

Supplemental Information

Figures S1-S8

Table S8

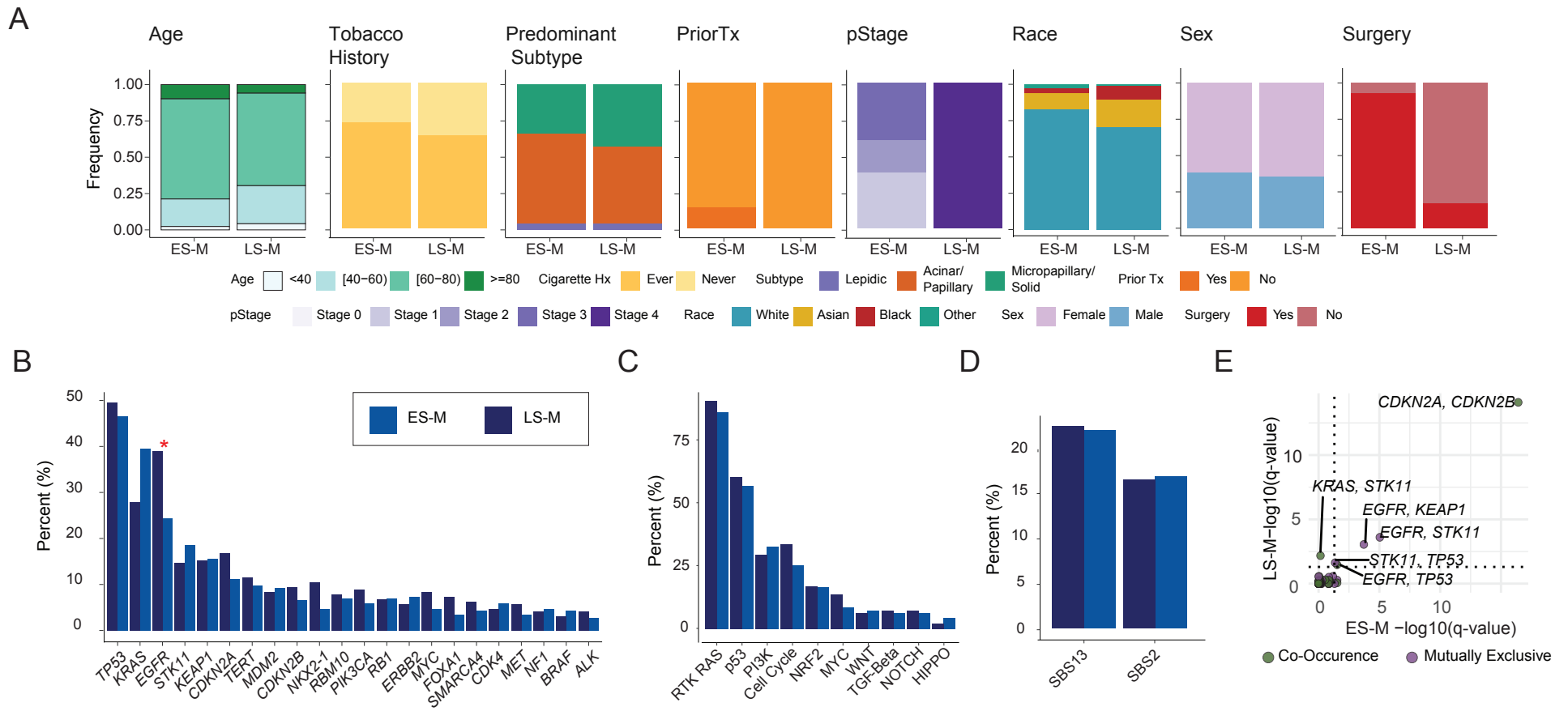
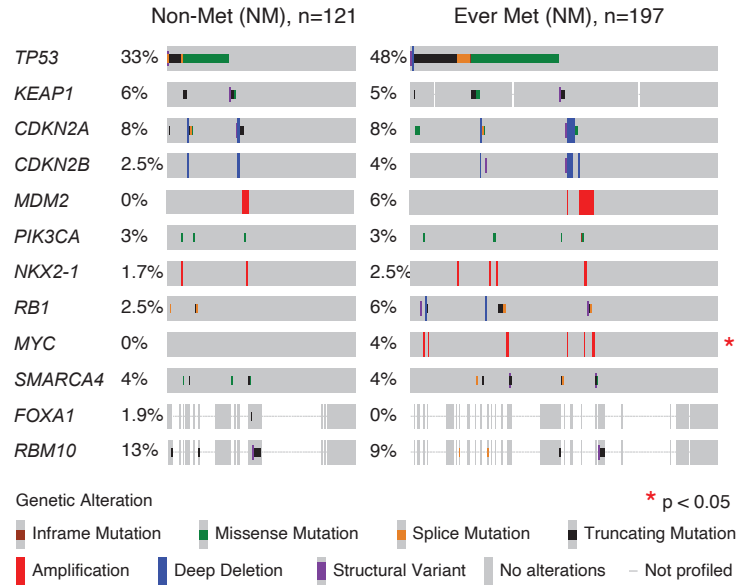


