SUPPLEMENTAL DATA

A small molecule inhibitor of PTP1B and PTPN2 enhances T cell anti-tumor immunity

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Supplementary Fig. 1. The chemical structures of ABBV-CLS-484 and Compound 182.



Supplementary Fig. 2. Synthetic Scheme for Compound 182^{*a*} and structure of clinical candidate ABBV-CLS-484. ^{*a*}Reagents and conditions: (a) (ii) Benzyl alcohol, Cs₂CO₃, DMF, r.t., 16 h, (ii) LiOH, MeOH/H₂O, 70 °C, 16 h, 94% over 2 steps; (b) (i) diphenylphosphoryl azide, Et₃N, *tert*-butanol / toluene, 80 °C, 16 h, (ii) diethylenetriamine, 130 °C, 16 h, 40% over 2 steps; (c) methyl 2-bromomethylacetate, K₂CO₃, 60 °C, 16 h, 77%; (d) NFSi, THF, r.t., 6 h, 62%; (e) *tert*-butanol, 0 °C, 1 h; (f) (i) Et₃N, 0 °C – r.t, 2 h, (ii) TFA/CH₂Cl₂, r.t., 1 h, 71% over 2 steps; (g) *t*BuOK, THF, r.t., 1 h, 90%; (h) (i) RockPhos Pd G3, Cs₂CO₃, DMF, H₂O, 80 °C, 3 h, (ii) 4-bromo-2-methylbutan-2-ol, Cs₂CO₃, r.t., 16 h, 53%; (i) H₂, Pd/C, MeOH, r.t., 4 h, 64%.

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Supplementary Fig. 3. HPLC Chromatogram of Compound 182 and Purity. HPLC chromatogram of XI (Compound 182; absorbance measured at wavelength 254 nm) and purity measurement. The compound had a retention time of 4.710 min and a purity of 95% based on peak area (area under the curve).



Supplementary Fig. 4. ¹H NMR spectrum of Compound 182. ¹H NMR spectrum of XI (Compound 182) obtained in deuterated DMSO- d_6 at 400 MHz.



Supplementary Fig. 5. ¹³C NMR spectrum of Compound 182. ¹³C-DEPTQ NMR spectrum of XI (Compound 182) obtained in deuterated DMSO-*d*₆ at 101 MHz.



Supplementary Fig. 6. Electron density for Compound 182. The region about the compound in the PTP1B/Compound 182 complex 2Fo-Fc electron density map is shown as a mesh and is contoured at 2σ. See Table 2 for crystallographic details.



Supplementary Fig. 7. PTP1B crystal structure. Example of the crystal lattice, with the asymmetric unit show in yellow.



Supplementary Fig. 8. Administration of Compound 182 every three days represses tumor growth. AT3-OVA mammary tumor cells were injected into the fourth inguinal mammary fat pads of 8 week-old C57BL/6 female mice. Mice were treated with Compound 182 (CMP-182; 10 mg/kg i.v.; n=5) or saline (n=5) on days (d) 9, 12, 15, 18, 21, 24 and 27 after tumor cell implantation. **a**) Tumor growth was monitored and tumor weights measured. **b**) Tumor-infiltrating lymphocytes or splenocytes including CD44^{hi}CD62L^{hi} CD8⁺ and CD4⁺ central/memory (CM) T cells and CD44^{hi}CD62L^{lo} CD8⁺ and CD4⁺ effector/memory (EM) T cells and CD4⁺CD25⁺FoxP3⁺ regulatory T cells (T_{regs}) were analysed by flow cytometry. **c**) Tumor-infiltrating T cells from (a) were assessed for intracellular granzyme B (GRZMB) and surface PD-1 and TIM-3 in unstimulated tumor-infiltrating CD8⁺ T cells. In (a-c) representative results (means \pm SEM) from at least two independent experiments are shown. Significance for tumor sizes in (a) was determined using a 2-way ANOVA Test and for tumor weights in (a) using a 2-tailed Mann-Whitney U Test. In (b-c) significances were determined using a 2-tailed Mann-Whitney U Test.



