SUPPLEMENTAL DATA

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S1. Facilitator Guides

Group 1

- 1. What methodology do you use when initially reviewing a peripheral blood smear?
 - a. How do slides come to you for review? (availability/consult patients vs intentional curriculum/slide collection)
 - b. What media do you use for review? (digital v light microscopy)
 - c. How much time do you typically spend reviewing each slide?
 - i. How much variability is there in time spent per slide?
 - d. How much of the slide do you typically review?
 - i. Is there a minimum amount of the slide you feel should be reviewed?
 - ii. If not, how would you recommend a novice "know" they have reviewed enough
- 2. What morphology do fellows struggle the most identifying?
- 3. What are the most crucial diagnoses a graduating fellow should be able to make on PBS review?
- 4. How do you balance education on common yet relatively benign conditions, and rare yet relatively morbid conditions?
- 5. What defines competency at graduation?

Group 2

Review of session 1

- 1. Method of Review
 - a. Most agree that trainees should learn a thorough, systematic method for review
 - b. Many report systematic approaches including specifics around of where to start on the slide, what power to use, how to systematically evaluate each cell line
 - c. A few note that they gestalt the smear more often using remote platforms when the setting is less formal, this may impact the degree of learner education.
- 2. Logistical Considerations
 - a. most utilize slides from patients for whom they have provided direct care
 - b. some use a "slide library" and argue that de-novo evaluation without vignette is important for early learners to pay attention to detail
 - c. many fondly recounted having these flashcards and still have them. However, this appears to not be a method currently used

- d. a few mention that virtual review is not good when large areas of slide require review
- e. some noted their time spent on review, and area of slide covered directly vary with their concern for an diagnosis that dictated urgent discovery (ie APL)
- 3. Disease specific discussion
 - a. Discussion of TMA and acute leukemia dominated the discussion
 - b. only 10% discussion on PLT disorders
 - c. Most focused on diagnosing TMA through identification of schistocytes
 - d. Many also commented that differentiating other causes of hemolytic anemia was important
 - e. Nearly all discussion of white blood cell morphology focused on either distinguishing reactive from malignant
- 4. Systems Based Practice
 - a. Most agreed that trainees should be able to identify diseases that would otherwise result in acute decompensation and death
 - b. Many noted that diagnosis is made on smear immediately, and before confirmatory send out testing results
 - c. most agree that distinguishing malignant from non malignant conditions with similar morphology is incredibly important (reactive cells)
 - d. Some mentioned there is difficulty assigning appropriate value to regional diseases (tick borne -USA) vs arboroinfections worldwide (Malaria)

Focus group 2 discussion

- 1. There is a general agreement trainees should use systematic evaluation. What methodologic tips or best practices should be used to minimize error?
- 2. What are the highest yield WBC morphologies AND disorders of WBCs to identify?
- 3. What are the highest yield RBC morphologies AND disorders of RBCs to identify?
- 4. What are the highest yield PLT morphologies AND disorders of PLTs to identify?
- 5. What are the common, but more benign diagnoses that trainees need to be able to identify? ie those that are low risk but commonly occurring
- 6. How do we reconcile the supposition that academic hematologists use PBS most when they are in the most resource rich environment with fastest turn around time? In contrast community physicians have generally slower turn around time, and less resources, are thought to not utilize this skill.

S2. List of pre-identified hematopathologic morphology

WBC

Atypical lymphocyte (reactive) Band Basophil Blast (undiferentiated) Dysplastic neutrophil Eosinophil Hairy Cell Hypersegmented neutrophil Hypogranular neutrophil LGL Lymphocyte (mature) Metamyelocyte Monocyte vs. monoblast Myeloblast with Auer Rod Myelocyte Neutrophil Neutrophil Neutrophil with toxic granulation Plasma Cell Promyelocyte Sezary Cell Dohle bodies Morula (inclusion)