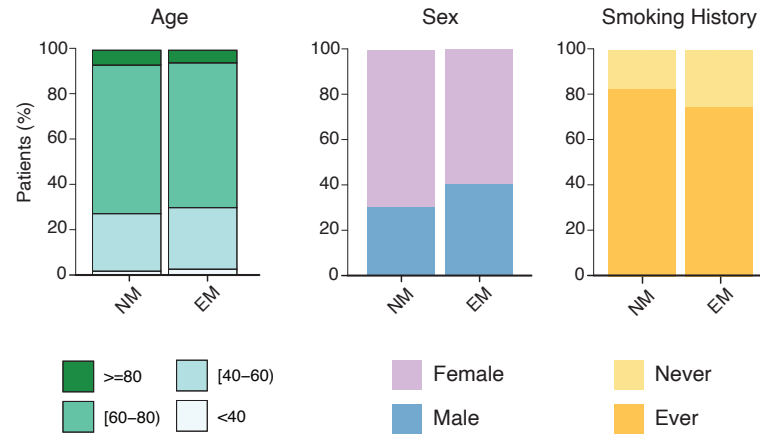
Figure S1. Clinicopathologic and genomic differences between early-stage metastatic (ES-M) and late-stage metastatic (LS-M) primary tumors, Related to Figure 1. (A) Bar plots comparing clinicopathologic features between ES-M and LS-M groups. (B) Bar plots showing the percentage of samples with gene alterations. (C) Bar plots depicting the percentage of samples with pathway alterations. (D) Bar plots displaying the percentage of samples with APOBEC signatures present. (E) Scatter plot showing q -value of gene pairs for co-occurrence and mutual exclusivity in ES-M and LS-M tumors. Dotted lines represent $q=0.05$; significant gene pairs labeled. Statistical analyses: (A-E) Fisher's exact test. $*q<0.05$, adjusted for false-discovery rate (FDR). Hx, history; pStage, pathologic stage; Tx, treatment.

Figure S2

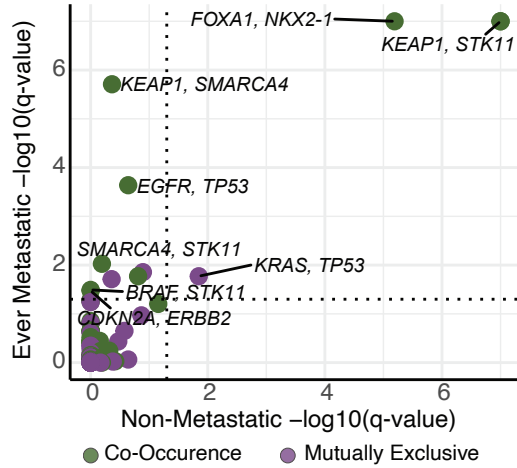
A



B



C



D

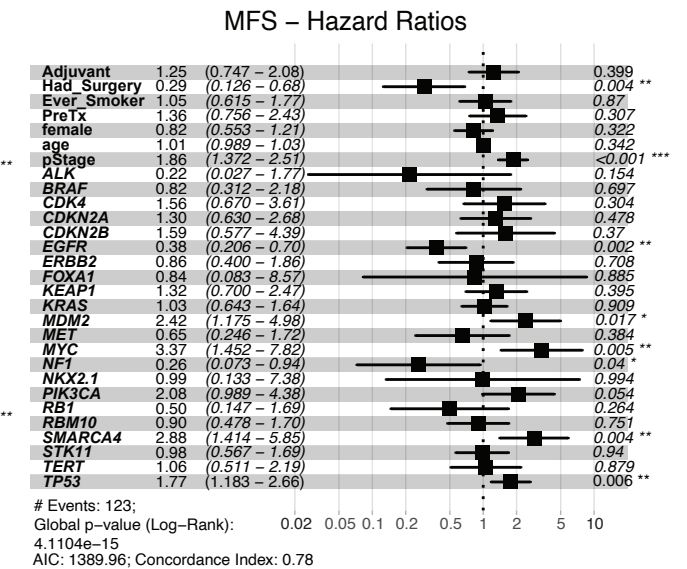
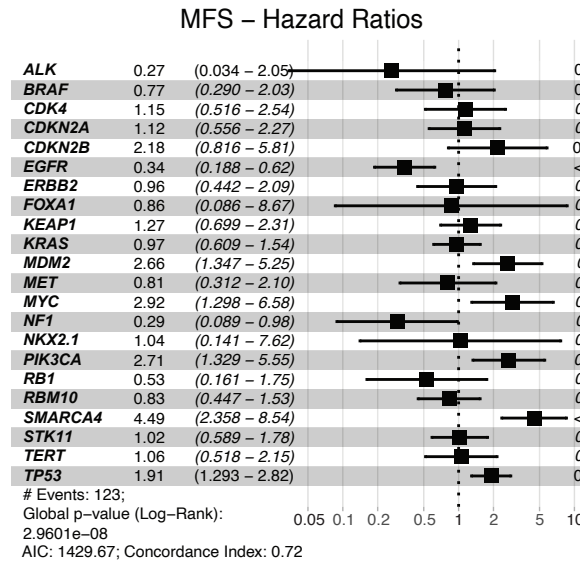
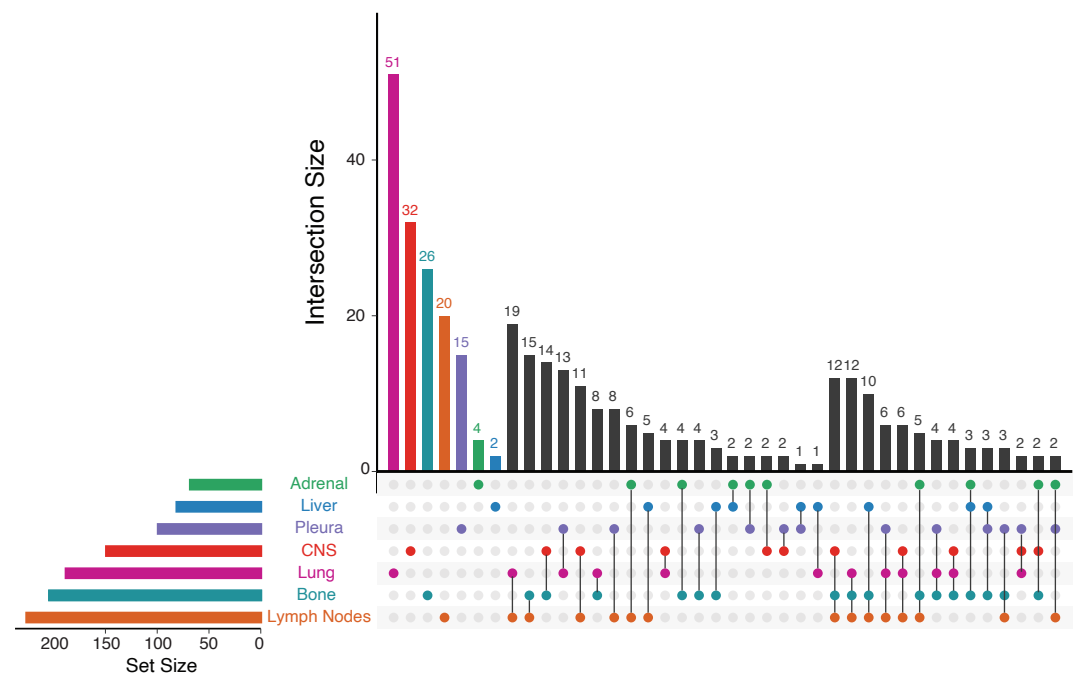


