

Figure S1. TCGA database analysis the expression of RBM33 in diverse solid tumors, related to Figure 1. Data shown as medians with SEM.

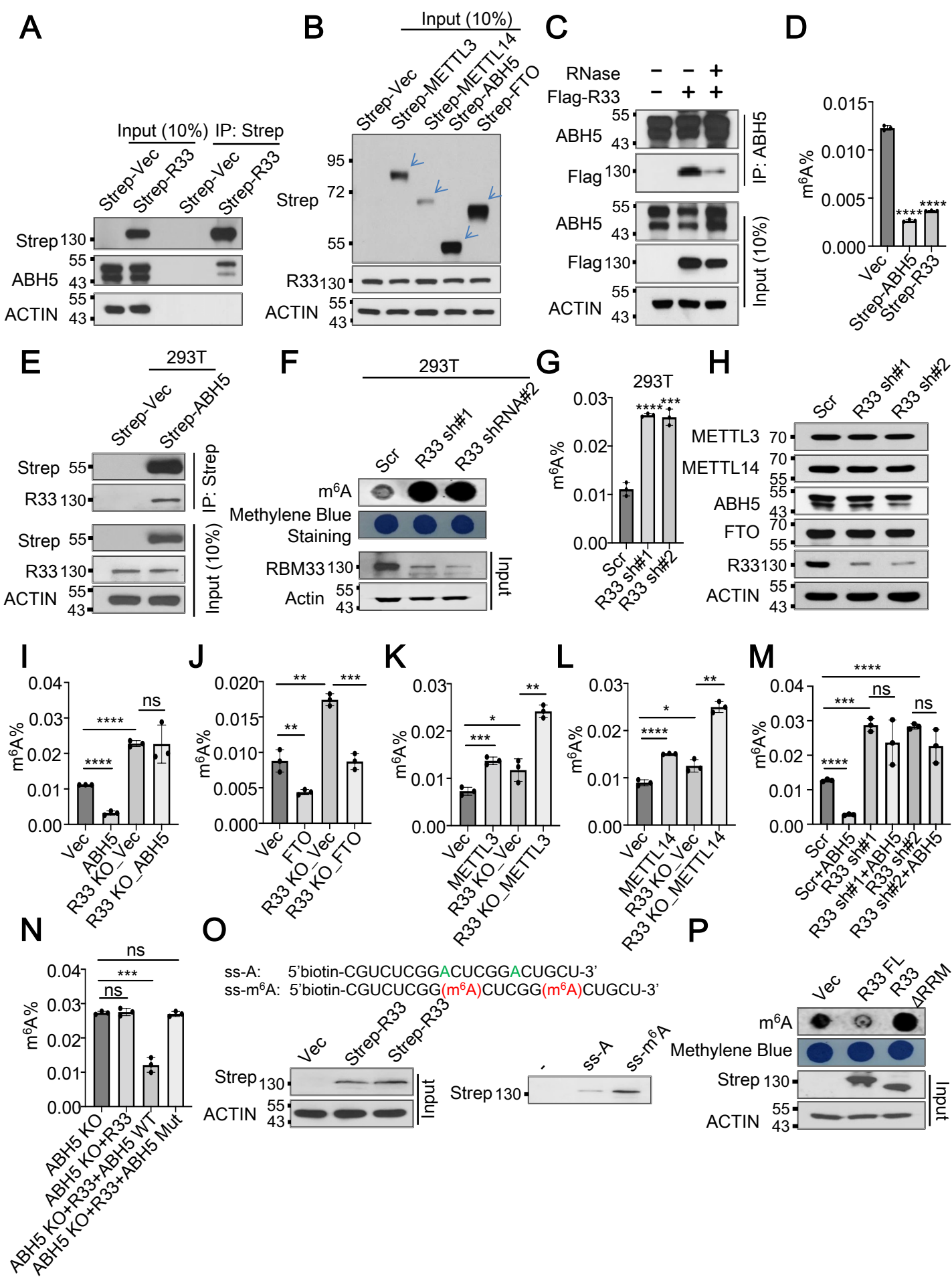


Figure S2 RBM33 depletion specifically inhibits ALKBH5 m⁶A demethylase activity in UM-SCC-1 cells, related to Figure 1 and Figure 2. (A) Co-IP analysis suggests exogenous RBM33 associated with endogenous ALKBH5. Co-IP analysis was performed in UM-SCC-1 cells stably expressing strep-tagged vector, and strep-tagged RBM33 by using Step-Tactin beads. (B) Input of Fig. 1D. (C) The interaction between RBM33 and ALKBH5 relies on RNA substrate. (D) RNA m⁶A methylation analysis suggests that RBM33 overexpression significantly inhibits global RNA m⁶A methylation in UM-SCC-1 cells. Data shown as medians with SEM. (E) Co-IP analysis suggests RBM33 interacts with ALKBH5 in HEK293T cells. (F and G) RNA m⁶A methylation analysis suggests that RBM33 knockdown significantly increases RNA m⁶A methylation in HEK293T cells by dot-blot and RNA m⁶A methylation quantification kit respectively. (H) Western blot analysis showing the effect of RBM33 knocking down on protein levels of m⁶A writers and erasers in UM-SCC-1 cells. (I-L) RNA m⁶A methylation analysis suggests that RBM33 knockdown selectively inhibits RNA m⁶A demethylase ALKBH5 activity in UM-SCC-1 cells. Data shown as medians with SEM. (M) RNA m⁶A methylation analysis suggests that RBM33 knockdown significantly inhibits ALKBH5 m⁶A demethylase activity in UM-SCC-1 cells. Data shown as medians with SEM. (N) RNA m⁶A methylation analysis showing the effect of ALKBH5 knockout on RBM33-mediated down-regulation of RNA m⁶A methylation in HEK293T cells. Data shown as medians with