

# Supplementary Information

## **Methane formation driven by light and heat prior to the origin of life and beyond**

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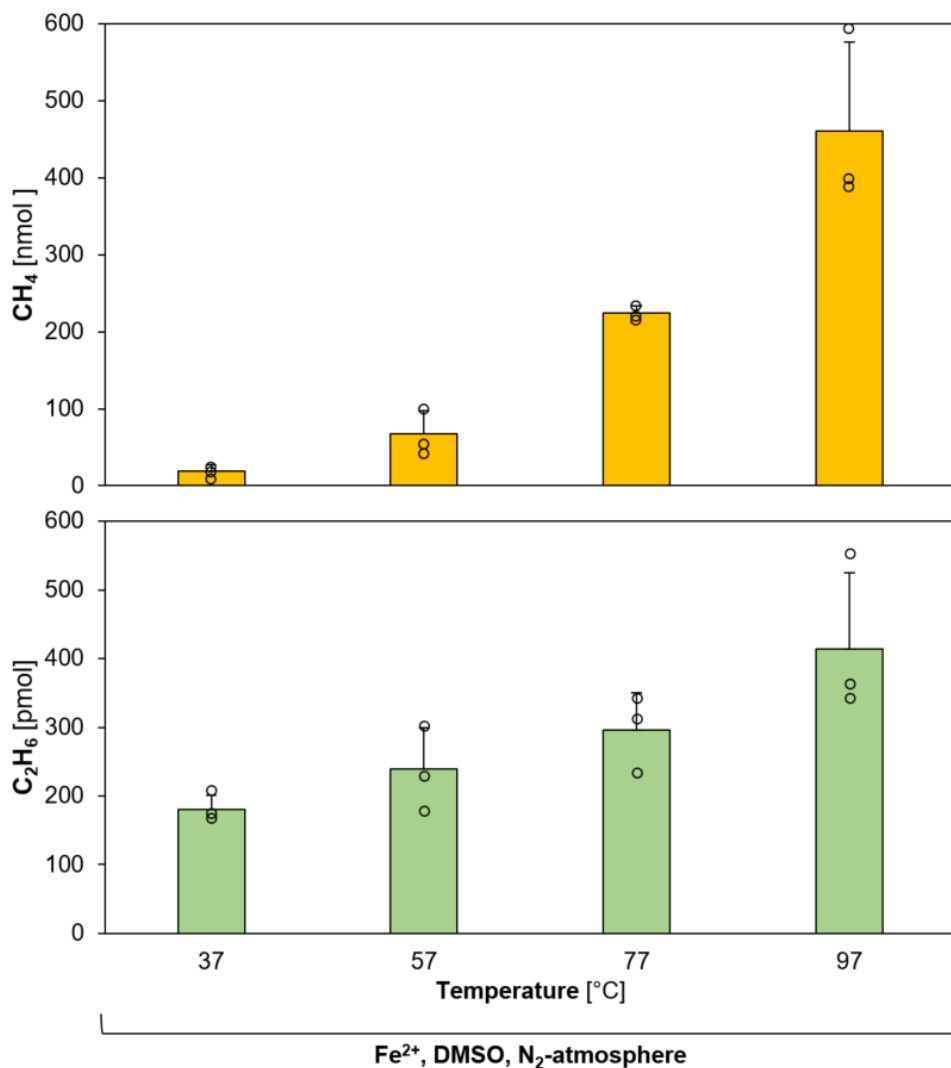
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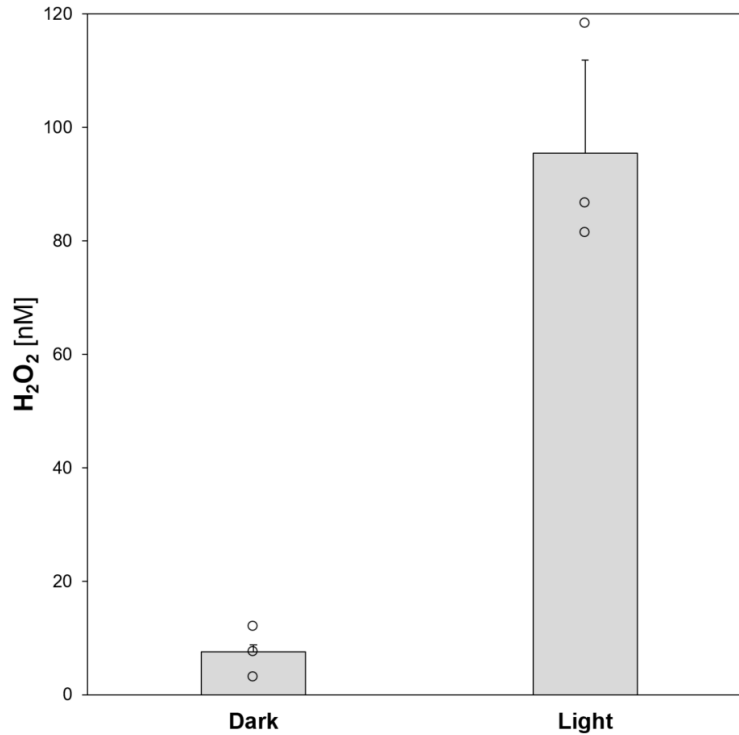
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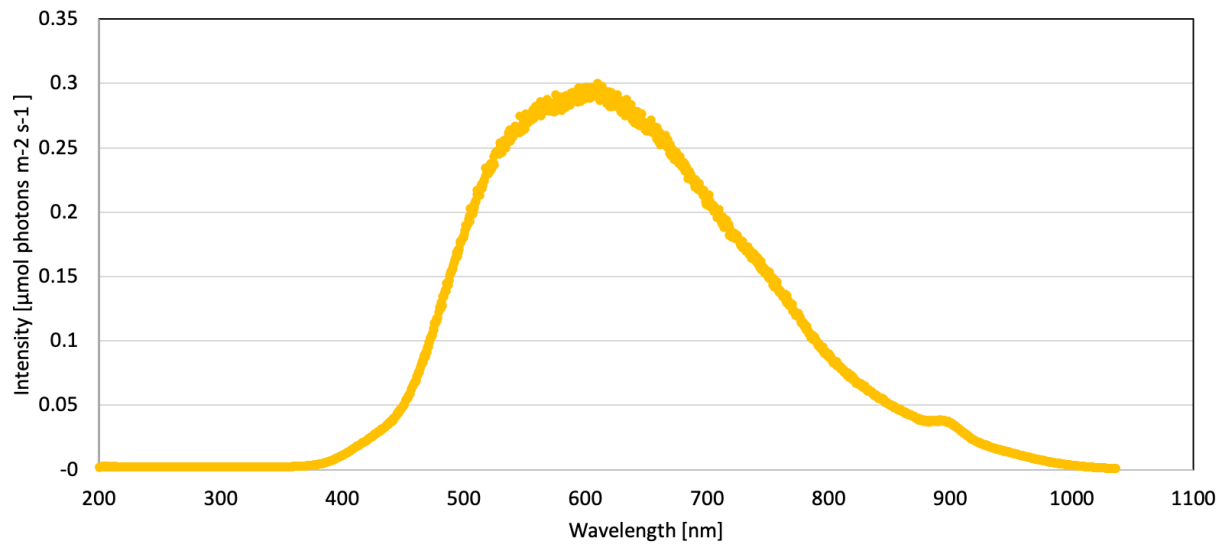
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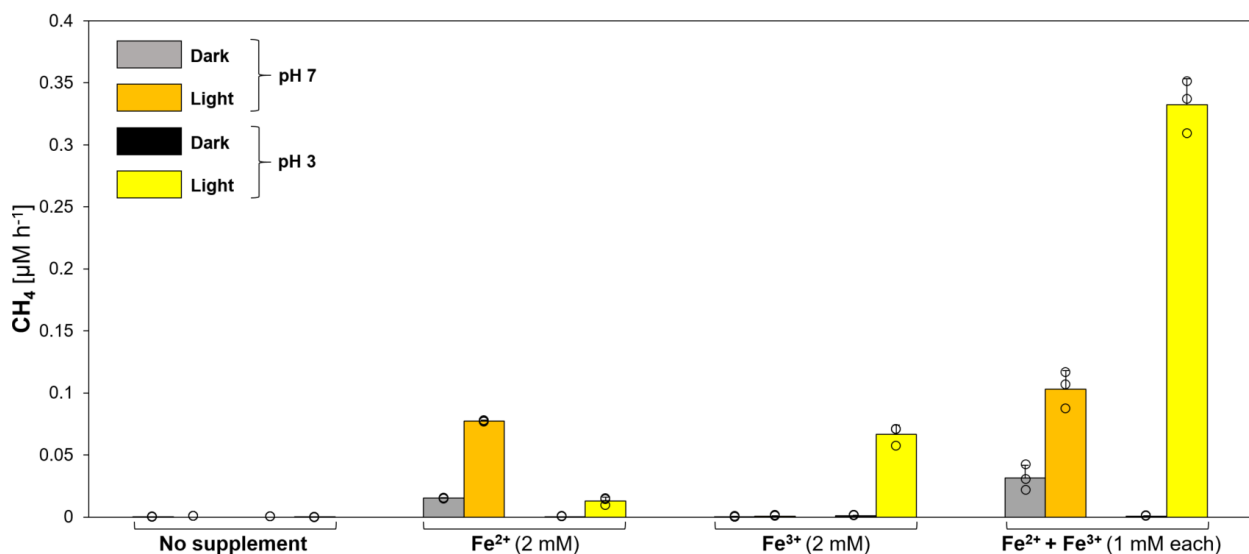
**Supplementary Fig. 1. Heat-driven CH<sub>4</sub> and C<sub>2</sub>H<sub>6</sub> formation from DMSO.** Formed CH<sub>4</sub> amounts increase from ~20 nmol (37 °C) to ~460 nmol (97 °C). Formed C<sub>2</sub>H<sub>6</sub> amounts increase from ~190 pmol (37 °C) to ~415 pmol (97 °C), thereby resulting in CH<sub>4</sub>:C<sub>2</sub>H<sub>6</sub> ratios from ~110 (37 °C) to ~1100 (97 °C). In a total volume of 4 mL, samples containing a 20 mM potassium phosphate buffer (pH 7), 1 M DMSO, 10 mM FeSO<sub>4</sub> were incubated at different temperatures in sealed 20 mL glass vials under N<sub>2</sub> for 4 days. The bars are the mean + standard deviation of triplicates, shown as circles.



