

Supplemental Figure 1: High density SNP array uncovers CNVs affecting coding genes in Chr6p22 and Chr9q33 in a stock of KOLF2.1J iPSCs and KOLF2.1J with doxycycline inducible NGN2 transgene. (A) Chromosome 6 cytoband schematics (top) and Log R Ratio (LRR) and B Allele frequency (BAF) plots (bottom) show reduction of signal intensity and a loss of heterozygosity in 6p22 region of KOLF2.1J iPSCs p3 and KOLF2.1J-NGN2. (B) Chromosome 9 cytoband schematics (top) and LRR and BAF plots (bottom) show reduction of signal intensity and a loss of heterozygosity in 9q33 region of KOLF2.1J iPSCs p3 and KOLF2.1J-NGN2.



**Supplemental Figure 2: CNVs detected by SNP array in Chr3 and Chr18 do not overlap with coding regions. (A-B)** Chromosome 3 cytoband schematics (top) and LRR and BAF plots (bottom) showing reduction of signal intensity and a loss of heterozygosity in 3p13 deleted region (A) and increased signal intensity and altered BAF in 3p14 duplicated region (B) of KOLF2.1J iPSCs compared to a control iPSC line. (C) Chromosome 18 cytoband schematics (top) and LRR and BAF plots (bottom) showing increased signal intensity and altered BAF in 18q22 duplicated region of KOLF2.1J iPSCs.



Supplemental Figure 3: Genes affected by Chr6p22 and Chr9q33 are expressed in neurons in the developing and adult brain. (A-D) DTNBP1 (A), JARID2 (B), ASTN2 (C) and ARID2 (D) expression in developing and adult human prefrontal cortex and in hPSC to iNeuron differentiation. Human prefrontal cortex expression data is retrieved from Brainspan (https://www.brainspan.org/rnaseg/search/index.html) and iNeuron differentiation expression plots from the web app created by Connor Ludwig, Kampmann Lab (https://kampmannlab.ucsf.edu/ineuron-rna-seq). Pcw; postconception week. Yrs; years.



## Supplemental Figure 4

Supplemental Figure 4: gDNA qPCR on an iPSC line with one copy of EZH1 exon 8-12 deletion demonstrates sensitivity of the technique for detection of hemizygous regions. (A) Schematic representation of *EZH1* exon 7-13. Red line illustrates the region deleted in a EZH1 patient iPSC line. Arrowheads (>, <) labeled with b-e indicate position of the primers used for gDNA qPCR. (B-E) gDNA qPCR results showing half levels amplification in regions deleted in EZH1 patient iPSCs compared to control and KOLF2.1Js that are expected to be diploid. (F) Randomly selected regions of the genome show similar amplification levels across all the hPSC lines.



**Supplemental Figure 5: Genome sequencing reanalysis confirms CNVs at Chr3p13, Chr3p14 and Chr18q22. (A-C).** Chromosome cytoband schematics with base resolution break points of the CNVs (top) and LRR and BAF plots (bottom) obtained from the KOLF2.1J iPSC genome sequencing reanalysis. LRR plots show reduction of signal intensity and BAF plots show loss of heterozygosity in Chr3p13 (A) and gain of signal intensity and altered BAF in Chr3p14 (B) and Chr18q22 (C) CNV regions compared to diploid up and downstream regions. Shadowed area represents the CNV defined by the SNP array and vertical lines the base-resolution breakpoints determined from the genome sequencing data.



Supplemental Figure 6: Chr3p13, Chr3p14 and Chr18q22 CNVs in KOLF2.1J were inherited from KOLF2 iPSC line. (A, B) LRR and BAF plots of the KOLF2 iPSC line SNP array deposited in HipSci, show that KOLF2 iPSC line carries two of the Chr3p14 (B) and Chr18q22 (C) CNVs in heterozygosity and is likely mosaic for Chr3p13 (A) deletion.