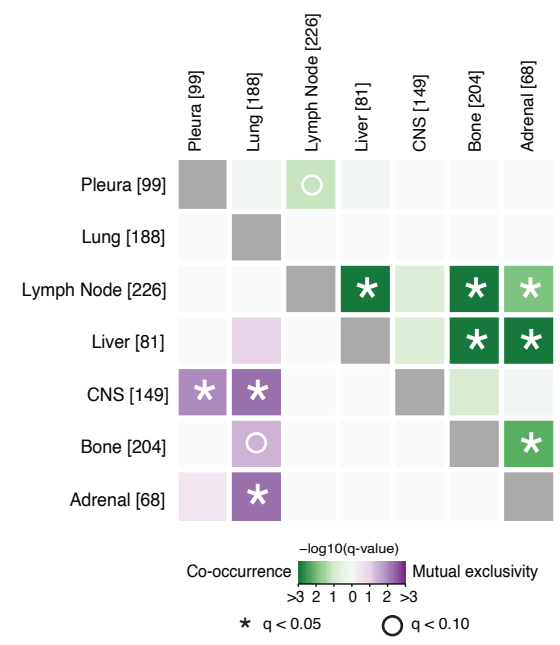
Figure S2. Clinicopathologic and genomic differences between nonmetastatic (NM) and ever-metastatic (EM) primary tumors, Related to Figure 2. (A) Oncoprint of NM and EM primary tumors from Dana-Farber Cancer Institute in the GENIE-Biopharma Collaborative data set. All genes altered at significantly different frequencies between the two groups in our cohort are displayed. (B) Comparisons of clinicopathologic features between nonmetastatic and metastatic patients in the GENIE validation cohort. (C) Scatter plot of the q -values of gene pairs for co-occurrence and mutual exclusivity between NM and EM tumors. The indicated gene pairs are those shared between EM and NM tumors and those private to NM tumors. Dotted lines represent $q=0.05$. (D) Forrest plot showing metastasis-free survival (MFS) for all genes altered in at least 3% of samples (*left*) and genes plus clinicopathologic features (*right*). Statistical analyses: (A-C) Fisher's exact test. (D) Cox-proportional hazards, log-rank test. p -values are as indicated. Squares represent hazard ratio (HR) and whiskers display 95% confidence interval (CI).

Figure S3

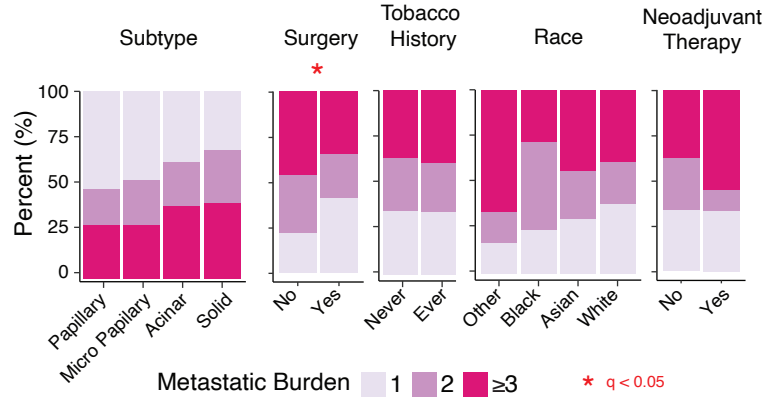
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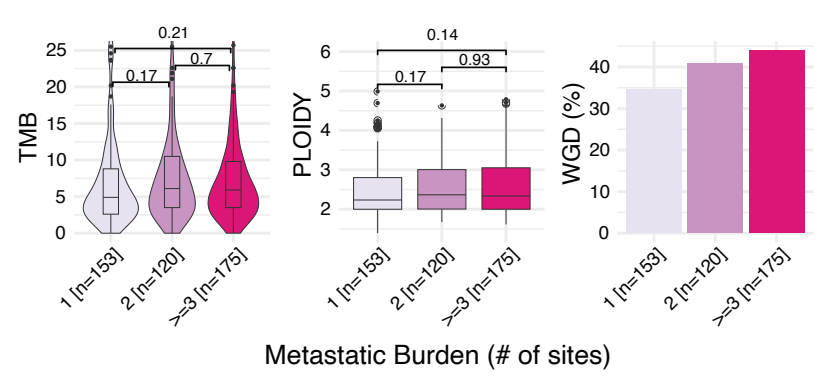
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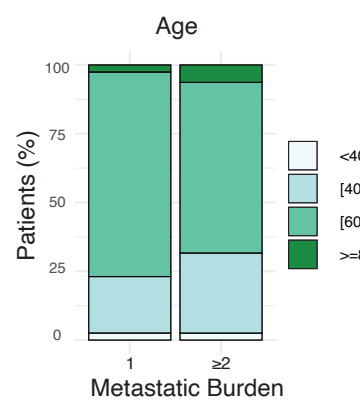
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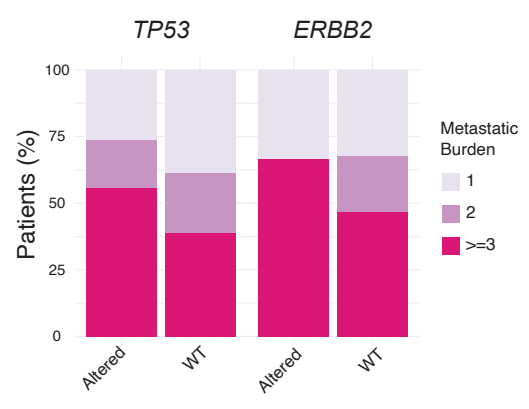
D



