

# Intestinal distension orchestrates neuronal activity in the enteric nervous system of adult mice.

Jean-Baptiste Cavin, Preedajit A Wongkrasant, Joel A Glover, Onesmo Begira Balemba, Wallace K. MacNaughton, and Keith A Sharkey **DOI: 10.1113/JP284171** 

Corresponding author(s): Keith Sharkey (ksharkey@ucalgary.ca)

The following individual(s) involved in review of this submission have agreed to reveal their identity: Ken D O'Halloran (Referee #1); Gemma Mazzuoli-Weber (Referee #3)

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Senior Editor: Harold Schultz

Reviewing Editor: Michel Neunlist

# **Transaction Report:**

(Note: With the exception of the correction of typographical or spelling errors that could be a source of ambiguity, letters and reports are not edited. Depending on transfer agreements, referee reports obtained elsewhere may or may not be included in this compilation. Referee reports are anonymous unless the Referee chooses to sign their reports.)

## **1st Editorial Decision**

Dear Dr Sharkey,

Re: JP-RP-2022-284171 "Intestinal distension orchestrates neuronal activity in the enteric nervous system of adult mice." by Jean-Baptiste Cavin, Preedajit A Wongkrasant, Joel A Glover, Onesmo Begira Balemba, Wallace K. MacNaughton, and Keith A Sharkey

Thank you for submitting your manuscript to The Journal of Physiology. It has been assessed by a Reviewing Editor and by 3 expert referees and we are pleased to tell you that it is acceptable for publication following satisfactory revision.

Please advise your co-authors of this decision as soon as possible.

The referee reports are copied at the end of this email.

Please address all the points raised and incorporate all requested revisions or explain in your Response to Referees why a change has not been made. We hope you will find the comments helpful and that you will be able to return your revised manuscript within 4 weeks. If you require longer than this, please contact journal staff: jp@physoc.org.

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We look forward to receiving your revised submission.

If you have any queries, please reply to this email and we will be pleased to advise.

Yours sincerely,

Harold D Schultz Senior Editor The Journal of Physiology https://jp.msubmit.net http://jp.physoc.org The Physiological Society Hodgkin Huxley House 30 Farringdon Lane London, EC1R 3AW UK http://www.physoc.org http://journals.physoc.org

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#### EDITOR COMMENTS

**Reviewing Editor:** 

All reviewers recognize the scientific quality of the study and the added value of the additional experiments performed.

The authors are to be congratulated for this important study.

#### Senior Editor:

Thank you for submission of your revised manuscript to the Journal of Physiology. The revision is a major improvement and deemed worthy for publication in the Journal with no further revision. However, the authors will need to submit the required Statistical Summary Document before it can be accepted for publication.

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#### REFEREE COMMENTS

Referee #1:

Thank you for submitting your manuscript to The Journal of Physiology. All issues relating to ethics and welfare are appropriately described in the manuscript.

#### Referee #2:

The authors have significantly revised their manuscript. They have added additional data which has greatly strengthened the study. The authors have addressed all reviewer concerns/comments adequately.

Referee #3:

#### General comment

I would like to make my compliments to the authors who have done a pioneering work and a very meaningful study in the field of neurogastroenterology. This study is describing, with state of the art live-cell confocal imaging, enteric jejunal and colonic neuronal activity, in terms of intracellular calcium trafficking in response to different stimuli such as KCl, veratridine, DMPP and luminal nutrients. The experiments have been with different mice reporter (Wnt1-, ChAT- and Calb1-GCaMP6). Results (very nice figures and explicative video material) show that not only the neuronal responses, but also their patterns and kinetics are dependent on the level of the intestinal wall distention. Mechanosensitive channels, in particular BK channels, regulating intracellular calcium level, are likely to be involved in this mechanism. The text is clear, good written, without redundancy. The discussion is well structured, linear, the main pitfall are argued well.

Thus, the authors demonstrated that the distention level of the intestine regulates the excitability of the enteric neurons, a novel concept and a very important piece of information to understand the physiology of the intrinsic neuronal circuits regulating motility.

END OF COMMENTS

## **Confidential Review**



Department of Physiology & Pharmacology Health Sciences Centre, Room 2037

> Telephone: (403) 220-4601 Email: ksharkey@ucalgary.ca

January 16, 2023

Harold Schultz Senior Editor Journal of Physiology

Dear Dr. Schultz:

Thank you for the provisional acceptance of our manuscript entitled "Intestinal distension orchestrates neuronal activity in the enteric nervous system of adult mice" for publication in the Alimentary Section of *The Journal of Physiology*.

We were delighted that the reviewers found the paper to be of merit with no further revision and we are grateful for the opportunity to publish our work in the *Journal*.

As requested, we have revised the manuscript to add all the supporting data into the paper. This has resulted in 2 additional Figs, and revision to some of the others and their respective figure legends (shown on the marked file). We have added a graphical abstract, the statistical summary and the first author's biosketch and photos. In addition, we revised the references to be in the correct format and we corrected a minor mislabelling on a 4 of the supplementary video files (uM, instead of um).

