

## Supplementary information

### Colistin resistance mutations in *phoQ* can sensitize *Klebsiella pneumoniae* to IgM-mediated complement killing

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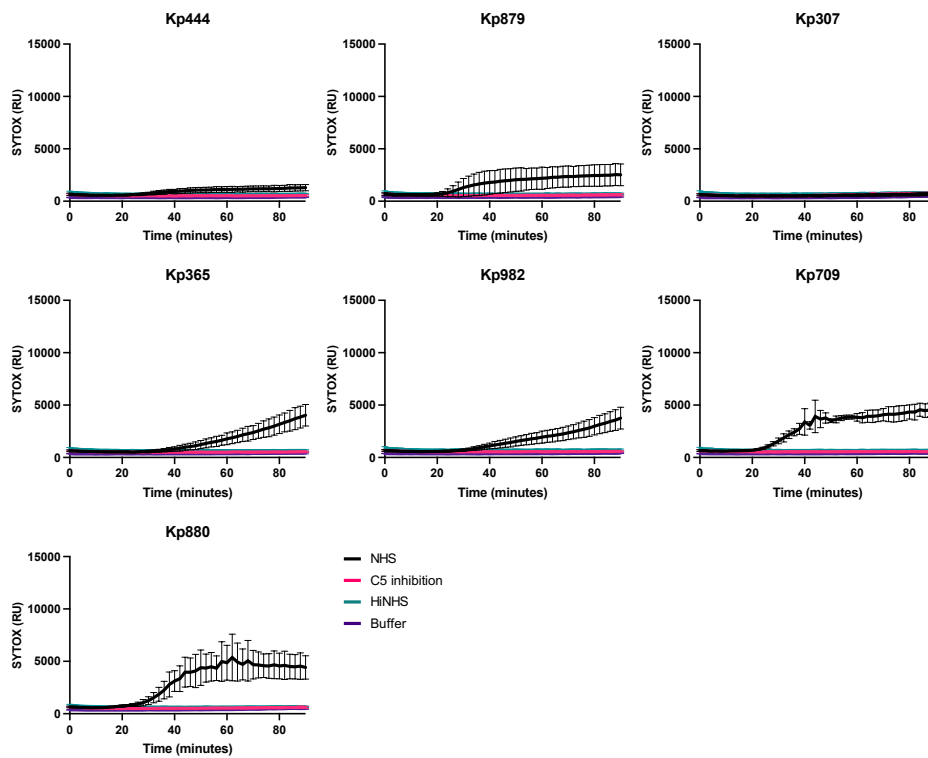
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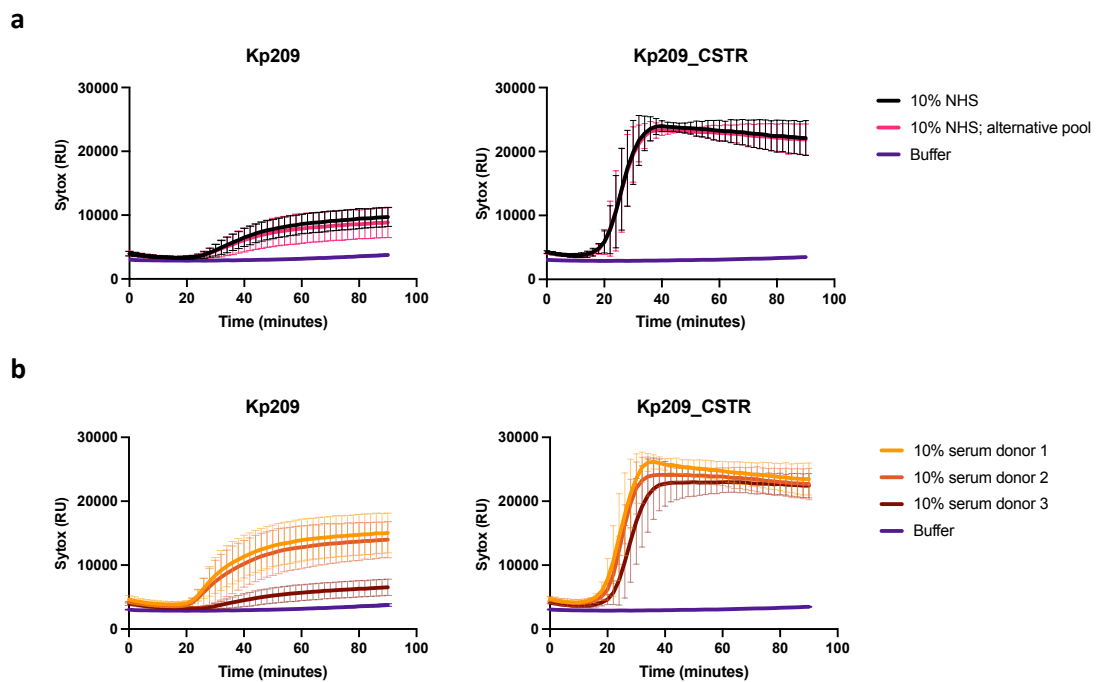
## Supplementary figure S11

