
fa_pair 0.452
fa_dun 0.569
ref 1
hbond_lr_bb 0.857
hbond_sr_bb 0.857
hbond_bb_sc 0.857
hbond_sc 0.857
p_aa_pp 0.915
dslf_ss_dst 0.5
dslf_cs_ang 2
dslf_ss_dih 5
dslf_ca_dih 5

bb_only.wts

METHOD_WEIGHTS ref 3.6 -0.000206398 -0.47402 -0.604935 1.15537 -
0.310674 0.954298 0.0 -0.442574 0.0 -0.204523 -0.74518 -0.654485 -
0.781173 -0.76836 -0.298617 -0.22266 0.025644 2.0 0.928496
fa_rep 0.44
fa_intra_rep 0.004
hbond_sr_bb 1.5
cart_bonded 0.5
rama 0.2
omega 0.5
ref 1

```

Execution: ./rosetta_scripts.default.linuxgccrelease -parser:protocol design.xml -s input_name.pdb -out:path:all /output_folder -nstruct 1 -overwrite

Script S3. Rosetta Remodel scripts

Example of parameters for TIP-99 loop modelling.

```

remodel.flags

-in:path input_files
-remodel:blueprint input_files/blueprint
-remodel:num_trajectory 1
-nstruct 500
-remodel:quick_and_dirty
-out:path:all output_files

```

```
-out:file:scorefile missing_loops.sc
-find_neighbors
-overwrite
-ex1
-ex2
```

Example of blueprint for TIP-99 loop modelling.

Residues that are remodeled to connect the loops of helices in bold. The two adjacent residues to the loop connection are also remodeled. Only TIP-97 required a 3 residue loop to connect loops. After loop closure in centroid mode the residues are Rosetta designed and neighboring residues to the loops are repacked. In the residue design, all amino acid identities are allowed except for cysteine.

Blueprint

```
1 D .
2 E .
3 E .
4 E .
5 E .
6 I .
7 K .
8 K .
9 A .
10 E .
11 K .
12 L .
13 V .
14 E .
15 D .
16 L .
17 L .
18 R .
19 E .
20 A .
21 K .
22 K .
23 I .
24 V .
25 E .
26 E .
27 I .
28 L .
29 R .
30 E .
31 L .
32 K .
33 K .
34 A .
35 V .
36 E .
37 E .
38 L .
39 R .
40 E .
41 R .
42 L .
```

43 E .
44 K L NOTAA C
0 x L NOTAA C
0 x L NOTAA C
45 D L NOTAA C
46 E .
47 E .
48 E .
49 A .
50 E .
51 R .
52 I .
53 E .
54 R .
55 E .
56 M .
57 K .
58 R .
59 L .
60 A .
61 E .
62 E .
63 A .
64 K .
65 K .
66 R .
67 M .
68 Q .
69 E .
70 T .
71 A .
72 E .
73 K .
74 L .
75 K .
76 K .
77 E .
78 L .
79 E .
80 R .
81 I .
82 E .
83 K .
84 E .
85 L .
86 R .
87 D .
88 R L NOTAA C
0 x L NOTAA C
0 x L NOTAA C
89 D L NOTAA C
90 E .
91 E .
92 E .
93 E .
94 I .
95 E .

```
96 K .
97 R .
98 A .
99 E .
100 E .
101 A .
102 K .
103 R .
104 R .
105 M .
106 E .
107 E .
108 L .
109 I .
110 R .
111 K .
112 A .
113 K .
114 E .
115 E .
116 M .
117 E .
118 K .
119 L .
120 I .
121 E .
122 K .
123 A .
124 E .
125 R .
126 E .
127 I .
128 K .
129 E .
130 K .
131 N .
132 K .
```

Execution: ./remodel.default.linuxgccrelease @remodel.flags

Script S4. Backbone RMSD calculation script