SEM. (O) Exogenous RBM33 preferentially binds to m⁶A-modified RNA oligos. Substrate pull-down analyses suggest both exogenous and endogenous RBM33 preferentially bind to m⁶A-modified RNA oligos. (P) Dot-blot analysis suggests that deletion RNA-recognition motif (RRM) of RBM33 blocks RBM33-mediated down-regulation of global RNA m⁶A methylation.

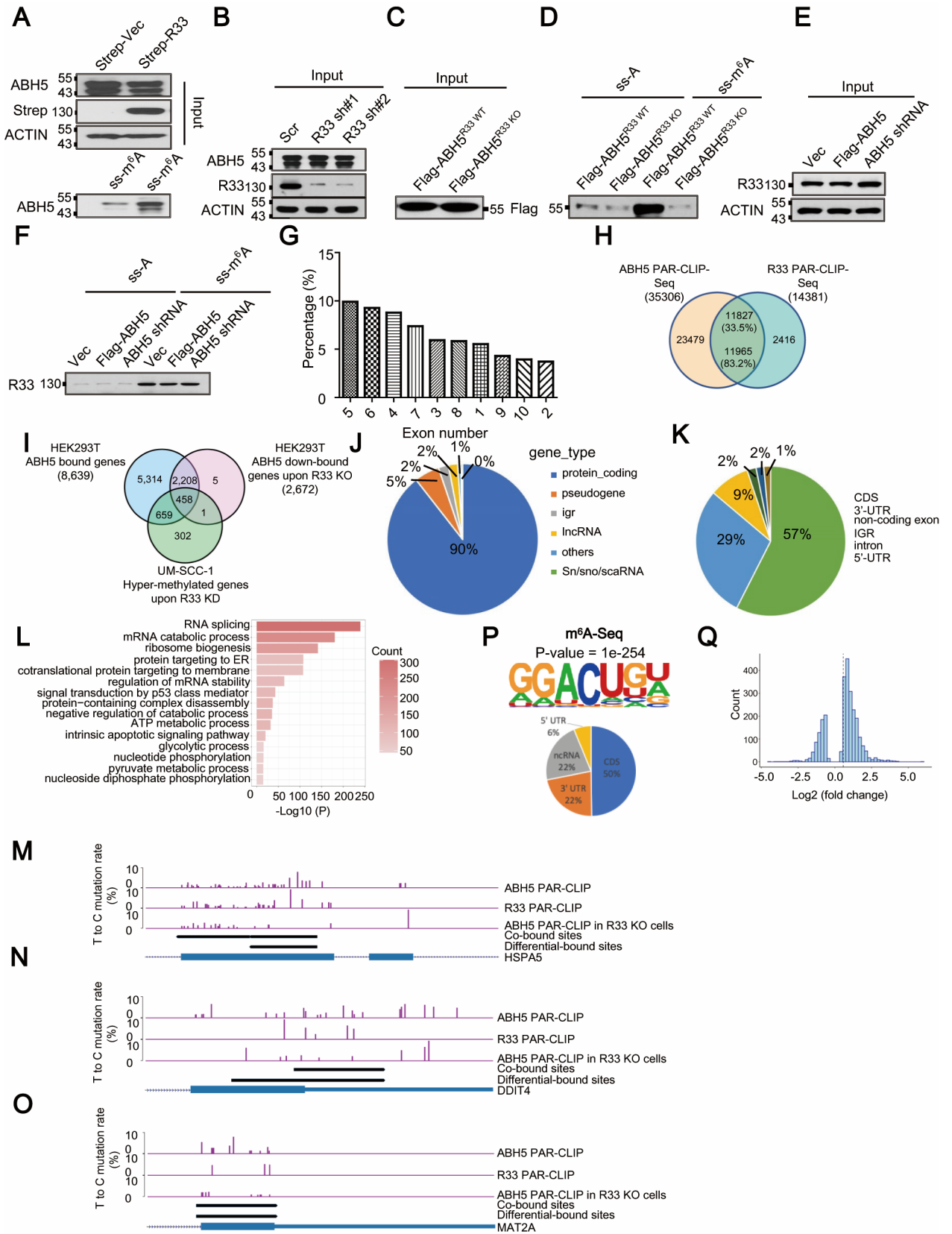


Figure S3 RBM33 binding sites are close to ALKBH5 binding sites, related to Figure 2. (A) m⁶A-modified RNA substrate pull-down analysis suggests that RBM33 overexpression significantly facilitates ALKBH5 substrate accessibility. (B) Input of Figure 2E. (C) Western blot analysis showing protein levels of Flag-tagged ALKBH5 purified from wild-type and RBM33 knockout 293T cells. (D) m⁶A-modified RNA substrate pull-down analysis suggests that RBM33 knockout significantly blocks ALKBH5 substrate accessibility. Flag-tagged ALKBH5 proteins purified from wild-type and RBM33 knockout 293T cells were subjected to RNA oligo pull-down analysis. (E and F) m⁶A-modified RNA substrate pull-down analysis suggests that RBM33 m⁶A-binding ability can not be regulated by ALKBH5. (G) Bar figure showing the exon content for RBM33 binding transcripts by RBM33 PAR-CLIP-seq analysis. (H) PAR-CLIP-seq analyses suggest that the majority of RBM33 binding sites could be overlapped with ALKBH5. (I) Overlap analysis for ALKBH5 bound genes, ALKBH5 down-bound genes upon RBM33 KO in 293T cells and hypermethylated genes upon RBM33 KD in UM-SCC-1 cells. (J) Pie charts show the distribution of the overlapped RBM33 and ALKBH5 PAR-CLIP-seq reads in RNA classes. (K) Pie charts show the distribution of the overlapped RBM33 PAR-CLIP peaks. (L) Gene