E



F



G

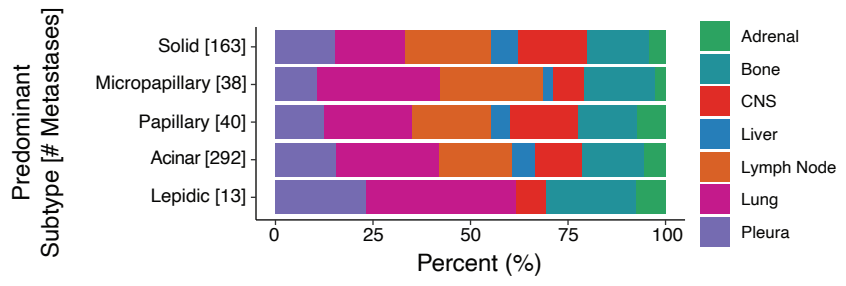
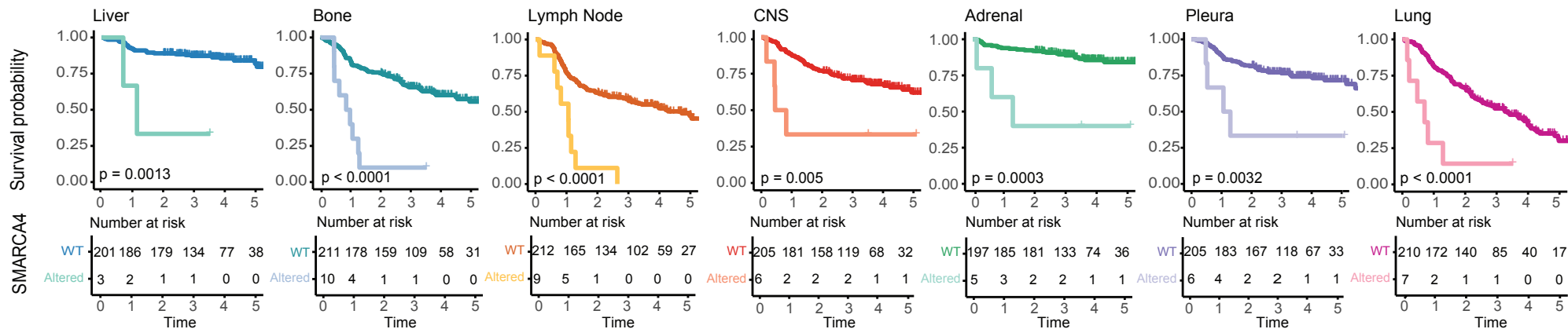


Figure S3. Comparisons of site-specific non- and ever-metastatic tumors and metastatic

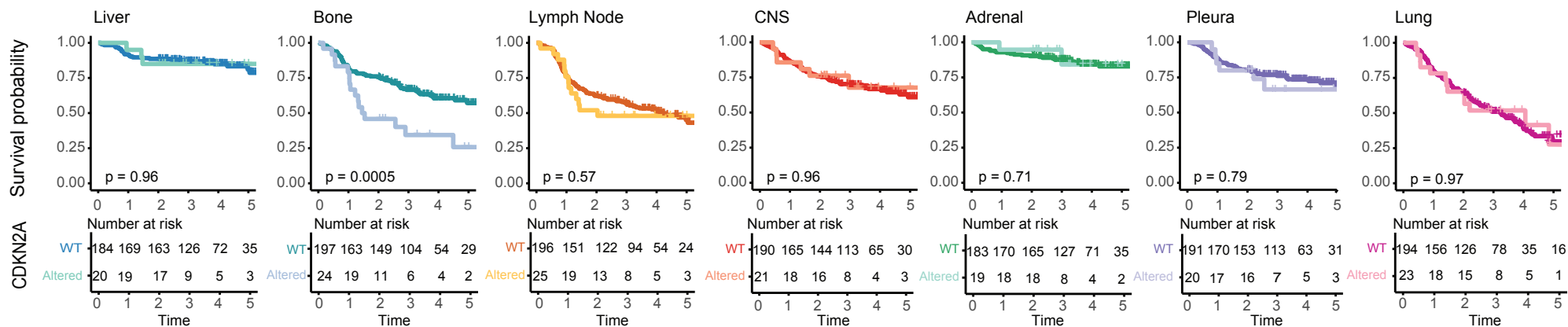
burden, Related to Figure 3. (A). Upset plot illustrating abundance of metastatic patterns across patients with at most 3 metastatic sites. (B) Heat map demonstrating co-occurrence and mutual exclusivity between metastatic sites. (C) Bar plots showing the distribution of metastatic burden for clinicopathologic features. (D) Violin plot of the distribution of tumor mutational burden (TMB) and boxplot showing ploidy for tumors with 1, 2, or ≥ 3 distinct metastatic sites. Boxplots display median values, interquartile range (IQR) boxes, and whiskers demonstrating 1.5 x IQR. *p*-values for pairwise comparisons between groups noted. Bar plots demonstrating proportion of patients with whole-genome duplication (WGD) by number of metastatic sites. (E) Bar plot showing the breakdown of age in patients, stratified by metastatic burden, for GENIE validation cohort. (F) Bar plots displaying the breakdown of metastatic burden for TP53 and ERBB2 altered and wild-type (WT) tumors for GENIE validation cohort. (G) Bar plots showing proportion of primary tumors with metastasis to a given site stratified by predominant histologic subtype. Statistical analyses: (B-C, E-G) Fisher's exact test. *q*-values correct for multiple comparisons using the false-discovery rate (FDR). (D) Wilcoxon rank-sum test for TMB and ploidy; Fisher's exact test for WGD. CNS, central nervous system; Met, metastatic.