Inner membrane permeabilization of Kp444, Kp879, Kp307, Kp365, Kp982, Kp709 and Kp880 in time. Bacteria were incubated in the presence of 10% NHS, 10% NHS in which C5 conversion was inhibited by addition of 20  $\mu\text{g}/\text{ml}$  OMCI and 20  $\mu\text{g}/\text{ml}$  Eculizumab (C5 inhibition), or 10% heat inactivated NHS (HiNHS), at 37  $^{\circ}\text{C}$  in the presence of 1  $\mu\text{M}$  SYTOX green nucleic acid stain, and inner membrane permeabilization (SYTOX fluorescence intensity) was detected every 2 minutes for 90 minutes in a microplate reader. Data represent mean  $\pm$  standard deviation of three independent experiments.



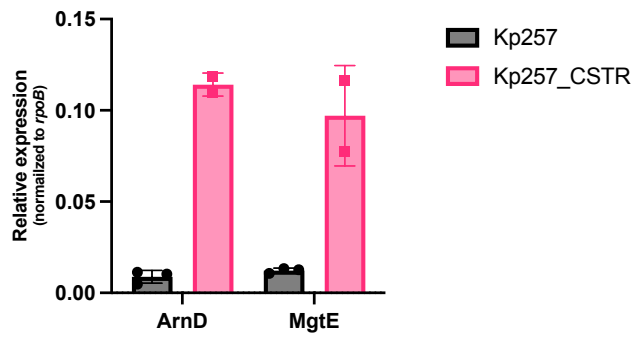
## Supplementary figure S12

(a) Inner membrane permeabilization of Kp209 and Kp209\_CSTR in the presences of (a) 10% NHS composed of sera from different pools of donors, or (b) 10% serum from different individual donors. (a&b) Bacteria were incubated at 37 °C in the presence of 1  $\mu$ M SYTOX green nucleic acid stain, and inner membrane permeabilization (SYTOX fluorescence intensity) was detected every 2 minutes for 90 minutes in a microplate reader. Data represent mean  $\pm$  standard deviation of three independent experiments.



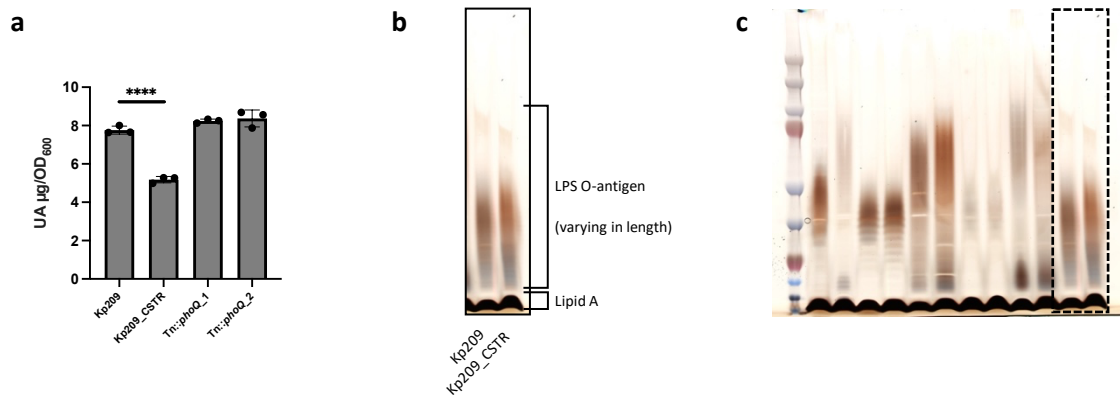
### Supplementary figure S13

RNA was isolated from log phase Kp257 and Kp257\_CSTR, and converted to cDNA. The relative expression of the *arnD* and *mgtE* genes compared to the *rpoB* control gene was determined by real-time PCR for both Kp257 and Kp257\_CSTR. Real-time PCR was performed in triplicates. Data represent mean  $\pm$  standard deviation of at least two independent biological replicates.



