# Supplementary Material

## **1 SUPPLEMENTARY FIGURES**



**Figure S1.** Schematic visualization of the eight  $(\chi_1, \chi_3)$  rotamer classes of a Gln–Gln pair, in which a side-chain–side-chain hydrogen bonding interaction could take place. The side-chains of the pair are located on neighbouring  $\beta$ -strands of an antiparallel  $\beta$ -sheet inside the polyQ amyloid core. Depending on the H-bond direction, the eight distinct structural classes are categorized into two forms:  $\downarrow$  and  $\uparrow$ .



**Figure S2.** The stabilities S(t) (see Eq. (1) in Methods of the main text) of the 30 candidate structures for the polyQ amyloid core examined with up-to-1- $\mu$ s molecular dynamics simulations using three distinct force fields: CHARMM36m (Huang et al. (2017)), AMBER14SB (Maier et al. (2015)) and OPLSAA/M (Robertson et al. (2015)). Strikingly, both **M1** and **M2** models demonstrated complete stability when simulated with AMBER14SB and OPLSAA/M, while in CHARMM36m none of the 30 candidates did.

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**Figure S3.** (A) Spatial arrangement of the side-chain and backbone atoms in the polyQ amyloid core model **M2**. Dashed lines represent the distances between the N<sub> $\epsilon$ </sub> atoms of the "a" and "b" types of Gln residues and their nearest backbone O atoms (brown), as well as the N<sub> $\epsilon$ </sub>–O<sub> $\epsilon$ </sub> distances between side-chains (black). (B) Side-chain–backbone distances (left panel), i.e., the distances between the N<sub> $\epsilon$ </sub> atoms of the "a" and "b" types of Gln residues and their closest backbone O atoms. Side-chain–side-chain distances (right panel), i.e., the N<sub> $\epsilon$ </sub>–O<sub> $\epsilon$ </sub> distances between the side chains, either from "a" to "b" or from "b" to "a". The calculations are presented for both the **M1** and **M2** models, using two force fields: OPLSAA/M (green) and AMBER14SB (orange); error bars show standard deviation.



**Figure S4.** Both polyQ<sub>15</sub> models **M1** (orange) and **M2** (green) remained stable throughout  $1-\mu$ s MD simulations in both the AMBER14SB (left) and OPLSAA/M (right) force fields. Shown are the mean  $\chi_1$  dihedral angles in type "a" (red) and "b" (blue) Gln side-chains, calculated over consecutive 20-ns windows; error bars show standard deviation.



**Figure S5.** Distributions of the (A)  $\chi_1$ , (B)  $\chi_2$ , and (C)  $\psi$  dihedral angles in the **M1** (orange lines) and **M2** (green lines) models of the D<sub>2</sub>Q<sub>15</sub>K<sub>2</sub> fibril obtained from 1- $\mu$ s MD simulations using the OPLSAA/M (left) and AMBER14SB (right) force fields. The upper panels show the type "a" conformers (red-shaded distributions) and the lower panels the type "b" (blue-shaded). The gray-shaded regions represent the ssNMR-informed constraints. The dashed vertical lines depict the mean values of the corresponding dihedral angles.



**Figure S6.** Illustration of the structural differences between the (A) **M1** and (B) **M2** models, visualized through views along the fibril axis z (top panels) and along the  $\beta$ -strand direction x (bottom panels). The yellow boxes highlight backbone atoms. (C) Sheet-to-sheet (left panel) and strand-to-strand (right panel) distances calculated for both the **M1** and **M2** models of polyQ<sub>15</sub> using the AMBER14SB (orange) and OPLSAA/M (green) force fields. The dashed black horizontal lines corresponds to the data obtained from X-ray experiments Perutz et al. (2002); Sikorski and Atkins (2005). Error bars represent standard deviation.



**Figure S7.** Distributions of the side-chain dihedral angles for Gln residues in the **M2** model of the poly $Q_{15}$  fibril. The water-facing side-chains (top) show more disorder than those internal to the polyQ amyloid core (bottom), but are nonetheless constrained to eight varyingly prominent specific rotamer states.