RBC

Acanthocyte agglutination (RBC) **Basophilic stippling** Bite Cell **Blister Cell** Burr Cell Heinz Body Howell-Jolly Body Hypochromia Macrocytic Microcyte Nucleated RBC Ovalocyte Pappenheimer Bodies Polychromatophil Ring form - malaria Schistocyte Sickle Cell Spherocyte Stomatocyte Target cell Teardrop tetrad - babesia

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Platelet (Giant) Platelet (Hyogranular) Platelet (normal) Platelet clumping (satellitism) Thrombocytopenia

S3. List of pre-identifed hematologic diagnoses

Diagnosis
Acute Leukemia
Acute Leukemia - AML vs ALL
Acute promyelocytic anemia
Anaplasmosis / Ehrlichiosis
Anemia - AIHA
Anemia - iron deficiency
Anemia - megaloblastic
Atypical Lymphocytosis
Babesiosis
CLL
CML
Dimorphic RBC population (prior transfusion)
Hairy Cell Leukemia
ITP
Malaria
Mantle Cell Lymphoma (Leukemic Phase)
May-Hegglin
MDS
Myelophthisic Anemia
Normal Monocytes (vs Monocytic AML)
Platelet Clumping
Reactive leukocytosis
Sickle Cell
T Cell Leukemia
Thalassemia
ТМА / МАНА

S4. Quantitative Statement Coding

Thematic Categories	Ν	% of statements*
Benefit provided by PBS review	31	23.7%
Cell lineage	43	32.8%
Medium used for PBS review	17	13.0%
Procedure of PBS Review	12	9.2%

Source of Slide	22	16.8%
Specific diagnoses	52	39.7%
Specific morphology	34	26.0%

* Does not sum to 100% as statements can be coded to multiple dimensions

1)	Benefit provided by PBS review	Ν	% of category		
	Total	31	-		
	Aid dx	10	32.3%	of benefit provided by PBS review	
	Cant miss (can result in death)	8	25.8%	11	
	Common	2	6.5%	11	
	Guides decision making	9	29.0%	11	
	Not Important	2	6.5%	11	
2)	Cell lineage	Ν	% of ca	tegory	
	Total	43	-		
	RBC	18	41.9%	of cell lineage	
	WBC	21	48.8%	п	
	PLT	4	9.3%	п	
3)	Medium used for PBS review	Ν	% of ca	tegory	
		17	-		
	Digital platorm used	17	N/A	N/A	
4)	Procedure of PBS review	Ν	% of ca	tegory	
		12	-		
	Abbreviated-systematic	1	8.3%	of procedure of PBS review	
	Gestalt	2	16.7%	11	
	Other	3	25.0%	11	
	Thorough - systematic	6	50.0%	1	
- 1					
5)	Source of slide	N	% of ca	tegory	
		22	-		
	Codex (collated slides)	3	13.6%	of source of slide	
	Flashcard	/	31.8%		
	Service patient (clinical context)	12	54.5%		
<i>c</i>)	Specific diagnoses	N	0/ cf cc	togon	
0)	Specific diagnoses	N 50	% 01 Ca	legory	
	Disorders of DDCs	52	-	of all specific diagrapses	
	Macroputia anomia	10	50.8%	of disorders of rod blood colle	
		L 2	×5.0	of disorders of red blood cells	
	iviegaloblastic anemia	3	18.8%		

