

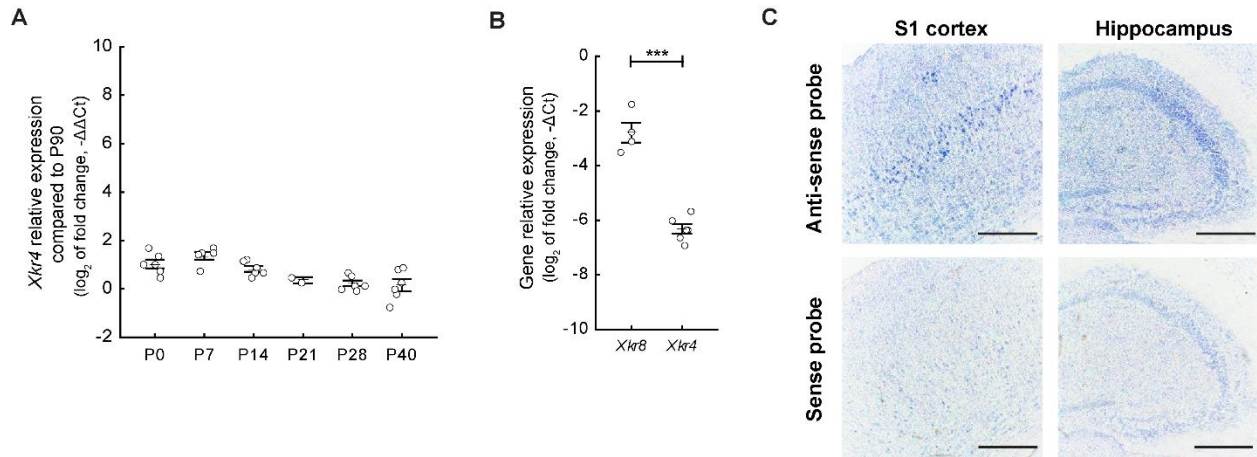
Phospholipid scramblase Xkr8 is required for developmental axon pruning

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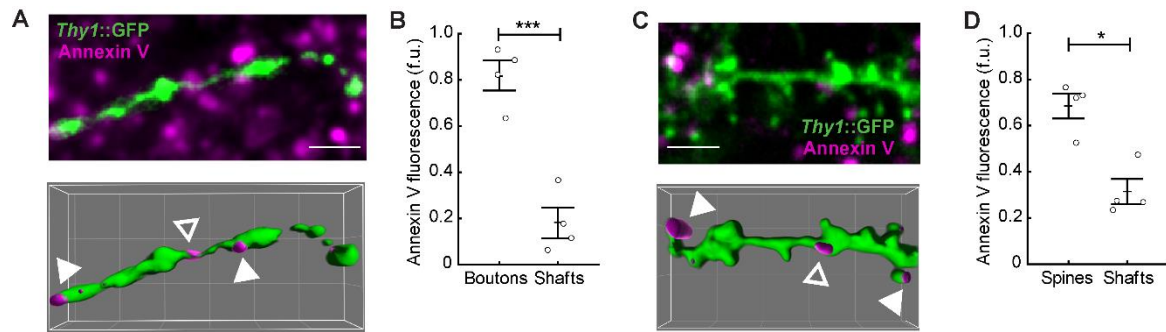
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Appendix Figure S1. Expression of *Xkr8* and *Xkr4* in developing brain. **A** Developmental profile of *Xkr4* mRNA expression in mouse brain from P0 to P40 was measured by quantitative RT-PCR and normalized to the expression of *Gapdh* at a relative time-point and to the expression of the gene of interest at P90. No significant changes in the expression of *Xkr4* were observed during the course of development. (one-way ANOVA, each dot represents an individual mouse, $n = 2-6$ per age group; mean \pm SEM).

B Expression of *Xkr8* and *Xkr4* mRNA in postnatal P0 brain by quantitative RT-PCR normalized to the expression of *Gadph* at P0. Significantly higher expression of *Xkr8* was observed in P0 brains compared to *Xkr4* (one-way ANOVA, each dot represents an individual mouse, $n = 2-6$ per age group; mean \pm SEM, *** $p < 0.001$).

C *Xkr8* mRNA expression in hippocampus and cortex of adult mice visualized by *in situ* hybridization; scale bar 200 μ m.



Appendix Figure S2. PtdSer is preferentially exposed on synaptic structures of pyramidal *Thy1::GFP* neurons.

