A STUDY OF DUST TOXICITY USING A QUANTITATIVE TISSUE CULTURE TECHNIQUE

BY

J. MARKS, M. ANN MASON, and G. NAGELSCHMIDT

From the Central Tuberculosis Laboratory, Cardiff, the Pneumoconiosis Research Unit, Cardiff, and the Safety in Mines
Research Establishment, Sheffield

(RECEIVED FOR PUBLICATION JANUARY 20, 1956)

A number of workers have employed tissue culture techniques in pneumoconiosis research since Fenn (1921) examined the phagocytosis of quartz and coal by rat exudate leucocytes. However, most attention has been directed to the degree of phagocytosis achieved rather than to evaluating accurately the toxicity of dust to the phagocytic cell. Moreover, the techniques used did not provide concordant results in the hands of different workers. example, Policard, Doubrow, and Boucharlat (1929) found that chick embryo macrophages were damaged by ingested quartz whereas Lauche (1931) observed no effect. Kasten (1939) and Belt, Friedmann, and King (1947) found coal was phagocytosed more readily than quartz whereas Buckup (1950) did not detect any difference. A quantitative study of the phagocytosis of quartz was undertaken by Marwyck and Fischer (1951).

Policard and Collet (1953) have injected dusts intraperitoneally into rats and examined the exudate phagocytes after five to 20 hours in an attempt to estimate dust toxicity. However, the phagocytes were not maintained in tissue culture and their observations were qualitative only.

A technique has been recently described (Marks and Mason, 1956) which permits a quantitative assessment of dust toxicity in tissue culture in three days. It has now been used to compare the toxicity of a number of dusts of interests in the field of pneumoconiosis and to examine the effect of protective agents. The chief purpose of the investigation was to determine whether the short-term toxicity of dusts as seen in tissue culture is related to their reported effect *in vivo* in animal experiments. In these, the amount and maturity of silicotic type lesions have usually been taken as an index of pathogenicity in experiments lasting several months or sometimes one or more years.

Methods and Materials

The technique employed in the present investigation was the same as that previously recorded using guinea-pig exudate cells but the explanation of the symbols used to record dust toxicity will be repeated for convenience:—

- + Considerably fewer cells adhere to the glass than in the controls. They are optically denser and less active than normal cells, a large proportion being partially or completely rounded off.
- ++ Less than 10% of the cells remain on the glass.

 Most of the survivors are optically dense whether rounded off or not.
- +++ Fewer than 50 cells remain (inoculum = 2×10^6 cells).

Intermediate degrees of damage designated trace, $+\pm$, and $++\pm$, were also recognized.

Experience showed that determinations of dust toxicity were most reproducible when a moderate degree of damage was taken as the end-point of titrations. The end-point, which can conveniently be used to express the degree of toxicity, was therefore defined as the smallest concentration of dust (in μ g. per 10° cells) required to produce in three days cell damage of the degree $+\pm$ or ++ (standard damage). A reference dust (tridymite 2612) was normally titrated in each experiment as a check on the experimental conditions.

With the exception of four dusts presented by Dr. B. M. Wright, the materials used in the present work were prepared at the Safety in Mines Research Establishment, Sheffield. Information available about their size is summarized in Table 1.

Most of the dusts were fractions graded by size primarily prepared for the animal experiments of Professor E. J. King and his colleagues. The particle size distribution was determined by counting and sizing with the optical or electron microscope. The specific surface was calculated from the size distribution and for several of the dusts checked by air permeability determinations. These gave good agreement for specific surfaces between 0.5 and 3 m.2/g. for isometric materials such as felspar, silica modifications, and coal. The plate-shaped minerals kaolin and mica were used in