Supplementary Fig. 9. Gating strategy for intratumoral lymphocytes. AT3-OVA mammary tumor cells were injected into the fourth inguinal mammary fat pads of C57BL/6 mice. Mice were treated with Compound 182 (10 mg/kg i.v.) or saline on days (d) 6, 8, 10, 12, 14, 16, 18 and 21 after tumor cell implantation. a) Tumor-infiltrating lymphocytes were stained with fluorochrome-conjugated antibodies for CD4, CD8, CD44, CD45, CD62L, PD-1, Tim-3 and Lag-3. Live PI⁻ (propidium iodide) CD45⁺ single cells were gated for CD4⁺ and CD8⁺ T cells and the proportions of CD4⁺ or CD8⁺CD44^{hi}CD62L^{hi} central/memory T cells and CD4⁺ or CD8+CD44hiCD62Llo effector/memory T cells and exhausted CD8+PD-1hiTIM-3hi or CD8⁺PD-1^{hi}Lag-3^{hi} T cells were determined by flow cytometry. **b**) Tumor-infiltrating T cells were stimulated with PMA/Ionomycin in the presence of Golgi Stop/Plug and stained for CD4, CD8, CD45 and intracellular IFN-y and TNF. Live L/D Fix⁻ (LIVE/DEAD[®] Fixable Near IRstain) CD45⁺ single cells were gated for CD8⁺ T cells and the proportions of activated CD8⁺IFN- γ^+ TNF⁺ TILs were determined by flow cytometry. c) Tumor-infiltrating lymphocytes were stained with fluorochrome-conjugated antibodies for CD4, CD8, CD25 and intracellular FoxP3 and GRZMB. Live L/D Fix⁻ (LIVE/DEAD® Fixable Near IR-stain) CD45⁺ single cells were gated for CD4⁺ and CD8⁺ T cells and the proportions of CD4⁺CD25^{hi}FoxP3⁺ T_{regs} and CD8⁺GRZMB⁺ cytotoxic T cell were determined by flow cytometry. d) Tumorinfiltrating lymphocytes were stained with fluorochrome-conjugated antibodies for CD45, CD19, CD11b, F4/80, Ly6C, Ly6G, NK1.1 and TCRβ. Live PI⁻ (propidium iodide) CD45⁺ single cells were gated for CD19⁺ and CD19⁻ cells and proportions of CD19⁺ B cells were determined by flow cytometry. CD19⁻ cells were gated for CD11b^{hi/lo} and F4/80^{hi/lo} cells and the proportions of CD11b^{hi/lo} F4/80^{hi/lo} monocytic (Ly6G^{lo}/Ly6C^{hi}) and granulocytic (Ly6G^{hi}/Ly6C^{hi}) myeloid derived suppressor cells (MDSCs) and were determined by flow cytometry. In (a, d) cells were quantified with Flow-Count Fluorospheres[®] (Beads).



Supplementary Fig. 10. Compound 182 does not affect splenic T cell numbers. a-c) AT3-OVA mammary tumor cells were injected into the fourth inguinal mammary fat pads of 8 weekold C57BL/6 female mice. Mice were treated with Compound 182 (CMP-182; 10 mg/kg i.v.; n=8) or saline (n=8) on days (d) 6, 8, 10, 12, 14, 16, 18 and 21 after tumor cell implantation. a) PD-1, TIM-3 and Lag-3 mean fluorescence intensities (MFI) on PD-1, TIM-3 and Lag-3 on PD-1^{hi} or PD-1^{hi}TIM-3^{hi} or PD-1^{hi}Lag-3^{hi} tumor-infiltrating CD8⁺ T cells were analysed by flow cytometry. b) Spleen weights from mice in (a) were determined. c) Splenocytes from mice in (a) including CD44^{hi}CD62L^{hi} CD8⁺ and CD4⁺ central/memory (CM) T cells and CD44^{hi}CD62L^{lo} CD8⁺ and CD4⁺ effector/memory (EM) T cells, CD4⁺CD25⁺FoxP3⁺ regulatory Т cells (T_{regs}) and granulocytic and monocytic CD11b⁺F4/80^{hi/lo}Ly6C⁺Ly6G^{+/-} (MDSCs) myeloid-derived suppressor cells were analysed by flow cytometry. d) AT3 (n=8) or AT3-OVA (n=5) mammary tumor cells were injected into the fourth inguinal mammary fat pads of 8 week-old C57BL/6 female mice or MC38 (n=7) colorectal tumor cells were xenografted into the flanks of 8 week-old C57BL/6 male mice. CD8⁺ and CD4⁺ EM T cells were analysed by flow cytometry. In (a-d) representative results (means \pm SEM) from at least two independent experiments are shown. In (d) significances were determined using a 1-way ANOVA Test.