A-D Organotypic hippocampal slices from *Thy1::GFP* (green) mice were labelled with PtdSer-binding fluorescently tagged Annexin V (magenta) at 16-19 DIV. Fluorescent confocal image z stacks were reconstructed on Imaris to obtain 3D surfaces of axons (**A**) and dendrites (**C**); scale bar 2 μm , grid cell 2 μm . The fluorescence of bound Annexin V within GFP+ was quantified on ImageJ by measuring the integrated density of fluorescent structures within *Thy1::GFP* surface. GFP+ axonal boutons (**B**) and dendritic shafts (**D**) were identified manually by their characteristic structure. Annexin V structures on boutons or spines are labelled with closed arrows, while those on shafts are labelled with open arrows. The data were compared by two-tailed Student's *t*-test, each dot represents OHSC preparation from one mouse, $n = 4$; mean \pm SEM, * $p < 0.05$, *** $p < 0.001$.

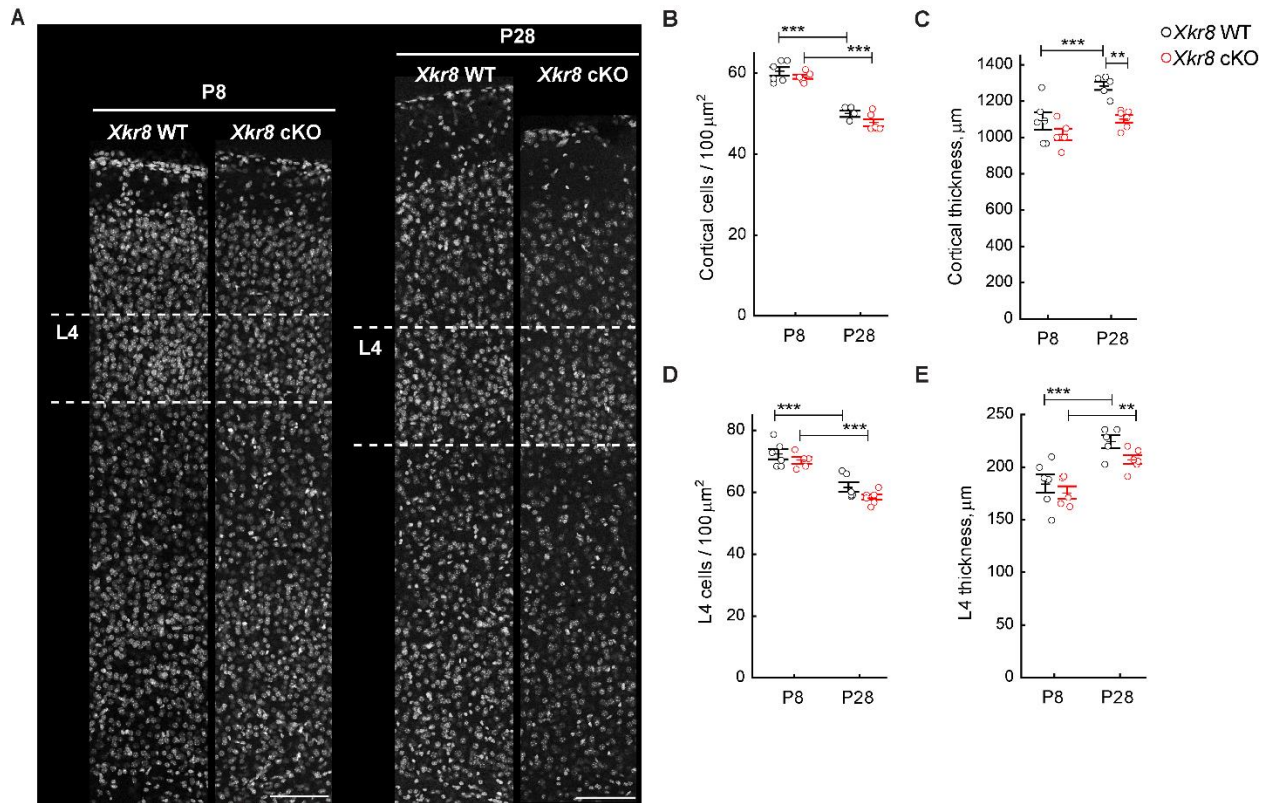
Appendix Table S1. Statistical analysis of data presented in Figure 1–3 using sex as an independent variable. (NOTE: Due to small sample sizes, these results are only indicative.)