ontology analysis for the co-bound genes by RBM33 and ALKBH5 via PAR-CLIP-seq analysis. (M-O) IGV browser tracks showing RBM33 and ALKBH5 PAR-CLIP reads in three selected representative genes in wild-type and RBM33 knockout HEK293T cells. (P) Consensus sequence motif identified after analysis of a common m⁶A peaks from three replicates. (Q) Histogram showing that the majority of genes co-regulated by ALKBH5 and RBM33 exhibit an increased m⁶A methylation in ALKBH5 depleted cells.

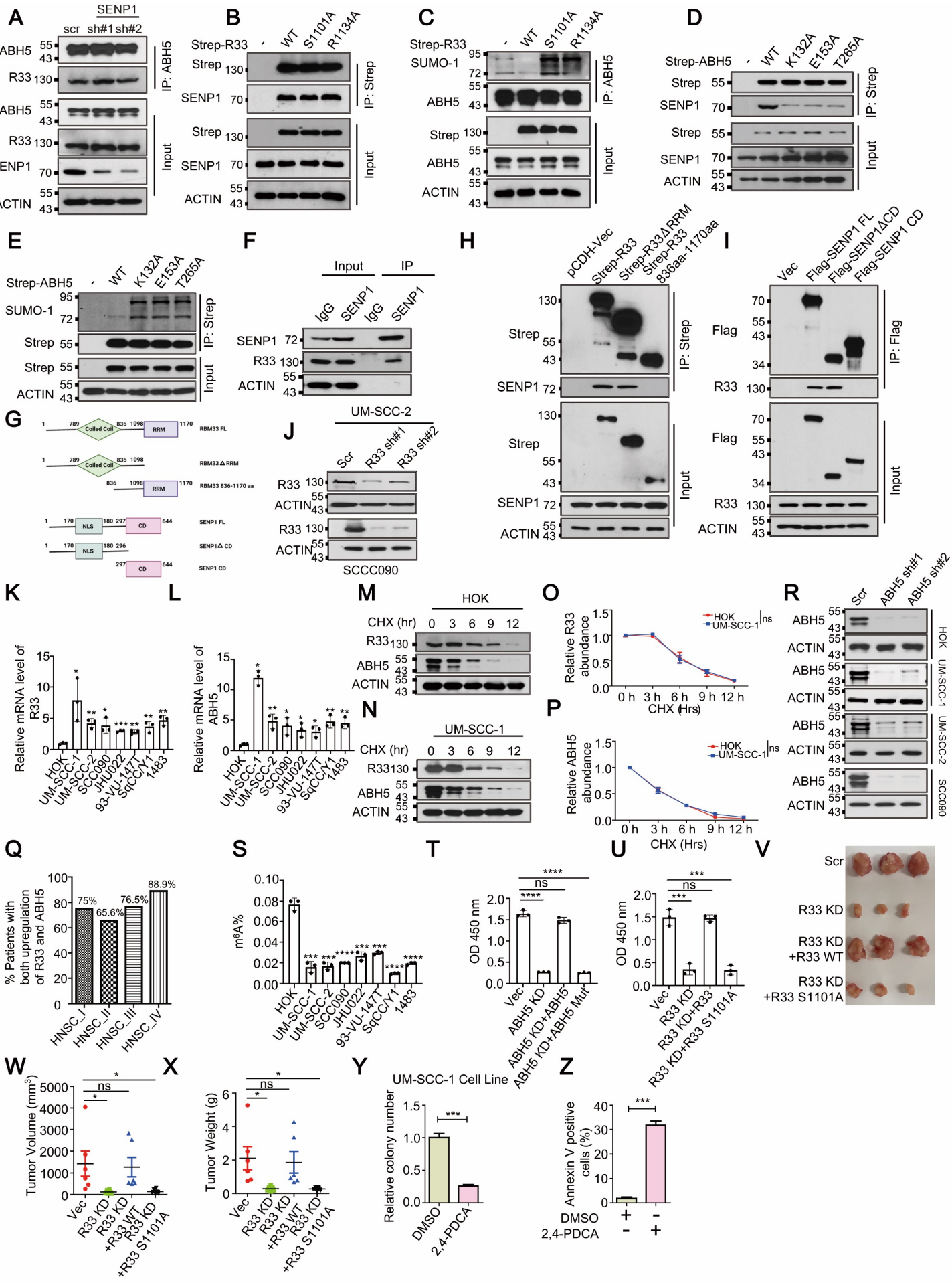


Figure S4 RBM33/ALKBH5 complex regulates HNSCC cell survival in an m⁶A dependent manner, related to Figure 4 and Figure 5. (A) Co-IP analysis suggests that the interaction between ALKBH5 and RBM33 can not be affected by KD of SENP1. (B) Co-IP analysis suggests that the interaction between RBM33 and SENP1 can not be affected by blocking the interaction between RBM33 and ALKBH5. (C) Denaturing IP assay suggests that SUMOylation of ALKBH5 can be extensively induced by overexpression of ALKBH5 interaction defective RBM33 mutant S1101A and R1134A. (D) Co-IP analysis suggests that the interaction between ALKBH5 and SENP1 can be significantly disrupted by blocking the interaction between RBM33 and ALKBH5. (E) Denaturing IP assay suggests that SUMOylation of ALKBH5 can be significantly induced by overexpression of RBM33 interaction defective mutant ALKBH5 K132A, E153A and T265A. (F) Co-IP analysis suggests that endogenous RBM33 interacts with endogenous SENP1. (G) Illustration of the regions for truncation mutants of both human RBM33 and ALKBH5. (H and I) Co-IP analyses suggest that the Coiled coil domain of RBM33 interacts with N-terminal of SENP1. (J) Western blot analysis