We are requesting to be allowed one supporting information file for the design of the imaging chamber and the associated 3D printer files. By including these purely technical files we allow for others to reproduce our work using similar equipment.

Finally, we are very pleased also to submit two potential cover art images (same image in different colours).

Thank you for accepting our paper for publication in *The Journal of Physiology*.

With best wishes

Yours sincerely,

Keith SC.

Keith A. Sharkey, Ph.D., CAGF, FCAHS Professor Dear Dr Sharkey,

Re: JP-RP-2023-284171R1 "Intestinal distension orchestrates neuronal activity in the enteric nervous system of adult mice." by Jean-Baptiste Cavin, Preedajit A Wongkrasant, Joel A Glover, Onesmo Begira Balemba, Wallace K. MacNaughton, and Keith A Sharkey

We are pleased to tell you that your paper has been accepted for publication in The Journal of Physiology.

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Yours sincerely,

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#### EDITOR COMMENTS

Thank you for your final edits to the manuscript. The supporting information files are fine. Congratulations on a very detailed and insightful study.

# **1st Confidential Review**

16-Jan-2023

30-Nov-2021

JP-RP-2021-282464 The Journal of Physiology Decision Letter

Dear Dr Sharkey,

Re: JP-RP-2021-282464 "Intestinal distension orchestrates neuronal activity in the enteric nervous system of adult mice" by Jean-Baptiste A Cavin, Joel A Glover, Wallace K. MacNaughton, and Keith A Sharkey

Thank you for submitting your manuscript to The Journal of Physiology. It has been assessed by a Reviewing Editor and by three Referees and the reports are copied below.

I regret to say that the manuscript has not been accepted for publication.

Some positive comments were made on the manuscript. Unfortunately, they did not outweigh the more serious criticisms which led the Reviewing Editor to recommend rejection.

I am sorry to have to pass on this disappointing news, and hope it will not discourage you from making future submissions of new work to The Journal of Physiology.

However, we believe your manuscript is worthy of further consideration and suggest that you transfer your manuscript to Physiological Reports

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# EDITOR COMMENTS

# **Reviewing Editor:**

Thank you for submitting your work to the Journal of Physiology. The manuscript has been reviewed by 2 expert referees. In this instance, while one referee found merit in your work, particularly the novel and integrated experimental design, another referee found serious concerning elements in the manuscript which upon review I concur with. In its current state, we are unable to proceed further with review of the manuscript for The Journal. At present, the manuscript represents an important methodological tool to explore ENS integration in response to various mechanical and chemical signals, this may be of interest as a preliminary methods paper in our sister journal Physiological Reports but the findings in the paper are too preliminary to be of high impact to warrant publication in The Journal of Physiology. In particular, the lack of identification of the specific neuron populations that are activated or inactivated during distention is of concern and a major gap of the study. In addition, while the authors speculate on the nature of the link between distention and Ca2+ regulation, this would need to be experimentally explored in some detail in order for the paper to be considered of sufficient impact.

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**REFEREE COMMENTS** 

Referee #1:

Comments for Author (Required):

Thank you for submitting your manuscript to The Journal of Physiology. There are some minor but important revisions to the text required pertaining to animal ethics and welfare.

Please start the Methods section with the sub-heading "Ethical approval". Thank you for providing full details of ethical approval, source of the animals and general housing conditions.

Please include the concentration of isoflurane used to induce deep anaesthesia. Please also include detail of the carrier gas.

Referee #2:

General comments:

The general aim of this study was to evaluate the changes in neuronal excitability (to KCl, veratramine, nutrient infusion) in response to distension. The authors utilized calcium imaging techniques in mice expressing GCaMP6s specifically in neurons (Wnt1-GCaMP mice) or IPANs (Calbindin1-GCaMP mice).

The rationale for the study is well thought out and the experiments that have been performed to test the hypothesis are appropriate. The novel techniques used to examine the hypothesis are eloquent and highly innovative. The findings are mostly discussed well, and appropriate references have been included. No major flaws are noted. The findings of this study will advance the field by providing important information regarding how neurons respond to distension. There are however some relatively minor concerns that need to be addressed to improve the overall quality of this study. These are outlined in the specific comments below.

Specific comments:

Methods:

- p.7, line 7. Does nicardipine do anything to this type of Ca2+ signaling in enteric neurons? Since L-type calcium channels can be activated by stretch/distention it's important to clarify this.

- p.7, description of preparation. It would be helpful to include "a piece of intestine" in the illustration in chamber design or could more easily refer to Fig. 1A in addition to this.

Results:

- p.13, line 17. It should be stated what veratridine is and why it was used specifically. Also, why was 75mM chosen as the concentration for KCl? Is this an increase to 75mM or is it an additional 75mM added to the solution? Does this alter the osmolarity or mucosal activity in any way that might affect the neuronal behavior? Are similar effects seen at a lower concentration such as 60mM? Please clarify.

- Fig. 1 and throughout. It is difficult to see the dashed red line, perhaps the authors could use a different color or thicken the line to make it more visible.