Figure	Two-way/Three-way ANOVA				Mean±SEM; n	
	Effect	Degr. of freedom	F	p	Male	Female
1a	SEX	<u>1, 13</u>	<u>0.61</u>	<u>0.45</u>	<u>2.17±0.79; 13</u>	<u>2.79±0.89; 12</u>
	AGE*SEX:	<u>5, 13</u>	<u>0.34</u>	<u>0.88</u>		
	P0				<u>5.87±2.13; 2</u>	<u>8.84±0.49; 2</u>
	P7				<u>2.48±2.58; 3</u>	<u>1.92±1.03; 2</u>
	P14				<u>2.07±1.04; 3</u>	<u>2.61±1.83; 2</u>
	P21				<u>1.20±NA; 1</u>	<u>1.22±0.11; 2</u>
	P28				<u>0.20±0.17; 2</u>	<u>1.72±0.80; 2</u>
	P40				<u>0.58±0.03; 2</u>	<u>0.46±0.05; 2</u>
1g	SEX	<u>1, 26</u>	<u>1.09</u>	<u>0.31</u>	<u>6.3±1.01; 18</u>	<u>8.04±1.51; 18</u>
	AGE*SEX:	<u>2, 26</u>	<u>0.80</u>	<u>0.46</u>		
	P8				<u>8.72±2.51; 6</u>	<u>13.17±3.48; 6</u>
	P15				<u>5.35±1.08; 6</u>	<u>6.50±1.49; 6</u>
	P28				<u>4.95±1.09; 6</u>	<u>4.45±0.87; 6</u>
2c	SEX	<u>1, 26</u>	<u>1.09</u>	<u>0.31</u>	<u>6.3±1.01; 18</u>	<u>8.04±1.51; 18</u>
	AGE*SEX:	<u>2, 26</u>	<u>0.80</u>	<u>0.46</u>		
	P8				<u>8.72±2.51; 6</u>	<u>13.17±3.48; 6</u>
	P15				<u>5.35±1.08; 6</u>	<u>6.50±1.49; 6</u>
	P28				<u>4.95±1.09; 6</u>	<u>4.45±0.87; 6</u>
	GENE*SEX:	<u>1, 26</u>	<u>0.20</u>	<u>0.66</u>		
	WT				<u>6.93±1.40; 9</u>	<u>9.34±1.22; 9</u>
	cKO				<u>5.76±1.51; 9</u>	<u>6.74±2.79; 9</u>
3b	SEX	<u>1, 26</u>	<u>11.59</u>	<u>0.002</u>	<u>0.13±0.006; 18</u>	<u>0.11±0.004; 18</u>
	AGE*SEX:	<u>2, 26</u>	<u>1.82</u>	<u>0.18</u>		
	P8				<u>0.15±0.006; 6</u>	<u>0.12±0.007; 6</u>
	P15				<u>0.12±0.008; 6</u>	<u>0.11±0.005; 6</u>
	P28				<u>0.13±0.011; 6</u>	<u>0.10±0.007; 6</u>
	GENE*SEX:	<u>1, 26</u>	<u>0.01</u>	<u>0.93</u>		
	WT				<u>0.12±0.009; 9</u>	<u>0.10±0.006; 9</u>
	cKO				<u>0.14±0.007; 9</u>	<u>0.12±0.004; 9</u>
3c	SEX	<u>1, 26</u>	<u>5.08</u>	<u>0.03</u>	<u>2.96±0.12; 18</u>	<u>2.66±0.09; 18</u>
	AGE*SEX:	<u>2, 26</u>	<u>0.76</u>	<u>0.48</u>		
	P8				<u>3.39±0.06; 6</u>	<u>2.91±0.11; 6</u>
	P15				<u>2.72±0.21; 6</u>	<u>2.37±0.15; 6</u>
	P28				<u>2.78±0.23; 6</u>	<u>2.69±0.11; 6</u>
	GENE*SEX:	<u>1, 26</u>	<u>0.49</u>	<u>0.49</u>		
	Wt				<u>2.93±0.17; 9</u>	<u>2.72±0.16; 9</u>
	cKO				<u>2.99±0.18; 9</u>	<u>2.60±0.07; 9</u>

