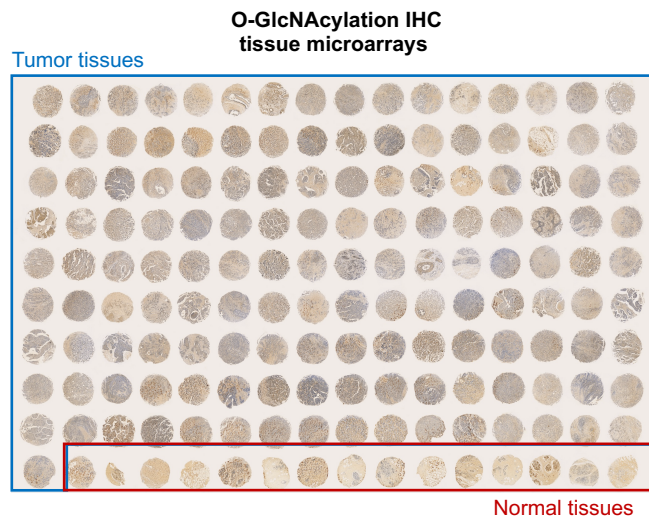
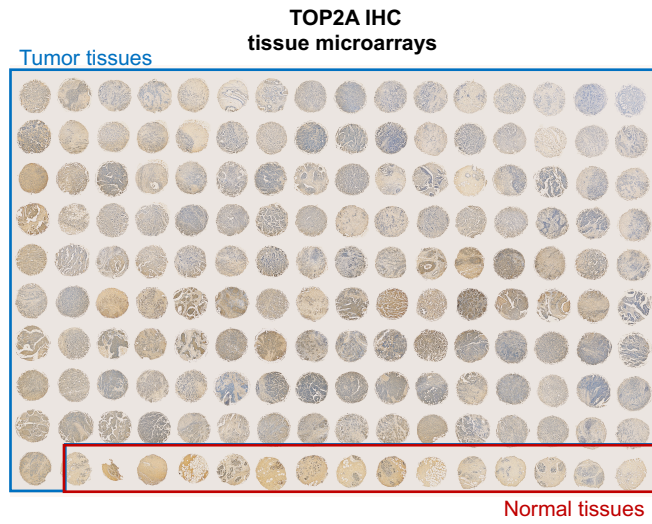


## Appendix

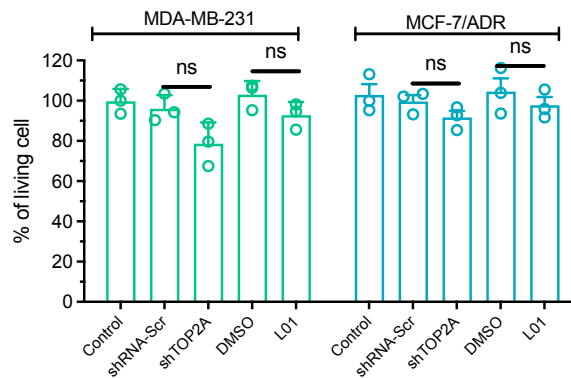
### ***O*-GlcNAcylation promotes topoisomerase II $\alpha$ catalytic activity in breast cancer chemoresistance**