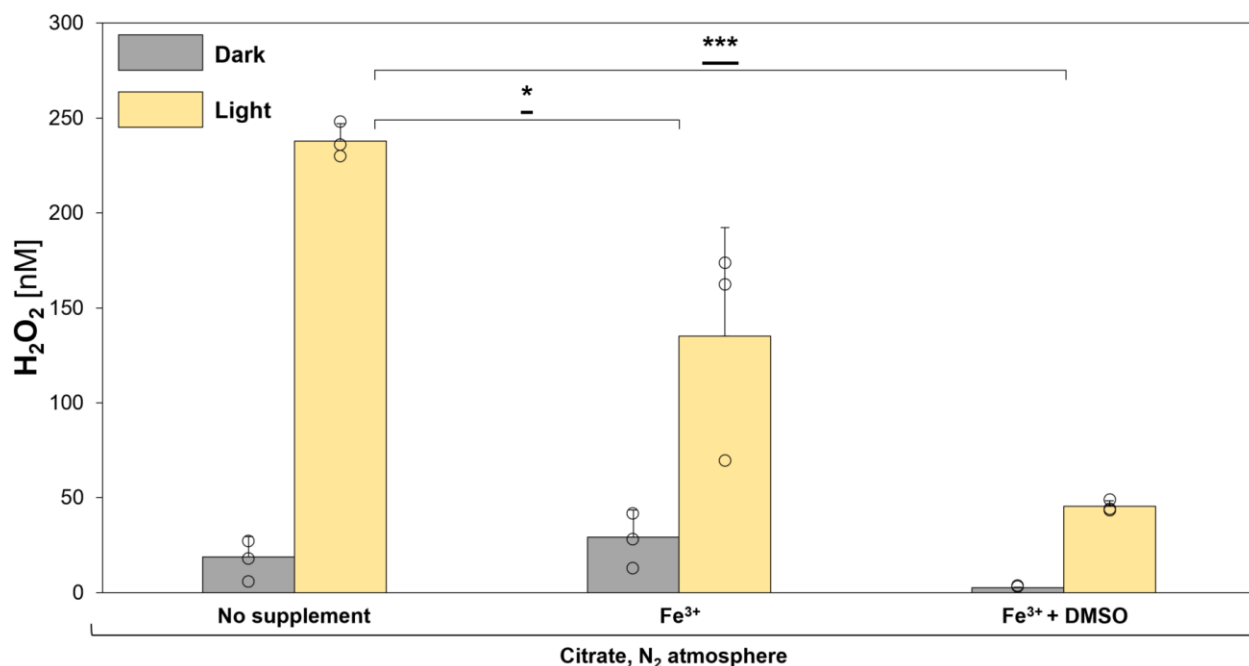
**Supplementary Fig. 2. Light-driven H<sub>2</sub>O<sub>2</sub> formation in pure buffer.** Formed H<sub>2</sub>O<sub>2</sub> concentrations increase from ~7 nM in the dark to ~95 nM under light. In a total volume of 4 mL, samples contained 20 mM potassium phosphate buffer (pH 7) that was bubbled with N<sub>2</sub> for 1 h and kept in an anoxic tent for 5 days. Samples were incubated in the dark or under illumination (Supplementary Fig. 3) for 20 h and analysed via fluorescence-based H<sub>2</sub>O<sub>2</sub> endpoint measurements (see Methods). The bars are the mean + standard deviation of triplicates, shown as circles.