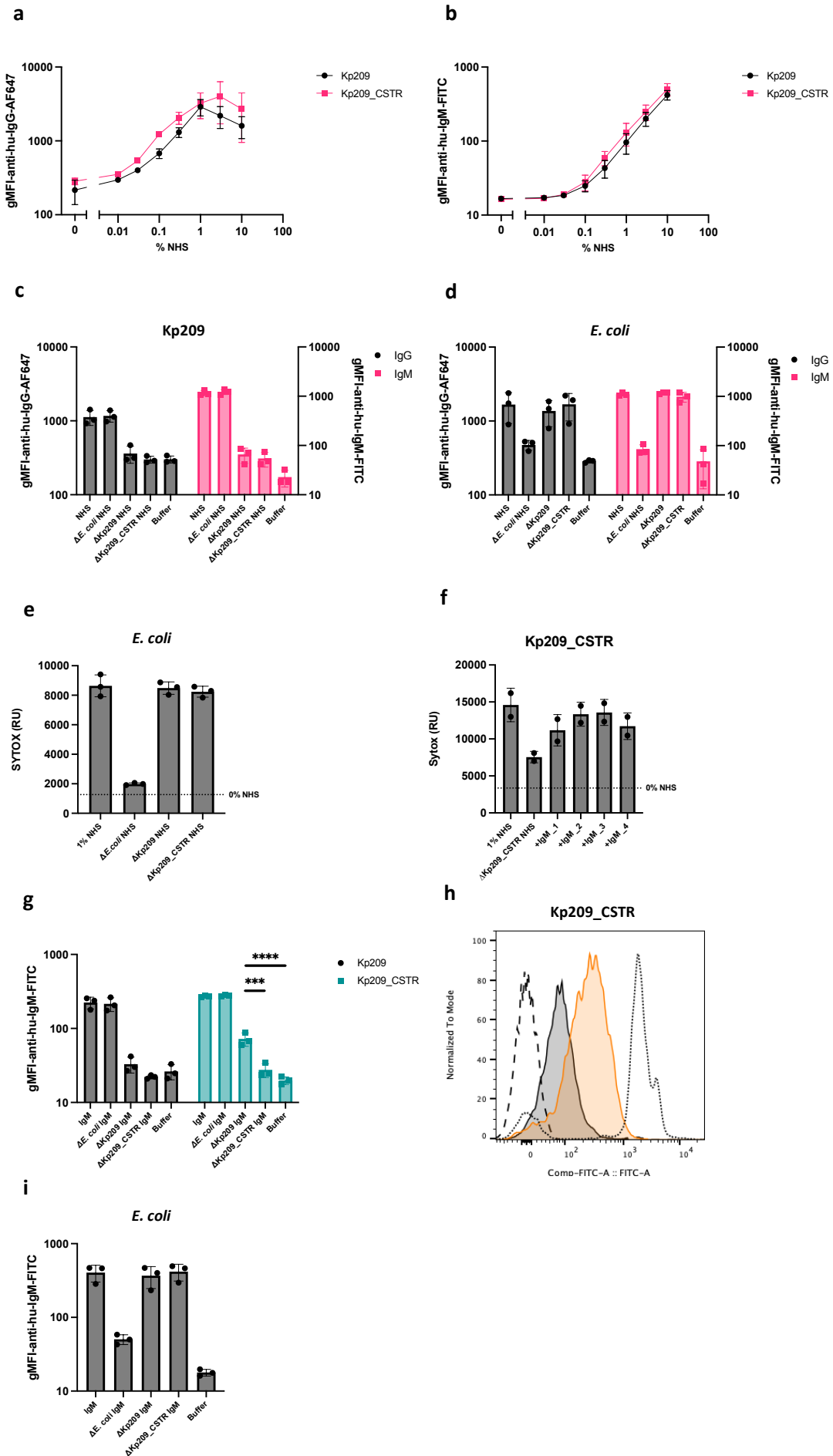
## Supplementary figure S14

(a) Capsule production of Kp209 and Kp209\_CSTR, and the Kp209\_CSTR Tn::*phoQ* mutants was determined by measuring uronic acid content. Absorbance was measured at 520 nm, the uronic acid content calculated using a glucuronolactone standard curve and normalized to the culture density at 600 nm. Data represent mean  $\pm$  standard deviation of three independent experiments. (F) Statistical analysis was performed using a paired one-way ANOVA with a Tukey's multiple comparisons test. Significance is shown as \*\*\*\* $p \leq 0.0005$ . (b) SDS-PAGE silver stain of Kp209 and Kp209\_CSTR LPS. Bacterial lysates were digested with protein K to remove all proteins and loaded on an SDS-PAGE gel. The LPS lipid A could, based on size, be distinguished from the O-antigen, which can have various lengths. Image is a representative of two independent experiments. Image is a representative of two independent experiments. Original gel is presented in supplementary Figure S2c. (c) Original gel of the SDS-PAGE silver stain containing of Kp209 and Kp209\_CSTR LPS, indicated with dashed lines.