- Fig. 3. How were the three different responses in proximal colon summarized in panel B? Were they added together? If so, it would be nice to have a summary of each type of activity independently as a supplemental figure/data point. Is there any correlation between the type of response and the distance from the cecum?

- Fig. 3 onwards. Why was KCl chosen over veratridine? A sentence indicating why now only looking at KCl responses would be a nice addition.

- Fig. 4C. At least one additional experiment should be carried out (currently n=2) for statistical evaluation ("virtually identical...compared to Fig. 3" is not quantitative or definitive).

p.21, last line and Fig. 6 legend. There are different values indicated for the average speed of the calcium wave propagating in the circumferential direction (i.e., 480 vs. 470 um.min-1).

- Fig. 7 and Table 2. It would be good if the authors were able to complete at least one additional experiment evaluating distension with nutrient administration since this is currently n=3 but n=4 for TTX and n=8 for control.

- Supplemental video files. In videos where veratridine or TTX are used, um is used instead of uM. Please correct. This also needs to be corrected for TTX in Figs. 4 and 7.

Discussion:

- General comment. The discussion is very well written but it would be good if the authors could include a short section speculating how these changes in neuronal activity with

distention translate to motility, absorption, etc. and why these changes may occur (are they more beneficial or detrimental?). A statement relating the findings to what occurs in patients with chronic constipation that have distended colons for example would be a valuable addition and a great way to translate this work to a focus for clinical studies.

Overall, this is an elegant study that has approached the topic in a novel and innovative fashion. The conclusions presented are valid based on the experimental evidence provided. This study adds to our knowledge and understanding of how mechanical changes are detected and regulated.

Referee #3:

I have some difficulties with this paper, including in the way the work is described. The authors make comparisons to empty segments. What do they mean by empty? Are the segments actually empty? Or do they mean undistended? If they mean undistended, that is the term they should use. If they mean empty in some other sense, this should be defined.

They also refer to the intestine with normal luminal contents. What do they mean? Do they mean containing oxygenated Krebs solution? This is not the normal content of the jejunum or the colon (for one thing, the lumen is anoxic in vivo). If they mean containing Krebs solution, this is what they should say. If they mean something different by "normal luminal contents", this should be defined.

They refer to "differences in neuronal intracellular Ca2+ homeostasis" (page 13). The study is hardly done under conditions in which homeostatic controls, in a physiological sense, are exerted by the organ. I suggest removing homeostatic.

In Fig 1C there are provided "Representative traces of neuronal Ca2+ fluorescence changes in the myenteric plexus". Are these traces from individual neurons, or averaged traces each from a region of myenteric plexus, as suggested by the description? It looks like there are about 7 traces for KCI. The description of Fig 2A has the same ambiguity. I apologize if this is made clear somewhere else in the manuscript. It would assist to have it in the figure legend.

Responses to KCl peak at 2 minutes in some cases, and sometimes last as long as 4 minutes. It is quite a long response to a high concentration of KCl, 75 mM, that neurons would not normally encounter. Perhaps the authors could consider whether there might be depolarization block, and failure of synaptic transmission.

Responses to veratridine, a toxin that blocks Na channel inactivation, causing a prolonged inward current and increased intracellular Ca, lasted over 5 minutes in some cases. The authors might comment on whether this may compromise neuronal function, including

activation of intrinsic sensory neurons and neuro-neuronal communication.

It would be valuable to know how long KCl or veratridine was present in the bath, whether they were flushed out, what concentrations they achieved and how the concentrations varied over time.

What was the composition of the Krebs' solution. Was it oxygenated on its way to the lumen?

The authors conclude that they have discovered that intestinal distension in the small and large intestines affects most of the neurons in the ENS by locally modulating their levels of intracellular Ca. This is written as if modulating intracellular Ca is the primary effect of distension. The authors might be encouraged to discuss whether there may be steps between distension and Ca responses, for example the release of active substances such as hormones from the mucosa or the release of neurotransmitters.

There is inadequate description of the nutrient solution, which is described as containing (in g per 100mL) 2 protein, 1.4 lipids, 7.4 sugar. Which sugar? Is this an absorbable sugar? Is it a sugar that is detected by sweet taste receptors in the jejunum or colon? What is the protein? What are the lipids? This information might help with the interpretation of the data of Table 2.

The authors state that "our study invites the development of more integrated research approaches to study how the ENS responds to the complex mechanical and chemical inputs it receives from the intestinal lumen". This would be more convincing if the neurons that responded were identified, for example as motor neurons, interneurons or putative intrinsic sensory neurons.

Kunze has described enteric neurons that are inhibited by mechanical forces (Kunze, J. Physiol 526: 375-385,2000).

In order to generate intraluminal distension, the authors transiently interrupted the withdrawal from the anal output, while keeping the oral input flowing. When the distension attained the desired amount (by visual estimation), the withdrawal on the anal end was resumed. Relaxation was attained by interrupting the perfusion flow while keeping the withdrawal on. It would be very important to know the luminal pressures achieved, their time-resolved changes and how the pressure relate to physiologically observed intraluminal pressures.

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