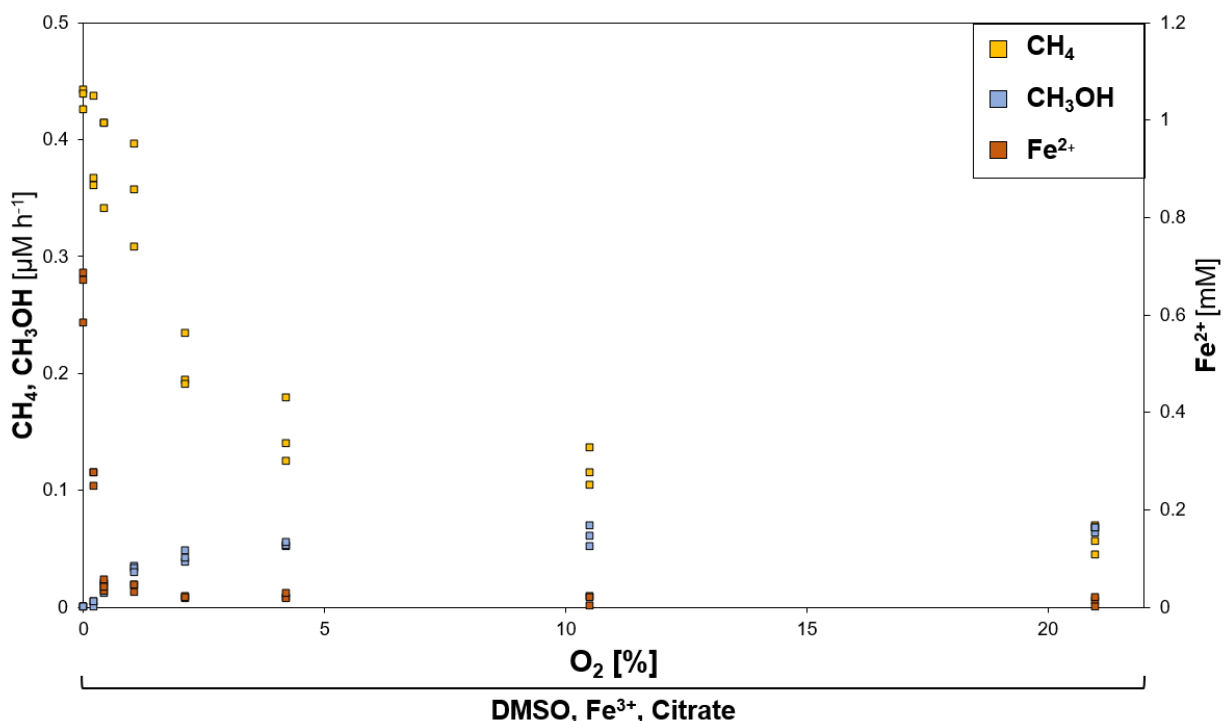
**Supplementary Fig. 3. Light spectrum of the used light bulbs.** For broad-spectrum sample illumination, Osram (Superlux, Super E SIL 60) light bulbs were used, with an intensity of  $82 \pm 4 \mu\text{mol photons m}^{-2} \text{s}^{-1}$ , and an energy flux of  $52 \pm 2 \text{ kJ m}^{-2} \text{h}^{-1}$ . The light spectrum was determined with a spectrometer (see Methods).