Identification		Specific Surface			Mass %	in Given Siz	e Ranges (μ)		
Identification		(m.²/g.)	< 0.23	0·23 to 0·45	0·45 to 0·9	0.9 to 1.8	1.8 to 3.6	3.6 to 7.2	> 7.2
Tridymite	2612	2.2	0.2	0.9	10.6	47.8	40.5	0	0
,,	2424	0.6	0	0	0	0.1	7.2	82.0	10.7
ristobalite	2571	2.3	0.2	1.4	15.8	42	40∙5	0	0
uartz	2531	8-3	13-1	47.0	39.8	0	0	0	0
	2866	2.3	0	0-1	6.6	84.9	8.4	0	0
itreous silica	2835	1.75	0.2	0.6	5.7	42.8	44.5	6.5	0
Bituminous coal	1849	3.4	0.1	1.7	13.6	42.9	27.5	14.3	Ó
	3832	1.1	0	0	0	2.4	16.6	81.0	Ó
	3968	2.5	0	0	0.2	36.8	63.0	0	Ó
team coal (Wright)	4311	0.9	0	0	1.4	5.7	6.5	26.2	60.1
nthracite	3731	1.2	0	0	0	2.3	32.9	48-1	16.7
	3965	2.2	0	0	0.5	25.3	66.6	7.6	0
", (Wright)	4312	1.3	0	Ō	1.7	7.2	21.1	44.4	25.6
elspar	109	3.7	0	8.3	41.2	43.8	6.7	0	0
IPO4	2518	2.9	1.4	4.7	18.7	47.7	27.4	0	Ó
NIPO.	2507	3.1	1.7	5.7	21.0	55.8	15.9	0	Ó
CaF,	2996	2.4	0.2	2.2	31.3	51.8	14.5	Ŏ	Ō
FePO₄	2267	2.3	0.2	3.2	21.1	43.6	31-9	0	0
Caolin	1876	10 approx.		'	!	Not available	· 	·	
lica	2431	10 ,,							
lumina	1342	100 "							
. Luiiiii.	1336	3 ,,				,, ,,			

 $\textbf{Table \ 1}$ Specific surface and size distribution of dust samples employed in the present work

fractions below 4 μ Stokes diameter, and the projected diameters of the particles did not exceed $10\,\mu$. However, as the ratio of diameter to thickness was unknown and variable, the specific surface could not be determined accurately and the value of $10~\text{m.}^2/\text{g.}$ cited is only a rough estimate.

Results

Different Forms of Silica.—The effect on replicate leucocyte cultures of vitreous silica and the three crystalline forms, tridymite, cristobalite, and quartz, was examined. The samples chosen were fairly similar in particle size, their specific surfaces approximating to 2 m.²/g. The toxicity titrations obtained in a representative experiment are shown in Table 2. Tridymite and cristobalite behaved similarly, producing the standard degree of cell damage in a

concentration of $7.5~\mu g$. per 10^{6} cells. Quartz and vitreous silica were eight times less toxic, the standard toxic dose being $60~\mu g$. per 10^{6} cells in both cases.

Particle Size.—The effect of dust samples with different ranges of particle size was studied using tridymite and quartz. The results of typical titrations presented in Table 3 show that toxicity increased with the specific surface of the dust sample. King, Mohanty, Harrison, and Nagelschmidt (1953a) observed a similar relation in vivo between the fibrogenic activity and specific surfaces of flint dust samples administered in equal weights.

Silicates.—One sample each of felspar, mica, and kaolin dusts was examined. The specific surfaces of

TABLE 2
FEFECT OF DIFFERENT FORMS OF SILICA ON EXUDATE CELL CULTURES

Dust Sample		Specific	Toxicity of Given Dust Concentrations (µg./106 cells)								
		Surface (m.²/g.)	120	60	30	15	7.5	3.7	1.8	0.9	
Tridymite Cristobalite Quartz Vitreous silica	2612 2571 2866 2835	2·2 2·3 2·3 1·65	+++ +++ ++± +++	+++ +++ +± ++	+++ +++ Trace +	++ ++ Trace Trace	+±* +± 0 0	+ + 0 0	Trace Trace 0	0 0 0 0	

^{*}Figures in bold type are the titration end-points.

Table 3 effect of specific surface on toxicity in exudate cell cultures

Dust Counts		Specific Surface	Toxicity of Given Dust Concentrations (µg./10 ⁶ cells)								
Dust Sample	(m.²/g.)	120	60	30	15	7.5	3.7	1.8			
Tridymite Tridymite Quartz Quartz	2612 2424 2531 2866	2·2 0·6 8·3 2·3	+ + + + + + + + + + +	+++ ++ ++± +±	+++ +± +± Trace	++ Trace Trace 0	+ ± *	+ 0	Trace 0		

^{*}Figures in bold type are the titration end-points.