showing the knockdown efficiency of RBM33 in UM-SCC-2 cells and SCC090 cells. (K and L) RT-qPCR analyses showing transcription levels of both RBM33 and ALKBH5 in normal control HOK cells and HNSCC cell lines. (M-P) Protein half-lives analyses for both RBM33 and ALKBH5 in normal control HOK cells and HNSCC cells. (Q) Immunohistochemical Staining (IHC) analysis suggests that RBM33 and ALKBH5 are simultaneously overexpressed in primary tumor tissues from different TNM stages of HNSC patients. In total, we analyzed 4 samples in stage I HNSCC TNM, 32 samples in stage II, 34 samples in stage III and 18 samples in stage IV. (R) Western blot analysis showing the knockdown efficiency of ALKBH5 in cell lines as indicated. (S) RNA m⁶A methylation levels in normal control HOK cells, and HNSC cell line such as UM-SCC-1, UM-SCC-2, SCC090, JHU022, 93-VU-147T, SqCC/Y1 and 1483. Data shown as medians with SEM. (T) CCK8 analysis showing the effect of wild-type and enzymatic mutant ALKBH5 overexpression on ALKBH5 knockdown-mediated UM-SCC-1 cell proliferation inhibition. (U) CCK8 analysis showing the effect of wild-type and ALKBH5 interaction defective mutant RBM33 (RBM33 S1101A) overexpression on RBM33 knockdown-mediated UM-SCC-1 cell proliferation inhibition. (V-X) Effect of wild-type and ALKBH5 interaction-defective mutant RBM33 overexpression on RBM33 knockdown-mediated tumor growth inhibition of UM-SCC-1 xenograft. (Y) Colony forming analysis showing the colony forming ability of UM-SCC-1 cells with or without 2,4-PDCA treatment. Data shown as medians with SEM. (Z) Annexin V staining analysis showing the cell apoptosis of UM-SCC-1 cells with or without 2,4-PDCA treatment. Data shown as medians with SEM.