## Supplementary figure S15

(a&b) Total IgG (a) and IgM (b) binding to Kp209 and Kp209\_CSTR in normal human serum (NHS). (c&d) IgG and IgM binding to Kp209 (c) and *E. coli* MG1655 (d) in NHS depleted using *E. coli* MG1655, Kp209 or Kp209\_CSTR ( $\Delta E. coli$  NHS,  $\Delta Kp209$  NHS, and  $\Delta Kp209\_CSTR$  NHS, respectively). (e&f) Inner membrane permeabilization of *E. coli* MG1655 (e) and Kp209\_CSTR (f) in the presence of 1% NHS,  $\Delta E. coli$  NHS,  $\Delta Kp209$  NHS, or  $\Delta Kp209\_CSTR$  NHS (f)  $\Delta Kp209\_CSTR$  NHS was supplemented with physiological concentrations of IgM (+IgM; 15  $\mu\text{g}/\text{ml}$  in 1% NHS) isolated from individual serum donors. (g) IgM binding to Kp209 and Kp209\_CSTR in IgM isolated from NHS, or isolated IgM depleted using *E. coli* MG1655, Kp209 or Kp209\_CSTR ( $\Delta E. coli$  IgM,  $\Delta Kp209$  IgM, and  $\Delta Kp209\_CSTR$  IgM, respectively). (h) IgM binding to Kp209 and Kp209\_CSTR with IgM isolated from NHS (dotted), IgM depleted with Kp209 (grey) or Kp209\_CSTR (orange). Slitted line corresponds to the buffer control. Geometric mean fluorescent intensity (gMFI) and number of events normalized to mode are depicted on the X- and Y-axis, respectively. The graph is representative for three independent repeats. (i) IgM binding to *E. coli* MG1655 in IgM isolated from NHS,  $\Delta E. coli$  IgM,  $\Delta Kp209$  IgM, or  $\Delta Kp209\_CSTR$  IgM. (a-d&g-i) IgG binding was performed in 0.3% NHS. IgM binding was performed in 10% (depleted) NHS or 45  $\mu\text{g}/\text{ml}$  (depleted) IgM isolated from NHS. Binding was detected using anti-hu-IgG-AF647 or anti-hu-IgM-FITC by flow cytometry. Flow cytometry data are represented by gMFI values of bacterial populations. Data represent mean  $\pm$  standard deviation of three independent experiments. (e&f) Bacteria were incubated at 37 °C in the presence of 1  $\mu\text{M}$  SYTOX green nucleic acid stain, and inner membrane permeabilization (SYTOX fluorescence intensity) was detected after (e) 60 or (f) 90 minutes. Data represents mean  $\pm$  standard deviation of a minimal of two independent experiments. (a&b) Statistical analysis was performed using a nonlinear regression with a variable slope comparing the logEC50 via sum-of squares F test, (c,d,g&i) a paired one-way ANOVA with a Tukey's multiple comparisons on  $\text{Log}_{10}$ -transformed gMFI data, (e) or a paired one-way ANOVA with a Tukey's multiple comparisons test on SYTOX fluorescence intensity data.



Supplementary table SI1 Differentially expressed genes in Kp209\_CSTR compared to Kp209

Kpn209S ID	Name/prediction	Log <sub>2</sub> (FC)	P <sub>adj</sub>
<b>Upregulated genes (Log<sub>2</sub>(FC)&gt;1, P<sub>adj</sub>&lt;0.05)</b>			
<i>Kpn209S_01234</i>	<i>mgtE.1</i>	5.25	1.06E-195
<i>Kpn209S_03326</i>	<i>mgtC</i>	4.15	3.81E-90
<i>Kpn209S_03165</i>	-	3.87	3.69E-57
<i>Kpn209S_03486</i>	<i>iolT</i>	3.79	1.12E-45
<i>Kpn209S_01244</i>	<i>arnC</i>	3.51	9.15E-146
<i>Kpn209S_03613</i>	-	3.42	1.46E-40
<i>Kpn209S_01243</i>	<i>arnB</i>	3.42	5.53E-98
<i>Kpn209S_01954</i>	-	3.39	5.43E-42
<i>Kpn209S_01095</i>	<i>clcB</i>	3.38	2.77E-69
<i>Kpn209S_03674</i>	-	3.35	1.09E-39
<i>Kpn209S_03723</i>	<i>yabl</i>	3.28	1.08E-51
<i>Kpn209S_00443</i>	<i>mgtA</i>	3.24	1.48E-51
<i>Kpn209S_01245</i>	<i>arnA</i>	3.22	5.16E-78
<i>Kpn209S_04650</i>	<i>ctpF</i>	3.16	2.08E-61
<i>Kpn209S_02782</i>	-	3.15	2.28E-52
<i>Kpn209S_01246</i>	<i>arnD</i>	3.11	2.67E-90
<i>Kpn209S_04711</i>	<i>scrB</i>	3.04	1.29E-21
<i>Kpn209S_04649</i>	<i>mdtA</i>	3.04	7.99E-46
<i>Kpn209S_04702</i>	<i>btuF</i>	3.03	2.00E-42
<i>Kpn209S_01247</i>	<i>arnT</i>	3.00	8.63E-102
<i>Kpn209S_02647</i>	<i>pmrD</i>	3.00	1.73E-56
<i>Kpn209S_03320</i>	<i>mamB</i>	2.98	1.40E-25