Dust Sample		Specific Surface		Toxicity of	Given Dust Cor	ncentrations (µg./	10 ⁶ cells)	
Dust Sam	pie	(m.²/g.)	120	60	30	15	7.5	3.7
Kaolin Felspar Mica Quartz Tridymite	1876 109 2431 2531 2612	10 3·7 10 8·3 2·2	+++ +++ +++ +++	+++ ++ + ++ +++	++* Trace 0 +± +++	Trace 0 0 Trace + + +	: : : ++	· · ·

TABLE 4 COMPARISON OF EFFECTS OF SILICATES AND QUARTZ AND TRIDYMITE ON EXUDATE CELL CULTURES

the latter two samples at an estimated 10 m.2/g. were probably considerably greater than those of the other dusts investigated. The silicates proved unexpectedly toxic, the effect of kaolin and felspar being as great as that of quartz although much less than that of tridymite. Mica was less toxic than the other two silicates. Details of an experiment in which silicates, quartz, and tridymite were compared are given in Table 4.

Coal.—Phagocytes appeared to suffer no harm from coal unless burdened with very large amounts. With such dosages it was uncertain whether the rounding off of the cells and their detachment from the glass should be considered as evidence of toxicity or merely of overloading. Many cells retained their activity despite the ingestion of very large amounts of coal but it should be noted that except in sample 1849 there were no very small particles. When coal was administered as a suspension in Ringer's solution the standard degree of cell damage was produced by all three samples of anthracite in a dosage of 240 μ g. per 10° cells but not by bituminous or steam coal. It was noted that the latter varieties, especially sample 3968, suspended poorly and appeared to be taken up by the phagocytes less well than anthracite. An attempt was made to improve the dispersion of coal particles by first suspending them in concentrations of 6 mg. per ml. in 66% serum or in a preparation of 1% (approximately) lecithin in Ringer's solution. Better coal suspensions were obtained in both cases and the phagocytosis of the bituminous varieties greatly increased. With these modifications the effects of the bituminous and steam coal samples approached those of the anthracite without quite equalling them. It appears likely, therefore, that differences observed between the effects of bituminous or steam coal and anthracite on leucocyte cultures were due only to the difficulty in dispersing the former in suspensions.

The toxicity of combinations of kaolin or felspar with different coals appeared to be additive.

Other Types of Dust.—The following dusts were examined which had previously been studied in animal experiments (Professor E. J. King, personal communication): ferric phosphate, calcium fluoride. and two types each of aluminium phosphate and alumina. They are described in Tables 1 and 5.

The highest concentration of these dusts tested was 120 μ g. per 10⁶ cells which was normally in excess of what could be phagocytosed. phosphate and α-alumina had a negligible effect on leucocyte cultures. The toxicity of aluminium phosphate was definite but not sufficient to produce standard cell damage and was unrelated to the crystalline form. Alumina which had been heated to 820° C. produced standard cell damage in a concentration of 120 μ g. per 10⁶ cells. Calcium fluoride produced the degree of damage designated $+ + in a concentration of 60 \mu g, per 10^6 cells, being$ the only dust of the group to approach quartz in its toxicity. The results obtained in a typical experiment are shown in Table 5. They demonstrate a correlation between toxicity for leucocytes in vitro and the

TABLE 5 EFFECT ON EXUDATE CELL CULTURES OF DUSTS WITH DIFFERENT FIBROGENIC ACTIVITIES

Dust Sample		Mineral Type	Fibrogenic Activity	Toxicity of Given Dust Concentrations (µg./10 ⁶ cells)			
			in vivo	120	60	30	15
AIPO ₄ AIPO ₄ FePO ₄ Alumina (HX1010 heated to 820° C. for 1 hr.)	2518 2507 2267 1342	Crystal structure like quartz Crystal structure like tridymite Crystal structure like quartz γ-alumina	Strong Strong None Strong	+ + Trace + ± *	Trace Trace 0 +	0 0 0 Trace	0 0 0
Alumina (HX1010 heated to 1220° C. for 5 hi	1336	α-alumina	Weak	Trace	0	0	0
CaF ₂	2996	Fluorspar	Weakt	+++	++	Trace	0

^{*}Figures in bold type are the titration end-points. †Policard and Collet (1952) found considerable activity in vivo.

^{*} Figures in bold type are the titration end-points.