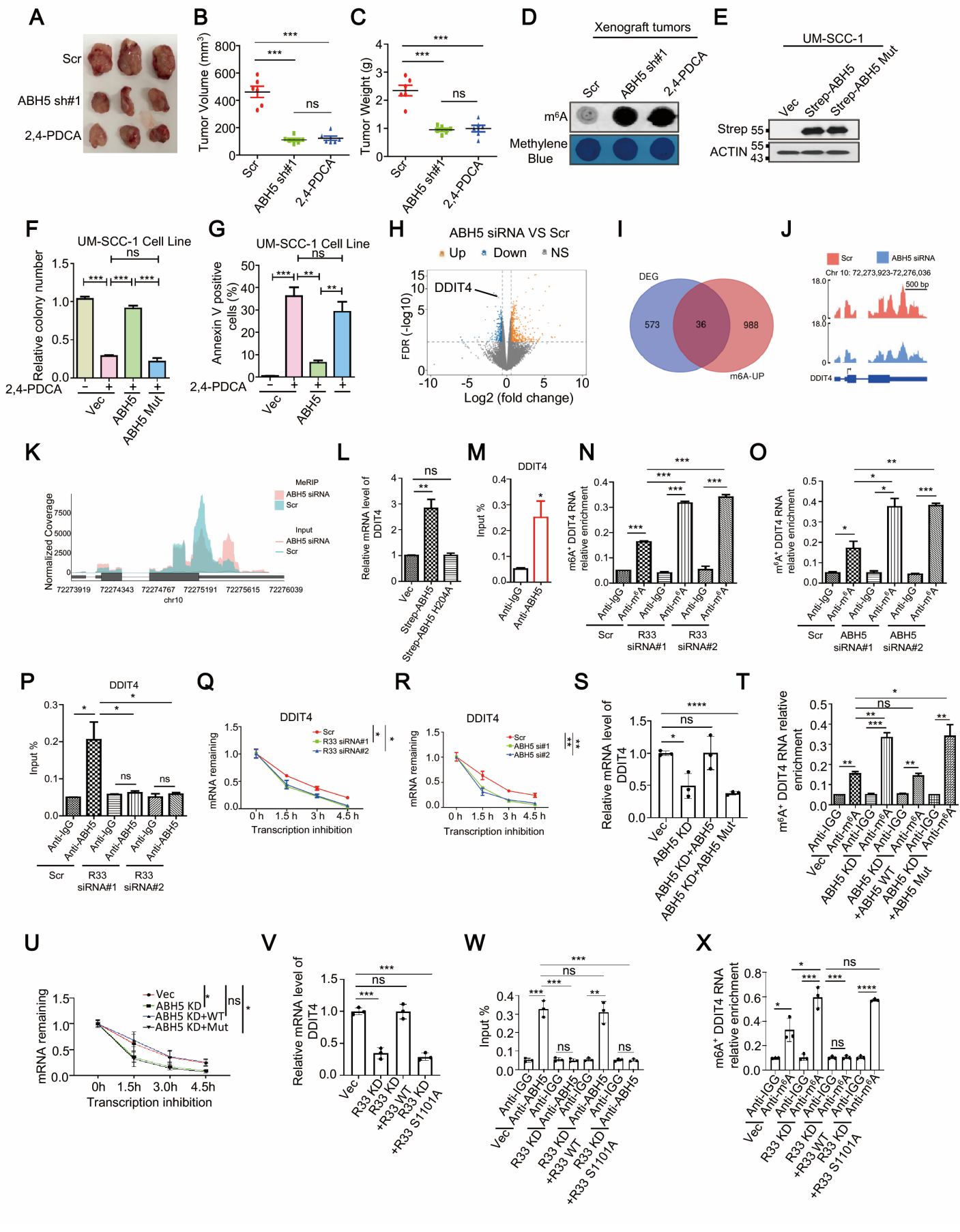


Figure S5 ALKBH5/RBM33 complex regulates gene expression of DDIT4 in an m⁶A-dependent manner, related to Figure 5, Figure 6 and Figure 7. (A-C) Effect of ALKBH5 depletion or inhibition on UM-SCC-1 xenograft growth *in vivo*. Data shown as medians with SEM. (D) Dot blot analysis showing that ALKBH5 depletion or inhibition significantly increases global mRNA m⁶A methylation *in vivo*. (E) Western blot analysis showing the protein expression of wild-type or enzymatic mutant ALKBH5 (ALKBH5 H204A). (F) Colony formation analysis showing that 2,4-PDCA-mediated inhibitory effect on UM-SCC-1 colony forming ability could be rescued by wild-type but not enzymatic mutant ALKBH5 overexpression. Data shown as medians with SEM. (G) Annexin V staining analysis showing the 2,4-PDCA-induced UM-SCC-1 cells apoptosis could be rescued by the wild-type but not enzymatic mutant ALKBH5 overexpression. Data shown as medians with SEM. (H) ALKBH5 knockdown leads to significant gene expression alterations. Differentially expressed genes shown in volcano figure in control or ALKBH5-depleted samples. (I) The Venn diagram shows a number of genes with both significant m⁶A increase and differential expression upon RBM33 and ALKBH5 knockdown. (J) RNA-Seq peak visualization of DDIT4 in control and ALKBH5-depleted cells. (K) m⁶A peak visualization of DDIT4 in control and ALKBH5-depleted cells. (L) RT-qPCR analysis showing the effect of ALKBH5 overexpression on transcripts of DDIT4 in UM-SCC-1 cells. (M) ALKBH5 RIP analysis suggests that ALKBH5 directly binds to transcripts of DDIT4 in UM-SCC-1 cells. Data shown as medians with SEM. (N and O) MeRIP analysis suggests that either RBM33 or