Supplementary Fig. 11. NK cell-specific deletion of PTP1B or PTPN2 does not alter NK cell development and tumor growth. **a-b**) Splenic CD3⁺CD19⁻CD49b⁻NK.1.1⁻ T cells and CD3⁻ CD19⁻CD49b^{hi}NK.1.1^{hi} NK cells from **a**) *Ptpn2*^{fl/fl} (n=5) and *Ncr1*-Cre;*Ptpn2*^{fl/fl} (n=5) or **b**) *Ptp1b*^{fl/fl} (n=5) and *Ncr1*-Cre;*Ptp1b*^{fl/fl} (n=5) C57BL/6 male mice were assessed for PTPN2 and PTP1B protein levels by flow cytometry; FMO staining is background staining. **c-d**) Proportions of NK cells (CD45⁺NK1.1⁺, CD49b⁺NKp46⁺CD3⁻TCRβ⁻CD19⁻F4/80⁻) from **c**) *Ptpn2*^{fl/fl} (n=4) and *Ncr1*-Cre;*Ptpn2*^{fl/fl} (n=4) or **d**) *Ptp1b*^{fl/fl} (n=4) and *Ncr1*-Cre;*Ptp1b*^{fl/fl} C57BL/6 male mice in each maturation subset [immature (Imm. CD27⁺, CD11b⁻), M1 (CD27⁺, CD11b⁺), M2 (CD27⁻, CD11b⁺)] and those staining for Ki67 (a marker of proliferation) or CD122 (IL2Rβ/IL15Rβ) were determined by flow cytometry. **e-f**) MC38 *B2m^{-/-}* tumor cells were xenografted (subcutaneous; s.c) into the flanks of **e**) 8 week-old *Ptp1b*^{fl/fl} (n=6) and *Ncr1*-Cre;*Ptp12*^{fl/fl} (n=6) C57BL/6 male mice, or **f**) 8 week-old *Ptp1b*^{fl/fl} (n=7) and *Ncr1*-Cre;*Ptp1b*^{fl/fl} (n=6) C57BL/6 male mice and tumor growth was monitored. In (a-f) representative results (means ± SEM) from at least two independent experiments are shown. In (a) significances were determined using a 1-way ANOVA Test.









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d

CMP-182 [0.1 µM]

CMP-182

[1 µM]



a

b

Supplementary Fig. 12. Compound 182 does not enhance NK cell cytotoxicity and proliferation in vitro. a-b) NK cells from C57BL/6 male mice (n=3 per group) were expanded *ex vivo* in the presence of IL-15 and then pre-treated with Compound 182 (CMP-182; 0.1 or 1 μ g/ml) or vehicle (1% v/v DMSO) in the absence of IL-15. a) NK cells were re-stimulated for 15 min with the indicated amounts of IL-15 and Y694 phosphorylated STAT-5 (p-STAT-5) mean fluorescence intensities (MFIs) determined by flow cytometry. b) Alternatively, NK cells were stained with carboxyfluorescein succinimidyl ester (CFSE, Invitrogen) and then restimulated with the indicated amounts of IL-15 for 96 h and NK cell proliferation (CFSE dilution) monitored by flow cytometry. c-d) NK cells from C57BL/6 male mice (n=3 per group) were expanded *ex vivo* in the presence of IL-15 and then pre-treated with CMP-182 (0.1 or 1 μ g/ml) or vehicle in the absence of IL-15. NK cells were re-stimulated with either IL-12 (10 ng/ml) or IL-18 (100 ng/ml) or with α -NK1.1 or α -Ly49H for 4 h. NK cells were maintained in 10 ng/ml IL-15 throughout the 4 h incubation time. Percentages of c) IFN- γ^+ and d) CD107A⁺ NK cells were determined by flow cytometry. In (a-d) results shown are means ± SEM and are representative of at least two independent experiments.



Supplementary Fig. 13. CRISPR/Cas9-mediated PTPN2 deletion in AT3 and AT3-OVA

tumor cells. PTPN2 was deleted in **a**) AT3 or **b**) AT3-OVA mammary tumor cells using control or *Ptpn2* sgRNAs and CRISPR RNP gene-editing. Cells were lysed and proteins resolved by SDS-PAGE and immunoblotted for PTPN2 (6F3) and re-probed for either tubulin or actin. Representative results from at least two independent experiments are shown.



Supplementary Fig. 14. PTPN2 deletion does not repress the growth of AT3-OVA mammary

tumors. AT3-OVA control cells (Ctl sgRNA; n=7) or those in which PTPN2 had been deleted by CRISPR RNP (*Ptpn2* sgRNA; n=7) were injected into the fourth inguinal mammary fat pads of 8 week-old female C57BL/6 mice and tumor growth was monitored. Representative results (means \pm SEM) from at least two independent experiments are shown.



Supplementary Fig. 15. Inhibition of PTP1B and PTPN2 by Compound 182 and ABBV-CLS-484. Effects of a) Compound 182 (CMP-182) versus b) ABBV-CLS-484 on PTP1B- and PTPN2-catalysed DiFMUP (10 μ M) hydrolysis. Results shown are means \pm SD and are representative of two independent experiments.