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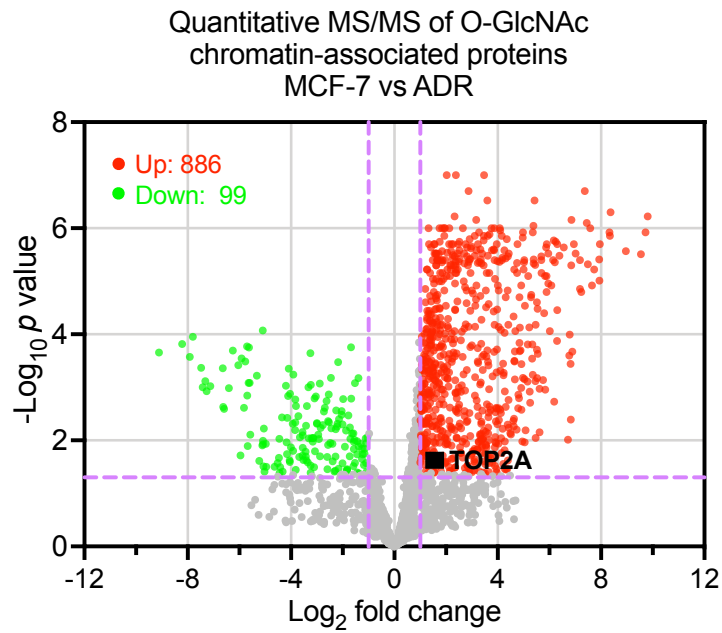
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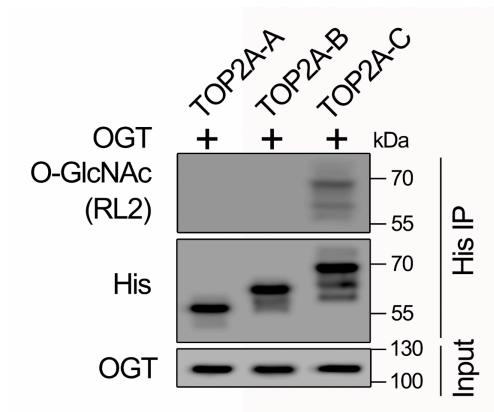
**Appendix Figure S1** Layout and IHC staining of TOP2A and cellular O-GlcNAcylation on a tissue microarray containing 145 breast tumor and 15 adjacent samples.



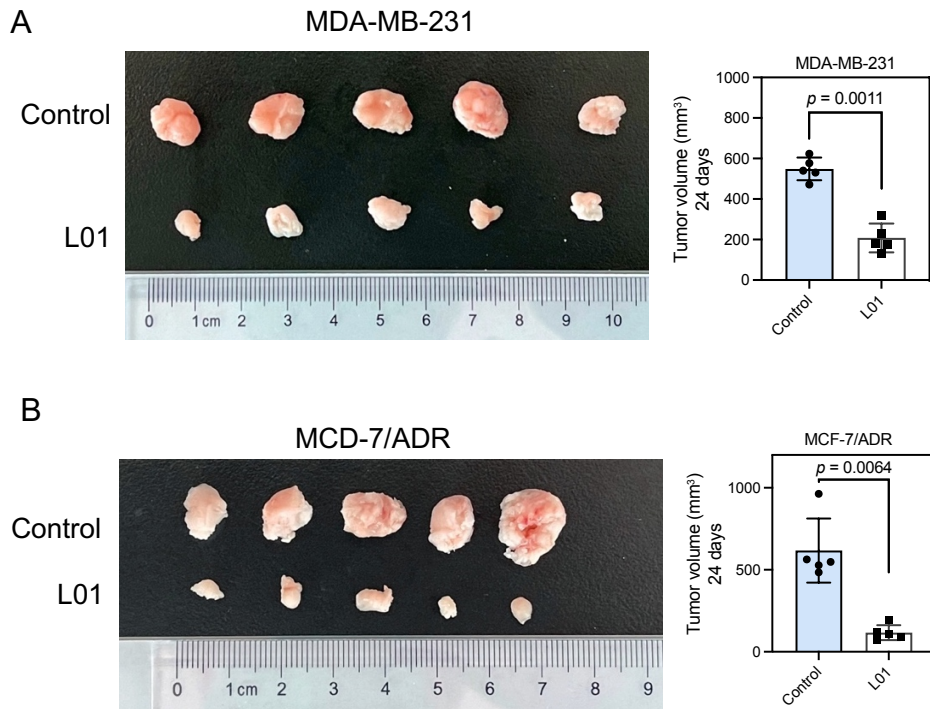
**Appendix Figure S2** MDA-MB-231 and MCF-7/ADR cells were transfected with scrambled shRNA (shRNA-Scr) or TOP2A shRNA (shTOP2A) for 48 h. Or the cells were treated with 50  $\mu$ M L01 for 48 h. The cell viability was assessed through a CCK-8 assay. Protein expression were shown in Figure 2D and E. OGT inhibition itself or TOP2A shRNA transfection itself did not have significant impact on the cell viability in MDA-MB-231 and MCF-7/ADR cells.  $n = 3$  biologically replicates. Unpaired t-test was used for statistical comparison.  $p$  value was indicted.