Figure S4

A *SMARCA4* Alterations and MFS



B *CDKN2A* Alterations and MFS



C *HIPPO* Alterations and MFS

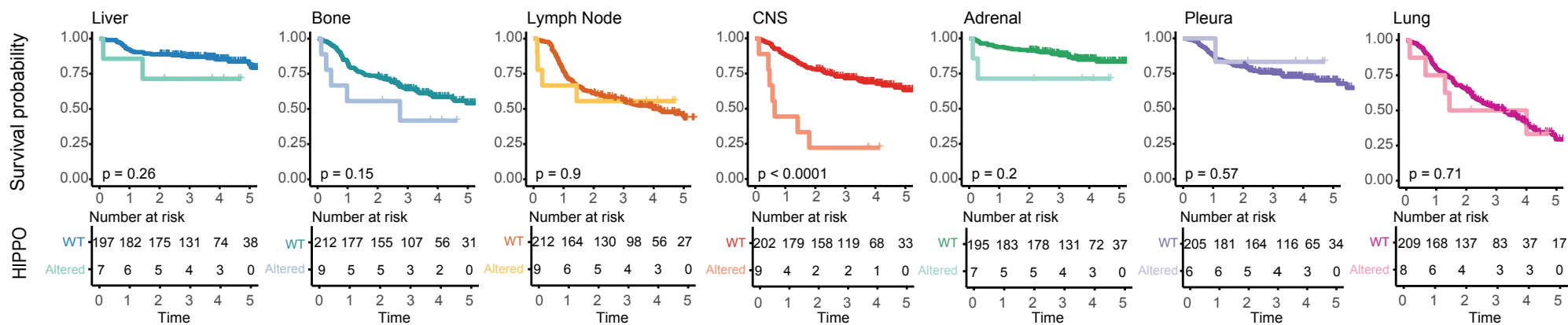


Figure S4. Kaplan-Meier Curves for Metastasis-Free Survival (MFS), Related to Figure 3.

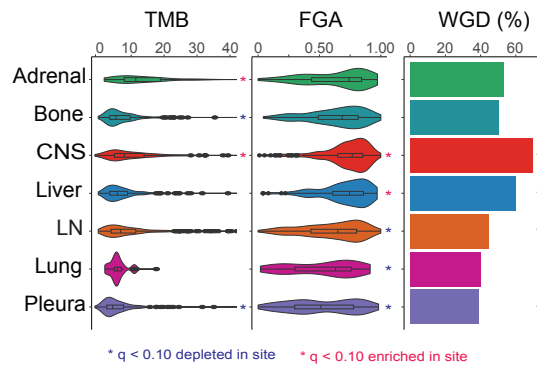
(A) Kaplan-Meier curves for MFS for each site stratified by SMARCA4 status. (B) Kaplan-Meier curves for MFS for each site stratified by CDKN2A status. (C) Kaplan-Meier curves for MFS for each site stratified by Hippo pathway status. CNS, central nervous system; WT, wild-type.

Figure S5

A



B



C

Metastatic Site	ALK	ARID1A	ATM	BRAF	CDK4	CDKN2A	CDKN2B	CTNNB1	EGFR	ERBB2	FOXA1	KEAP1	KRAS	MDM2	MET	MGA	MYC	NF1	NKX2-1	PIK3CA	RB1	RBM10	SETD2	SMAD4	SMARCA4	STK11	TERT	TP53	Cell Cycle	HLIPPO	MYC	NOTCH1	NRF2	TGF- β 1	PI3K	RTK-RAS	P53	WNT		
Adrenal	Y	0	15	2	6	2	29	21	3	19	2	6	11	19	37	5	6	6	11	10	11	10	6	0	2	6	13	24	15	63	44	5	21	15	19	44	6	79	69	6
Bone	Y	1	6	4	3	5	24	20	2	38	1	4	11	23	30	7	8	6	8	6	12	1	4	12	1	4	6	18	8	59	41	5	15	5	23	27	8	86	69	7
CNS	Y	2	8	4	4	7	32	24	5	32	3	5	11	21	32	6	7	5	9	6	16	7	8	5	5	4	21	17	68	50	7	16	12	23	40	9	86	74	12	
Liver	Y	7	4	3	2	5	32	28	5	45	4	4	11	14	18	7	9	3	12	4	14	7	9	7	3	5	4	10	12	63	53	5	17	6	15	28	9	84	73	10
LN	Y	4	3	6	7	3	19	12	5	25	2	4	9	17	29	6	6	5	8	6	13	4	7	8	5	4	8	16	11	59	36	5	14	9	18	31	6	81	68	10
Lung	Y	0	12	0	0	12	12	8	8	52	0	0	4	16	36	8	0	8	12	8	4	12	4	8	0	4	0	16	8	52	32	0	20	0	20	32	8	88	60	12
Pleura	Y	3	5	4	5	5	25	19	4	32	2	4	10	18	29	6	7	5	9	6	13	5	7	8	3	4	7	17	12	61	42	5	15	8	19	31	7	84	70	10

Figure S5. Genomic comparisons between metastatic lesions from different sites, Related to

Figure 4. (A) Oncoprint of metastatic lesions (ML) stratified by anatomic site displaying genomic features and all genes altered in $\geq 3\%$ of cohort. $*q < 0.10$ for at least one metastatic site. (B) Violin plots of the distribution of tumor mutational burden (TMB) and fraction of genome altered (FGA) for seven metastatic sites. Boxplots display median values, interquartile range (IQR) boxes, and whiskers demonstrating $1.5 \times$ IQR. *Right:* Bar plots showing frequency of whole genome duplication (WGD) across each site. (C) Heat map listing percentage of samples with a given gene or pathway alteration, stratified by metastatic site. $*q < 0.10$, red * indicates alteration enriched in metastatic site, blue * indicates alteration depleted in metastatic site. Y, yes - metastatic lesions from indicated anatomic site; N, no - metastatic lesions from all other anatomic sites. Statistical analyses: (A, C) Fisher's exact test comparing site of interest to all other sites. q -values correct for multiple comparisons using the false-discovery rate (FDR). $*q < 0.10$. (B) Wilcoxon rank-sum test for TMB and FGA; Fisher's exact test for WGD. CNS, central nervous system; LN, lymph node.