<b><i>Kpn209S_04708</i></b>	<i>scrK</i>	2.91	9.08E-19
<b><i>Kpn209S_03612</i></b>	-	2.91	2.84E-23
<b><i>Kpn209S_01631</i></b>	<i>ribB</i>	2.91	1.21E-49
<b><i>Kpn209S_04552</i></b>	<i>pagP</i>	2.83	3.30E-52
<b><i>Kpn209S_02224</i></b>	-	2.81	9.56E-50
<b><i>Kpn209S_04703</i></b>	<i>hmuU</i>	2.76	1.72E-35
<b><i>Kpn209S_00310</i></b>	-	2.75	1.24E-28
<b><i>Kpn209S_02223</i></b>	-	2.72	2.16E-52
<b><i>Kpn209S_02797</i></b>	-	2.66	2.38E-45
<b><i>Kpn209S_03487</i></b>	<i>fruB</i>	2.62	1.69E-11
<b><i>Kpn209S_01249</i></b>	<i>arnF</i>	2.60	2.38E-43
<b><i>Kpn209S_04337</i></b>	<i>ybjG</i>	2.56	1.08E-51
<b><i>Kpn209S_04704</i></b>	<i>fhuC</i>	2.53	5.59E-30
<b><i>Kpn209S_03568</i></b>	-	2.50	1.38E-15
<b><i>Kpn209S_02225</i></b>	-	2.48	2.67E-29
<b><i>Kpn209S_04648</i></b>	<i>bepG</i>	2.42	3.12E-32
<b><i>Kpn209S_04492</i></b>	<i>kdpA</i>	2.38	8.00E-22
<b><i>Kpn209S_02460</i></b>	<i>apbE</i>	2.36	4.22E-44
<b><i>Kpn209S_04416</i></b>	<i>bioB</i>	2.34	4.79E-28
<b><i>Kpn209S_04298</i></b>	<i>macA</i>	2.34	1.63E-55
<b><i>Kpn209S_04087</i></b>	<i>phoP</i>	2.34	1.41E-29
<b><i>Kpn209S_05121</i></b>	-	2.34	1.16E-22
<b><i>Kpn209S_02783</i></b>	-	2.34	1.54E-13
<b><i>Kpn209S_03672</i></b>	<i>zntB</i>	2.32	1.10E-19
<b><i>Kpn209S_02461</i></b>	<i>ada</i>	2.27	1.97E-46