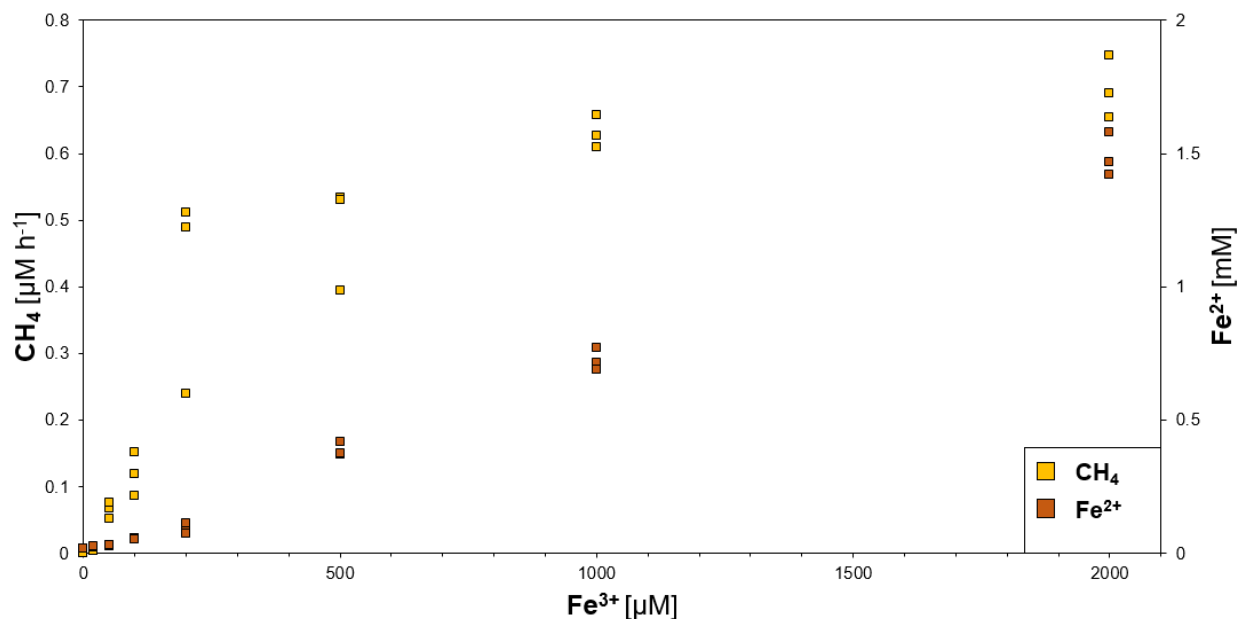
**Supplementary Fig. 4. Light-driven CH<sub>4</sub> formation is enhanced upon [Fe(H<sub>2</sub>O)<sub>6</sub>]<sup>3+</sup> photolysis under acidic conditions.** Only trace CH<sub>4</sub> levels (<0.0002 μM h<sup>-1</sup>) were measured without iron addition. Upon Fe<sup>2+</sup>-supplementation, CH<sub>4</sub> formation rates increased at pH 7 from ~0.015 μM h<sup>-1</sup> in the dark to ~0.077 μM h<sup>-1</sup> under light and at pH 3 from <0.0002 μM h<sup>-1</sup> in the dark to ~0.013 μM h<sup>-1</sup> under light, suggesting a light-driven ·OH formation from OH<sup>-</sup>. Conversely, ~0.67 μM CH<sub>4</sub> h<sup>-1</sup> are formed upon Fe<sup>3+</sup>-supplementation in the light under acidic conditions, while only trace amounts of CH<sub>4</sub> were detected under pH-neutral conditions or in the dark. At pH 7, a stoichiometric 1:1 ratio of Fe<sup>3+</sup> and Fe<sup>2+</sup> increased CH<sub>4</sub> formation rates by ~1.3-fold to ~0.1 μM h<sup>-1</sup> in comparison to Fe<sup>2+</sup>-supplemented samples, while the corresponding CH<sub>4</sub> formation rates at pH 3 increased ~25-fold to ~0.316 μM h<sup>-1</sup>. Samples consisting of buffered solutions (pH 3 or 7) were incubated in the dark or under broad spectrum light (Supplementary Fig. 3) for 24 h under N<sub>2</sub>. The bars are the mean + standard deviation of triplicates, shown as circles.



**Supplementary Fig. 5. Iron and DMSO reduce H<sub>2</sub>O<sub>2</sub> concentrations generated by light.** Under light, H<sub>2</sub>O<sub>2</sub> concentrations decrease from ~238 nM (buffer + citrate) to ~135 nM in the presence of iron (buffer + citrate + Fe<sup>3+</sup>). This decrease in H<sub>2</sub>O<sub>2</sub> concentration indicates the reaction of H<sub>2</sub>O<sub>2</sub> with Fe<sup>2+</sup> formed by LMCT. Upon addition of DMSO, H<sub>2</sub>O<sub>2</sub> levels further decrease to ~45 nM under light, underlining the ROS-scavenging effect of DMSO. In a total volume of 4 mL, samples contained 20 mM potassium phosphate buffer (pH 7), 10 mM citrate and, optionally, 2 mM FeCl<sub>3</sub> and 500 mM DMSO that were previously degassed with N<sub>2</sub> and kept in an anoxic tent overnight. Samples were incubated in the dark or under illumination (Supplementary Fig. 3) for 3 h and directly analysed via fluorescence-based H<sub>2</sub>O<sub>2</sub> endpoint measurements (see Methods). Statistical analysis was performed using paired two-tailed *t*-tests, \*:  $p \leq 0.05$ , \*\*\*:  $p \leq 0.001$ . The bars are the mean + standard deviation of triplicates, shown as circles.