	Hemoglobinopathies	1	6.3%	н
	Other Hemolytic anemias	4	25.0%	н
	ТМА	7	43.8%	н
	Infections	7	13.5%	of all specific diagnoses
	Anaplasmosis	2	28.6%	of infections
	Babesiosis	1	14.3%	н
	EBV	1	14.3%	н
	Leukocytosis	2	28.6%	н
	Monocytosis	1	14.3%	н
	Malignancies	23	44.2%	of all specific diagnoses
	CML	1	4.3%	of malignancies
	Hairy Cell Leukemia	1	4.3%	н
	CLL	1	4.3%	н
	Burkitt Lymphoma	3	13.0%	н
	APL	4	17.4%	н
	Acute Leukemias	9	39.1%	н
	MPN/MDS	2	8.7%	н
	Mantle Cell Lymphoma	2	8.7%	н
	Pancytopenia	2	3.8%	of all specific diagnoses
	PLT disorders	4	7.7%	of all specific diagnoses
	ITP	1	25.0%	of malignancies
	May-Hegglin	1	25.0%	н
	TP NOS	2	50.0%	н
7	Specific morphology	Ν	% of cat	egory
		34	-	
	RBC Morphology	18	48.6%	of all specific morphology
	Agglutination	1	2.7%	of all RBC morphology
	Bite cell	1	2.7%	п
	Dimorphism	2	5.4%	11
	Hypochromia	2	5.4%	11
	Macrocytosis	3	8.1%	11
	Microcytosis	1	2.7%	11
	Schistocyte	4	10.8%	11
	Spherocyte	4	10.8%	н
	WBC Morphology	13	35.1%	of all specific morphology
	Atypical lymphocyte	2	5.4%	of all WBC morphology
	Auer Rod	1	2.7%	н

Blast	3	8.1%	11
Cup Cell	1	2.7%	11
Promyelocyte	2	5.4%	11
Dysplasia	1	2.7%	11
Pelger-Huet	3	8.1%	11
Maturation (WBC)	2	5.4%	11
Dohle bodies	1	2.7%	11
PLT Morphology	3	8.1%	of all specific morphology
Clumping (PLT)	2	66.7%	of all PLT morphology
Large (Giant) PLT	1	33.3%	11

S5. Participant rating – level of training at skill acquisition

TITLE	1	2	3	4	5	Mean	STD	High	Low	Spread
Atypical lymphocyte (reactive)	2	1	5	0	0	2.38	0.86	3	1	2
Band	8	0	0	0	0	1.00	0.00	1	1	0
Basophil	7	0	1	0	0	1.25	0.66	3	1	2
Blast (undiferentiated)	6	0	2	0	0	1.50	0.87	3	1	2
Dysplastic neutrophil	1	1	5	0	1	2.88	1.05	5	1	4
Eosinophil	7	1	0	0	0	1.13	0.33	2	1	1
Hairy Cell	1	0	7	0	0	2.75	0.66	3	1	2
Hypersegmented neutrophil	5	1	2	0	0	1.63	0.86	3	1	2
Hypogranular neutrophil	1	1	5	1	0	2.75	0.83	4	1	3
LGL	1	0	6	0	1	3.00	1.00	5	1	4
Lymphocyte (mature)	8	0	0	0	0	1.00	0.00	1	1	0
Metamyelocyte	3	0	3	0	2	2.75	1.56	5	1	4
Monocyte vs. monoblast	0	0	5	0	3	3.75	0.97	5	3	2
Myeloblast with Auer Rod	5	1	2	0	0	1.63	0.86	3	1	2
Myelocyte	3	0	3	0	2	2.75	1.56	5	1	4
Neutrophil	8	0	0	0	0	1.00	0.00	1	1	0
Neutrophil with toxic granulation	2	4	2	0	0	2.00	0.71	3	1	2
Plasma Cell	5	0	2	0	1	2.00	1.41	5	1	4
Promyelocyte	4	0	3	0	1	2.25	1.39	5	1	4
Sezary Cell	1	0	5	0	2	3.25	1.20	5	1	4
Dohle bodies	1	3	4	0	0	2.38	0.70	3	1	2
Morula (inclusion)	1	2	3	0	2	3.00	1.32	5	1	4
Acanthocyte	3	2	3	0	0	2.00	0.87	3	1	2
agglutination (RBC)	6	0	2	0	0	1.50	0.87	3	1	2
Basophilic stippling	1	1	5	0	1	2.88	1.05	5	1	4
Bite Cell	5	1	2	0	0	1.63	0.86	3	1	2
Blister Cell	3	0	4	0	1	2.50	1.32	5	1	4

