Supporting information

pH-tunable membrane-active polymers, NCMNP2a-x, and their potential membrane protein applications

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I. Additional figures

Fig. S1 Structural characterizations of NCMNP2a-x polymers. (A) ¹H-NMR spectra of nonmodified SMA polymer, Tris and NCMNP2a-x representative in D_2O , 300 MHz. The SMA spectrum has no singlet signal of methylene protons from Tris (δ = 3.53 ppm). In contrast, it appears and shifts to 3.67 ppm in NCMNP2a-x, suggesting that the Tris grafts on individual MAnh units instead of free-form presence. (B) FT-IR spectra of SMAnh and its derivatives altered with different amounts of Tris, NCMNP2a-x. Observation of the broad peaks in $3000 - 3600$ cm⁻¹ region of NCMNP2a-x spectra indicates the target polymers obtained; this attributed to a combination of N-H stretching frequency from amide and O-H from the carboxylic acid and alcoholic group pendants. Moreover, the shift of peaks at 1856 and 1781 cm⁻¹ (C=O anhydride stretching) to lower wavenumbers 1708 cm⁻¹ (C=O acid stretching) and 1650 cm⁻¹ (C=O amide stretching) illustrates for non-residue of unreacted MAnh and formation of amides on the polymer backbone. (C) Deconvolution of the FTIR data for NCMNP-2a-x (where x= 25, 50 and 70). The original curve, baseline and its substraction is represented by $C(a)$ for NCMNP2a-25, $C(c)$ for NCMNP2a-50 and C(e) for NCMNP2a-50 where the corresponding decolvolution were represented by C(b) for NCMNP2a-25, C(d) for NCMNP2a-50 and C(f) for NCMNP2a-50.

Fig. S2¹H-NMR spectra of commercial SMAnh and its hydrolysis SMA in CDCl₃ and D₂O, respectively.

Fig. S3 ¹H-NMR spectra of NCMNP2a-x copolymers with full integration: (A) NCMNP2a-5, (B) NCMNP2a-25, (C) NCMNP2a-50 and (D) NCMNP2a-70.

Fig. S4 Effect of Tris incorporation percentages on the protein purification efficiency from the native membranes. Purification profiles of AcrB extracted with (A) non-modified SMA polymer, SMA2000 (2.5 % w/v), and (B) NCMNP2a-70 (5 % w/v).

Fig. S5 Cryo-EM analysis of purified AcrB-NCMN particles. (A) Representative 2D classes of AcrB particles extracted from various polymers at different pH conditions. (B) The Fourier shell correlation (FSC) resolution curves for 3D reconstruction using the gold standard with $FSC =$ 0.143.

Fig. S6 Comparison of TM7, TM12, and Iα helix in protomer O of AcrB extracted by SMA2000 $(6BAJ)^1$ and our NCMN polymers (from current work) at same pH conditions, pH = 7.8.

Fig. S7 Distances between F386 residues of wide-type AcrB in this work. Owing to the lipids preserved, the central cavity of AcrB-NCMN particles remains unchanged with distances of F386-

F386 residues similar to AcrB structure in SMA2000 (6BAJ). Whereas, in a detergent system where all the supporting lipids were removed, such distances were reduced to around 6.7 Å.³

Fig. S8 Effect of Tris incorporation percentages on the protein purification efficiency from the native membranes. (A) Comparative purification profiles of *Bc*TSPO solubilized by NCMNP2a-5 and NCMNP2a-50. (B) Visualization of purified *Bc*TSPO particles on Coomassie-stained SDS-PAGE compared to protein ladder (M). Lanes 1 and 2 correspond to *Bc*TSPO particles extracted by NCMNP2a-5 and NCMNP2a-50, respectively.

Fig. S9 Fluorescence emission spectra of PpIX upon excitation at 410 nm. (A) Spectra of PpIX only at different laser pulses. Quenching of fluorescence was undetectable, suggesting that the PpIX was stable under such experimental conditions. (B) Spectra of PpIX/ *Bc*TSPO-NCMNP2a-50 solutions at different pH upon excitation at 410 nm. At pH 3 – pH 5, the fluorescence intensity of PpIX solution displays a less intense signal compared to $pH = 7.8$, indicating higher-order aggregates of PpIX.²

Fig. S10 Photodegradation of PpIX in the presence of *Bc*TSPO upon excitation at 410 nm. Decays of PpIX at 632 nm as a function of the light pulse catalyzed by *Bc*TSPO extracted by NCMNP2a-5 (A) and NCMNP2a-50 (B) at $pH = 7.8$.