ALKBH5 depletion significantly increases DDIT4 mRNA m⁶A methylation. Data shown as medians with SEM. (P) ALKBH5 RIP analysis suggests that ALKBH5 enrichment at DDIT4 transcripts could be blocked by RBM33 KD. Data shown as medians with SEM. (Q) mRNA half-life analysis indicates that RBM33 KD significantly promotes mRNA decay of DDIT4. Data shown as medians with SEM. (R) mRNA half-life analysis suggests that ALKBH5 depletion significantly promotes mRNA decay of DDIT4. Data shown as medians with SEM. (S) RT-qPCR analysis showing the effect of wild-type and enzymatic mutant ALKBH5 (ALKBH5 H204A) overexpression on ALKBH5 knockdown-mediated down-regulation of DDIT4. Data shown as medians with SEM. (T) MeRIP analysis suggests that ALKBH5 depletion-induced up-regulation of DDIT4 mRNA m⁶A methylation can be rescued by wild-type ALKBH5 overexpression. Data shown as medians with SEM. (U) mRNA half-life analysis suggests that ALKBH5 depletion-induced mRNA decay of DDIT4 can be rescued by wild-type but not enzymatic mutant ALKBH5 overexpression. Data shown as medians with SEM. (V) RT-qPCR analysis showing the effect of wild-type and ALKBH5 interaction defective mutant RBM33 (RBM33 S1101A) overexpression on RBM33 knockdown-mediated down-regulation of DDIT4. Data shown as medians with SEM. (W) ALKBH5 RIP analysis suggests that only wild-type RBM33 overexpression can rescue the decreased ALKBH5 enrichment at DDIT4 transcripts mediated by RBM33 knockdown. Data shown as medians with SEM. (X) MeRIP analysis suggests that RBM33 depletion induced up-regulation of DDIT4 mRNA m⁶A methylation can be rescued by wild-type but not ALKBH5 interaction defective mutant RBM33 overexpression. Data shown as medians with SEM.