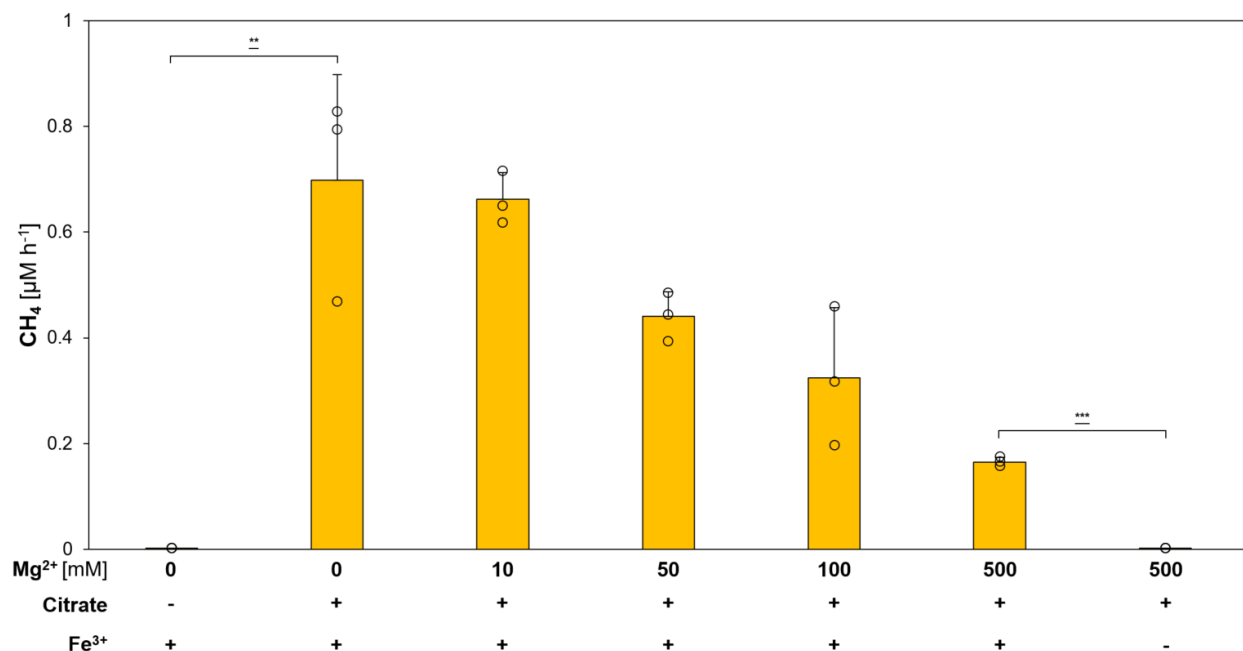


**Supplementary Fig. 6. Light-driven formation of CH<sub>4</sub>, CH<sub>3</sub>OH and Fe<sup>2+</sup> from DMSO and Fe<sup>3+</sup> under varying O<sub>2</sub> concentrations.** In a total volume of 4 mL, samples containing 20 mM potassium phosphate buffer (pH 7), 500 mM DMSO, 2 mM FeCl<sub>3</sub> and 10 mM citrate were incubated for one day in sealed 20 mL glass vials under constant broad-spectrum illumination (Supplementary Fig. 3). Mixtures of air and N<sub>2</sub> were generated and adjusted to 1 bar, ranging from ~21 % O<sub>2</sub> to pure N<sub>2</sub> (0 % O<sub>2</sub>). From 0 % O<sub>2</sub> to ~1 % O<sub>2</sub>, CH<sub>4</sub> rates decreased from ~0.44 μM h<sup>-1</sup> to ~0.35 μM h<sup>-1</sup>, while Fe<sup>2+</sup> levels decreased from ~1 mM to ~0.07 mM. From ~1 % O<sub>2</sub> to 21 % O<sub>2</sub>, CH<sub>4</sub> rates further decreased by an additional ~0.2 μM h<sup>-1</sup>, while Fe<sup>2+</sup> levels further decreased from ~0.07 mM to ~0.04 mM, indicating an immediate Fe<sup>2+</sup> oxidation by O<sub>2</sub> or the Fenton reaction at high O<sub>2</sub> concentrations and a formation of excess Fe<sup>2+</sup> under suboxic and anoxic conditions. Apart from the anoxic sample (0 % O<sub>2</sub>), formation of CH<sub>3</sub>OH was detected in the presence of O<sub>2</sub>, ranging from ~0.003 μM h<sup>-1</sup> (~0.2 % O<sub>2</sub>) to ~0.07 μM h<sup>-1</sup> (21 % O<sub>2</sub>). Experiments were conducted in triplicates; individual values are shown.