**Figure S8.** Ramachandran plot for the  $\beta$ -turn residues (Q38 and Q39) in the Q44-HttEx1 polyQ amyloid core. The distribution was obtained over the last 1  $\mu$ s of the 5- $\mu$ s MD simulation. The  $\beta$ -turn was initially prepared as a type I' conformer (whose canonical dihedrals are indicated with the red/white plus-signs); however, during the simulation it transitioned to the type II (black/green crosses).



**Figure S9.** Ramachandran plots of the N17,  $Q_{44}$ , and PRD domains elucidate the characteristic secondary structures present in these three disparate domains of the HttEx1 protein:  $\alpha$ -helical,  $\beta$ -sheet, and PPII-helical, respectively. The distributions were obtained over the last 1  $\mu$ s of the 5- $\mu$ s MD simulation. For clarity,  $\log_{10}$ (probability density) values below 1.3, characterized primarily by noise, have been excluded.

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**Figure S10.** Ramachandran plots for residues 2–36 in the Q44-HttEx1 fibril. The plots showcase the conformational space explored by each individual residue, revealing variations in backbone torsion angles across the protein structure. The colorbar is calibrated such that a normalized value of 1 corresponds to the bin with the highest frequency among all protein residues. Each bin spans one degree in both  $\phi$  and  $\psi$ . The distributions were obtained over the last 1  $\mu$ s of the 5- $\mu$ s MD simulation.

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**Figure S11.** Ramachandran plots for residues 37–71 in the Q44-HttEx1 fibril. The plots showcase the conformational space explored by each individual residue, revealing variations in backbone torsion angles across the protein structure. The colorbar is calibrated such that a normalized value of 1 corresponds to the bin with the highest frequency among all protein residues. Each bin spans one degree in both  $\phi$  and  $\psi$ . The distributions were obtained over the last 1  $\mu$ s of the 5- $\mu$ s MD simulation.

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**Figure S12.** Ramachandran plots for residues 72–106 in the Q44-HttEx1 fibril. The plots showcase the conformational space explored by each individual residue, revealing variations in backbone torsion angles across the protein structure. The colorbar is calibrated such that a normalized value of 1 corresponds to the bin with the highest frequency among all protein residues. Each bin spans one degree in both  $\phi$  and  $\psi$ . The distributions were obtained over the last 1  $\mu$ s of the 5- $\mu$ s MD simulation.



**Figure S13.** Ramachandran plots for residues 107–110 in the Q44-HttEx1 fibril. The plots showcase the conformational space explored by each individual residue, revealing variations in backbone torsion angles across the protein structure. The colorbar is calibrated such that a normalized value of 1 corresponds to the bin with the highest frequency among all protein residues. Each bin spans one degree in both  $\phi$  and  $\psi$ . The distributions were obtained over the last 1  $\mu$ s of the 5- $\mu$ s MD simulation.

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## REFERENCES

- Huang J, Rauscher S, Nawrocki G, Ran T, Feig M, De Groot BL, et al. Charmm36m: an improved force field for folded and intrinsically disordered proteins. *Nature methods* **14** (2017) 71–73.
- Maier JA, Martinez C, Kasavajhala K, Wickstrom L, Hauser KE, Simmerling C. ff14sb: improving the accuracy of protein side chain and backbone parameters from ff99sb. *Journal of chemical theory and computation* **11** (2015) 3696–3713.
- Robertson MJ, Tirado-Rives J, Jorgensen WL. Improved peptide and protein torsional energetics with the opls-aa force field. *Journal of chemical theory and computation* **11** (2015) 3499–3509.
- Perutz MF, Finch JT, Berriman J, Lesk A. Amyloid fibers are water-filled nanotubes. *Proceedings of the National Academy of Sciences* **99** (2002) 5591–5595.
- Sikorski P, Atkins E. New model for crystalline polyglutamine assemblies and their connection with amyloid fibrils. *Biomacromolecules* **6** (2005) 425–432.