Final Concentration of KAI (SO ₄) ₂ ×10 ⁻⁵ M	Equivalent Al Concentration (μg./10° cells)	Toxic Effect with Given Tridymite Concentration (µg./10 ⁶ cells)					
KAI (504)2 × 10 · M	(μg./10° cens)	30	15	7.5	Nil.		
Nil	Nil	+++	+++	++			
4.25	0.5	+++	++	+	Ò		
8∙5	1	++	++	+	Ō		
17	2	+ +	+ +	Trace	ŏ		
34	4	1 + +	· +	Trace	ŏ		
68	8	+++	1 4	+	Тгас		

Table 6 Protection of exudate cell cultures by potassium alum against toxic effect of silica as tridymite

fibrogenic activity reported for the same dusts in vivo.

Inhibition of Silica Toxicity.—Experiments of the type shown in Table 6 were set up in which different concentrations of silica (tridymite 2612) were added to leucocyte cultures along with varying amounts of a potential antagonist. Antagonists were used in concentrations found by preliminary experiment to have little or no effect themselves on the cells. The most potent of those examined was potassium aluminium sulphate which protected cells very considerably against the effect of silica, presumably by virtue of aluminium hydroxide formed at the pH of the culture medium (7.2 - 7.4). Alumina HX 1010 (unheated), the preparation used by Gardner, Dworski, and Delahant (1944) to depress silica activity in vivo, provided only a slight degree of protection in tissue culture. A mixture of aluminium metal dust and alumina was very slightly more protective than alumina alone. Iron in the form of ferric chloride had a weak protective effect against silica, not greater than that of alumina. Mica and felspar dusts were also slightly protective but kaolin was not. The concentrations used of these three dusts were 30, 15, and $7.5 \mu g$. per 10^6 cells respectively, the highest possible concentrations without causing confusion by their own toxicity.

Discussion

The relative toxicity of different forms of silica to leucocytes in tissue culture found in the present work agreed well with their pathogenicity and fibrogenic qualities in vivo as described by Gardner (1938) and by King and others (1953b). The high toxicity of cristobalite found in vitro is of interest since this type of silica has been reported to be responsible for a particularly severe form of silicosis in man (Vigliani and Mottura, 1948). Fairly close agreement was also found between the toxicity in tissue culture and the effects in vivo of aluminium and ferric phosphate and alumina heated to different degrees reported by King. King has found calcium fluoride weakly fibrogenic but the tissue culture results accord better with those of Policard and

Collet (1953) who found calcium fluoride as pathogenic as quartz when injected intraperitoneally into rats. Coal dust which is relatively harmless *in vivo* had little effect on leucocyte cultures unless the cells were grossly overloaded with it. No clear difference related to type of coal could be demonstrated.

It appears probable from the correlation now demonstrated between fibrogenic activity and toxicity to leucocytes that injury to the phagocytic cell by dust is the stimulus for fibrous tissue formation in pneumoconiosis. In the intact animal this stimulus is presumably maintained for long periods by the continuous recruitment of fresh macrophages replacing those that die. In addition, no doubt, many of the cells containing low concentrations of dust survive for considerable periods during which time they might suffer damage and promote fibrosis.

The inhibition of silica toxicity by alumina in animals reported by Gardner and others (1944) and that by iron reported by Kettle (1932) have been reproduced in tissue culture although in neither case was the protection in vitro striking. However, the concentrations of iron used in the present work (10⁻³ to 10⁻⁴ M) are difficult to compare with those used by Kettle who coated silica particles with two-thirds of their weight of iron oxide. Gardner found great differences between the protective powers of different batches of alumina and it is possible that his most potent sample, HX 1010, which was used in the present work, has deteriorated. In contrast to the relatively weak actions of alumina and iron, considerable protection against silica toxicity was afforded to leucocytes by potassium alum, no doubt by means of aluminium hydroxide formed in the culture medium. The toxicity of silica was also found to be reduced in tissue culture by felspar and mica dusts, recalling the unexpectedly slight effect in vivo of quartz accompanied by these minerals in the form of granite as reported by Gardner (1938) and King, Ray, Harrison, and Nagelschmidt (1950).