Burr Cell	4	2	2	0	0	1.75	0.83	3	1	2	
Heinz Body	2	1	5	0	0	2.38	0.86	3	1	2	
Howell-Jolly Body	3	2	3	0	0	2.00	0.87	3	1	2	
Hypochromia	5	1	2	0	0	1.63	0.86	3	1	2	
Macrocytic	6	1	1	0	0	1.38	0.70	3	1	2	
Microcyte	6	1	1	0	0	1.38	0.70	3	1	2	
Nucleated RBC	8	0	0	0	0	1.00	0.00	1	1	0	
Ovalocyte	2	2	4	0	0	2.25	0.83	3	1	2	
*** Pappenheimer Bodies	1	0	4	1	2	3.38	1.22	5	1	4	
Polychromatophil	4	1	2	1	0	2.00	1.12	4	1	3	
Ring form - malaria	2	5	1	0	0	1.88	0.60	3	1	2	
Schistocyte	7	1	0	0	0	1.13	0.33	2	1	1	
Sickle Cell	8	0	0	0	0	1.00	0.00	1	1	0	
Spherocyte	7	0	1	0	0	1.25	0.66	3	1	2	
Stomatocyte	2	1	4	0	1	2.63	1.22	5	1	4	
Target cell	6	1	1	0	0	1.38	0.70	3	1	2	
Teardrop	5	1	2	0	0	1.63	0.86	3	1	2	
tetrad - babesia	1	4	2	0	1	2.50	1.12	5	1	4	
Platelet (Giant)	2	0	6	0	0	2.50	0.87	3	1	2	
*** Platelet (Hyogranular)	0	0	4	2	2	3.75	0.83	5	3	2	
Platelet (normal)	8	0	0	0	0	1.00	0.00	1	1	0	
Platelet clumping (satellitism)	6	1	0	0	1	1.63	1.32	5	1	4	
Thrombocytopenia	5	1	2	0	0	1.63	0.86	3	1	2	
Acute Leukemia	5	1	2	0	0	1.63	0.86	3	1	2	
*** Acute Leukemia - AML vs ALL	1	0	4	0	3	3.50	1.32	5	1	4	
Acute promyelocytic anemia	3	0	5	0	0	2.25	0.97	3	1	2	
Anaplasmosis / Ehrlichiosis	0	3	3	1	1	3.00	1.00	5	2	3	
Anemia - AIHA	3	2	3	0	0	2.00	0.87	3	1	2	
Anemia - iron deficiency	6	1	1	0	0	1.38	0.70	3	1	2	
Anemia - megaloblastic	6	1	1	0	0	1.38	0.70	3	1	2	
Atypical Lymphocytosis	2	1	5	0	0	2.38	0.86	3	1	2	
Babesiosis	1	3	3	0	1	2.63	1.11	5	1	4	
CLL	4	0	4	0	0	2.00	1.00	3	1	2	
CML	3	0	5	0	0	2.25	0.97	3	1	2	
Dimorphic RBC population (prior											
transfusion)	0	2	5	0	1	3.00	0.87	5	2	3	
Hairy Cell Leukemia	1	0	6	1	0	2.88	0.78	4	1	3	
ITP	3	0	5	0	0	2.25	0.97	3	1	2	
Malaria *** Mantle Cell Lymphoma	1	6	1	0	0	2.00	0.50	3	1	2	
(Leukemic Phase)	1	0	5	1	1	3.13	1.05	5	1	4	
*** May-Hegglin	0	0	6	1	1	3.38	0.70	5	3	2	
*** MDS	1	0	5	1	1	3.13	1.05	5	1	4	

Myelophthisic Anemia *** Normal Monocytes (vs	2	0	6	0	0	2.50	0.87	3	1	2
Monocytic AML)	0	0	5	0	3	3.75	0.97	5	3	2
Platelet Clumping	6	1	1	0	0	1.38	0.70	3	1	2
Reactive leukocytosis	2	3	3	0	0	2.13	0.78	3	1	2
Sickle Cell	7	1	0	0	0	1.13	0.33	2	1	1
*** T Cell Leukemia	1	0	3	1	3	3.63	1.32	5	1	4
Thalassemia	5	1	2	0	0	1.63	0.86	3	1	2
TMA / MAHA	6	0	2	0	0	1.50	0.87	3	1	2
*** Denotes attending level skill										