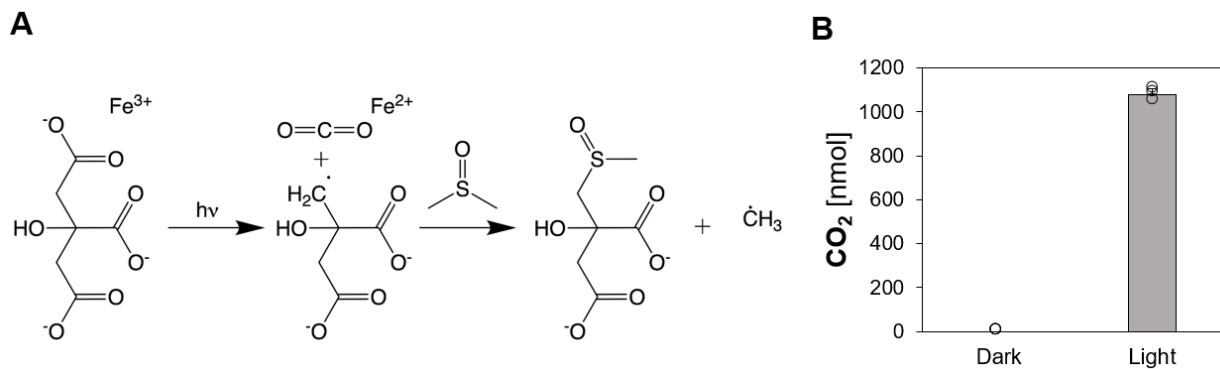


**Supplementary Fig. 7. Light-driven CH<sub>4</sub> formation from DMSO correlates with the iron concentration.** In a total volume of 4 mL, samples containing 20 mM potassium phosphate buffer (pH 7), 500 mM DMSO and 10 mM citrate were incubated for one day under N<sub>2</sub> in sealed 20 mL glass vials under constant broad-spectrum illumination (Supplementary Fig. 3). Different amounts of Fe<sup>3+</sup> (as FeCl<sub>3</sub>) were added, ranging from 0 μM to 2000 μM. Only trace amounts of CH<sub>4</sub> (<0.01 μM h<sup>-1</sup>) were formed upon supplementation with 0 or 20 μM Fe<sup>3+</sup>. Methane amounts then increased from ~0.06 μM h<sup>-1</sup> CH<sub>4</sub> (50 μM Fe<sup>3+</sup>) to ~0.4 μM h<sup>-1</sup> (200 μM Fe<sup>3+</sup>), with Fe<sup>2+</sup> levels also increasing from ~0.03 mM to ~0.11 mM. From 200 μM Fe<sup>3+</sup> to 2000 μM Fe<sup>3+</sup> supplementation, CH<sub>4</sub> levels increased by ~0.3 μM h<sup>-1</sup>. In the same range, Fe<sup>2+</sup> levels increased from ~0.11 mM to ~1.5 mM, indicating that LMCT-driven Fe<sup>2+</sup> formation occurred at higher rates as its consumption to Fe<sup>3+</sup> via the photochemical Fenton reaction. Experiments were conducted in triplicates; individual values are shown.