**Appendix Figure S3** TOP2A was previously reported to be a O-GlcNAcylated chromatin-associated protein (*Liu Y, Chen Q, Zhang N, et al. Proteomic profiling and genome-wide mapping of O-GlcNAc chromatin-associated proteins reveal an O-GlcNAc-regulated genotoxic stress response[J]. Nature communications, 2020, 11, 5898*). Volcano plot of label-free relative quantitative proteomics data of O-GlcNAc chromatin-associated proteins in MCF-7 and MCF-7/ADR cells (n = 9 biologically replicates, unpaired t-test). TOP2A was labelled.



**Appendix Figure S4** His-tagged TOP2A-A, -B and -C expression plasmids were transfected with the OGT expression plasmid in HEK-293T cells. After 48 h, His-tag IP was performed, and the immunoprecipitated fractions were analyzed by Western blot for the indicated proteins.



**Appendix Figure S5** The effects of L01 on tumor xenografts in nude mice. (A) MDA-MB-231 cells were injected subcutaneously into the axillae of nude mice ( $n = 5$  for each group). 1 mg/kg L01 was administered by tail vein injection every other day for 22 days. Control group was treated with DMSO. Volumes of tumors were monitored with caliper twice a week until 24 days. Unpaired t-test was used for statistical comparison.  $p$  value was indicated. (B) MCF-7/ADR cells were injected subcutaneously into the axillae of nude mice ( $n = 5$  for each group). 4 mg/kg Adm and 1 mg/kg L01 was administered by tail vein injection. Control group was treated with DMSO. Volumes of tumors were monitored with caliper twice a week until 24 days. Unpaired t-test was used for statistical comparison.  $p$  value was indicated.

### Appendix Table S1

The relationship between *TOP2A* mRNA expression and clinicopathological factors in 1072 TCGA breast cancer patients (female).

Variables	No. of patients	TOP2A		Chi-Square	<i>p</i> value
		low	high		
<b>Age</b>					
< 50	292	139	153	0.9225	0.3368
≥ 50	780	397	383		
<b>Stage</b>					
I	188	115	73	11.38	0.0007***
II + III + IV + X	884	421	463		
<b>T classification</b>					
T1	278	168	110	16.34	< 0.0001****
T2 + T3 + T4	794	368	426		
<b>N classification</b>					
NX + N0	523	265	258	0.1829	0.6689
N1 + N2 + N3	549	271	278		

The cutoff for high and low TOP2A expression were  $\geq$  median or  $<$  median, \*\*\* $p < 0.001$ , \*\*\*\* $p < 0.0001$

**Appendix Table S2**

The relationship between TOP2A protein expression and clinicopathological factors in 145 breast tumor samples on a tissue microarray.

Variables	No. of patients (female)	TOP2A		Chi-Square	<i>p</i> value
		low	high		
<b>Age</b>					
< 50	70	30	40	0.6109	0.4344
≥ 50	75	37	38		
<b>Grade</b>					
1	31	20	11	5.318	0.0211*
2 + 3	114	47	67		
<b>Stage</b>					
0 + I	10	6	4	0.3341	0.5633
II + III	135	61	74		
<b>T classification</b>					
Tis + T1 + T2	93	50	43	5.957	0.0147*
T3 + T4	52	17	35		
<b>N classification</b>					
N0 + N1	119	56	63	0.1938	0.6598
N2 + N3	26	11	15		
<b>Lymph node metastasis</b>					
No	96	50	46	3.947	0.047*
Yes	49	17	32		
<b>PR</b>					
Negative	107	46	61	1.699	0.1924
Positive	38	21	17		
<b>ER</b>					
Negative	110	52	58	0.2083	0.6481



Positive	35	15	20		
HER2					
Negative	65	33	32	0.9866	0.3206
Positive	80	34	46		

High group: IHC score  $\geq 4$  (median score), low group: IHC score  $< 3$ ,  $*p < 0.05$ .

**Appendix Table S3**

The relationship between cellular O-GlcNAcylation expression and clinicopathological factors in 145 breast tumor samples on a tissue microarray.