<b><i>Kpn209S_01787</i></b>	<i>fimA</i>	2.27	6.99E-06
<b><i>Kpn209S_01248</i></b>	<i>arnE</i>	2.27	9.90E-21
<b><i>Kpn209S_03301</i></b>	<i>maeA</i>	2.22	1.23E-42
<b><i>Kpn209S_03166</i></b>	<i>slyB</i>	2.17	2.38E-19
<b><i>Kpn209S_01782</i></b>	<i>fimH</i>	2.15	2.94E-05
<b><i>Kpn209S_04297</i></b>	<i>macB</i>	2.12	5.55E-33
<b><i>Kpn209S_04415</i></b>	<i>bioF</i>	2.11	1.50E-17
<b><i>Kpn209S_00268</i></b>	<i>mdoB</i>	2.11	1.69E-32
<b><i>Kpn209S_04710</i></b>	<i>sacX</i>	2.10	3.80E-05
<b><i>Kpn209S_02462</i></b>	<i>alkB</i>	2.06	9.26E-22
<b><i>Kpn209S_03599</i></b>	<i>speG</i>	2.06	2.33E-35
<b><i>Kpn209S_00627</i></b>	<i>nhaK</i>	2.05	5.59E-43
<b><i>Kpn209S_01786</i></b>	<i>fimC</i>	2.05	8.39E-05
<b><i>Kpn209S_04088</i></b>	<i>phoQ</i>	2.04	2.00E-24
<b><i>Kpn209S_01785</i></b>	<i>fimD</i>	2.03	9.59E-05
<b><i>Kpn209S_04814</i></b>	<i>yheI</i>	1.98	9.73E-24
<b><i>Kpn209S_04414</i></b>	<i>bioC</i>	1.98	9.80E-15
<b><i>Kpn209S_01784</i></b>	<i>fimF</i>	1.96	1.91E-04
<b><i>Kpn209S_01788</i></b>	<i>fim</i>	1.96	1.97E-04
<b><i>Kpn209S_04709</i></b>	<i>scrY</i>	1.95	2.35E-04
<b><i>Kpn209S_03142</i></b>	<i>mdtK</i>	1.93	3.93E-16
<b><i>Kpn209S_02206</i></b>	<i>pgpC</i>	1.89	4.07E-19
<b><i>Kpn209S_04101</i></b>	<i>dmdB</i>	1.88	2.68E-16
<b><i>Kpn209S_04100</i></b>	<i>betB</i>	1.87	8.48E-13
<b><i>Kpn209S_01783</i></b>	<i>fimG</i>	1.86	5.10E-04

<b><i>Kpn209S_03183</i></b>	<i>ydgT</i>	1.86	2.63E-17
<b><i>Kpn209S_03598</i></b>	-	1.85	5.05E-17
<b><i>Kpn209S_00017</i></b>	<i>yaeQ</i>	1.79	3.31E-27
<b><i>Kpn209S_01781</i></b>	-	1.74	1.55E-03
<b><i>Kpn209S_02799</i></b>	<i>ftsl</i>	1.72	5.67E-09
<b><i>Kpn209S_04413</i></b>	<i>bioD</i>	1.69	2.68E-08
<b><i>Kpn209S_04493</i></b>	<i>kdpB</i>	1.66	1.54E-10
<b><i>Kpn209S_03300</i></b>	-	1.61	5.59E-06
<b><i>Kpn209S_04249</i></b>	<i>dpaL</i>	1.60	7.22E-04
<b><i>Kpn209S_03185</i></b>	<i>ydgJ</i>	1.60	8.07E-16
<b><i>Kpn209S_03327</i></b>	<i>zapE</i>	1.58	1.39E-14
<b><i>Kpn209S_00693</i></b>	-	1.56	5.13E-17
<b><i>Kpn209S_04813</i></b>	<i>mdlB</i>	1.56	5.91E-14
<b><i>Kpn209S_01938</i></b>	<i>ppnN</i>	1.54	9.77E-13
<b><i>Kpn209S_04677</i></b>		1.52	9.18E-05
<b><i>Kpn209S_04467</i></b>	<i>ybgC</i>	1.50	3.59E-12
<b><i>Kpn209S_03728</i></b>	<i>ompN</i>	1.49	9.90E-05
<b><i>Kpn209S_04757</i></b>	<i>fetA</i>	1.49	3.69E-15
<b><i>Kpn209S_01476</i></b>	<i>miaF</i>	1.48	1.74E-16
<b><i>Kpn209S_03182</i></b>	<i>ydgK</i>	1.47	1.84E-08
<b><i>Kpn209S_03933</i></b>	<i>pfeA</i>	1.44	6.22E-09
<b><i>Kpn209S_01523</i></b>	<i>osmY</i>	1.41	1.08E-17
<b><i>Kpn209S_01538</i></b>	<i>gatY</i>	1.40	8.90E-03
<b><i>Kpn209S_04466</i></b>	<i>tolQ</i>	1.38	6.13E-11
<b><i>Kpn209S_01477</i></b>	<i>miaE</i>	1.36	3.59E-16