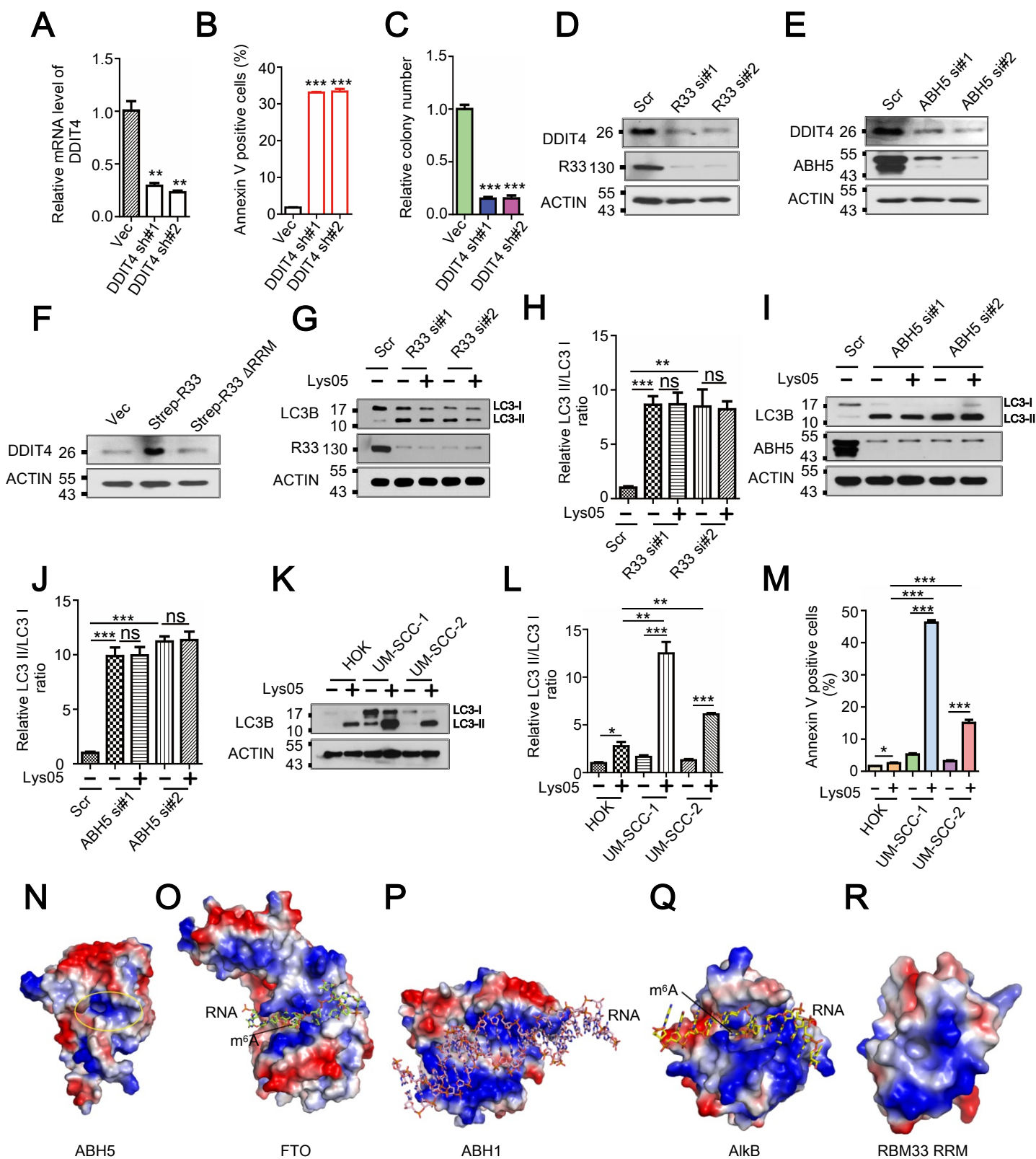


Figure S6 HNSCC cells are more sensitive to autophagy inhibitor treatment as compared to the normal control HOK cells, related to Figure 7 and discussion section. (A) qRT-PCR analysis showing the KD efficiency of DDIT4 in UM-SCC-1 cells. Data shown as medians with SEM. (B) Annexin V staining analysis showing the effect of DDIT4 KD on cell apoptosis of UM-SCC-1. Data shown as medians with SEM. (C) Colony-forming analysis showing the effect of DDIT4 KD on colony-forming ability of UM-SCC-1. Data shown as medians with SEM. (D and E) Western blot analysis showing the effect of RBM33 (D) or ALKBH5 depletion (E) on protein levels of DDIT4 in UM-SCC-1 cells. (F) Western blot analysis showing the effect of wild-type and RBM33 depleting RRM domain overexpression on protein levels of DDIT4 in UM-SCC-1 cells. (G-J) Either RBM33 or ALKBH5 depletion significantly inhibits autophagy in UM-SCC-1 cells. Data shown as medians with SEM. (K-L) Western blot analysis showing the effect of autophagy inhibitor treatment on protein levels of LC3B in the cell lines as indicated. Data shown as medians with SEM. (M) Cell apoptosis analysis showing that HNSC cells, UM-SCC-1 and UM-SCC-2 are more sensitive to autophagy inhibitor-induced cell apoptosis than the normal control HOK cells. Data shown as medians with SEM. (N-R) Comparison of electrostatic surfaces among ALKBH5, FTO, ALKBH1 and AlkB. Positively charged surface is colored in *blue*, negatively charged surface is colored in *red*, and neutral surface is colored in *white*. Alkbh5 (PDB 4O7X) is less positively charged compared with FTO (PDB 5ZMD), ALKBH1 (PDB 6KSF) and AlkB (PDB 4NID). The active center is shown in a *yellow ellipse*.