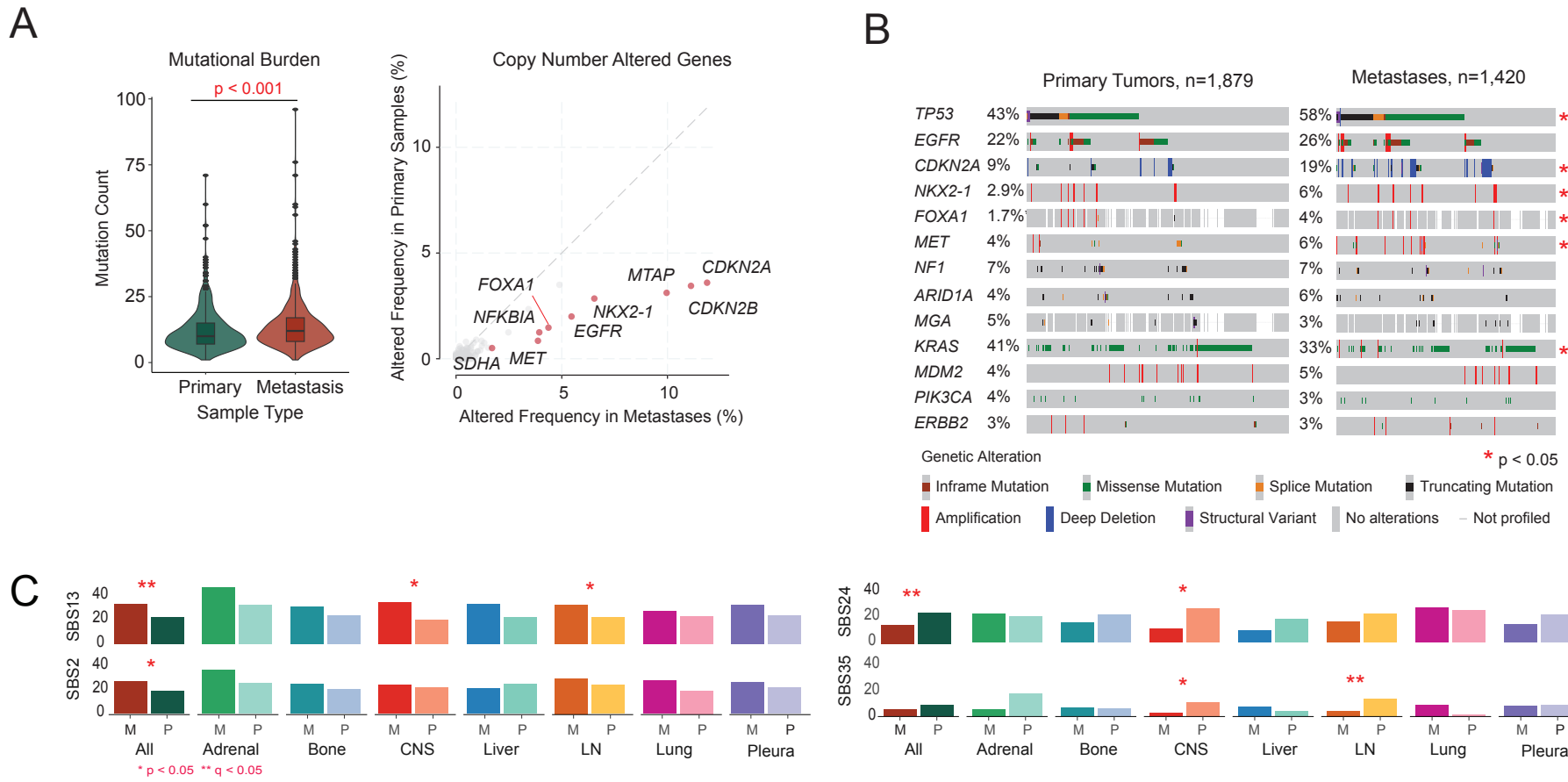


Figure S6

Figure S6. Comparison of alterations between primaries and metastases in GENIE validation cohort and mutation signatures for MSK-IMPACT samples, Related to Figure 4.

(A) GENIE validation cohort. *Left*: Violin plot showing differences in mutational burden between primary and metastatic samples. Boxplots display median values, interquartile range (IQR) boxes, and whiskers demonstrating 1.5 x IQR. *Right*: Dot plot comparing differences in gene-level copy number alterations called between primary and metastases. (B) Oncoprint of primary and metastatic tumors in GENIE validation cohort. (C) Frequencies of IMPACT samples with APOBEC-related signatures present according to metastatic site. Statistical analyses: (A) Wilcoxon rank-sum test. (B-C) Fisher's exact test. q -values account for multiple tests through false-discover rate (FDR) correction.

