

Additional File 1:

Table: Primers and probes used to determine viral abundance through qRT-PCR and qPCR.

Target virus	Primer/Probe sequence 5'-3'		
Bovine Nidovirus	F	TCGCCAACGCTTAACGA	
	R	GGCATTGAGCAGCGAYTCA	
	Pr	CCAAATTAAAGGCGATGCTTGCTTCCG FAM-BHQ1	
Bovine narmovirus-1 (Bulang virus)	F	GGATTGTTACCAAGAGGATCATGAT	
	R	TATCCAGCATCCCTGCTATGC	
	Pr	CCATAATAATACCTCGCGCTGCACATGAGA Qusar 670 BHQ2	
Influenza D virus	F	CCCCRATAGGATCAATGTCGAT	
	R	TTCTCCCCCTATGATTCCATTYTC	
	Pr	CAATTGAAGCCAAAATYCTTGACCAGGG TAMARA BHQ2	
Bovine Coronavirus	F	GCGTCCAAGGCTATATTGCTAA	
	R	CCCAACATTGGATTCTGACATAA	
	Pr	TGCCTTCAACAGGTATT FAM-MGB	
Bovine Rhinitis A (adapted from Kishimoto et al., 2017) [1]	F	CGGTGTAGCGACGACAACA	
	R	TGGGCATCCTTAGTCTCCCCA	
	Pr	GGCCTTGGGACRCCCCTCTCCA Quasar670/BHQ2	
Bovine Rhinitis B (adapted from Kishmitot M et al., 2017) [1]	F	GAACCGCGATTGTGTYYTAGGG	
	R	CGCMACTGAGGTTAGCTTCTCC	
	Pr	CTGTCCCTTGACGGCGTGGC Fam/BHQ1	
Bovine Viral Diarrhea From [2]	F	BVD190-F GRA GTC GTC ART GGT TCG AC	
	R	un_labelled_Primer	
	Pr	V326 TCA ACT CCA TGT GCC ATG TAC un_labelled_Primer	
Bovine alphaherpesvirus-1. From [3]	F	TQ-pesti TGC YAY GTG GAC GAG GGC ATG C Fam-BHQ1	
	R	gB-F TGT GGA CCT AAA CCT CAC GGT un_labelled_Primer	
	Pr	gB-R A GTC GAG CAG ACC CGT GTC un_labelled_Primer	
		BoHV1-P AGG ACC GCG AGT TCT TGC CGC FAM-BHQ1	

Cycling parameters Reverse transcription step involving one cycle at 45°C for 10 min.

The PCR cycling conditions included an RT inactivation/initial denaturation of 95°C for 10 min followed by amplification with 45 cycles of 95°C for 15 sec and 60°C for 45 sec.

References:

1. Kishimoto M, Tsuchiaka S, Rahpaya SS, Hasebe A, Otsu K, Sugimura S, Kobayashi S, Komatsu N, Nagai M, Omatsu T *et al*: **Development of a one-run real-time PCR detection system for pathogens associated with bovine respiratory disease complex.** *J Vet Med Sci* 2017, **79**(3):517-523.
2. Hoffmann B, Depner K, Schirrmeier H, Beer M: **A universal heterologous internal control system for duplex real-time RT-PCR assays used in a detection system for pestiviruses.** *J Virol Methods* 2006, **136**(1-2):200-209.
3. Wang J, O'Keefe J, Orr D, Loth L, Banks M, Wakeley P, West D, Card R, Ibata G, Van Maanen K *et al*: **Validation of a real-time PCR assay for the detection of bovine herpesvirus 1 in bovine semen.** *J Virol Methods* 2007, **144**(1-2):103-108.