<b><i>Kpn209S_03932</i></b>	<i>yciB</i>	1.33	8.40E-10
<b><i>Kpn209S_01121</i></b>	-	1.33	2.00E-07
<b><i>Kpn209S_04756</i></b>	<i>fetB</i>	1.32	1.22E-12
<b><i>Kpn209S_01524</i></b>	<i>diaA</i>	1.31	9.16E-15
<b><i>Kpn209S_02488</i></b>	<i>lpxT</i>	1.31	9.59E-05
<b><i>Kpn209S_00015</i></b>	<i>nlpE</i>	1.30	6.10E-15
<b><i>Kpn209S_01790</i></b>	-	1.29	6.40E-09
<b><i>Kpn209S_04812</i></b>	<i>glnK</i>	1.25	2.66E-03
<b><i>Kpn209S_00337</i></b>	-	1.25	1.87E-08
<b><i>Kpn209S_01636</i></b>	<i>tolC</i>	1.25	7.20E-11
<b><i>Kpn209S_04729</i></b>	<i>lpxL</i>	1.24	6.57E-09
<b><i>Kpn209S_01478</i></b>	<i>miaD</i>	1.24	6.74E-14
<b><i>Kpn209S_04846</i></b>	<i>yajR</i>	1.22	1.74E-07
<b><i>Kpn209S_04464</i></b>	-	1.20	4.50E-08
<b><i>Kpn209S_02092</i></b>	-	1.20	1.52E-05
<b><i>Kpn209S_01480</i></b>	<i>miaB</i>	1.19	3.77E-13
<b><i>Kpn209S_04338</i></b>	<i>deoR</i>	1.19	1.50E-08
<b><i>Kpn209S_04465</i></b>	<i>tolR</i>	1.18	5.96E-08
<b><i>Kpn209S_00016</i></b>	<i>arfB</i>	1.18	6.49E-13
<b><i>Kpn209S_03847</i></b>	-	1.17	1.63E-02
<b><i>Kpn209S_04469</i></b>	<i>cydX</i>	1.12	1.16E-02
<b><i>Kpn209S_00761</i></b>	<i>tatA</i>	1.11	8.31E-11
<b><i>Kpn209S_03299</i></b>	-	1.11	4.30E-02
<b><i>Kpn209S_05125</i></b>	<i>umuC</i>	1.11	1.37E-03
<b><i>Kpn209S_01479</i></b>	<i>miaC</i>	1.11	2.31E-12

<b><i>Kpn209S_04468</i></b>	-	1.08	7.62E-05
<b><i>Kpn209S_04470</i></b>	<i>cydB</i>	1.05	1.10E-07
<b><i>Kpn209S_01122</i></b>	-	1.04	1.31E-05
<b><i>Kpn209S_00758</i></b>	<i>tatD</i>	1.04	4.59E-08
<b><i>Kpn209S_04712</i></b>	<i>cra_</i>	1.03	1.59E-07
<b>Down regulated genes Log<sub>2</sub>(FC)&lt;-1, P<sub>adj</sub>&lt;0.05)</b>			
<b><i>Kpn209S_04410</i></b>	-	-3.28	2.33E-35
<b><i>Kpn209S_04407</i></b>	<i>eptA</i>	-2.78	2.52E-23
<b><i>Kpn209S_02806</i></b>	<i>yoaE</i>	-2.60	7.03E-34
<b><i>Kpn209S_00354</i></b>	-	-2.47	7.68E-24
<b><i>Kpn209S_00355</i></b>	-	-2.26	1.78E-21
<b><i>Kpn209S_04408</i></b>	<i>pmrA</i>	-2.16	3.29E-16
<b><i>Kpn209S_00356</i></b>	-	-1.97	4.50E-08
<b><i>Kpn209S_02517</i></b>	<i>oprB</i>	-1.81	1.16E-06
<b><i>Kpn209S_04409</i></b>	<i>pmrB</i>	-1.80	7.02E-12
<b><i>Kpn209S_04419</i></b>	<i>proY</i>	-1.72	1.04E-05
<b><i>Kpn209S_04252</i></b>	<i>yedS</i>	-1.70	1.25E-19
<b><i>Kpn209S_04420</i></b>	<i>hutH</i>	-1.69	8.22E-05
<b><i>Kpn209S_03684</i></b>	<i>cycA</i>	-1.67	2.39E-05
<b><i>Kpn209S_00357</i></b>	-	-1.66	1.30E-06
<b><i>Kpn209S_04421</i></b>	<i>hutU</i>	-1.64	2.94E-05
<b><i>Kpn209S_02966</i></b>	<i>rutC</i>	-1.60	7.18E-04
<b><i>Kpn209S_02520</i></b>	<i>bglB</i>	-1.59	4.59E-08
<b><i>Kpn209S_01312</i></b>	<i>feoA</i>	-1.57	8.04E-05
<b><i>Kpn209S_02514</i></b>	<i>yohK</i>	-1.57	6.19E-05