```
calculate_rmsd.py

##### START OF SCRIPT #####
from pyrosetta import init
from pyrosetta.io import pose_from_pdb
from pyrosetta.rosetta.core.scoring import CA_rmsd
import matplotlib.pyplot as plt

init()
# top_pdb.list contains a list of file names without pdb extention
f = open("top_pdbs.list","r")
out=open("rmsd_pdbs.dat","w+")

list_pdbs = []
```

```

list_rmsd = []
for line in f:
    native = line.splitlines()[0]+".pdb"
    relaxed = line.splitlines()[0]+"_0001.pdb"
    pose_native = pose_from_pdb("/input/"+native)
    pose_relaxed = pose_from_pdb("/output/"+relaxed)
    rmsd = CA_rmsd(pose_native,pose_relaxed)
    list_pdbs.append(native)
    list_rmsd.append(rmsd)
    out.write(native+"\t"+str(rmsd)+"\n") #write to file
f.close()
out.close()
##### END OF SCRIPT #####

```

Script S5. Rosetta Ab initio structure prediction

3-mer and 9-mer were generated via the Robetta web server (12). The input.fasta contains the designed amino acid sequence in fasta format. Typically >3000 AbinitioRelax simulations are performed to observe a folding funnel with the following flags:

```

abinitio.flags

-in:file:native input_files/design.pdb
-in:file:fasta input_files/input.fasta
-in:file:frag3 input_files/frags.200.3mers
-in:file:frag9 input_files/frags.200.9mers
-nstruct 50
-abinitio:relax
-abinitio::increase_cycles 10
-abinitio::rg_reweight 0.5
-abinitio::rsd_wt_helix 0.5
-abinitio::rsd_wt_loop 0.5
-relax::fast
-out:file:silent output_files/fold_silent_${SLURM_ARRAY_TASK_ID}.out
-overwrite

```

Execution: ./AbinitioRelax.default.linuxgccrelease @abinitio.flags

SI References

1. P. S. Huang, *et al.*, High thermodynamic stability of parametrically designed helical bundles. *Science (80-.).* **346**, 481–485 (2014).
2. W. Sheffler, D. Baker, RosettaHoles: Rapid assessment of protein core packing for structure prediction, refinement, design, and validation. *Protein Sci.* **18**, 229–239 (2009).
3. H. Park, *et al.*, Simultaneous Optimization of Biomolecular Energy Functions on Features from Small Molecules and Macromolecules. *J. Chem. Theory Comput.* **12**, 6201–6212 (2016).
4. P. S. Huang, *et al.*, Rosettaremodel: A generalized framework for flexible backbone protein design. *PLoS One* **6** (2011).
5. R. Das, D. Baker, Macromolecular modeling with Rosetta. *Annu. Rev. Biochem.* **77**, 363–382 (2008).
6. M. Bansal, S. Kumart, R. Velavan, HELANAL: A Program to Characterize Helix Geometry in Proteins. *J. Biomol. Struct. Dyn.* **17**, 811–819 (2000).
7. J. Jumper, *et al.*, Highly accurate protein structure prediction with AlphaFold. *Nature* **596**, 583–589 (2021).
8. M. Mirdita, *et al.*, ColabFold - Making protein folding accessible to all. *bioRxiv*, 2021.08.15.456425 (2022).
9. M. Mirdita, M. Steinegger, J. Söding, MMseqs2 desktop and local web server app for fast, interactive sequence searches. *Bioinformatics* **35**, 2856–2858 (2019).
10. M. Mirdita, *et al.*, Uniclust databases of clustered and deeply annotated protein sequences and alignments. *Nucleic Acids Res.* **45**, D170–D176 (2017).
11. A. L. Mitchell, *et al.*, MGnify: The microbiome analysis resource in 2020. *Nucleic Acids Res.* **48**, D570–D578 (2020).
12. D. E. Kim, D. Chivian, D. Baker, Protein structure prediction and analysis using the Robetta server. *Nucleic Acids Res.* **32**, 526–531 (2004).