Fig. 11 Mapping schemes used to model the three residues - A) **SMA**, B) **SMT**, and C) **TRI**, combinations of which were used to generate different Tris-substituted polymer models. D) MARTINI 3.0 beads used in the simulations to model polymers.

Fig. 12 A) Schematic showing the generation of polymer chains using the three modeled residues, B) Snapshots of relaxed polymer chains, C) Representative snapshot of simulation box for selfassembly simulations.

Fig. S13 Averaged RDF between water and polymer components in charged simulations including A) TC1 (backbone) beads attached to maleic acid, B) TC1 beads attached to styrene, C) TC5 (styrene) beads, D) SQ5n (charged COO-) beads, E) P2 (amide group from Tris) beads, and F) TN4a (hydroxyl group from Tris) beads. Also shown is the RDF between water and the entire polymer chains to identify the contribution of these individual groups on the overall RDF. The legends indicate different polymers based on their percentage of Tris substitution for easy understanding: Tris-5 % corresponds to NCMNP2a-5, Tris-25 % corresponds to NCMNP2a-25, and so on.

Fig. S14 Averaged RDF between polymer components from multiple chains including A) TC1 (backbone) beads, B) TC5 (styrene) beads, C) SQ5n (charged COO-) beads, D) P2 (amide group from Tris) beads, and E) TN4a (hydroxyl group from Tris) beads. Also shown is the RDF between the whole polymer chains to identify the contribution of these individual groups on the overall RDF. The legends indicate different polymers based on their percentage of Tris substitution for easy understanding: Tris-5 % corresponds to NCMNP2a-5, Tris-25 % corresponds to NCMNP2a-25, and so on.

Fig. S15 Averaged RDF between polymer components including A) TC1 (backbone) beads, B) TC5 (styrene) beads, and C) SQ5n (charged COO-) beads, with lipid head (column 1), glycerol (column 2), and tail (column 3) groups. The legends indicate different polymers based on their percentage of Tris substitution for easy understanding: Tris-5 % corresponds to NCMNP2a-5, Tris-25 % corresponds to NCMNP2a-25, and so on.

Fig. S16 Averaged RDF between polymer components including A) P2 (amide group from Tris) beads, and B) TN4a (hydroxyl group from Tris) beads, with lipid head (column 1), glycerol

(column 2) and tail (column 3) groups. The legends indicate different polymers based on their percentage of Tris substitution for easy understanding: Tris-5 % corresponds to NCMNP2a-5, Tris-25 % corresponds to NCMNP2a-25, and so on.

Fig. S17 Averaged RDF calculated using uncharged simulation trajectories between A) polymerwater, B) polymer-polymer, and C) polymer-lipids. The legends indicate different polymers based on their percentage of Tris substitution for easy understanding: Tris-5 % corresponds to NCMNP2a-5, Tris-25 % corresponds to NCMNP2a-25, and so on.

II. Additional tables

*^a*Molar ratio. *^b*Experimental grafting percentage of amine calculated based on ¹H-NMR characterization. *^c*Stability of NCMNP2a-x polymers in various buffer environments determined based on turbidity experiments. ^d Morphology of AcrB-NCMN particles visualized using negative-TEM analysis.

Table S2. Cryo-EM data collection and processing

Table S3. Structure refinement and validation statistics

Table S4. Backbone RMSD comparison of AcrB structures

Table. S5. List of all CG MD simulations performed

Table S6. Number of sodium ions within 5 Å of the polymer chains when simulated individually or with other polymer chains in the absence of lipids. Data obtained by averaging the values from all replica simulations for the last 1 μs of the simulation trajectory.

Table S7. Radii of Gyration (in Angstroms) for polymer chains (and polymer backbones where specified) in various simulated systems. Data obtained by averaging the values from all replica simulations for the last 1 μs of the simulation trajectory.

Table S8. Solvent accessible surface areas (SASA) for hydrophobic beads in different polymers when simulated individually in the absence of lipids. Data obtained by averaging the values from all replica simulations for the last 1 μs of the simulation trajectory.

Table S9. Solvent-accessible surface areas (SASA) for benzenes comprising N1-N2-N3 and N4- N5-N6 beads in different polymers when simulated individually and in the presence of lipids. Data obtained by averaging the values from all replica simulations for the last 1 μs of the simulation trajectory.

Table S10. Quantification of the Tris grafting on the polymer using FTIR data.