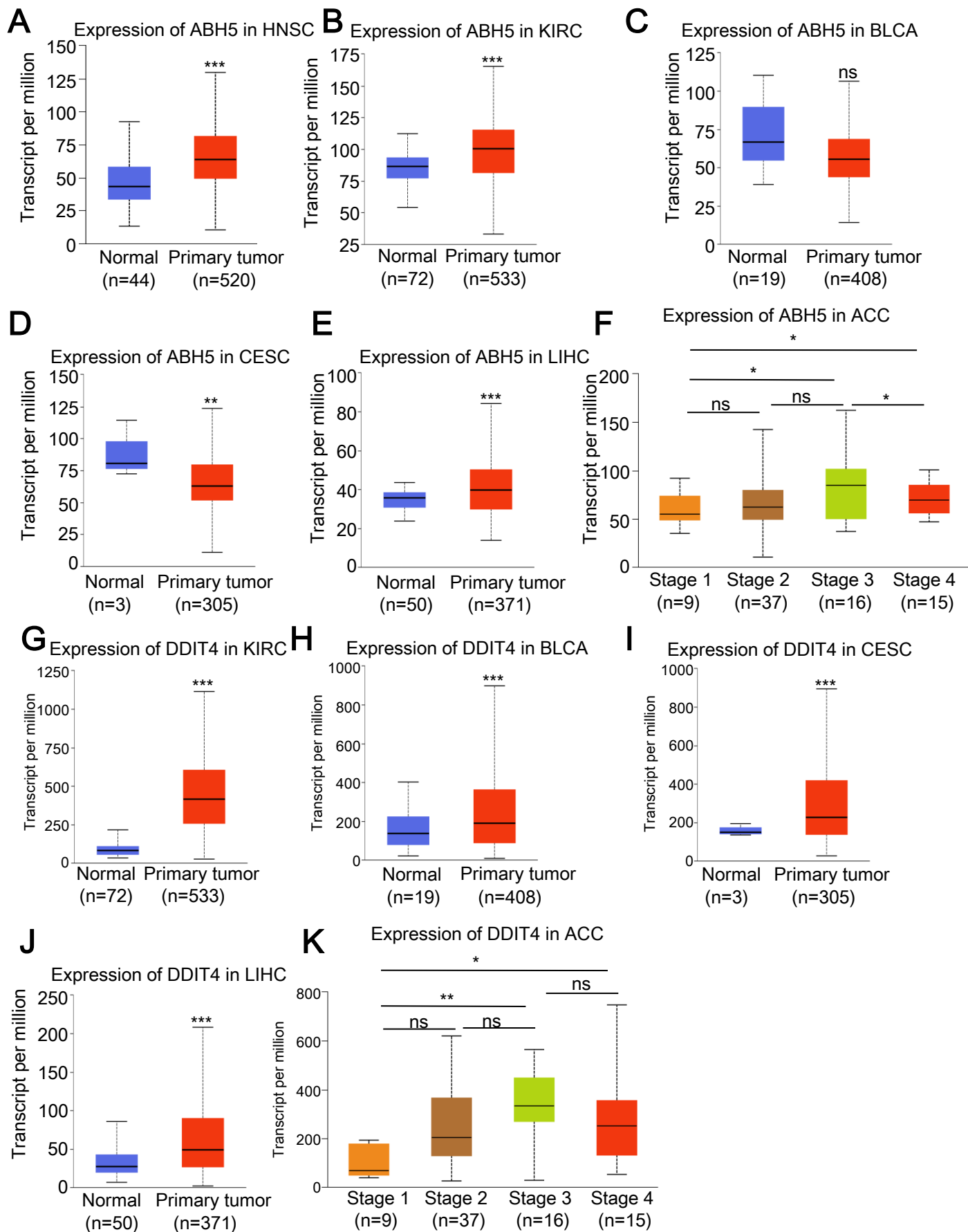


Figure S7 TCGA database analysis the expression of ALKBH5 and DDIT4 in RBM33 overexpressed solid tumors, related to Figure 7 and discussion section. Data shown as medians with SEM.