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Supplemental information

Understanding heterogeneity of human bone marrow

plasma cell maturation and survival

pathways by single-cell analyses

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Figure S1. Integrated single-cell transcriptomic datasets. A) UMAP plots of single-cell transcriptomic profiles from 5 subjects before 2-step cell quality control and colored by sorted subject id, FACS-sorted labels, gene number group and cell subsets, which is mainly grouped by the location of cell population on UMAP, proliferating status and whether they are contaminating cells. HighGN: high gene number, indicating cells have greater than 800 detected genes, which is correspond to the gene cutoff in Fig. 1B; LowGN: low gene number, representing cells have greater than 200 detected genes but less or equal than 800 genes; contam: contaminating cell populations. B) and C) Contaminated cell population (B) and plasma cell (C) associated master gene expression. The redder the dot indicates the higher log-normalized gene expression.



Figure S2. Stability and reliability of cell clusters. A) Expression of contaminated cell markers in current UMAP. B) Cell clusters (left) when not removing AC233755.1 and AC233755.2 genes before detecting cell clusters in the first run. Expression of AC genes in the previous UMAP (right). C) Current UMAP colored by sampled subjects, FACS-sorted cell populations and defined stages

of BMPC maturation. D) Two-way hierarchical clustering of RI by comparing cell clustering results from running different combinations of key cell clustering parameters (left), the parameter information was shown in the middle labels and scatter plot showed the average RI of each row of heatmap (right). Dashed red line labeled the RI value 0.85. (dim: dimension; pc: PC; res: resolution.) E) Expression of AC genes in current UMAP. F) Dot plots for expression of marker genes in each subgroup of bone marrow plasma cell defined in Fig. 1C. Here and in later figures, colors represent Min-Max normalized mean expression of marker genes in each cluster, and dot sizes indicate the proportion of cells expressing each marker gene. G) Fraction of cells in current scRNA-seq data defined cell clusters by sampled subjects. H) Log-normalized gene expression of SPN in cell clusters.



Figure S3. The features and pathways characterize the IgM dominant cell populations. A) Boxplots showing the expression of indicative IgM highly expressed genes as well as three Ig genes associated with somatic recombination of immune receptors in (C). B) Heatmap showed the row-scaled gene expression of CCR10 highly correlated genes (|Pearson correlation coefficient| \geq 0.6, '||' implies absolute value).



of PRDM1 in cell clusters. B) and C) Violin plots show the percentage of transcripts coming from ribosomal genes (ribo), mitochondrial-encoded mitochondrial genes (mito), and immunoglobulin genes (Ig) (B) and the distribution of total detected gene numbers (nGene), total UMIs (nUMI) (C). D) X-axis is regrouped cell populations when taking both scRNA-seq defined cell subgroups and

FACS-sorted cell labels into account. Y-axis is the log-normalized gene expression. Arrow points out the FACS-sorted popA cells starting CD138 (encoded by SDC1) transcripts.



Figure S5. Patterns of Hallmark pathway enrichment analysis. A) Heatmap showed the two-

way hierarchical clustering of enrichment scores in each pathway. X-axis is the cell cluster id. The redder the color, the higher of enrichment scores. Numbers in the left-side of the heatmap showed the defined patterns. B) Heatmap showed the row hierarchical clustering of enrichment scores in each pathway. C) Heatmap showed the two-way hierarchical clustering of enrichment scores in IgM dominant cell lineage using DEGs are specifically differentially expressed in IgM lineage compared with DEGs detected by comparisons made between IgG1 dominant clusters.



6756756

Cluster id

Figure S6. Genes driven the difference of TNFα signaling via NFKB pathway in late LLPC.

A) Heatmap showed the scaled corresponding TNF α singaling via NF κ B pathway enriched maturation-associated DEG expressions in cluster 5, 6 and 7 in each individual. B) GSEA prerank analysis results using preranked genes between cluster 6vs7 (left); enrichment score, FDR qvalue, and leading-edge genes were listed (right). Gene colored in red were DEGs from Table S4 and in orange were cluster 6vs7 specific DEGs included in Table S6. C) Expression of differentially expressed leading-edge genes (|avgLogFC| > 0.25, Bonferroni adjusted p value < 0.05) from (B) in cluster 5, 6 and 7, the order of genes was corresponding to the gene rank in preranked gene list in Table S6.