Variables	No. of patients (female)	O-GlcNAc		Chi-Square	<i>p</i> value
		low	high		
<b>Age</b>					
< 50	70	32	38	0.002119	0.9633
≥ 50	75	34	41		
<b>Grade</b>					
1	31	19	12	3.956	0.0467*
2 + 3	114	47	67		
<b>Stage</b>					
0 + I	10	6	4	0.3895	0.5326
II + III	135	60	75		
<b>T classification</b>					
Tis + T1 + T2	93	48	45	3.886	0.0487*
T3 + T4	52	18	34		
<b>N classification</b>					
N0 + N1	119	62	57	11.6	0.0007***
N2 + N3	26	4	22		
<b>Lymph node metastasis</b>					
No	96	51	45	6.63	0.01*
Yes	49	15	34		
<b>PR</b>					
Negative	107	43	64	4.678	0.0306*
Positive	38	23	15		
<b>ER</b>					
Negative	110	47	63	1.43	0.2317

Positive	35	19	16		
HER2					
Negative	65	29	36	0.03864	0.8442
Positive	80	37	43		

High group: IHC score  $\geq 3$  (median score), low group: IHC score  $< 3$ , \* $p < 0.05$ , \*\*\* $p < 0.001$ .

### Appendix Table S4

Clinical characteristics of breast cancer patient samples.

Patient no.	Gender	Age	Grade	Stage	ER	PR	HER2	Chemotherapy-resistant /relapsed
1#	Female	67	2	II	Positive	Positive	Positive	No
2#	Female	52	2	II	Negative	Negative	Positive	No
3#	Female	44	3	III	Negative	Negative	Positive	Yes
4#	Female	48	3	III	Negative	Negative	Negative	Yes
5#	Female	58	3	III	Positive	Positive	Positive	Yes

**Appendix Table S5**

Full-length human wild type and O-GlcNAcylation site mutants TOP2A	Forward	Reverse
Subclone into pLVX-IRES-Neo vector	TTCCTCGAGGCCACCAT GTACCCATACGATGTTCC AGATTACG	ATCCGCGGCCGCTTA AAACAGATCATCTTC ATCTGAC
<i>O</i> -GlcNAcylation site mutant (Ser1469 → Ala) TOP2A	GAATCGCCGCAAAGG AAGCCAGCAACTTCTGA TGATTCTGACTC	GAGTCAGAATCATCA GAAGTTGCTGGCTTC CTTTTGC GGCGATTC
Truncated TOP2A-A	Forward	Reverse
Subclone into pET28a(+) vector	TTCGAGCTCATGGAAGT GTCACCATTGCAG	CTTGTCGACAATACC ACAGCCAATGGCAG
Subclone into pCMV vector	CGGGTCGACATGGAAGT GTCACCATTGCAG	CTTGGGCCCTCAATG GTGATGGTGATGATG AATACCACAGCCAAT GGCAG
Truncated TOP2A-B	Forward	Reverse
Subclone into pET28a(+) vector	TTCGAGCTCGTAGAAAG CATACTAAACTGGG	CTTGTCGACGGTTGT AGAATTAAGAATAGC TAC
Subclone into pCMV vector	CGGGTCGACATGGTAGA AAGCATACTAAACTGGG	CTTGGGCCCTCAATG GTGATGGTGATGATG GGTTGTAGAATTAAG AATAGCTAC
Truncated TOP2A-C	Forward	Reverse
Subclone into pET28a(+) vector	TTCGAGCTCATTGAAAT CTCAGAGCTTCCC	CTTGTCGACAAACA GATCATCTTCATCTG AC
Subclone into pCMV vector	CGGGTCGACGCCACCAT GATTGAAATCTCAGAGC TCCC	CTTGGGCCCTCAATG GTGATGGTGATGATG AAACAGATCATCTTC ATCTGAC
Flag-tagged human full-length OGT	Forward	Reverse
Subclone into pcDNA3.1 vector	CTTGGTACCATGGCGTC TTCCGTGGGC	AGTGGATCCTCACTT ATCGTCGTCATCCTT GTAATCTGCTGACTC AGTGA CTCAAC