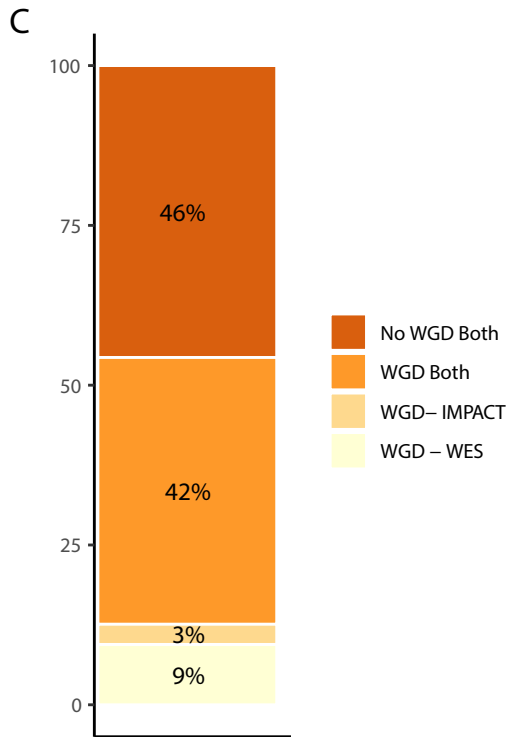
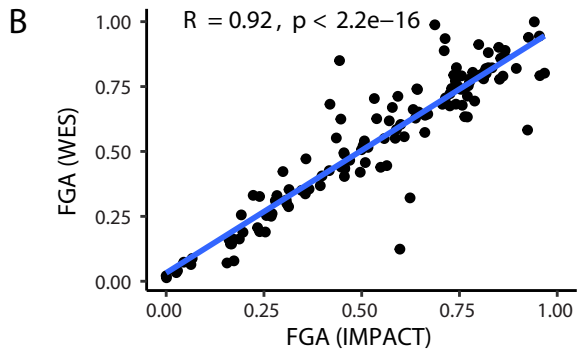
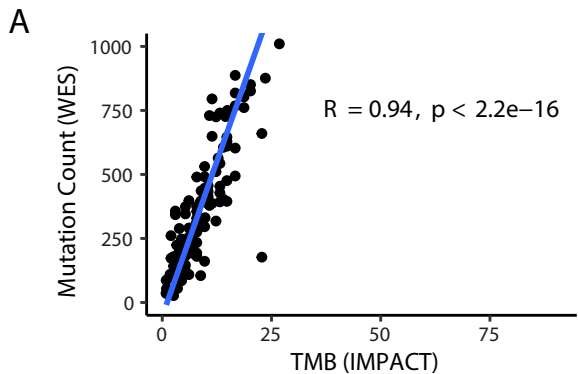


Figure S7

Figure S7. Comparison of sample-level genomic features for samples sequenced with MSK-IMPACT and whole-exome sequencing (WES), Related to Figure 4. (A) Scatter plot highlighting correlation between tumor mutational burden (TMB) reported by MSK-IMPACT (x-axis) and number of mutations detected by WES (y-axis). (B) Fraction of genome altered (FGA) estimated from MSK-IMPACT (x-axis) versus FGA estimated from WES (y-axis). (C) Percentage of samples with similar or differing calls for whole-genome duplication (WGD) based on data from MSK-IMPACT and WES. In our cohort, 121 samples were sequenced by next-generation sequencing (MSK-IMPACT) and WES. We observed agreement between the two methods in ~88% of whole-genome duplication calls. Statistical analyses: (A-B) Pearson correlation with correlation coefficients and p-values as indicated.