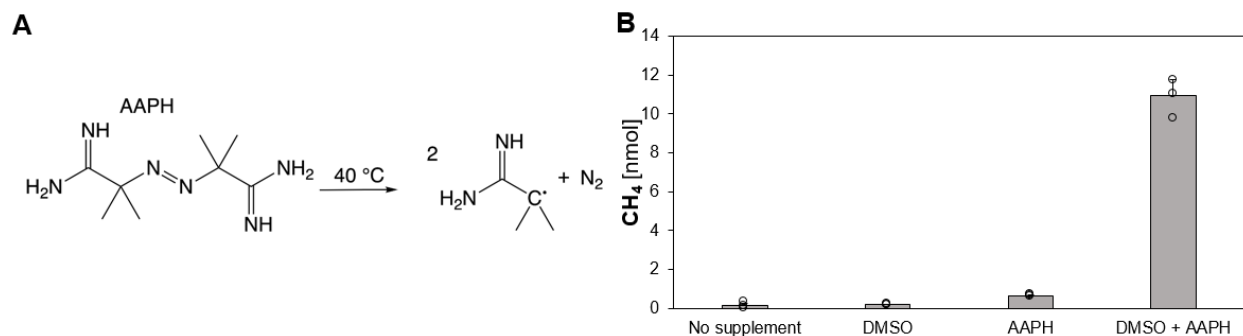




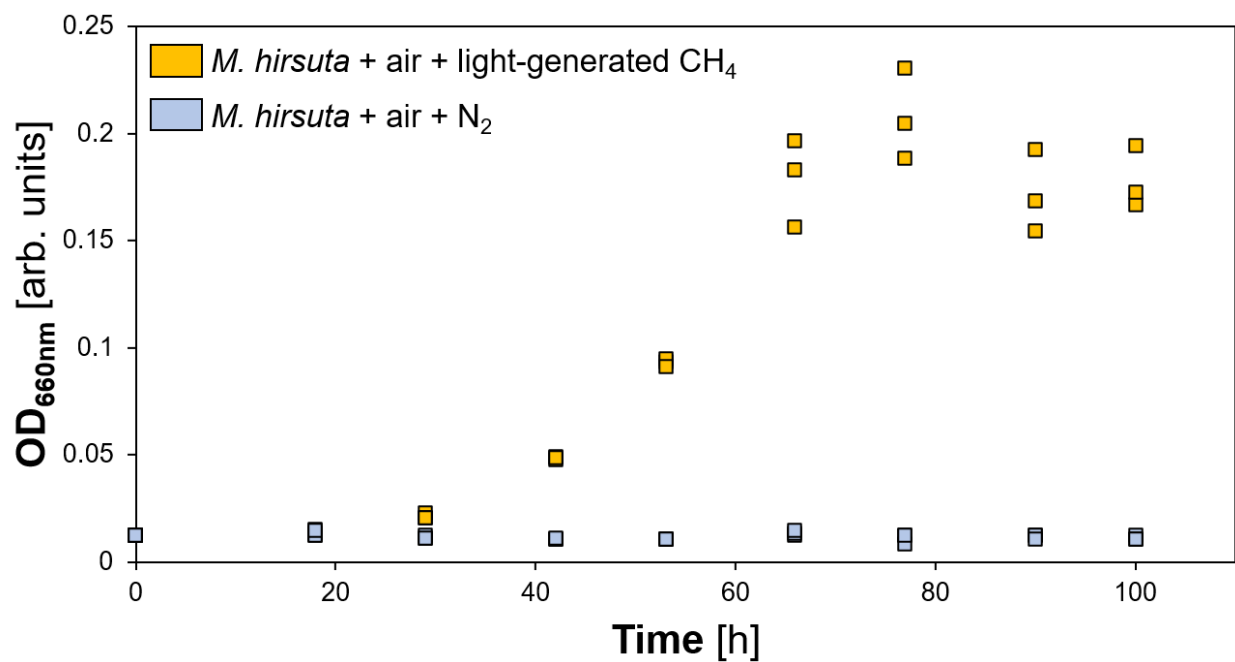
**Supplementary Fig. 8. Magnesium decreases light-driven CH<sub>4</sub> formation from DMSO.** In a total volume of 4 mL, samples containing 20 mM potassium phosphate buffer (pH 7), 500 mM DMSO and, optionally, 2 mM Fe<sup>3+</sup> (as FeCl<sub>3</sub>) and 10 mM citrate were incubated for one day under N<sub>2</sub> in sealed 20 mL glass vials under constant broad-spectrum illumination (Supplementary Fig. 3). Different amounts of Mg<sup>2+</sup> (as MgCl<sub>2</sub>) were added, ranging from 0 mM to 500 mM. Only trace amounts of CH<sub>4</sub> (<0.01 μM h<sup>-1</sup>) were formed in the presence of Mg<sup>2+</sup> and citrate. Upon supplementation with Fe<sup>3+</sup> without addition of citrate trace amounts of CH<sub>4</sub> (<0.01 μM h<sup>-1</sup>) were detected. Upon citrate and Fe<sup>3+</sup> supplementation, CH<sub>4</sub> rates increased to ~0.7 μM h<sup>-1</sup>, while increasing Mg<sup>2+</sup> amounts decreased CH<sub>4</sub> rates until ~0.16 μM h<sup>-1</sup> at 500 mM Mg<sup>2+</sup>. Experiments were conducted in triplicates; individual values are shown as circles.



**Supplementary Fig. 9. Decomposition of the LMCT-induced citrate radical drives  $\cdot\text{CH}_3$  formation from DMSO.** (A) Illumination of a  $\text{Fe}^{3+}$ -citrate complex results in  $\text{Fe}^{2+}$  and a citrate radical, disassembling into  $\text{CO}_2$  and a carbon-centered radical that further reacts with DMSO by cleaving off a  $\cdot\text{CH}_3$ . (B) While no  $\text{CO}_2$  is formed in the dark,  $\sim 1080$  nmol  $\text{CO}_2$  is detected from illuminated samples containing 2 mM  $\text{FeCl}_3$ , 10 mM citrate, 500 mM DMSO in 20 mM potassium phosphate buffer (pH 7) under  $\text{N}_2$ , after 24 h. The bars are the mean + standard deviation of three independent measurements, shown as circles.



**Supplementary Fig. 10. Organic radicals drive CH<sub>4</sub> formation from DMSO.** (A) At 40 °C, the compound 2,2'-azobis(2-methylpropionamidine) dihydrochloride (AAPH) decomposes into 2 carbon-centered organic radicals and N<sub>2</sub>. (B) CH<sub>4</sub> is formed upon the interaction between DMSO and AAPH. While only ~0.2 nmol or ~0.6 nmol CH<sub>4</sub> are formed from DMSO or AAPH alone, respectively, ~11 nmol CH<sub>4</sub> are formed upon combining both substances, indicating CH<sub>4</sub> formation driven by carbon-centered radicals. 500 mM DMSO was mixed with 20 mM AAPH and incubated for 3 h in N<sub>2</sub>-saturated DPBS buffer at 40°C under N<sub>2</sub>. The bars are the mean + standard deviation of triplicates, shown as circles.



**Supplementary Fig. 11. Light-driven CH<sub>4</sub> formation sustains methanotrophic growth of *Methylocystis hirsuta*.** Methanotrophic growth of *M. hirsuta* is sustained by light-generated CH<sub>4</sub>. While CH<sub>4</sub>-supplemented samples (yellow squares) grew, N<sub>2</sub>-supplemented samples (blue squares) did not exhibit bacterial growth (see Methods). Experiments were conducted in triplicates; individual values are shown.