Primer Pair	Targeted Region (GRCh37/hg19)	Notes	Primer Orientation	Sequence (5'->3')	Product Length (bp)
1	chr6:15372726+15372836	Upstream deleted	Forward	gactacaggcgtgcaccaccac	111
		Chr6 region	Reverse	aaggcggacggatcacaaggga	
2	chr6:15270358+15270456	Upstream deleted	Forward	tctgtccgtccgtccttccgtc	99
			Reverse	aggaggtggaggctgcagtgag	
3	chr6:15487546+15487631	Chr6 region in	Forward	ggttccagcaggtcaacacggg	86
		JARID2: Exon 6	Reverse	tcgctgtgcttctcctttgcgg	
4	chr6:15500050+15500126	Within deleted	Forward	gctgtgtccccatccctggtct	77
		Chr6 region in JARID2	Reverse	gaaccacctcggcagacacagc	
	chr6:15514462+15514592	Within deleted	Forward	acacactctgggtagggacgcc	131
5		Chr6 region in JARID2	Reverse	gcactgtggaaagggacgaggc	
•		Within deleted	Forward	acctctcaggctctcacgtttccc	150
6	chr6:15/23401+15/23550	Chr6 region	Reverse	agcaggagttggagccacagga	
7	chr6:15803765+15803892	Downstream Chr6	Forward	tcccttccccccccgacata	128
		deleted region	Reverse	ttggttgggctgtccaccttgc	
Q	cbr6-15880438+15880552	Downstream Chr6	Forward	gtctgagcgggactggaccctt	- 115
0	CIII0.15869458+15869552	deleted region	Reverse	cgtttggcgtccctgtggagac	
9	chr0.110220407+110220617	Upstream deleted	Forward	tgcagccagcacacagcaaact	121
5	6113.113223437 113223617	Chr9 region	Reverse	gccaagatcgcaccactgcact	
		Within deleted	Forward	ccgtcgggctccagacacttga	77
10	chr9:119249647+119249723	Chr9 region: Exon 20	Reverse	gcagctggtcgaagccatggag	
11	chr9:119302810+119302918	Within deleted	Forward	gcttgggtgacagctttggcct	109
		Chr9	Reverse	tagatggcctgcttggtgccct	
12	chr9:119366867+119366974	Within deleted Chr9	Forward	ccatggcccacactgcatgagg	108
			Reverse	agctgagaggatggtggcagca	
40		Downstream	Forward	ccctccccaggagccacaatga	80
13	cnr9:119382905+119382984	region	Reverse	ggagtgccaagcctaggggaca	
		Within EZH1	Forward	ccaacatttgcccaacccagca	109
14	chr17:40870724-40870832	Intron between Exon 8 and 9	Reverse	ggtctgccaagggaggatgggt	
15	chr17:40870536-40870611	Within EZH1 Exon	Forward	agaccccaatgcacttccccct	76
15	cm17.40070530-40070011	9	Reverse	ttgctcccgctgcacagacttg	10
		Within EZH1	Forward	ggaacacaagccacgtgagcca	107
16	chr17:40869846-40869952	Intron between Exon 10 and 11	Reverse	gagcggtcccacagtaactggg	
17	chr17:40864313 40864437	Within EZH1 Exon	Forward	tagtggaagcaccctcggagcc	125
17	CIII 17.40804313-40804437	12	Reverse	gcacgtcttggtccccagaagc	120
18	cbr6:170863510+170863606	Chr6 control	Forward	cttcgcttccgctggcccatag	97
10	0110.1700000101170000000	region within TBP	Reverse	ctcagtgcagtggtggccttcg	57
19	chr7:5596654-5596787	Chr7 control	Forward	tgctcttctctgtgccccctcc	134
		region within ACTB	Reverse	acgggttcctactgtctggccc	
	chr12:6644756+6644900	Chr12 control	Forward	ttccaccgcaaaatggcccctc	145
20		region within			
		GAPDH (used for normalization)	Reverse	ccagacacccaatcctcccggt	
21	chr6:15481823+15729819	Chr6 Breakpoint	Forward	taccttgtctttggctctgtcg	
		Sanger Seq	Reverse	atctgagccagcatggtagagg	
22	abr0:110210800 110272070	Chr9 Breakpoint	Forward	ttctttcacctggactagaccc	
~~	0113.1132400337113312019	Sanger Seq	Reverse	gcagaatttcgcttttgtcg	