Figure S8

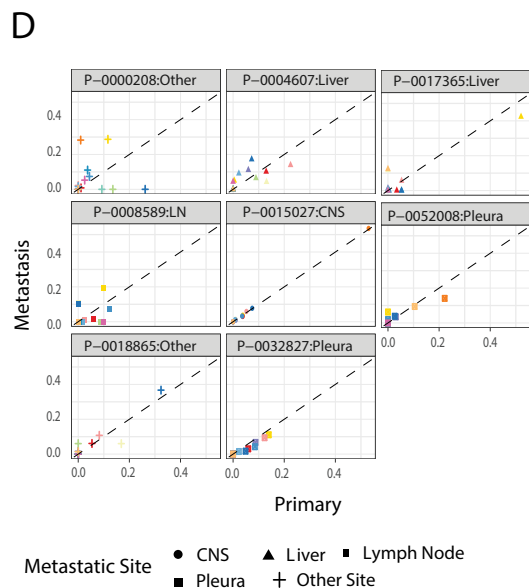
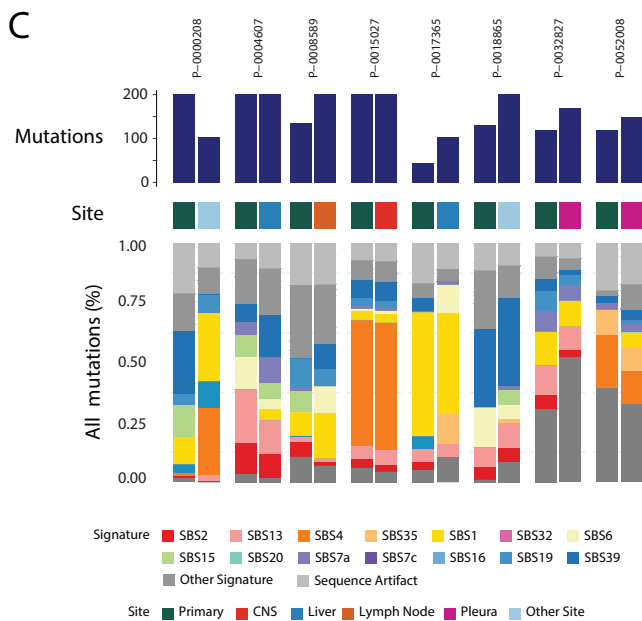
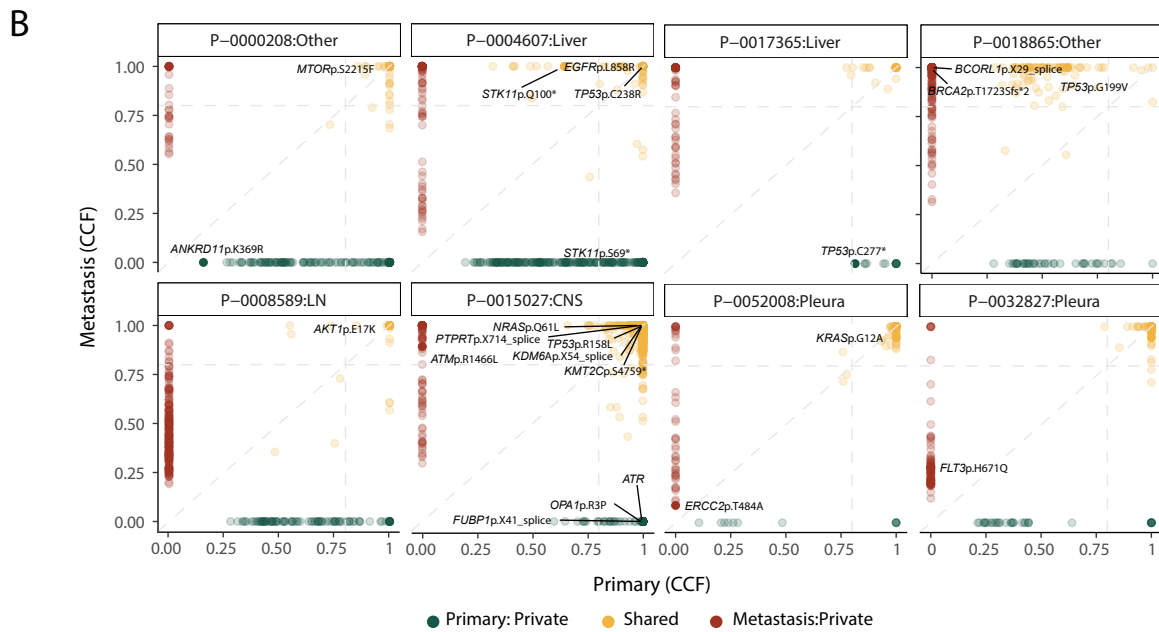
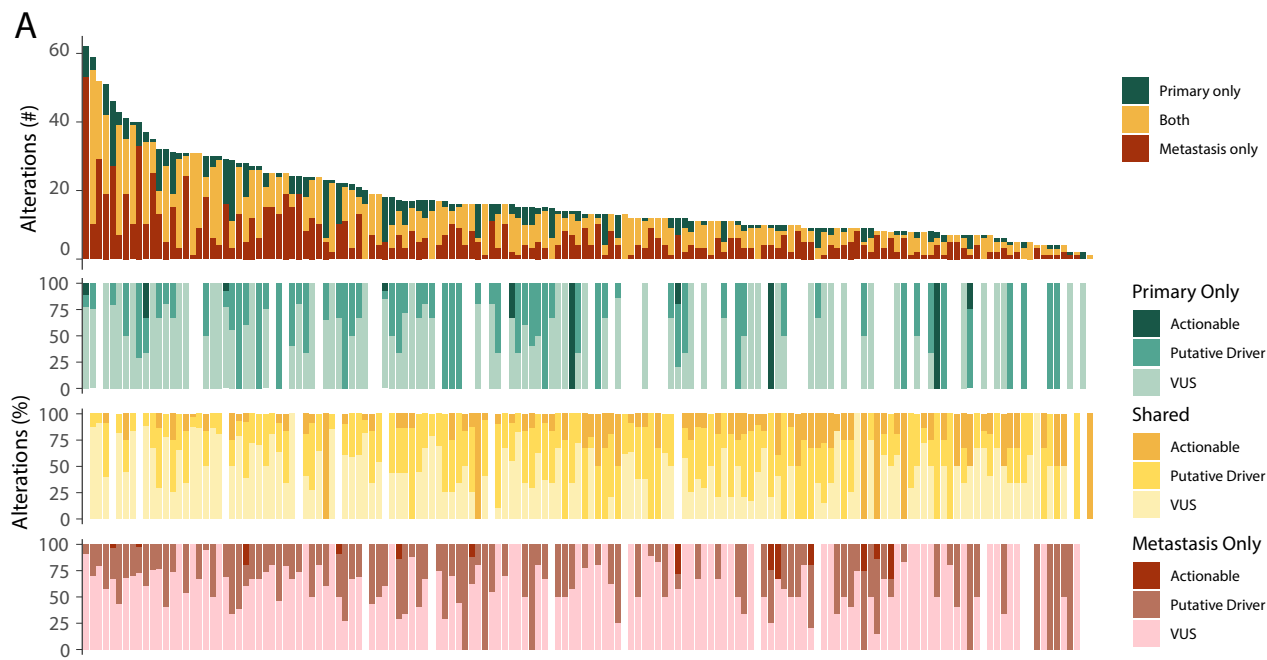


Figure S8. Patient-level comparisons in paired primary and metastatic samples in

IMPACT samples and whole-exome sequencing (WES), Related to Figure 5. (A) *Top:* Bar

plot illustrating fraction of alterations shared and private between matched primary and

metastases. *Bottom:* Bar plots showing the distribution of actionable, oncogenic, and variants of

unknown significance (VUS) separated by alterations private to primary, shared alterations, and

alterations private to metastasis. (B) Comparison of cancer cell fraction (CCF) for individual

mutations in paired WES primary and metastatic samples. Each point represents an individual

mutation. Mutations considered oncogenic per OncoKB are highlighted by displaying name of

their associated gene. (C) Comparison of mutational signatures for paired WES primary and

metastatic samples. (D) Quantification of differences in selected mutational signatures from

panel C. CNS, central nervous system.

Table S8. Overview of cohorts used in specific analyses, Related to STAR Methods

Analysis	Cohort					
	NM	ES-M	LS-M	ML	MP-M	WES
1. Features associated with metastasis in primary tumors	X	X	X			
2. Features associated with MFS in surgical patients	X	X				
3. Patterns of metastasis and metastatic burden		X	X			
4. Features associated with site-specific metastasis		X	X			
5. Time-to-event site-specific metastasis		X				
6. Comparisons of metastases across sites				X		
7. Analysis of mutational signatures	X	X	X	X		X
8. Comparisons of unmatched metastases and primary tumors		X	X	X		
9. Comparisons of matched primary tumors and metastases					X	X

ES-M, early-stage metastatic; LS-M, late-stage metastatic; MFS, metastasis-free survival; ML, metastatic lesion; MP-M, matched primary-metastasis; NM, nonmetastatic; WES, whole-exome sequencing.