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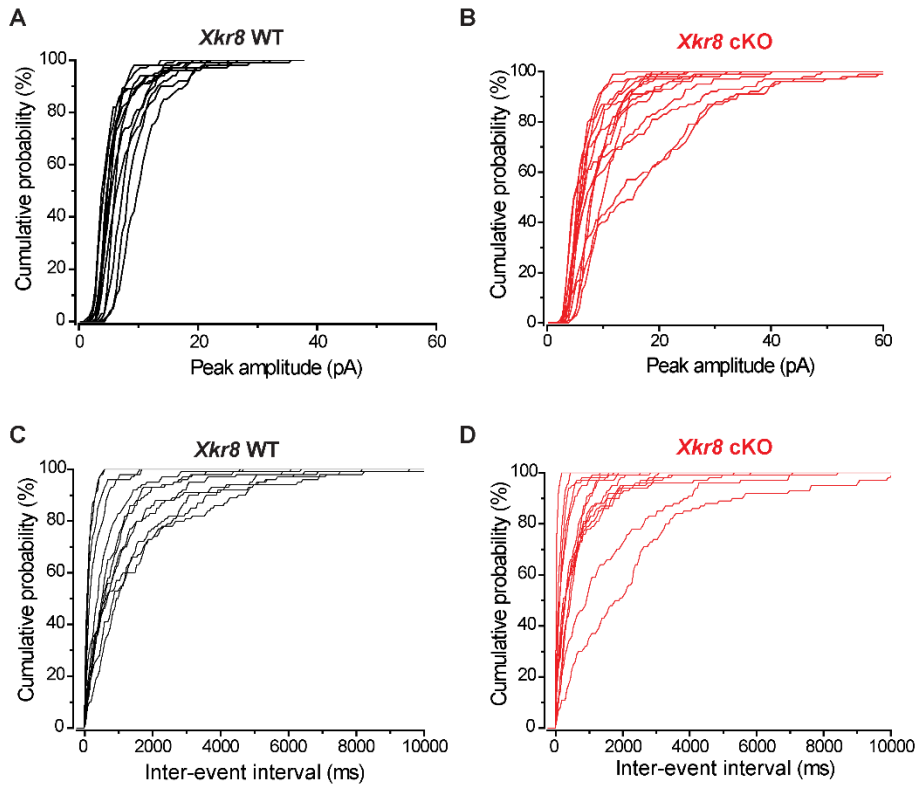
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3e	SEX AGE*SEX: P8 P15 P28 GENE*SEX: WT cKO	1, 26 2, 26 1, 26	2.41 3.23 0.11	0.13 0.06 0.78	13.29±1.63; 18 3.97±0.51; 6 17.07±0.36; 6 18.83±0.57; 6 13.04±2.38; 9 13.54±2.36; 9	12.69±1.83; 18 2.25±0.44; 6 16.34±0.36; 6 19.49±0.47; 6 12.56±2.50; 9 12.83±2.83; 9
3f	SEX GENE*SEX: WT cKO	1, 8 1, 8 	0.72 0.01 	0.42 0.93 	1059±59; 6 1002±70; 3 1116±96; 3	980±68; 6 932±58; 3 1029±131; 3
3j	SEX AGE*SEX: P8 P28 GENE*SEX: WT cKO	1, 17 1, 17 1, 17	0.58 4.75 2.53	0.46 0.04 0.13	48.1±6.4; 12 33.1±9.6; 6 63.0±1.4; 6 39.7±11.6; 6 56.4±4.5; 6	52.3±6.6; 12 49.5±12.1; 6 55.1±6.6; 6 35.1±8.6; 6 69.4±1.7; 6
3k	SEX AGE*SEX: P8 P28 GENE*SEX: WT cKO	1, 17 1, 17 1, 17	0.04 2.46 2.86	0.84 0.14 0.11	23.9±4.1; 12 15.7±3.3; 6 32.1±6.3; 6 27.5±8.1; 6 20.3±2.5; 6	24.7±2.5; 12 22.7±5.0; 6 26.8±1.4; 6 21.7±4.6; 6 27.8±1.7; 6
3l	SEX AGE*SEX: P8 P28 GENE*SEX: WT cKO	1, 17 1, 17 1, 17	3.16 0.48 6.86	0.09 0.50 0.02	2.52±0.44; 12 2.32±0.66; 6 2.72±0.65; 6 1.25±0.16; 6 3.78±0.45; 6	2.02±0.19; 6 2.01±0.31; 6 2.03±0.27; 6 1.49±0.16; 6 2.56±0.18; 6