Figure S7. Visualization of indicative genes. A) Light and dark blue bar plots showed the proportion of cells in each cluster not expressed (NotExp) and expressed (Exp) gene TNFRSF17, TNFRSF14 and TNFRSF13B. B) Violin plots showed the expression of remaining expressed TNF family genes. C) Dotplot visualized the gene expression of pro-apoptotic, anti-apoptotic and ER-stress associated genes based on the FACS-sorted cell labels. The bluer the dots, the lower the gene expression. The size of dots represented the percentage of cells from associated cell population expressed the corresponding genes. D) Dotplot showed the gene expression from CCR, CD, CXCR ad interleukin (IL) families.



size. X-axis is the number of cells contained in detected cell lineages. Y-axis is the number of cell lineages. Orange and purple boxes summarize the total number of lineages that have at least 2 or 5 cells. B) The distribution of overlapped cell clusters of each non-singleton lineages. Orange boxes summarize the number of lineages having at least 2 cells and overlapped with at least 2 cell clusters. C) and D) Count the total number (C) and percentage (D) of cells in each cell cluster by if cells have matched scVDJ-seq information, if they are from singleton lineages (yellow) or not (dark grey). Cells with no matched scVDJ information were colored in ash.



cluster_id	count	50	100	500	1000	2000	3000	4000	5000
1	462	99.6	97.6	86.8	66.9	31.2	16.7	8.9	3
2	1021	99.8	99.3	87.3	66.4	33.3	15.8	6.7	2.5
3	1330	99.2	98	87.1	74.3	49.9	33.6	20.1	11.7
4	2454	99.9	99.5	92	79.6	58.5	41.4	29.5	20.6
5	1947	99.4	98.8	88.5	76.8	55	39	27.7	19.3
6	2034	99.5	99.1	94.4	83.3	63.9	46.8	35.1	26.2
7	2815	99.6	99.3	91.5	79.1	53.9	36.1	23.8	15.1
8	473	98.9	98.1	90.1	79.5	58.4	42.1	27.5	20.9
9	1070	99.6	99.2	87.4	72.2	45.6	27.9	18.2	11.3
10	65	96.9	96.9	89.2	76.9	52.3	38.5	18.5	13.8
11	1241	98	94.8	76.1	57.9	36.8	23.5	14.6	9.2
12	117	100	99.1	72.6	40.2	6.8	0.9	0	0
13	350	99.7	99.7	97.7	83.7	59.1	36.9	24	12.6
14	1441	99.7	99.4	95.2	85.7	64.1	45.2	29.8	18.9
15	527	100	100	93	80.3	56.9	41.7	26.6	18

Figure S9. Summary of Ig heavy chain gene transcript numbers in each cell cluster. Circos plot summarizes the proportion of cells in each cell cluster that pass the criterion by using different cutoffs to gate Ig heavy chain genes. The most inner circle is colored as Fig. 1C and numbers indicate corresponding cell cluster id. From inner to outer circle, the cutoffs are 50, 100, 500, 1000, 2000, 3000, 4000 and 5000. The table summarizes the percentage of cells in each cell cluster that passed cutoffs. The count column shows the total number of cells in each cell cluster.



Figure S10. Permutation results for the mutation frequency test. A) Barplot showing the distribution of p values from ANOVA test for the global heavy chain variable region (Global), CDR and FR mutation frequency using real data and shuffled data from permutation test. Orange bars and grey bars indicated the p values from using real and randomly shuffled datasets. B) – D) The distribution of p values from Tukey's HSD (honestly significant difference) tests for the Global (B), CDR (C) and FR (D) mutation frequency between cluster 6 and other 14 clusters by using real

and randomly shuffled datasets. Red arrows pointed out the adjusted p value from Tukey's HSD test using real data, left upper side showed the p value from permutation test and significant results were colored in red. Titles indicated the objects of comparison.



Figure S11. Clonal connectivity visualization in all cell clusters. A) Circos plot from main IgG1 dominant cell clusters 1 to 15 in a connecting detected lineages in each donor. Lineages were

colored by the latest cell cluster. B) - C) Percentage of connected clones between any two cell clusters when only considering lineages across at least two cell clusters (B) or lineages include singletons (C). Numbers in each block showed the percentage of lineages from each row cluster that were connected with other. The numbers showing in the grey boxes recorded the total number of lineages and cells showing in bracket in associated cell cluster. Yellow boxes highlighted the highly connected cell blocks and dashed orange box highlighted those clones showing high connectivity in one direction but not in another.



Figure S12. Clonal connectivity of nearly identical clones in IgG1-dominant clusters 1 to 8. A homology of 98% was used within the CDR3 region to cluster clones within each of the 5 subjects. Cell clusters are represented by the outer ring colors and sequences are grouped into clones in size descending order from counterclockwise to clockwise. Lines connecting cell clusters indicate a clone that was found in multiple clusters.



clusters. A) Clones were clustered by identical V, J, CDR3 length and 98% CDR3 homology to identify near identical clones and plotted on UMAP. Cells highlighted in red are members of the same clone. B) V region genes from the same clones were aligned against germline sequences. Blue squares indicate mutations in the individual sequences when compared to germline sequences and pink squares indicate matching nucleotides to the germline sequence. The cluster of each cell where the transcript was identified to be from is shown on the left.