The silicates, particularly kaolin and felspar, produced effects in vitro for which there have been no clear-cut equivalents in vivo. In experimental

animals the silicates produce far less fibrosis than does quartz (Cummins, 1937; King, Harrison, and Nagelschmidt, 1948; Mohanty, Roberts, King, Harrison, and Nagelschmidt, 1953). However, the published experimental work is not altogether consistent. For example, Kettle (1932) produced necrotic lesions in mice by the subcutaneous injection of kaolin, and Rüttner, Bovet, Weber, and Willy (1952) fibrous nodules by intraperitoneal injection. Mica was found lethal to phagocytes by Lemon and Higgins (1935) and by Policard (1934). A complicating factor has been introduced by the observation of Belt and King (1945) and King, Gilchrist, and Rae (1947) that the fibrogenic activity of mica was greatly increased if the dust had been treated with acid. This finding suggests that the effect of silicates in tissue culture may have depended to some extent on the experimental conditions and that investigation into the influence of these would be desirable.

It is not intended to suggest on the basis of the present work that tissue culture could be used to estimate the fibrogenic activity of different dusts. A much more comprehensive investigation than the present one would be necessary to define the utility of the technique in this respect. It does appear, however, that the rapidity and sensitivity of the quantitative tissue culture technique described above should make it useful for screening substances for protective action against dust toxicity and for investigating the mechanism of such toxicity. the latter case a useful line of approach would appear to be the physical and chemical modification of the surface of dust particles, and a biochemical study of the mechanism of cell damage. The difference between the biological effects of tridymite and cristobalite on one hand and quartz and vitreous silica on the other requires investigation in this manner, as does the augmenting action of fluorine etching on the pathogenicity of quartz (King and others, 1953c).

Summary

A fairly close agreement was found between the toxicity to leucocytes *in vitro* of a number of dusts of interest in the field of pneumoconiosis and their fibrogenic activity as reported by other workers. Silicates proved more toxic in tissue culture, however, than experiments on animals would have suggested. Silica toxicity to leucocytes *in vitro* is antagonized by certain aluminium compounds.

We are indebted to Professor J. Gough for helpful comments on the paper.

REFERENCES

Belt, T. H., and King, E. J. (1945). Spec. Rep. Ser. med. Res. Coun. (Lond.), No. 250, p. 29.

—, Friedmann, I., and King, E. J. (1947). J. Path. Bact., 59, 159. Buckup, H. (1950). Arch. Hyg. (Berl.), 132, 273. Cummins, S. L. (1937). Brit. J. exp. Path., 18, 395. Fenn, W. O. (1921). J. gen. Physiol., 3, 439.

Gardner, L. U. (1938). In Silicosis and Asbestosis, ed. A. Lanza, p. 257. Oxford Univ. Press, London.

—, Dworski, M., and Delahant, A. B. (1944). J. industr. Hyg., 26, 211.

Kasten, W. (1939). Arch. Gewerbepath. Gewerbehyg., 9, 337. Kettle, E. H. (1932). J. Path. Bact., 35, 395. King, E. J., Gilchrist, M., and Rae, M. V. (1947). Ibid., 59, 324.

—, Harrison, C. V., and Nagelschmidt, G. (1948). Ibid., 60, 435.

—, Mohanty, G. P., Harrison, C. V., and Nagelschmidt, G. (1953a). British Journal of Industrial Medicine, 10, 76.

—, —, —, (1953b). Ibid., 10, 9.

—, —, —, (1953b). Ibid., 10, 9.

—, —, —, (1953b). Ibid., 10, 9.

—, —, —, (1953b). Arch. industr. Hyg., 7, 455.

—, Ray, S. C., Harrison, C. V., and Nagelschmidt, G. (1950). British Journal of Industrial Medicine, 7, 37.

Lauche, A. (1931). Verh. disch. path. Ges., 26, 107. Lemon, W. S., and Higgins, G. M. (1935). Amer. Rev. Tuberc., 32, 243.

Marks, J., and Mason, M. A. (1956). British Journal of Industrial Medicine, 13, 38.

Marwyck, C. van, and Fischer, E. (1951). Arch. Hyg. (Berl.), 135, 161. Mohanty, G. P., Roberts, D. C., King, E. J., Harrison, C. V., and Nagelschmidt, G. (1953). J. Path. Bact., 65, 501.

Policard, A. (1934). J. industr. Hyg., 16, 160.

—, and Collet, A. (1953). Rev. Hémat., 8, 132.

—, Doubrow, S., and Boucharlat, M. (1929). C.R. Acad. Sci. (Paris), 189, 593.

Rüttner, J. R., Bovet, P., Weber, R., and Willy, W. (1952). Naturwissenschaften, 39, 332.

Vigliani, E. C., and Mottura, G. (1948). British Journal of Industrial Medicine, 5, 148.