<b><i>Kpn209S_00358</i></b>	<i>ufaA</i>	-1.56	9.23E-07
<b><i>Kpn209S_03046</i></b>	-	-1.46	3.37E-03
<b><i>Kpn209S_02515</i></b>	-	-1.44	5.42E-06
<b><i>Kpn209S_03019</i></b>	-	-1.42	2.42E-03
<b><i>Kpn209S_03686</i></b>	<i>astB</i>	-1.40	1.28E-03
<b><i>Kpn209S_03685</i></b>	<i>astE</i>	-1.38	4.42E-03
<b><i>Kpn209S_02519</i></b>	<i>bglF</i>	-1.34	2.49E-08
<b><i>Kpn209S_01311</i></b>	<i>feoB</i>	-1.33	2.72E-03
<b><i>Kpn209S_02189</i></b>	<i>grcA</i>	-1.32	4.25E-03
<b><i>Kpn209S_02830</i></b>	<i>dadX</i>	-1.32	5.04E-05
<b><i>Kpn209S_00849</i></b>	<i>zinT</i>	-1.30	2.80E-06
<b><i>Kpn209S_03595</i></b>	<i>por</i>	-1.30	7.66E-03
<b><i>Kpn209S_04342</i></b>	-	-1.28	1.74E-02
<b><i>Kpn209S_03687</i></b>	<i>astD</i>	-1.28	7.21E-03
<b><i>Kpn209S_03635</i></b>	<i>ttdT</i>	-1.27	2.02E-03
<b><i>Kpn209S_01194</i></b>	-	-1.27	1.74E-07
<b><i>Kpn209S_00616</i></b>	<i>acs</i>	-1.25	7.97E-04
<b><i>Kpn209S_02967</i></b>	<i>dadA</i>	-1.24	9.09E-03
<b><i>Kpn209S_02984</i></b>	-	-1.18	7.67E-03
<b><i>Kpn209S_00254</i></b>	-	-1.18	1.80E-02
<b><i>Kpn209S_01240</i></b>	<i>nikA</i>	-1.16	7.75E-04
<b><i>Kpn209S_00625</i></b>	<i>cidA</i>	-1.16	9.95E-04
<b><i>Kpn209S_02511</i></b>	<i>mgIC</i>	-1.14	8.95E-03
<b><i>Kpn209S_02084</i></b>	<i>nrdH</i>	-1.13	9.03E-03
<b><i>Kpn209S_03634</i></b>	<i>namA</i>	-1.11	2.34E-03

<b><i>Kpn209S_05120</i></b>	<i>scrK</i>	-1.10	4.96E-04
<b><i>Kpn209S_04178</i></b>	<i>putA</i>	-1.10	5.78E-04
<b><i>Kpn209S_02491</i></b>	<i>uxuA</i>	-1.09	5.87E-04
<b><i>Kpn209S_02422</i></b>	<i>nuoG</i>	-1.08	5.10E-07
<b><i>Kpn209S_02421</i></b>	<i>nuoF</i>	-1.08	5.66E-05
<b><i>Kpn209S_05135</i></b>	-	-1.08	9.26E-06
<b><i>Kpn209S_01195</i></b>	<i>bcsQ</i>	-1.07	4.63E-04
<b><i>Kpn209S_00362</i></b>	-	-1.07	1.05E-02
<b><i>Kpn209S_05141</i></b>	-	-1.06	5.51E-03
<b><i>Kpn209S_01175</i></b>	<i>kdgK</i>	-1.06	3.85E-02
<b><i>Kpn209S_04476</i></b>	<i>sdhB</i>	-1.06	7.56E-03
<b><i>Kpn209S_03607</i></b>	<i>bioD</i>	-1.05	9.26E-03
<b><i>Kpn209S_02450</i></b>	<i>glpQ</i>	-1.05	1.37E-04
<b><i>Kpn209S_04789</i></b>	<i>ykgO</i>	-1.05	6.90E-03
<b><i>Kpn209S_03594</i></b>	<i>uspG</i>	-1.01	2.78E-02
<b><i>Kpn209S_02049</i></b>	<i>srIE</i>	-1.01	1.79E-02

**Supplementary table SI2. Real-time PCR primers**

Primer	Sequence (5'→ 3')	T <sub>M</sub> (°C)	Amplicon size (bp)	Reference
arnD-F	AACTACTGACCATGGCGGCG	61.5	116	12*
arnD-R	GCCAGCCAGTTCACCACGAA	62.5		12*
mgtE-F	TGGTGTGCATTACCCTGTG	57.0	100	This study
mgtE-R	ATAAACGGCGCGGAAACTA	56.3		This study
rpoB-F	GATCCGTGGCGTGACTTATT	55.0	118	54
rpoB-R	GCCCATGTAGACTTCTTGTCT	54.6		54

\*Reference 12 used the alternative gene name *pgbP* for *arnD*