

Fig S1. Sex-specific RNA splicing of SEPARATOR.

Fifty GFP-positive larvae and fifty GFP-negative larvae at the L1 stage were carefully sorted, and total RNA was extracted from each group. To determine the splicing patterns, RT-PCR was performed using specific primers targeting the 3' end of the Hr5Ie1 promoter sequence and the 5' end of the EGFP coding sequence. (A) The relative locations of the primer target sites are indicated by blue arrows. (B) The PCR products were subsequently purified and subjected to sequencing in order to validate the splicing junctions. The resulting splicing patterns are depicted in the right panel. (C) The relative levels of non-sex-specifically regulated exons (exon4 and exon6) and female-specific exons (exon5a, exon5b) of SEPARATOR were determined through RNA sequencing (RNAseq) analysis. The FPKM (fragments per kilobase per million mapped reads) values of each exon were normalized using the average FPKM of the non-sex-specifically regulated exons (exon4 and exon6). The bar plot displays the means and \pm SD (standard deviation) for triple biological replicates. Statistical significance of mean differences was assessed using a Tukey's multiple comparisons test, with p-values denoted as follows: $p < 0.01^{**}$ and $p < 0.0001^{****}$.



Fig S2. Sex-specific RNA splicing patterns verified through RNA sequencing analysis.

The splicing patterns of SEPARATOR were verified through RNAseq analysis in both GFPpositive and GFP-negative mosquitoes, with triple biological replicates for each condition. The RNAseq reads for the different genotypes were aligned, and the location of exons is indicated at the bottom in blue.



Fig S3. The transgene copy number for SEPARATOR was determined using Oxford Nanopore genome sequencing.

A standard box plot is used to illustrate the coverage distributions of three chromosomes (Chr1, Chr2 and Chr3) and the SEPARATOR transgenes (1174D) in SEPARATOR mosquitoes. The center line represents the median, while the first and third quartiles define the boundaries of the box. The upper and lower whiskers extend from the box to the highest and lowest observed values, respectively, but no further than 1.5 times the Interquartile Range (IQR) from the box. Based on the sequencing depths, the coverage for chromosomes 1, 2, and 3 were 6.31, 6.30, and 6.08, respectively, while the coverage for the SEPARATOR transgenes was 16.14. From the coverage analysis, it suggests that the SEPARATOR transgene (1174D) is present in three copies.



Fig S4. COPAS data processing.

The COPAS raw data was initially filtered for the larvae using size and optical density criteria (Ext/Tof). Next, the particles that exhibited fluorescence (GFP/RFP) were gated. Subsequently, DBSCAN clustering was used to automatically cluster and denoise the data. Finally, the larvae that were GFP-positive were selected.



Fig S5. Transcription profiling and expression analysis of GFP-positive and GFP-negative larvae at the L1 stage in SEPARATOR mosquitoes.

(A) PCA analysis and (B) hierarchical clustering of six samples used for RNA sequencing.

(C) MA-plots were generated to visualize the differential expression patterns between GFPpositive and GFP-negative larvae at the L1 stage in SEPARATOR mosquitoes. In the plot, significantly upregulated (male-enriched) genes (FDR < 0.05 and fold-change > 2) are indicated by red dots, significantly downregulated (female-enriched) genes (FDR < 0.05 and fold-change > 2) are indicated by blue dots, and non-significantly differentially expressed genes are represented by gray dots (FDR > 0.05 or fold-change < 2). Additionally, five well-known sex-enriched genes were marked in the plot. A network visualization was created to illustrate the relationship among enriched Gene Ontology (GO) terms for the upregulated (D) and downregulated (E) genes.



Fig S6. The RNA sequencing depth in SEPARATOR mosquitoes is greater than that observed in previous Matthews's RNA-seq datasets.

In the transcriptome comparison analysis, we employed GFP-positive (Male, L1M) and GFPnegative (Female, L1F) larvae at the L1 stage from SEPARATOR mosquitoes. Additionally, we utilized larvae at the L3 (L3M and L3F) and L4 (L4M and L4F) stages, as well as early pupae (EPM and EPF), mid pupae (MPM and MPF), late pupae (LPM and LPF), and carcass of adult mosquitoes (CM and CF) from Matthews's RNA-seq datasets. The sequencing depth of the RNA-Seq data was achieved through the utilization of an integrated web application known as iDEP.



Fig S7. Male-enriched genes from different developmental stages were identified in the transcriptome comparison analysis.

In our transcriptome comparison analysis, we included L1 stage larvae from SEPARATOR mosquitoes. Furthermore, we incorporated L3 and L4 stage larvae, along with early pupae (EP), mid pupae (MP), late pupae (LP), and adult mosquito carcass (Adult) from Matthews's RNA-seq datasets. The correlation of male-enriched genes in this analysis was visualized using an UpSet plot, facilitated by the integrated web application iDEP.



Fig S8. Female-enriched genes from different developmental stages were identified in the transcriptome comparison analysis.

In our transcriptome comparison analysis, we included L1 stage larvae from SEPARATOR mosquitoes. Furthermore, we incorporated L3 and L4 stage larvae, along with early pupae (EP), mid pupae (MP), late pupae (LP), and adult mosquito carcass (Adult) from Matthews's RNA-seq datasets. The correlation of female-enriched genes in this analysis was visualized using an UpSet plot, facilitated by the integrated web application iDEP.



Fig S9. Conducting a gene ontology (GO) analysis on sex-enriched genes throughout various developmental stages

In our transcriptome comparison analysis, we included L1 stage larvae from SEPARATOR mosquitoes. Furthermore, we incorporated early pupae (EP), mid pupae (MP), and late pupae (LP), from Matthews's RNA-seq datasets. The correlation of GO terms in sex-enriched genes within this analysis was identified and facilitated by the integrated web application iDEP.



Fig S10. Gene expression analysis and clustering methods are used to identify and isolate a distinct set of genes associated with the larvae stage.

Through mfuzz clustering analysis using comprehensive developmental stage data from a previous study, specific genes associated with either L1 or L2-L4 stages were identified. Notably, cluster 17 predominantly consisted of genes expressed in L1, while cluster 1 exhibited gene expression primarily in L2-L4 stages.

Supplementary Tables

- table S1. The sex sorting of SEPARATOR mosquitoes during the initial 15 generations
- table S2. Summary of results from COPAS sorting experiments
- table S3. FPKM of each exons in SEPARATOR mosquitoes
- table S4. Combined annotations exons count

table S5. Enriched Gene Ontology (GO) terms for GFP-positive versus GFP-negative larvae in SEPARATOR mosquitoes

- table S6. Library info_matthews
- table S7. Read count_matthews
- table S8. Read count_matthews
- table S9. Upset Result-Male enriched
- table S10. Upset Result-Female enriched
- table S11. Upset Result-GO term
- table S12. Akbari samples pruned
- table S13. Akbari combined count
- table S14. deseq2 1174D_pos_vs_neg.annotations.cluster_17.significantly_changed
- table S15. deseq2 1174D_pos_vs_neg.annotations.cluster_1.significantly_changed
- table S16. eseq2_1174D_pos_vs_neg.annotations.1st_instar_1tpm.significantly_changed
- table S17. eseq2_1174D_pos_vs_neg.annotations.1st_instar_10tpm.significantly_changed
- table S18. Sequences of primers and gBlock fragment used in this study