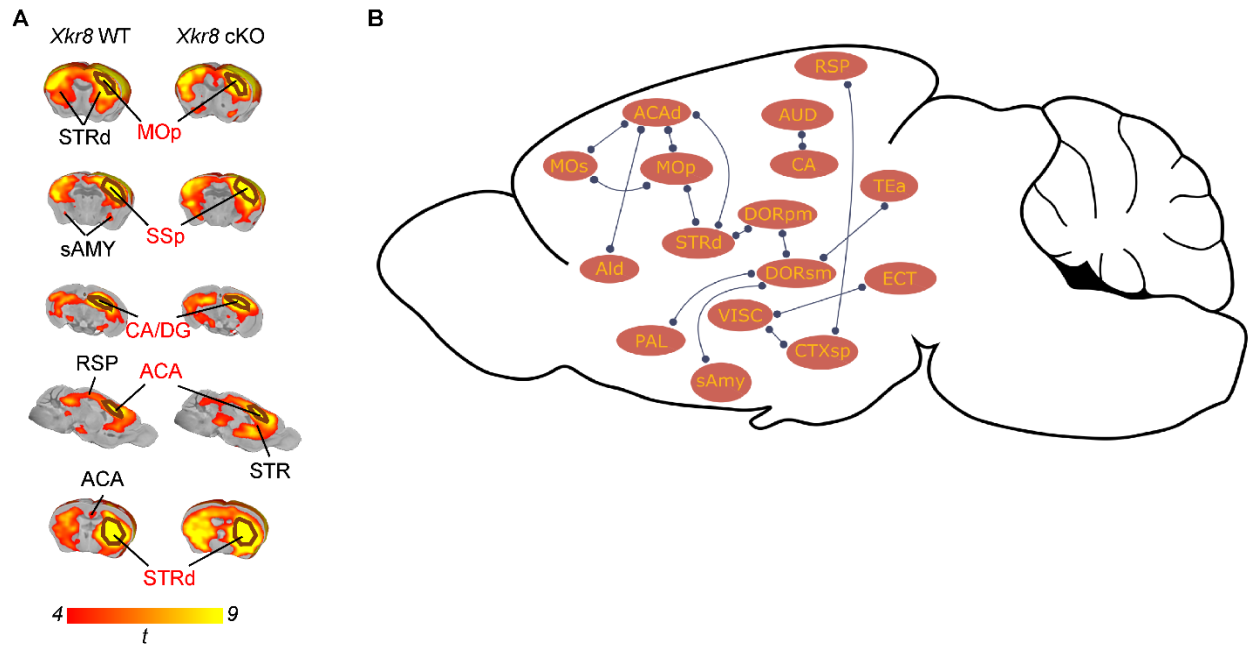
Appendix Figure S3. No decrease in developmental cell loss in *Xkr8* knockout.

A Cortex of *Xkr8* WT and cKO mice was labelled with DAPI to identify nuclei; scale bar 100 μm . **B-E** (B, D) Quantification of cortical and layer 4 (L4) cell density revealed no difference between *Xkr8* WT and cKO animals. (C, E) Cortical thickness and layer 4 (L4) thickness did not increase in *Xkr8* cKO animals (nested design and mixed model ANOVA, each dot represents an individual mouse, $n = 4-6$ per age and genotype group; mean \pm SEM, ** $p < 0.01$, *** $p < 0.001$).



Appendix Figure S4. Synaptic activity of individual neurons in *Xkr8* cKO and WT hippocampus.

A-D (A, B) Individual traces of sEPSC peak amplitudes of *Xkr8* WT and cKO neurons. (C, D) Individual traces of sEPSC inter-event intervals of *Xkr8* WT and cKO neurons. A subset of *Xkr8* cKO neurons with particularly high synaptic activity (B, D) compared to *Xkr8* WT neurons (A, C) was observed.



Appendix Figure S5. Seed-based correlation maps of functional connectivity in *Xkr8* WT and cKO mice.

A Seed-based correlation maps in control and *Xkr8* cKO mice (seed area delineated in black) with red/yellow heat-maps indicating regions exhibiting significant rsfMRI connectivity with the seed volume. The corresponding between-group voxel-wise differences have been reported in **Figure 7**.

B Inter-group comparison of intra-hemispheric rsfMRI connectivity in regionally-parcellated brains using network-based statistics (NBS) showed regions exhibiting increased intra-hemispheric connectivity ($t > 2.1$, FWE corrected at $p < 0.05$). Only pairs of regions characterized by significantly increased intra-hemispheric connectivity independently in both hemispheres were retained for display.

Data information: ACA: anterior cingulate cortex, ACAd: anterior cingulate cortex, dorsal, Ald: agranular insular area, dorsal, sAMY: striatum-like amygdalar nuclei, AUD: auditory areas, CA: Ammon's horn, CTXsp: cortical subplate, DG: dentate gyrus, DORpm: poly-modal association cortex-related region, DORsm: sensory-motor cortex-related region, ECT: ectorhinal area, MOp: primary motor area, MOs: secondary motor area, PAL: pallidum, RSP: retrosplenial area, SSp: primary somatosensory area, STR: striatum, STRd: striatum dorsal region, SUB: subiculum, TEa: temporal association areas, VIS: visual areas, VISC: visceral area.